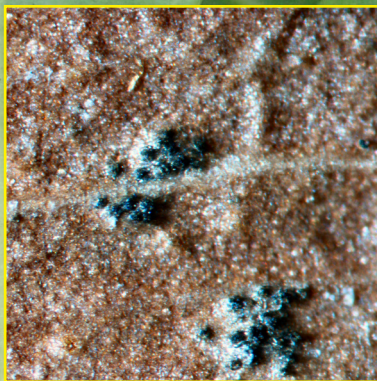


Mycosphaerella and its anamorphs:
2. Conspectus of *Mycosphaerella*

André Aptroot



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Abstract

A revision of the species described in *Mycosphaerella* and *Sphaerella* is presented, together with observations on the types of most species or their disposition. The genus *Stigmidium* is expanded to encompass fungicolous species and internal parasites of algae, and includes the genus *Mycophycias*.

The following new names or combinations in *Mycosphaerella* have been made for taxa that only have names in *Sphaerella* or were otherwise invalid: ***Mycosphaerella aiacu*** (Speg.) Aptroot, ***Mycosphaerella aristoteliae*** (Cooke) Aptroot, ***Mycosphaerella bubakii*** Aptroot, ***Mycosphaerella cleidionii*** (Berk. & Broome) Aptroot, ***Mycosphaerella goodiaefolia*** (Cooke) Aptroot, ***Mycosphaerella lasiana*** (Sacc.) Aptroot, and ***Mycosphaerella piperis*** Sawada ex Aptroot.

The following new combinations in other genera are proposed as a result of type studies: ***Arecophila advena*** (Syd.) Y.Z. Wang, Aptroot & K.D. Hyde, ***Arthopyrenia callunae*** (De Not.) Aptroot, ***Arthopyrenia picconii*** (De Not.) Aptroot, ***Arthopyrenia welwitschii*** (A.L. Sm.) Aptroot, ***Collemopsisidium pneumatophorae*** (Kohlm.) Aptroot, ***Davidiella acrocomiicola*** (Bat.) Aptroot, ***Davidiella allicina*** (Fr. : Fr.) Crous & Aptroot, ***Davidiella ammophilae*** (Durieu & Mont.) Aptroot, ***Davidiella ariadna*** (Sacc.) Aptroot, ***Davidiella carinthiaca*** (Jaap) Aptroot, ***Davidiella cecropiae*** (Bat., J.L. Bezerra & Matta) Aptroot, ***Davidiella cepharanthae*** (Sawada) Aptroot, ***Davidiella chrysobalani*** (Miles) Aptroot, ***Davidiella clandestina*** (Niessl) Aptroot, ***Davidiella craterispermi*** (Hansf.) Aptroot, ***Davidiella decidua*** (Ellis & Kellerm.) Aptroot, ***Davidiella dioscoreicola*** (Syd. & P. Syd.) Aptroot, ***Davidiella dircae*** (Ellis & Everh.) Aptroot, ***Davidiella disseminata*** (De Not. & Carestia) Aptroot, ***Davidiella ephedricola*** (Butin) Aptroot, ***Davidiella lactucae*** (Ellis & Kellerm.) Aptroot, ***Davidiella liabi*** (Petr.) Aptroot, ***Davidiella mappiae*** (Petch) Aptroot, ***Davidiella myrticola*** (Speg.) Aptroot, ***Davidiella nemorosa*** (Sacc. & Speg.) Aptroot, ***Davidiella pogostemonis*** (Khokhr.) Aptroot, ***Davidiella pontederiae*** (Peck) Aptroot, ***Davidiella populorum*** (G.E. Thomps.) Aptroot, ***Davidiella rosigena*** (Ellis & Everh.) Aptroot, ***Davidiella sapindi*** (Ellis & Everh.) Aptroot, ***Davidiella spilota*** (Syd.) Aptroot, ***Davidiella stephanorossiae*** (Duke) Aptroot, ***Davidiella woronichinii*** (G. Woron.) Aptroot, ***Dothidotthia melanococca*** (Lév. ex Triana & Planchon) Aptroot, ***Glomerella tetraspora*** (Seaver) Aptroot, ***Gnomonia leightonii*** (Berk. & Broome) Aptroot, ***Montagnula melanorhabdos*** (Petr.) Aptroot, ***Montagnula perforans*** (Roberge) Aptroot, ***Planistromella cattleyae*** (Cash & A.M.J. Watson) Aptroot, ***Planistromella conglomeratiformis*** (Bubák & Vleugel) Aptroot, ***Planistromella majuscula*** (Cooke) Aptroot, ***Planistromella operculata*** (Sacc.) Aptroot, ***Planistromella zonata*** (Ellis & Everh.) Aptroot, ***Stigmidium apophlaeae*** (Kohlm.) Aptroot, ***Stigmidium ascophylli*** (Cotton) Aptroot, ***Stigmidium parasiticum*** (G. Winter) Aptroot, and ***Wettsteinina plantaginicola*** (Höhn.) Aptroot.

INTRODUCTION AND HISTORY

The genus *Mycosphaerella* Johanson was first circumscribed in more or less the present form as *Sphaerella* Sacc. (1882). No type was indicated, as this was not obligatory or even common practice at the time. A genus was thought to be better characterised by the whole suite of included species than by one “typical” species. Saccardo assigned to this genus the species in *Sphaeria* Haller with presumably 1-septate, hyaline ascospores. As he wanted to deal with all taxa hitherto described, he based these combinations not only on species of which he had personal experience by having seen fresh or herbarium specimens, but also those known to him from the literature. As the descriptions of many older *Sphaeria* species are far from accurate, this resulted in taxa being included that had multiseptate, aseptate or brown ascospores, or of which no ascospores were known, but which were in fact coelomycetes. In some cases the conidia of the

coelomycetes were mistaken for ascospores, but in many cases no conidia or ascospores were ever observed, and many fungi have been described in the genera *Sphaeria* or *Sphaerella* based on the fact that their fruiting bodies were spherical in outline or “spherical and small”.

The first species of *Mycosphaerella* was described already in the 18th century by Persoon (1794). It was found on dead leaves of *Corylus*, and therefore described as *Sphaeria corylea* Pers. Soon after, Persoon (1801) found the same species on dead leaves of various other trees, and therefore changed the (now inappropriate) name to *Sphaeria maculiformis* Pers. Although such a name change to a more appropriate, but younger name is not allowed any more under the International Code for Botanical Nomenclature, it was perfectly sound at that time. In the same publication in which the name *Sphaeria corylea* was introduced, Persoon described *Sphaeria punctiformis* Pers. from dead leaves of various different trees and differing

from the preceding species by the dispersed rather than aggregated ascomata. Over the centuries to follow, mycologists have struggled with this character of ascomatal aggregation, which is not correlated to any other character. In fact, all species mentioned thus far are very homogenous in every other character, and it is often difficult to judge whether ascomata are aggregated or not, as they may be aggregated only on a portion of a leaf. Close observation revealed that the degree of ascomatal aggregation is only correlated with discontinuities in the substratum, and can therefore be regarded as behaviour rather than an intrinsic character. Consequently, these taxa have often been synonymised, and in recent times they are treated as the same taxon. Many morphological species have been described numerous times under different names, partly as forms, varieties and subspecies of *M. punctiformis*, but in over 100 cases also as separate species, usually based on an aberrant host. Many of these species have already been synonymised, notably by Tomilin (1979), but many others are reported here to be morphologically indistinguishable, usually based on the examination of the original type material. As indicated above, the name *Sphaeria corylea* is the oldest available, and would take priority were it not that the name *Sphaeria maculiformis* has been sanctioned by Fr., as is *Sphaeria punctiformis*. Therefore these two names are equally eligible for this species, and the choice for taking up *Sphaeria punctiformis*, as was done by e.g. Tomilin (1979) and Barr (1972), is followed here.

Although Persoon already saw and reported hyaline, 1-septate ascospores in his material in the 18th century, most consecutive authors in the beginning of the 19th century worked without microscope. When they described a *Sphaeria* or (later) *Sphaerella*, they meant literally a “spherical” fungus, nothing more or less. It soon became fashionable to describe a new *Sphaerella* species from each host genus or even species. Examination of the original type material of many of these species shows that these specimens often contain no ascospores, and probably were never mature. Many of the older names are inapplicable for this reason, notably many of those from Auerswald, Desmazières, Fries, Fuckel, Roumeguère and Schweinitz, who all described dozens of species.

The following is typical of the collections of each of these authors:

- Auerswald: many names invalid as they are cited as synonymes already when first mentioned; most types destroyed during WW II in Berlin;
- Desmazières: most material immature;
- Fries: most material belongs to other ascomycetes;
- Fuckel: most material contains the coelomycetous anamorph only;
- Roumeguère: material usually very sparse and immature or coelomycetous;
- Schweinitz: most material overmature.

In the beginning of the 20th century, mycologists became aware that the genus name *Sphaerella* had been unfortunate, because it was already in use for some green algae. Later on, even a grass would be described in *Sphaerella*. By then, hundreds of fungi were described in *Sphaerella*, but only a couple of algae, and mycologists argued that the name would be reserved for the fungi. However, this was not allowed by the Code as it was formulated at the time, although the present Code would allow (and even encourage) conservation of such a widely used name. The new genus *Mycosphaerella* Johanson was erected for the fungal *Sphaerella* species, with *Mycosphaerella vulgaris* (P. Karst.) Johanson as type. This is based on a new name for *Sphaeria punctiformis* (now regarded to be illegitimate as it is superfluous) introduced by Karsten to circumscribe the combination of *S. maculiformis* and *S. punctiformis* in one species, which is a perfectly sound idea, but once again not acceptable according to the present Code. As most of the Code, including the typification sections, is supposed to be retrospectively active, *Mycosphaerella punctiformis* (Pers. : Fr.) Starbäck should be considered the type of the genus *Mycosphaerella*. In the beginning of the 20th century, some mycologists refused to accept the change from *Sphaerella* to *Mycosphaerella*, and persevered in describing new species in *Sphaerella*, and even combined newly described *Mycosphaerella* species in *Sphaerella*.

In the 19th and 20th centuries, thousands of species and infraspecific taxa were described in the genus *Sphaerella*, and in the 20th century about a thousand more were combined into *Mycosphaerella*, and a further thousand new species described. The total number of taxa to be considered is thus in the order of 3000, and the total number of names close to 10,000. Many species were described only because the host represented a genus from which no species of *Mycosphaerella* was reported before. Knowledgeable authors like Petrak fell into the trap of lumping at home, while splitting abroad. He confidently identified specimens from Central Europe, also from hosts they were not known from, but described the same species over and over again as new from non-European material. In general, all specimens from Australia, Asia, Africa, South America and the Pacific were described by the authors to whom these materials were sent regardless whether they were even fertile. In the 20th century, species were increasingly described by resident scientists on these continents, sometimes after careful studies, but often just after having sent part of the material to specialists and having received the identification “*Mycosphaerella* sp.; no species known from this host” or similar annotations.

Phytopathologists also contributed to the taxonomy of *Mycosphaerella*, but sometimes also to the confusion, e.g. by incubating leaves with a known phytopathogenic anamorph of a *Mycosphaerella* in the

soil in order to obtain the teleomorph. Teleomorphs were often obtained, but not necessarily of the respective anamorph species, as can be deduced from analysing the type materials, which often contain many different fungi (even sometimes several different species of *Mycosphaerella*), most of which surely invaded the leaves after incubation. Phytopathologists often identify the observed plant diseases (e.g. brown leaf spots on *Fragaria*) with the name of the teleomorph of the fungus (e.g. *Mycosphaerella fragariae* author) which is assumed to cause the disease.

Von Arx (1949), based on his personal experience with *Mycosphaerella* species in nature and in culture, was the first to synonymise many saprobic species, usually based on the descriptions, not on examination of original type material. Tomilin (1979) monographed the genus, and provided many additional synonyms for saprobic species, usually based on examination of original type material. Barr (1972), although only a regional treatment, provided a recent overview not restricted to one host family and classified the respective species into a system of sections, some of which are accepted here. Partial treatments of host-based groups (e.g. Crous, 1998, 1999) abound. More recently, Kuijpers & Aptroot (2002) published a revision of the species of the section *Longispora*, and Braun *et al.* (2003) disposed some species from Barr's sections *Tassiana* and *Didymellina* in the new genus *Davidiella* Crous & U. Braun, shown to be a sister group to *Mycosphaerella*.

ANAMORPHS

Over the years *Mycosphaerella* has been linked to nearly 50 anamorph genera (Crous *et al.* 2000, 2004a, b). von Arx (1983) treated several generic names that he regarded as anamorphs, or potential anamorphs of *Mycosphaerella*, many based on the different morphs colonising the same substratum. Sutton & Hennebert (1994), who studied the different types of conidiogenesis that occurred in these genera, found that many had been prematurely reduced to synonymy by von Arx (1983), or had teleomorph affinities elsewhere. Twenty-three genera were eventually accepted as anamorphs of *Mycosphaerella* by Sutton & Hennebert (1994). *Asteromella* Pass. & Thüm. is now commonly accepted as the spermatial state that occurs with most species of *Mycosphaerella* (Crous *et al.*, 2000), and it is possible that spermatial states have also been described as anamorphs in genera such as *Ascochyta* Lib., *Asteroma* DC. and *Phoma* Sacc. In contrast, genera sometimes cited as anamorphs of *Mycosphaerella* have teleomorphs in other genera (Sivanesan, 1984; Barr, 1996; Crous, 1998, 1999; Crous & Corlett, 1998), while new genera such as *Cercostigmia* U. Braun (= *Pseudocercospora* Speg.),

and *Xenostigmia* Crous have been erected for *Stigmia*-like species with *Mycosphaerella* teleomorphs (Braun, 1995; Crous, 1998; Crous & Corlett, 1998).

Most of the reported (possible) anamorphs of *Mycosphaerella* species are based on older literature citations reporting a species of *Mycosphaerella* occurring in close proximity to an anamorph, and these relations remain to be confirmed. It is not the subject of the present study. Anamorphs have been mentioned where they were found in the literature and the names have sometimes been changed into the currently accepted names, but no evaluation is undertaken. This is envisaged to be the subject of further studies such as that by Crous & Braun (2003) in the same series.

GENERIC SUBDIVISION

In the past, several segregates from *Mycosphaerella* have been proposed, often based on the associated anamorphs. Examples include *Cercosphaerella* Kleb. (with *Cercospora* Fresen. anamorphs), *Ovosphaerella* Laib. (with *Ovularia* Sacc.), *Ramisphaerella* Laib. (with *Ramularia* Unger), *Septisphaerella* Laib. (with *Septoria* Sacc.), *Ramularisphaerella* Kleb. (with *Ramularia*) and *Septorisphaerella* Kleb. (with *Septoria*). These genera, described by Klebahn (1918) and Laibach (1922) respectively, have never been widely used. Moreover, all of these generic names were not validly introduced, as they were not accepted by the respective authors themselves at the moment they are described (as can be read from the context in German, and from the fact that in no instance a combination is made, and the species are in all cases cited as belonging to *M.* = *Mycosphaerella*), and are therefore not yet available for a potential future segregation of *Mycosphaerella*.

Barr (1972) provides an overview of the genus and classifies the species known from America into a system of sections. Some of these sections are accepted here, as they seem to single out morphologically recognisable groups that share the same biology and are even found in most phylogenetic analyses. Her section *Tassiana* (incl. *Didymellina* and *Stigmia*) is now accepted as the genus *Davidiella* (Braun *et al.*, 2003), and her section *Cymadothea* has been resurrected to genus level as well. The genus *Davidiella* was originally restricted to a few phylogenetically related species with *Cladosporium* anamorphs. Subsequently it was realized that there are strong morphological characters supporting its separation, notably the sole-shaped ascospores with usually (depending on preservation) irregularly angular lumina. Therefore all accepted species with sole-shaped ascospores with angular lumina and pyriform asci are classified here in the genus *Davidiella*. Her section *Longispora* has been the subject of a separate

study (Kuijpers & Aptroot, 2002). Her sections *Caterva*, *Fusispora*, and *Mycosphaerella* are accepted below and throughout the book. The remaining sections, viz. *Macula* and *Plaga*, encompass all the parasitic *Mycosphaerella* species, which are extremely variable in anamorph relations, phylogenetically diverse, and do not form well-defined clades.

SPECIES CONCEPT: THE THEORY

The genus *Mycosphaerella* as it is presently conceived, is remarkable among ascomycete genera in that it encompasses both saprobic and parasitic life forms. Presently, the parasitic species are supposed to be host-specific. However, in some cases experimental evidence exists of the contrary. The saprobic species, although in the past often described repeatedly from different hosts, are generally accepted to be less host-specific.

Special attention is drawn here to the actual life strategy of the saprobic species. When examining a collection, it is often striking that the whole leaf or stem is covered with just the respective *Mycosphaerella* species, with or without its respective anamorph(s). It becomes immediately obvious that the species did not invade the plant when it was dead, because in that case all kinds of fungi would have a more or less equal chance to invade the substratum, leading to mosaics of different species, as is often observed on plant materials not colonised by *Mycosphaerella*. The *Mycosphaerella* should therefore be present already in the living tissue, presumably as endophyte, and start sporulating some time after the tissue died. This fully agrees with observations on some specimens growing on fern fronds, where first growth and secondly sporulation starts around so-called hydatodes, in fact the open ends of vessels. As endophytes generally live in the vessels, they need no adaptations for intimate host-interactions, as they are only in contact with the inert apoplast.

CLASSIFICATION AND PHYLOGENY

The genus *Mycosphaerella* is currently classified in a separate order, the *Mycosphaerellales* (Nannf.) P.F. Cannon (Kirk *et al.*, 2001), in the class *Dothideomycetidae* of the *Ascomycotina*.

In recent years, several phylogenetic studies have been published on various sets of *Mycosphaerella* species or their anamorphs (e.g. Crous *et al.*, 2000, 2001; Verkley *et al.*, 2004), partly in close collaboration with me. The major observation still stands that *Mycosphaerella* is monophyletic.

Of note are the observations that morphologically similar species may exist on the same host (Crous *et al.* 2004a, b; Verkley *et al.*, 2004) and that, on the

other hand, morphologically and phylogenetically identical parasitic species may occur on widely different host families, showing that the current host-based species concept for parasitic *Mycosphaerella* species is not universally applicable. Recent DNA studies have confirmed these observations: the parasitic species *Mycosphaerella citri*, thus far known from the dicotyledonous *Citrus*, has been found also on the totally unrelated host *Acacia* and even on the monocotyledonous host *Musa* (Crous *et al.*, 2004b).

The phylogenetic classification of the hosts (mostly higher plants) has seen a considerable amount of change in recent years, due to concerted DNA sequence-effort. Even though the system is far from stable yet, the most recent conspectus (Bremer *et al.*, 2003) is followed here for the assignment of host genera to phanerogam families.

SPECIES CONCEPT: THE PRACTICE

The scope of the current research was to resolve the status of all names in *Mycosphaerella* and *Sphaerella*, to provide the correct reference and host, to check the names against the Code, and to either cite its current disposition or morphologically study material, preferably type material, of as many names as possible.

The first part of this was tremendously helped by Corlett (1988), although examination of some of the publications, especially older ones in German or Latin sometimes led to different interpretations. To evaluate the differences, one could e.g. compare the two entries for *Sphaerella vulgaris* in both publications: they were seen as valid species by Corlett, but shown to be both illegitimate (superfluous) by me. New (and many old) names are added too, but not much effort is put into incorporating the most recently (after 2000) described species.

Much effort was put into checking the names against the ICBN Code (Greuter *et al.*, 2000). Many errors have been made over the years, with some still persisting. The number of homonyms is relatively high in this group, because many species were named after their host, and scientists could thus come up with identical names for different taxa, obviously without taking notice of previous work. The change from *Sphaerella* to *Mycosphaerella* was also a source of confusion: some authors preferred the name *Sphaerella* over *Mycosphaerella*, and would combine new species described in *Mycosphaerella* into *Sphaerella*. The result can be that one homonym can have priority (in *Sphaerella*), but cannot be used in *Mycosphaerella*, as another name has priority in *Mycosphaerella*, as in the case of e.g. *M. acanthopanacis*. Another example of a rule which is often ignored is Art. 60.12, which rules that the spelling of epithets of fungi that refer to their host

should be in congruence with the spelling of the host name. This means that it should be *pyri* and *pyricola* after *Pyrus*, not *piri* or *piricola*, irrespective of what the original author wrote, and *pirolae* after *Pirola* (not *pyrolae*). Incidentally, this does not rule the spelling of *pyriformis* (because it does not refer to the host): *piriformis* is to be retained if the original author wrote it like that.

The examination of the type material is the main contribution of the current book. Types have been examined for most names; those not examined were either not available, or concern species which were already treated elsewhere. Many non-type specimens were also examined. Only a minority of those are cited here, as the information-content was usually low: either they agree fully with the type or they do not and often diverge noticeably from the protologue, as many specimens were apparently identified solely on the basis of the host, without examination of microscopic details.

The studied type specimens turned out to belong to dozens of different genera, but most were either *Mycosphaerella* or contained only a coelomycete. In the latter case, no further information is provided here, so as to prevent potential use of the respective epithets for (partly common and well-known) coelomycetes. In general, I have been reticent in the number of new combinations, and only actually formally proposed those in genera I have a profound experience with. Still, this includes some unexpected re-dispositions, like to the genera *Collemopsidium* Nyl. and *Stigmidium* Trevis. A key to those genera is provided.

MORPHOLOGY

The morphology of the teleomorphs of the genera *Davidiella* and *Mycosphaerella* is extremely uniform in many details.

The ascomata are invariably carbonized, uniformly rather large-cellular with parenchymatous cells. The wall extends also below the hamathecium. No clypeus formation is present. The only noticeable variation in the ascomata is the general size, but this character was not found to be highly correlated with any other character and is largely omitted from the observations. The only commonly found species with consistently large ascomata is *M. subradians*. Ascomata can be regularly dispersed or aggregated (Fig. 962). This is not currently seen as a valuable character, as it is shown to vary on a single leaf.

The ostiole is always apical, and the cells are more regularly arranged around it.

Hamathecium filaments (periphyses, paraphyses, pseudoparaphyses, paraphysoids) are always absent.

The asci occur in three main types, viz. pyriform (Fig. 978), cylindrical (Fig. 986), or cylindrico-clavate (Fig. 985). The ascus type is always constant within a

specimen and can usually be determined, even in old material. The size is not an independent character: it depends on the shape and the size of the ascospores it contains, as there is never a lot of empty space. The thickness of the ascus wall is dependent on the size of the ascospores, and moreover not very constant within even one ascus. The fine internal structure of the ascus (especially the base and the tip) has been studied in detail using many different mounting fluids, e.g. IKI (Lugol), KOH (10%), Cresyl and lactophenol, all of which clarified variation/structures in similar genera like *Stigmidium* *fide* Roux & Triebel (1994). However, no variation was detected and all type slides were subsequently investigated in tap water, and all dried slides are preserved in the CBS herbarium.

The asci always contain 8 ascospores, which are arranged irregularly uniseriate to irregularly biseriate or multiseriate (in one bundle). The latter state occurs only within the section *Longispora* where it is a distinguishing character. In all other groups the ascospores are more biseriate whenever the asci are pyriform, and tend to be more uniseriate when the asci are cylindrical; with the clavate asci holding an intermediate position. Therefore no extra information is gained by noting this character in addition to the ascus type.

The ascospores contain the most useful morphological characters in the group. They are always hyaline and remain uncoloured after discharge in most species, though exceptions do occur. All are uniseptate, but the septum can be median or conspicuously suprmedian (e.g. Fig. 4), or slightly submedian. A few taxa are known that have 3-septate ascospores. Ascospore shape varies from pyriform (e.g. Fig. 987, in all species of the section *Mycosphaerella*) to sole-shaped (e.g. Fig. 981, in the genus *Davidiella*) to cylindrical (e.g. Fig. 8, in the section *Longispora*) to irregularly long-ellipsoid (e.g. Fig. 988, in the remaining sections). The upper and lower ends of the ascospores can be rounded or slightly pointed, and this is often characteristic within a species or section (e.g. section *Mycosphaerella*: upper end rounded, lower end pointed; genus *Davidiella*: all rounded). The wall is thin, but the lumina of the ascospores in the genus *Davidiella* are often large and angular (Fig. 981), giving the ascospores a characteristic appearance. However, this character is somewhat influenced by the preservation conditions and should be used with care.

As can be seen from the key, genera that have often been confused with *Mycosphaerella* often have either some kind of interascal hamathecium filaments or pigmented ascospores.

Anamorphs are rather variable in the group, but are the subject of other studies and have not been investigated in detail. Sometimes anamorphs were present on the same material as the (type) specimens of the teleomorph, sometimes even on the same leaf spot.

In that case a note is often made, but a biological relation between the two is not guaranteed.

HOW TO USE THIS BOOK

To identify a *Mycosphaerella*, first check the generic key. In the case your species belongs to *Davidiella*, use that key. If it really is a *Mycosphaerella*, evaluate whether the species is saprobic (occurring on dead leaves or dead parts of leaves, or even waste paper) or parasitic (occurring on well-defined spots on living leaves). If your species is saprobic use the key to *Mycosphaerella* species. This will lead to one name for each group of morphologically indistinguishable specimens, most of which are currently accepted as species, but some of which may turn out in future to consist of various cryptic species (Crous *et al.* 2004a, b; Verkley *et al.*, 2004). If your species is presumably parasitic, use the host index at the back. This lists also the saprobic species entries, so it could also be used for saprobic species, at least to find potential alternative names for your species.

Due to recent name changes, a slightly different nomenclature is accepted here than was annotated on the actual specimens. The main difference is that all species now placed in *Davidiella* have been annotated as *Mycosphaerella*, usually with the same epithet, with the following exceptions: *Davidiella ammophilae* was partly annotated as *Mycosphaerella minor* and partly as *Mycosphaerella longissima* and *Davidiella allicina* and *D. clandestina* together as *Mycosphaerella tassiana*. Also, in the text the more prudent wording “is morphologically indistinguishable from” is often chosen instead of the more final “is a new synonym of”.

Acknowledgements

The completion of this work was only possible with the collaboration of many colleagues. I am especially grateful to my wife, Mariëtte Teeuwen, for supporting me, and for inking the drawings, and to John David, Harrie Sipman and Gary Samuels for their help and hospitality during my stays at IMI, B and BPI respectively. The curators of all herbaria mentioned are warmly thanked for their often prompt collaboration. Vadim Melnik, Simeon Vanev and Pedro Crous made detailed comments on various versions of the text. Finally, my colleagues at CBS, especially Gerrit Stegenhuis, Walter Gams, Arien van Iperen, Angelina Kuijpers and Gerard Verkley, have helped considerably with the completion of this work.

Key to some genera and species often confused with *Mycosphaerella*, including those into which some are newly combined below:

- 1 ascospores not septate..... *Glomerella*
- 1 ascospores 1-septate (3-septate only in *Mycosphaerella sphaerulinae*)
 - 2 ascospores brown
 - 3 ascospores ornamented..... *Arecophila*
 - 3 ascospores not ornamented
 - 4 hamathecium absent..... *Dothidotthia*
 - 4 hamathecium present..... *Montagnula*
 - 2 ascospores hyaline
 - 5 ostiole with a long neck..... *Gnomonia*
 - 5 ostiole punctiform or schizolytic
 - 5 ostiole punctiform or schizolytic
 - 6 ascospores with incomplete pseudosepta..... *Wettsteinina*
 - 6 ascospores with 1 true septum
 - 7 hamathecium filamentous
 - 8 periphyses present in the ostiole..... *Stigmatidium*
 - 8 periphyses absent
 - 9 hamathecium anastomosing above asci..... *Collemopsidium*
 - 9 hamathecium branched between asci..... *Didymella*
 - 9 hamathecium simple, moniliform..... *Arthopyrenia*
 - 7 hamathecium absent or parenchymatous..... *Planistromella*
 - 10 hamathecium parenchymatous..... *Planistromella*
 - 11 ascospores 9.5-10 µm long, on trees..... *conglomeratiformis*
 - 11 ascospores 8.5-11 µm long, on palms..... *zonata*
 - 11 ascospores 11-12.5 µm long, ascomata simple..... *operculata*
 - 11 ascospores 11-13 µm long, ascomata aggregate..... *majuscula*
 - 11 ascospores 22-30 µm long..... *acervata*
 - 10 hamathecium absent
 - 12 ascospores apiosporous, 9-11 µm long..... *Omphalospora stellariae*
 - 12 ascospores with median septum
 - 13 ascomata in stromata
 - 14 only upper half of ascoma carbonized..... *Oligostroma proteae*
 - 14 ascoma wall fully carbonized..... *Cymadothea*
 - 15 ascospores 11-13 µm long..... *lenticula*
 - 15 ascospores 22-25 µm long..... *trifolii*
 - 15 ascospores 13-16 µm long..... *Scirrhiachora groveana*
 - 13 ascomata simple
 - 16 asci cylindrical to clavate..... *Mycosphaerella*
 - 16 asci pyriform, ascospores sole-shaped..... *Davidiella*

Key to the morphological species of *Davidiella*:

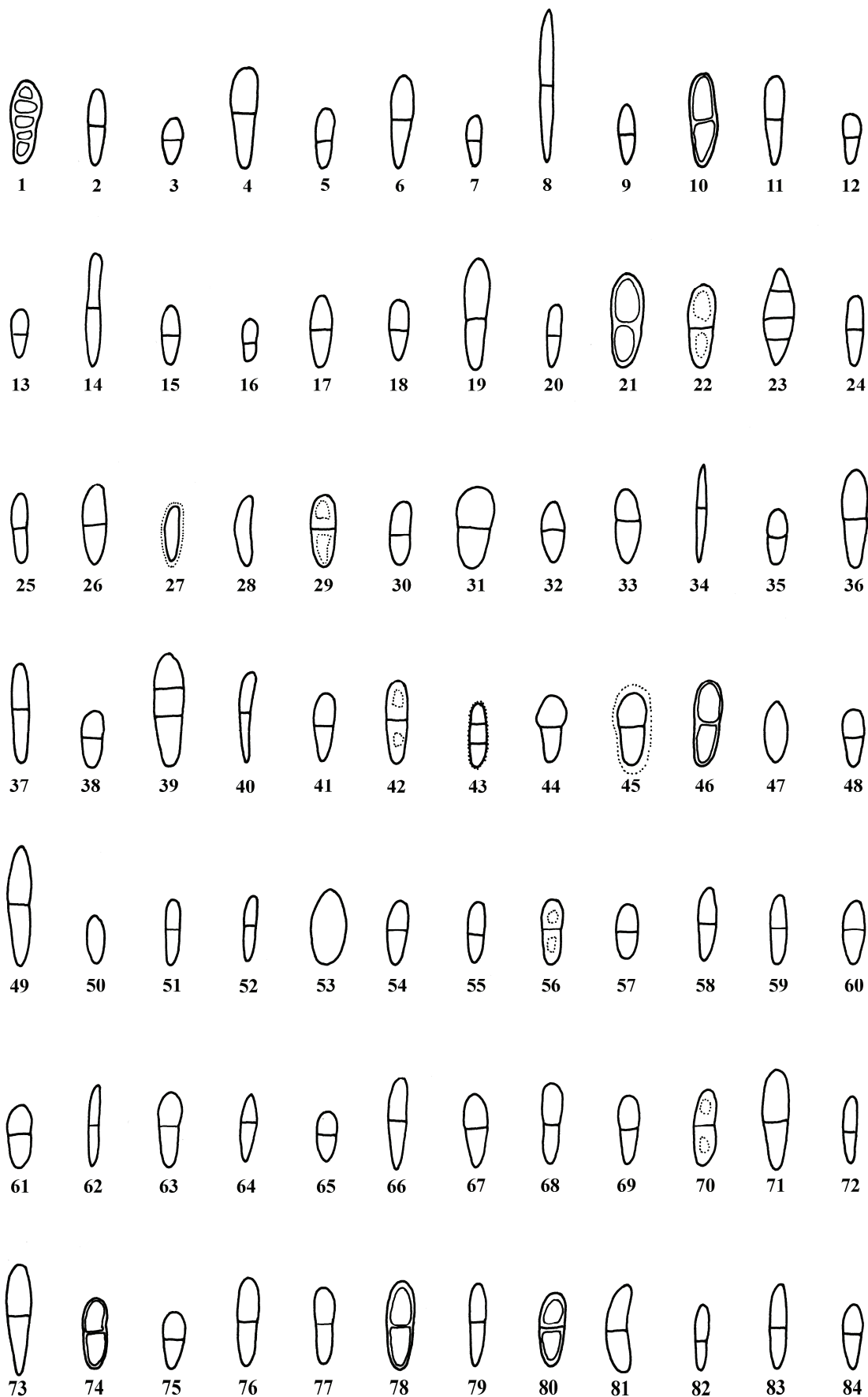
- 1 on spots on living leaves
 - 2 ascospores 8-11 µm long, host Leguminosae..... *ariadna*
 - 2 ascospores 8.5-10.5 µm long, host Menispermaceae..... *cepharantha*
 - 2 ascospores 9.5-11 µm long, host Arecaceae..... *acrocomiicola*
 - 2 ascospores 10-12 µm long, host Rosaceae..... *rosigena*
 - 2 ascospores 11-14 µm long, host Leguminosae..... *carinthiaca*
 - 2 ascospores 12-14 µm long, host Dioscoreaceae..... *dioscoreicola*
 - 2 ascospores 13-15 µm long, host Asteraceae..... *lactucae*
 - 2 ascospores 14-16 µm long, host Caryophyllaceae..... *woronichinii*
 - 2 ascospores 14-17 µm long, host Myrtaceae..... *myrticola*
 - 2 ascospores 14-17 µm long, host Asteraceae..... *decidua*
 - 2 ascospores 14-19 µm long, host Pontederiaceae or Nymphaeaceae..... *pontederiae*
 - 2 ascospores 14-20 µm long, host Fabaceae..... *nemorosa*

APTROOT

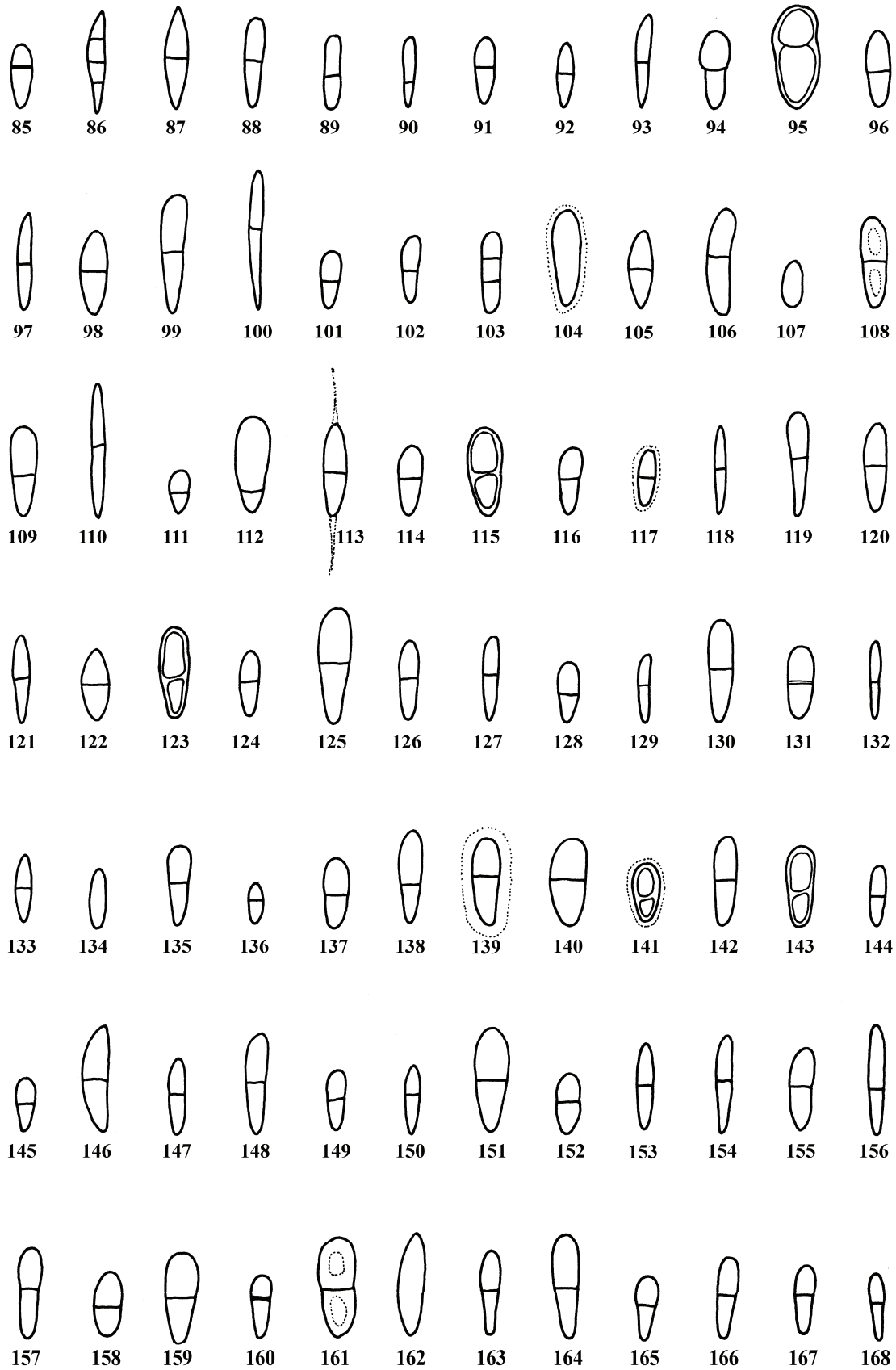
2 ascospores 15-17 µm long, host Oleaceae	<i>sapindi</i>
2 ascospores 15-19 µm long, host Thymelaeaceae.....	<i>dircae</i>
2 ascospores 15-19 µm long, host Moraceae.....	<i>cecropiae</i>
2 ascospores 16-19 µm long, host Lamiaceae	<i>pogostemonis</i>
2 ascospores 17-22 µm long, host Poaceae	<i>spilota</i>
2 ascospores 18-21 µm long, host Icacinaceae.....	<i>mappiae</i>
2 ascospores 18-23 µm long, host Chrysobalanaceae	<i>chrysobalani</i>
2 ascospores 23-35 µm long, host Rubiaceae.....	<i>craterispermi</i>
1 on dead tissues, including dead leaf tips etc.	
3 ascospores usually wider than 4.5 µm	
4 ascospores mostly below 22 µm long.....	<i>allicina</i>
4 ascospores mostly 22-30 µm long.....	<i>clandestina</i>
4 ascospores 30-35 µm long, on Asteraceae leaves.....	<i>liabi</i>
4 ascospores 26-36 µm long, on Apiaceae leaves	<i>stephanorossiae</i>
3 ascospores usually thinner than 4.5 µm	
5 ascospores (6-)10-13(-17) µm long.....	<i>ammophilae</i>
5 ascospores 10-15 × 3-5 µm long	<i>disseminata</i>
5 ascospores 12.5-13.5 × 3-3.5 µm, with sheath, on <i>Populus</i>	<i>populorum</i>
5 ascospores 15-18 × 3-4 µm, on <i>Ephedra</i> branches.....	<i>ephedricola</i>

Key to the morphological species of *Mycosphaerella*:

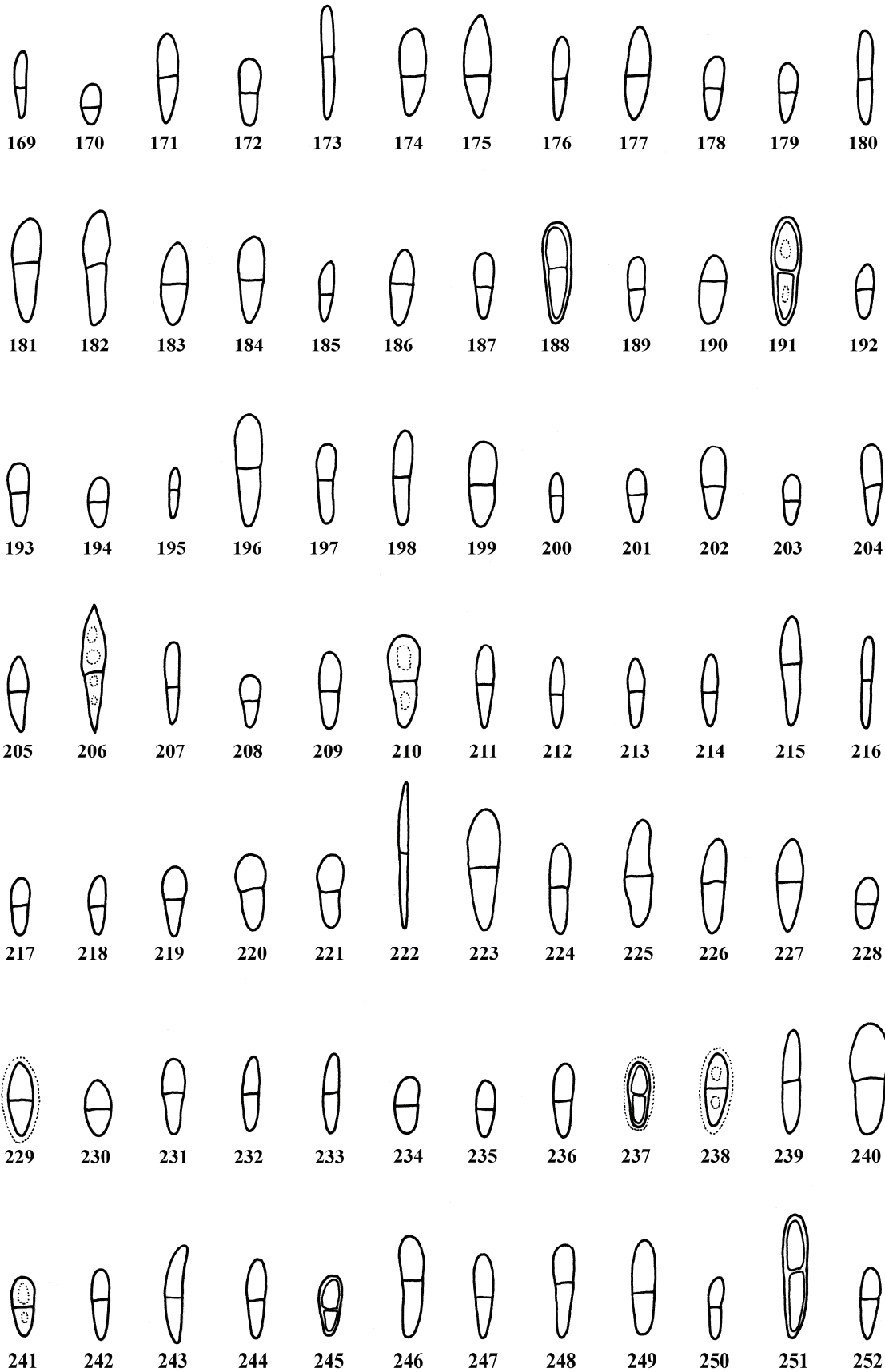
- 1 on spots on living leaves, asci cylindrical to clavate: many species; identify by using the host index and comparing the comments under the species listed
- 1 on dead tissues, including dead leaf tips etc.
 - 2 ascomata schizolytic, ascospores 7-12 µm long, on conifers *juniperina*
 - 2 ascomata opening with a preformed punctiform ostiole
 - 3 asci cylindrical to clavate
 - 4 ascospores linear (ratio over 6:1) sect. ***Longispora***
 - 6 ascospores lower ends pointed, 25-60 µm long *populi*
 - 6 ascospores lower end rounded
 - 7 ascospores 12-22 µm long *millegrana*
 - 7 ascospores over 20 µm long
 - 8 ratio 6-7:1 *topographica*
 - 8 ratio over 8:1
 - 9 ascospores 20-39 µm long *latebrosa*
 - 9 ascospores 36-40 µm long, on ferns *pteridis*
 - 4 ascospores pyriform to ovoid, asci strictly cylindrical sect. ***Mycosphaerella***
 - 10 ascospores 6-7 × 1.5-2 µm *nigromaculans*
 - 10 ascospores 7-11 × 2-3 µm *punctiformis*
 - 10 ascomata grouped, ascospores 8-9 × 1.5-2 µm *bumeliae*
 - 10 ascospores 12-14 × 3.5-5 µm *catesbeyi*
 - 4 ascospores ellipsoid to irregular, asci clavately cylindrical sect. ***Caterva***
 - 11 ascospores 7.5-10 µm long *arachnoidea*
 - 11 ascospores 11-18 × 3-4 µm long, on ferns etc *filicum*
 - 11 ascospores (8-)10-13(-15) µm long, ascomata over 0.2 mm *subradicans*
 - 11 ascospores 11-13 µm long, ascomata tiny, under 0.2 mm *asensioi*
 - 11 ascospores 13-17 µm long, turning stems red *rubella*
 - 11 ascospores 15-17 µm long, on grasses *graminicola*
 - 11 ascospores (13-)15-19 µm long *superflua*
 - 11 ascospores 21-24 µm long *molluginis*
 - 11 ascospores 18-21 µm long, on conifers *cunninghamiae*
 - 3 asci pyriform, ascospores fusiform sect. ***Fusispora***
 - 12 ascospores 15-18 µm long, on Cyperaceae *caricicola*
 - 12 ascospores 16-19 µm long, on Leguminosae *mimosicola*
 - 12 ascospores 16-19 µm long, on Moraceae *arachnoidea*
 - 12 ascospores 17-20 µm long, on Poaceae *panicicola*
 - 12 ascospores 21-25 µm long, on Ericaceae *munyangica*
 - 12 ascospores 24-32 µm long, on Cyperaceae etc *alpina*



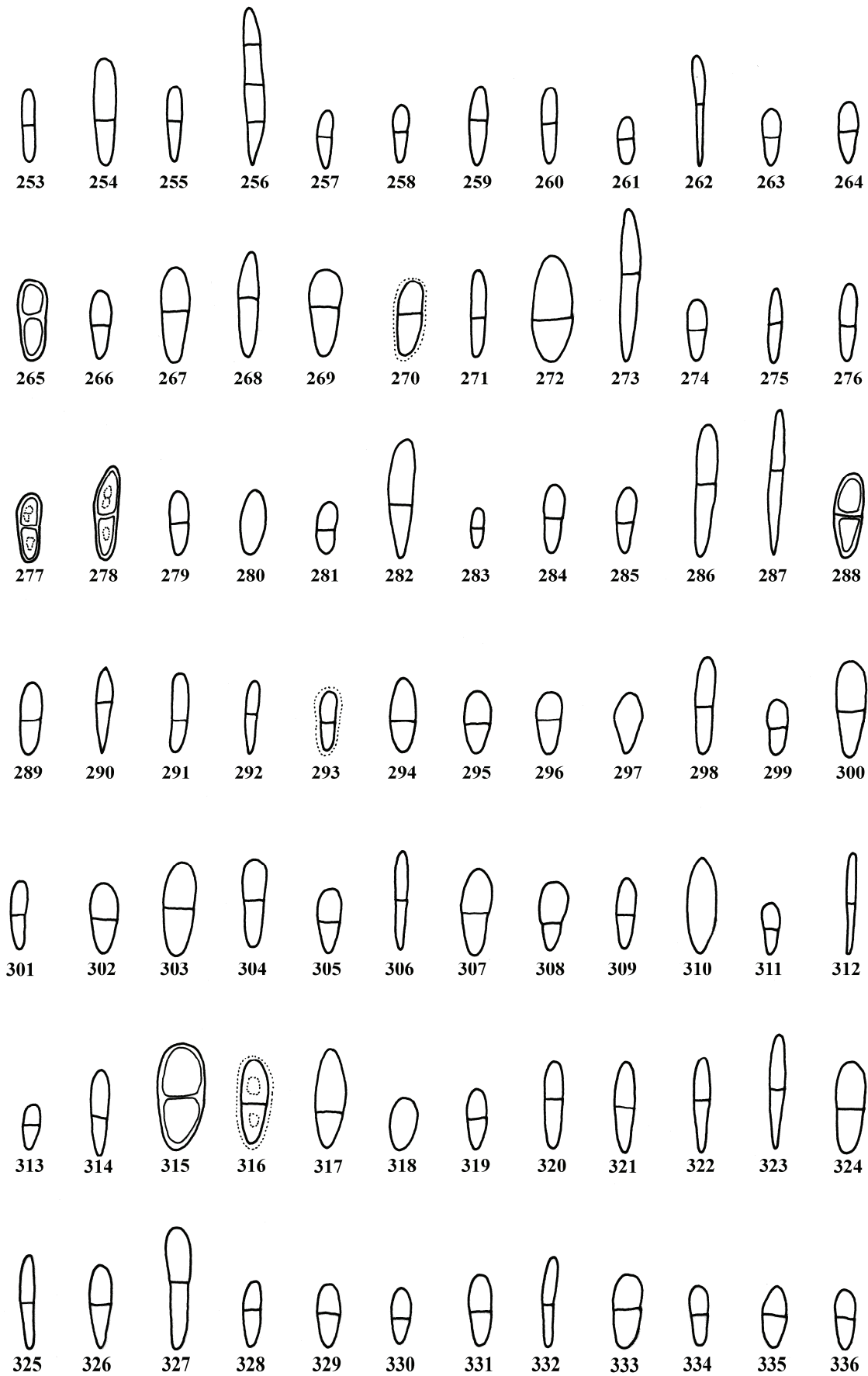
Figs 1-84. Outlines of ascospores, in water, not to scale. See main body of text for explanation. In general type specimens are illustrated; and if they were not available, the most characteristic other specimen.



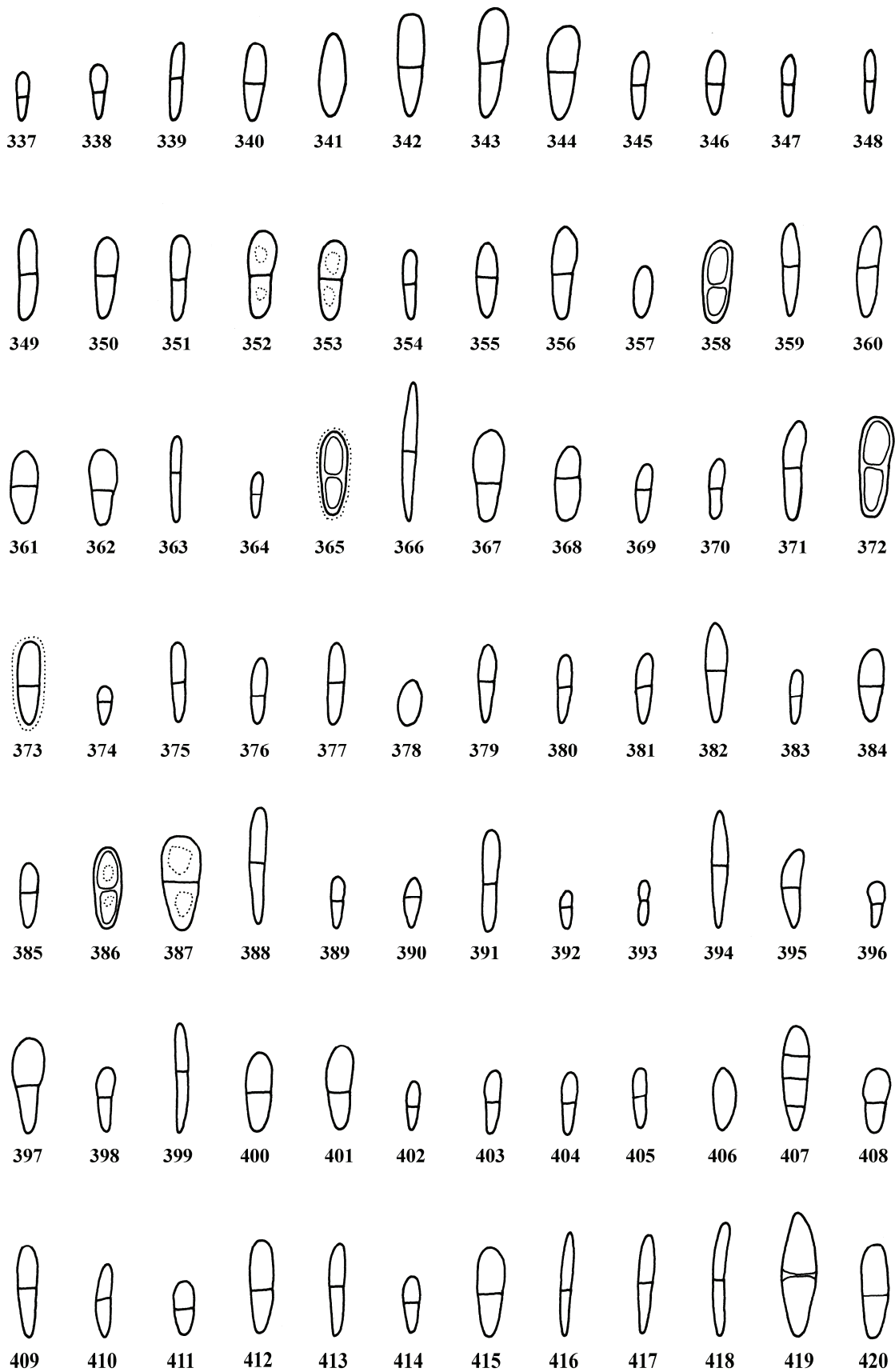
Figs 85-168. Outlines of ascospores, in water, not to scale. See main body of text for explanation. In general type specimens are illustrated; and if they were not available, the most characteristic other specimen.



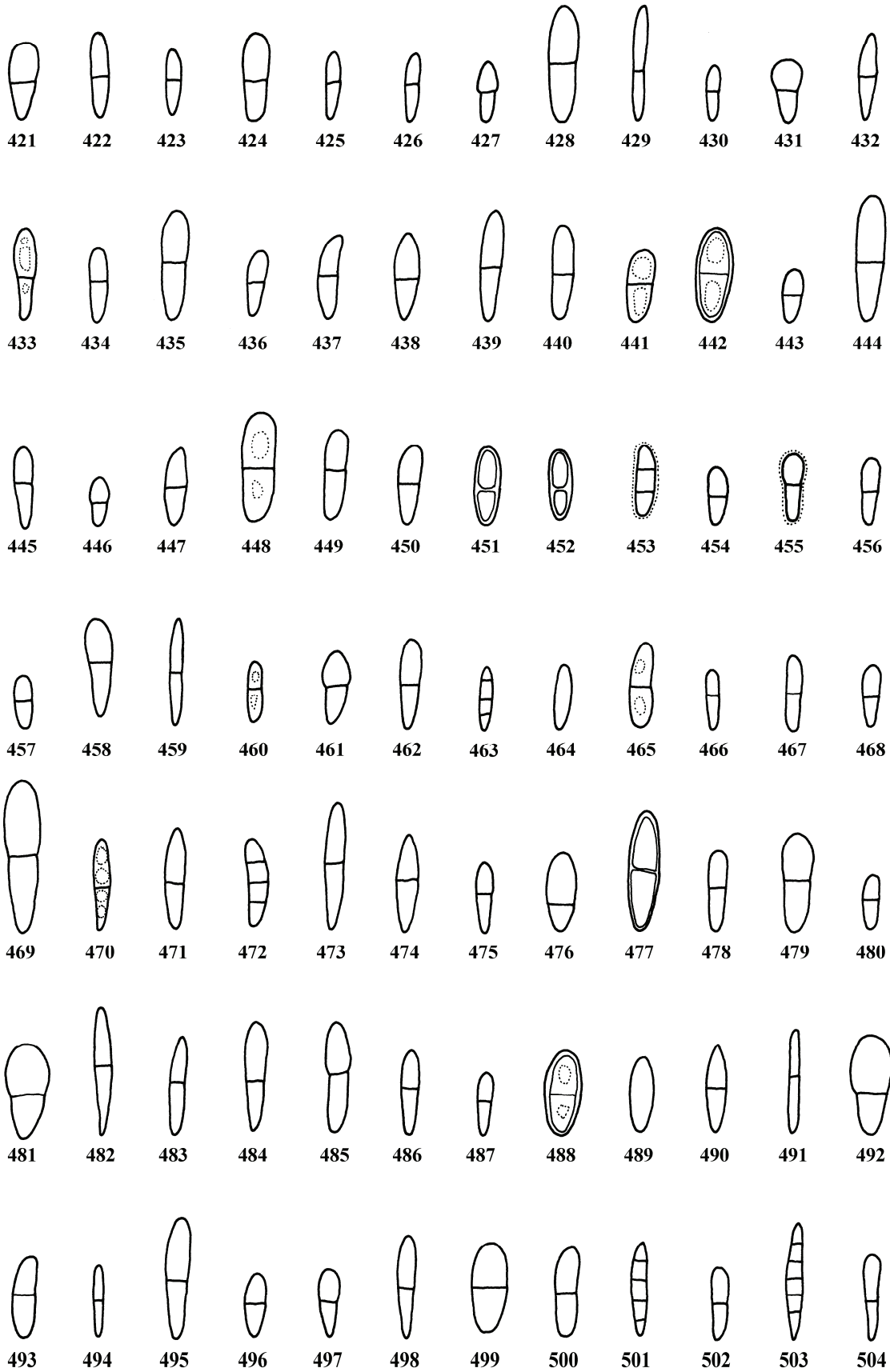
Figs 169-252. Outlines of ascospores, in water, not to scale. See main body of text for explanation. In general type specimens are illustrated; and if they were not available, the most characteristic other specimen.



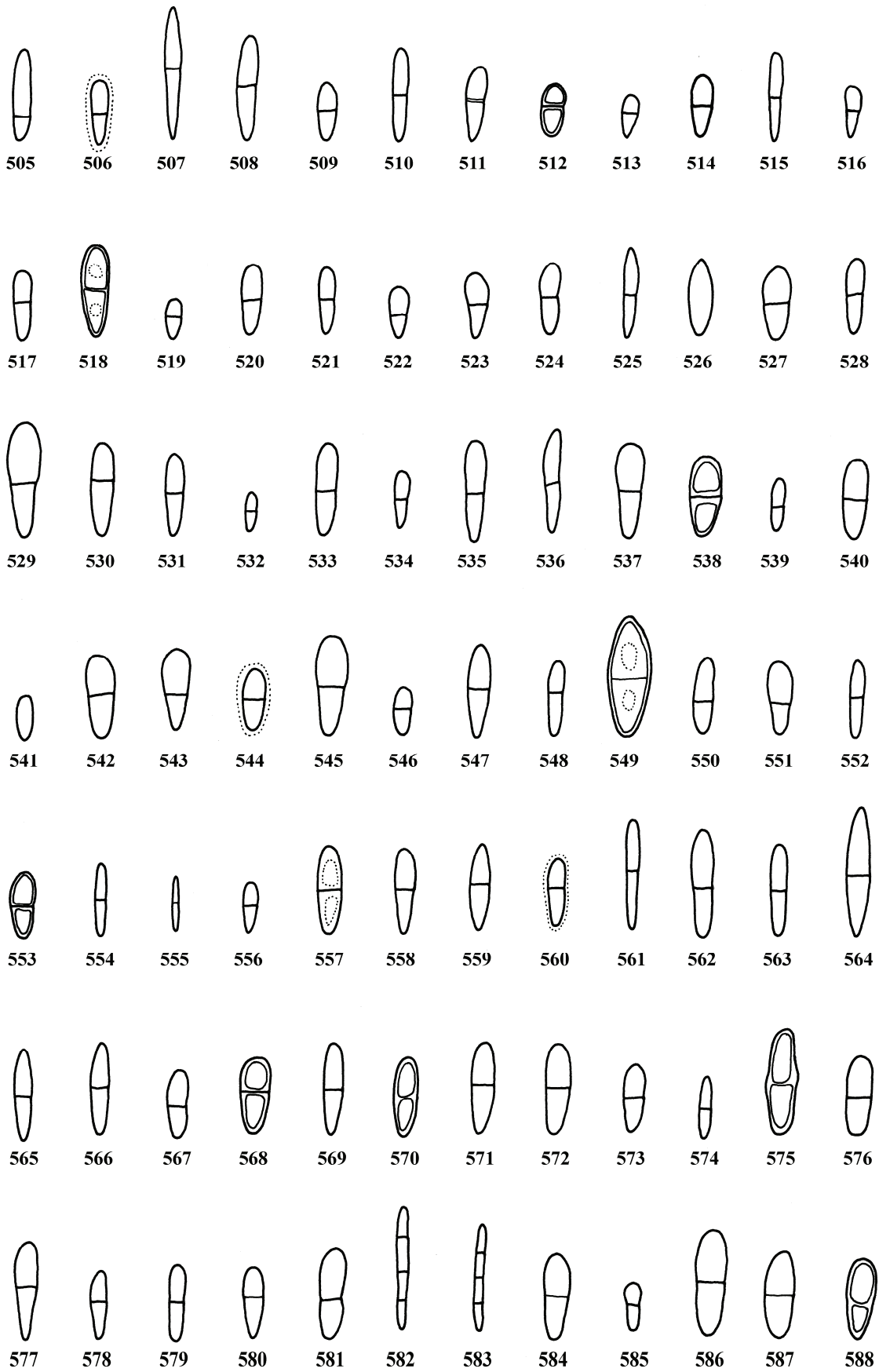
Figs 253-336. Outlines of ascospores, in water, not to scale. See main body of text for explanation. In general type specimens are illustrated; and if they were not available, the most characteristic other specimen.



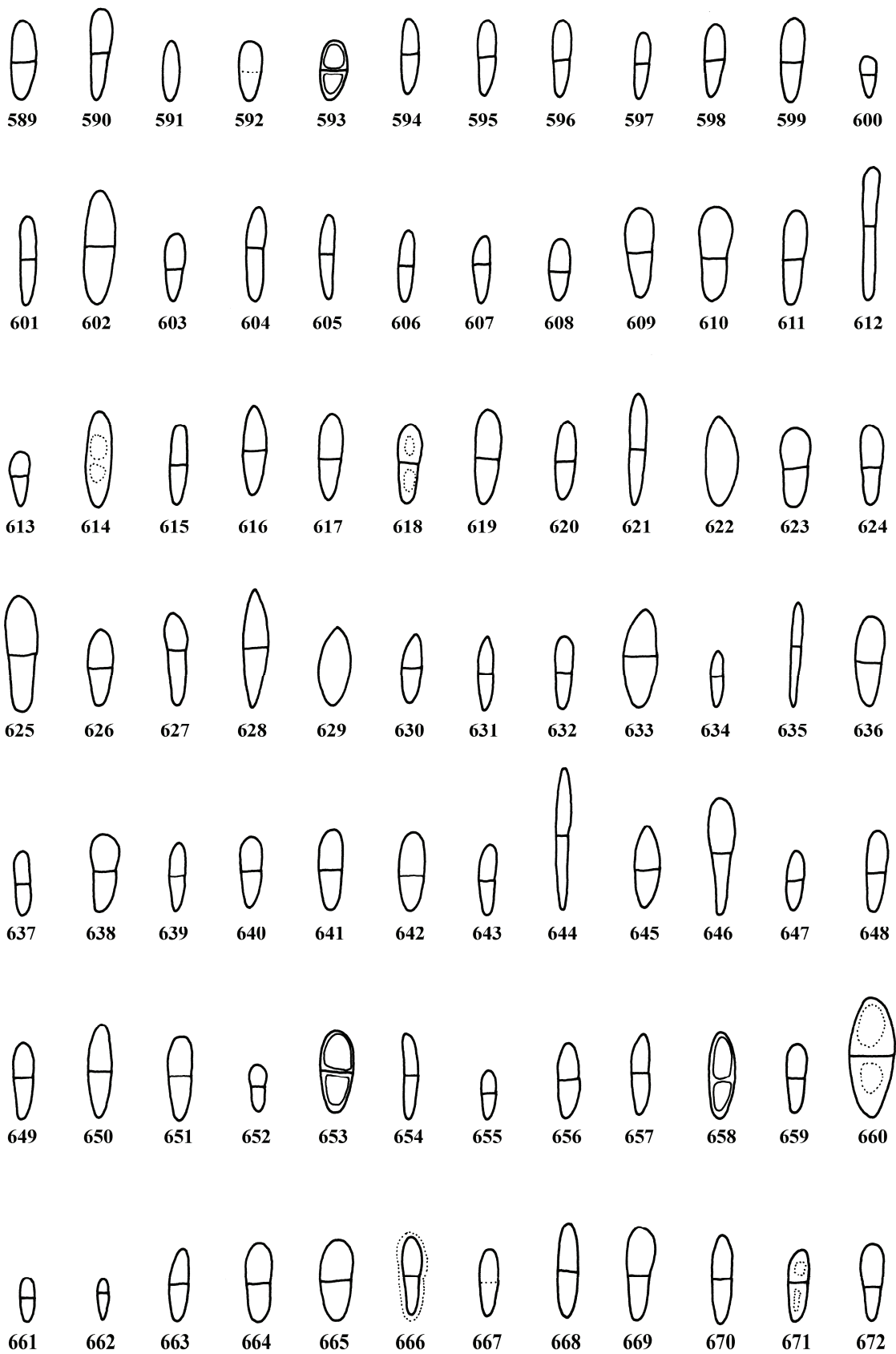
Figs 337-420. Outlines of ascospores, in water, not to scale. See main body of text for explanation. In general type specimens are illustrated; and if they were not available, the most characteristic other specimen.



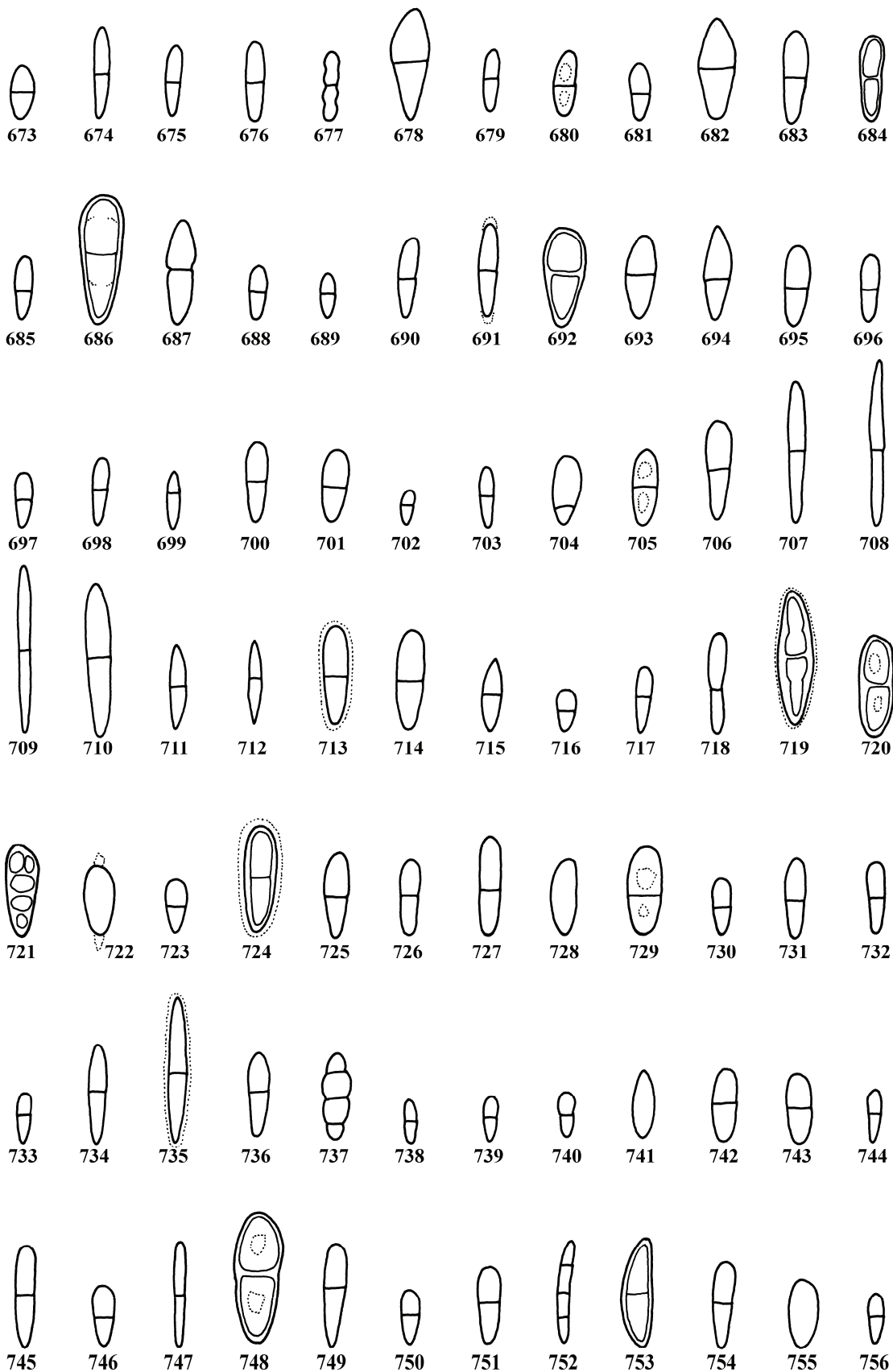
Figs 421-504. Outlines of ascospores, in water, not to scale. See main body of text for explanation. In general type specimens are illustrated; and if they were not available, the most characteristic other specimen.



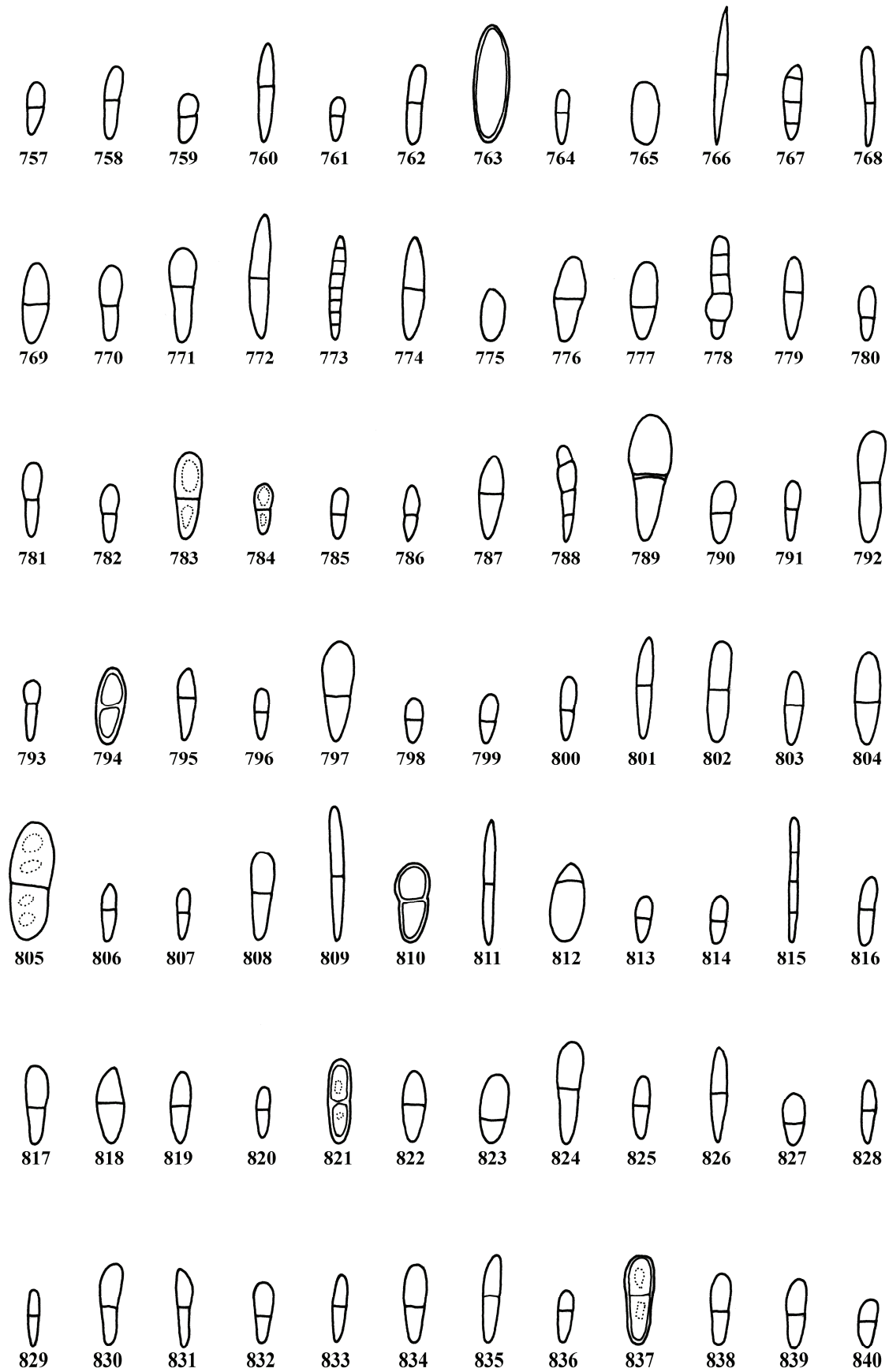
Figs 505-588. Outlines of ascospores, in water, not to scale. See main body of text for explanation. In general type specimens are illustrated; and if they were not available, the most characteristic other specimen.



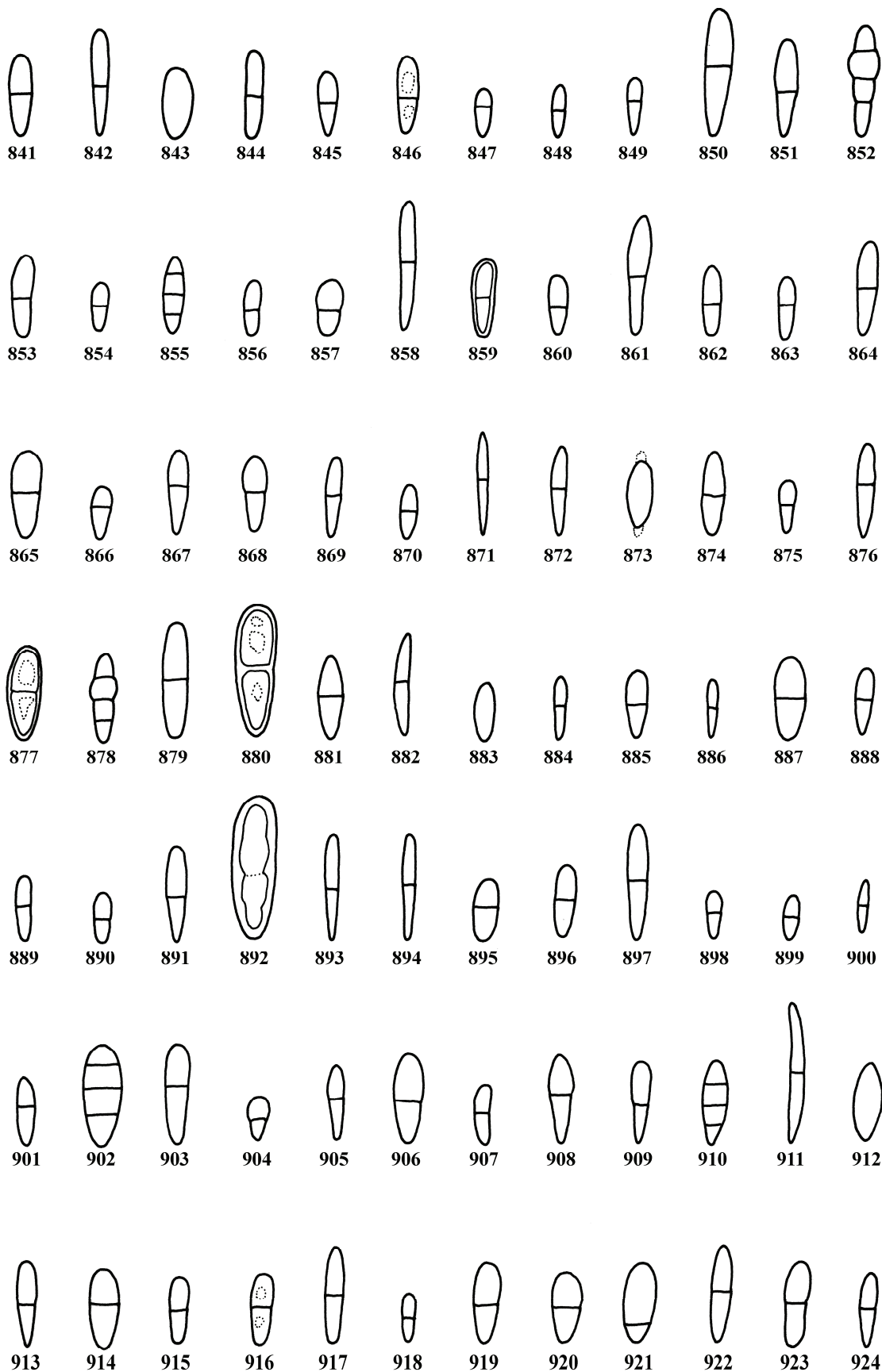
Figs 589-672. Outlines of ascospores, in water, not to scale. See main body of text for explanation. In general type specimens are illustrated; and if they were not available, the most characteristic other specimen.



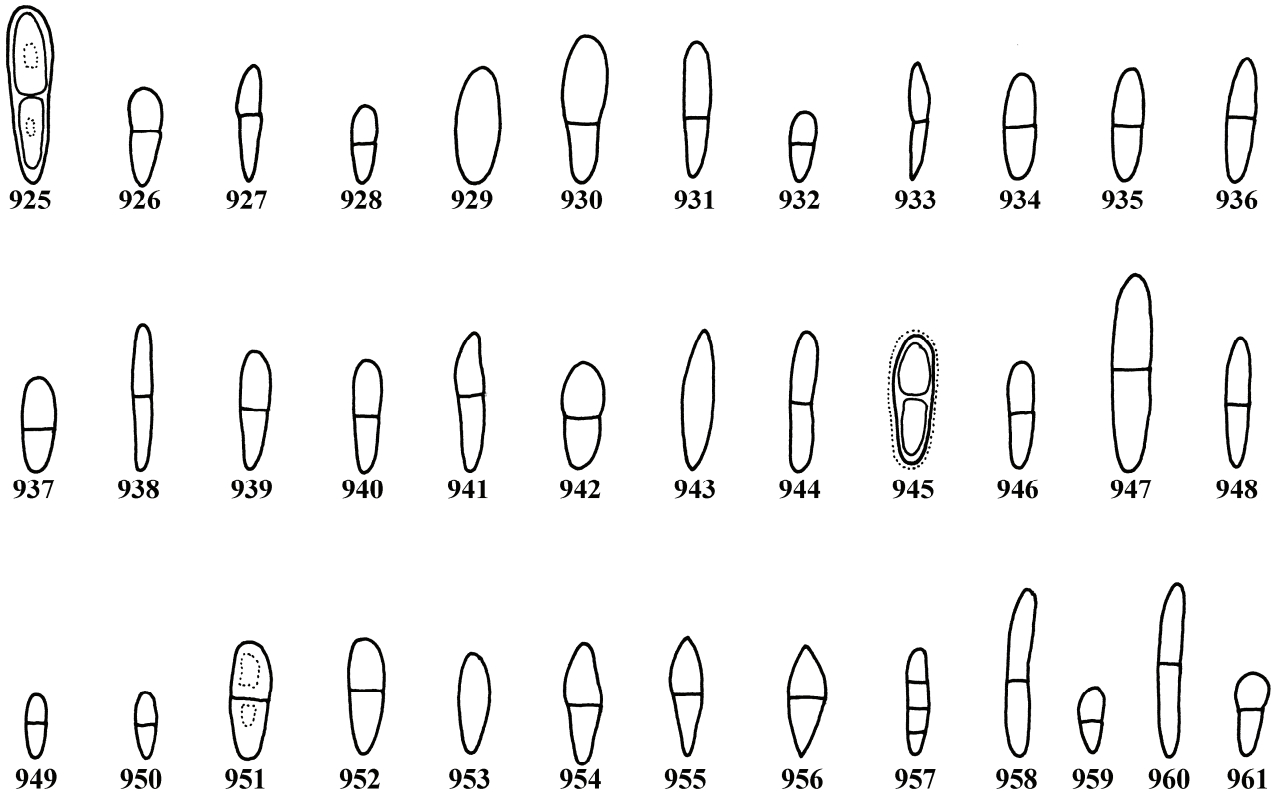
Figs 673-756. Outlines of ascospores, in water, not to scale. See main body of text for explanation. In general type specimens are illustrated; and if they were not available, the most characteristic other specimen.



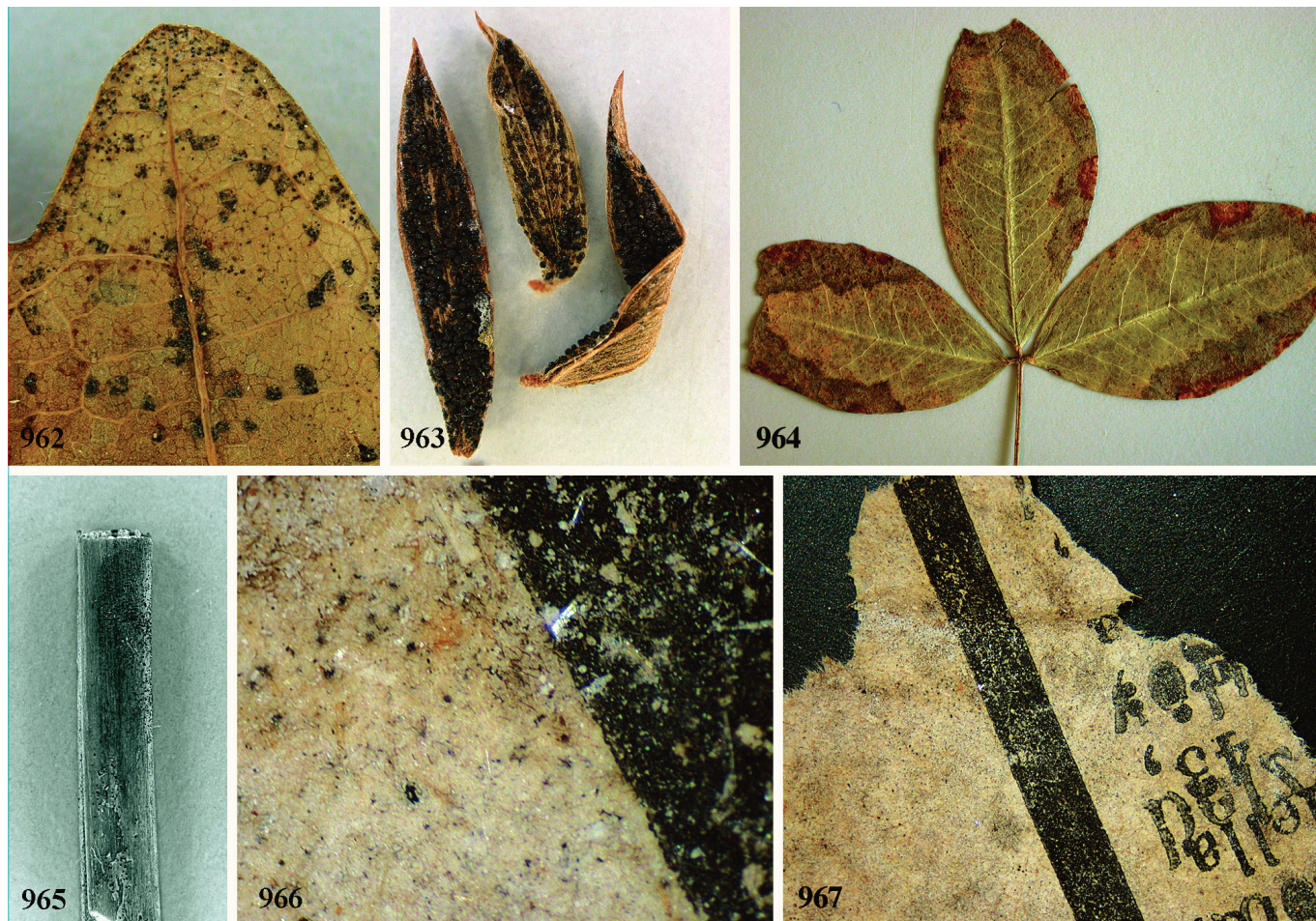
Figs 757-840. Outlines of ascospores, in water, not to scale. See main body of text for explanation. In general type specimens are illustrated; and if they were not available, the most characteristic other specimen.



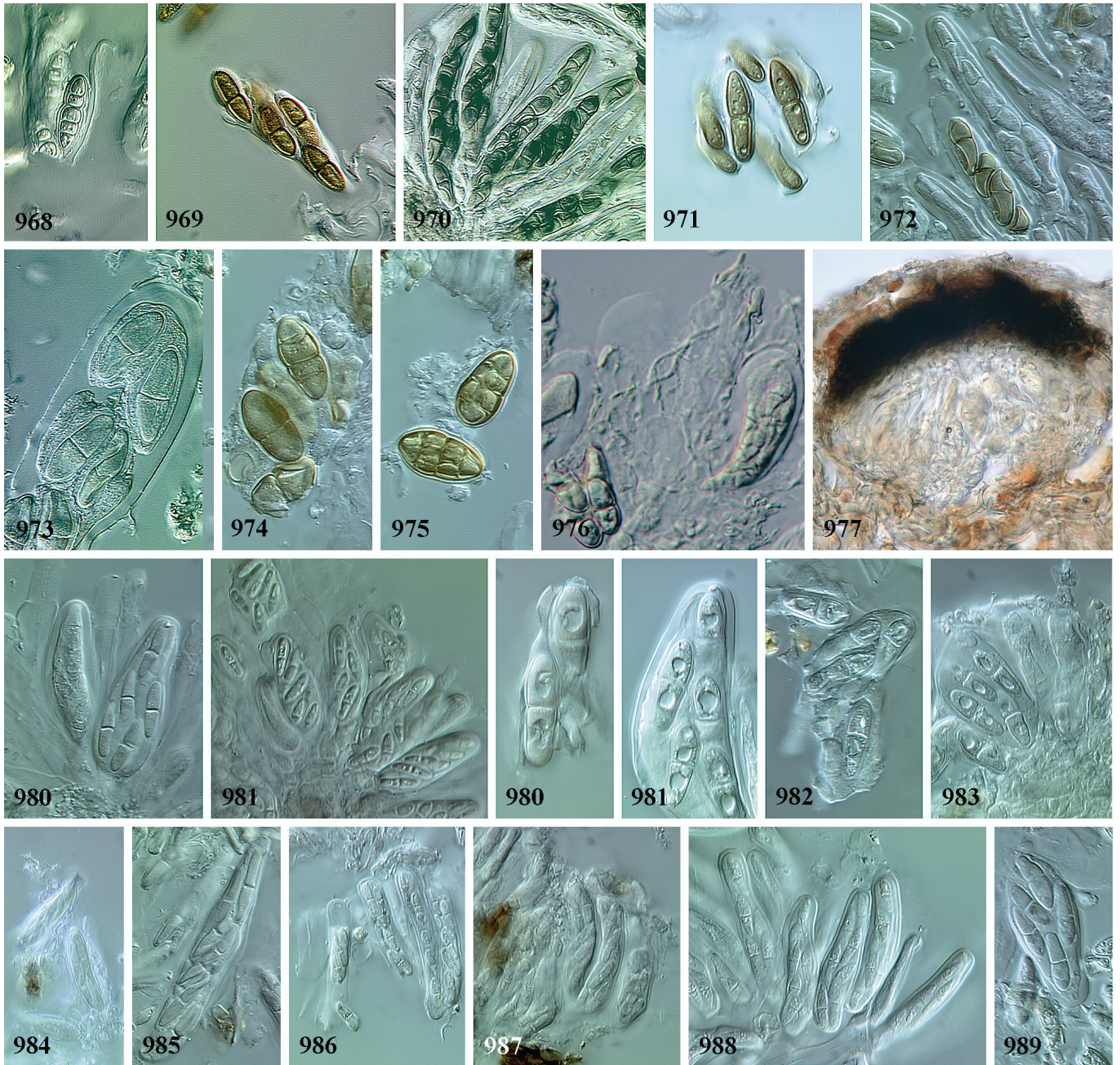
Figs 841-924. Outlines of ascospores, in water, not to scale. See main body of text for explanation. In general type specimens are illustrated; and if they were not available, the most characteristic other specimen.



Figs 925-961. Outlines of ascospores, in water, not to scale. See main body of text for explanation. In general type specimens are illustrated; and if they were not available, the most characteristic other specimen.



Figs 962-967. Habitus illustrations; 962-963 aggregated ascomata on lower surface of decaying leaves; 964 infected margins of living leaf; 965 stem with ascomata; 966-967 ascomata on waste paper.



Figs 968-989. Asci and ascospores, in water, not to scale. See main body of text for explanation. 968-977 various excluded taxa; 978-983 *Davidiella* species; 984-989 *Mycosphaerella* species.

ALPHABETICAL LIST OF MYCOSPHAERELLA AND SPHAERELLA NAMES

Note: this list is alphabetic on epithet. When a combination in *Mycosphaerella* exists, this is the first name given. If not, the entry usually starts with a combination in *Sphaerella*. The remaining names are given in chronological order, with full reference. Abbreviations follow the standard systems for journals (BPH/S, Bridson & Smith, 1991), books (TL-2, Stafleu & Cowan, 1976-1988) and herbarium acronyms (Holmgren *et al.*, 1990). Authors are following the standard system (Brummit & Powell, 1992).

+ Each entry consist of three or four lines. The first line gives all homotypic names. In case that the taxon was published in an exsiccate, the date given is the publication date. Note that a species and a heterotypic subspecific taxon can be published on the same label.

+ The second line gives the type country and host name and family (according to Farr *et al.*, 1979, but using when possible the phylogenetic classification of Bremer *et al.*, 2003) for each name; full details are given usually only when the type has actually been studied. In case that the collection was published in an exsiccate, the date given is the collection date.

+ The optional third line gives the anamorphs. Anamorphs mentioned (if any) are those cited in the literature.

+ The last line gives synonymies and dispositions reported in the literature, original observations by the author, preferably on type materials, additional materials studied if relevant and often a new disposition or an indication from which species it is morphologically indistinguishable.

In the case that different epithets exist for one homotypic species, the entry is alphabetized under the accepted or at least usable name. Epithets that merely are superfluous new names for valid homotypic taxa (e.g. the two entries sub *vulgaris*) are mentioned with only one line: the name and after "See" the name under which the full entry is given.

Mycosphaerella abietis (Rostr.) Lindau, Handb. Pflanzenkrankh., ed. 3, 2: 534. 1908 ["1906"] = *Sphaerella abietis* Rostr., Tidsskr. Planteavl. 1902: 597. 1902 = *Rehmiellopsis abietis* (Rostr.) O. Rostr., Dansk Bot. Ark. 2(5): 15. 1916 = *Delphinella abietis* (Rostr.) E. Müll., in E. Müll. & Arx, Beitr. Kryptogamenfl. Schweiz 11(2): 26. 1962.

Type — Denmark: Jutland, Horsens. On upper and lower surface of young, living needles of *Abies alba* ["*pectinata*"] (Pinaceae). Lind, V 1902 (BPI, isotype).

Anamorphs: *Dothiorella* *vide* Müller (op. cit.), *Phoma abietis* Briard and *Toxosporium abietinum* Vuill. *vide* Rostrup (op. cit.).

Accepted as *Delphinella abietis* (Rostr.) E. Müll. by Müller & Von Arx (op. cit.). In the isotype studied only a coelomycete could be found.

Mycosphaerella abutilontidicola Miura, Industr. Contr. 27: 166. 1928.

Type — China: *Abutilon avicennae* (Malvaceae).

No material has been seen and the location of the type is unknown.

Mycosphaerella acaciae (Cooke & Harkn.) Tomilin, Novosti Sist. Nizsh. Rast. 1968: 162. 1968 = *Sphaerella acaciae* Cooke & Harkn., Grevillea 9: 9. 1880.— Fig. 1, 968.

Type — USA: California, San Francisco, Golden Gate Park. On upper and lower surface of dead leaves of *Acacia* (Fabaceae). Harkness no. 1415, IV 1880 (BPI, holotype).

This is a new synonym of *Oletheriostrigula papulosa* (Durieu & Mont.) Huhndorf & R.C. Harris, with asci clavate, surrounded by 1-2 µm wide, mostly simple pseudoparaphyses, ascospores consistently 4-septate and sole-shaped, 19-22 × 4-5.5 µm, surrounded by a gelatinous sheath.

Mycosphaerella acaciigena Crous & M.J. Wingf., Stud. Mycol. 50: 463. 2004.

Type — Venezuela: Acarigua, on leaves of *Acacia mangium* (Fabaceae). Wingfield, V 2000 (CBS 9873, holotype); cultures ex-type CBS 115432, 112515, 112516 = CPC 3836-3838.

Anamorph: *Pseudocercospora acaciigena* Crous & M.J. Wingf. *vide* Crous *et al.* (op. cit.).

No material was studied of this recently described species.

Sphaerella acaenae Henn., Deutsche Südpolar-Exped. 1901-1903, Bot., Pilze, 8: 9. 1907 ["1906"].

Type — Kerguelen Island: *Acaena ascendens* (Rosaceae). No material could be studied, as the type and only material in B has been destroyed by fire.

Mycosphaerella acanthopanacis Syd. & P. Syd., Ann. Mycol. 11: 59. 1913 = *Sphaerella acanthopanacis* (Syd. & P. Syd.) Trotter, Syll. Fung. 24: 890. 1928, later homonym (illegitimate, Article 53).— Fig. 2.

Type — Japan: Mino, Kawanye-mura. On upper and lower surface of dead leaves of *Acanthopanax ricinifolium* (Araliaceae). Hara, 1912 (S, holotype).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradians*, with tiny ascomata, asci cylindrical, ascospores 13-15 × 3-3.5 µm. Additional material (Latvia, Vidzeme, Gulbene, on *A. sessilis*, Starcs no. 48, IV 1933, B) is postmature.

Mycosphaerella acanthopanacis (Cif. & Gonz. Frag.) Cif., Quaderno 19: 230. 1961, later homonym (illegitimate, Article 53) = *Sphaerella acanthopanacis* Cif. & Gonz. Frag., in Gonz. Frag. & Cif., Bol. Soc. Esp. Hist. Nat. 25: 447. 1925.

Type — Dominican Republic: *Acanthopanax aculeatum* (Araliaceae).

No material could be found in either the Ciferri herbarium in BPI, nor in MA, where the material of Gonz. Frag. is kept.

Sphaerella acerifera Cooke, J. Bot. 4: 248. 1866 = *Laestadia acerifera* (Cooke) Sacc., Syll. Fung. 1: 423. 1882 = *Plagiostomella acerifera* (Cooke) Petr. & Syd., Ann.

Mycol. 22: 362. 1924 = *Apioplagiostoma acerifera* (Cooke) M.E. Barr, Mycol. Mem. 7: 103. 1978.

Type — United Kingdom: Kent, Darenth. On upper and lower surface of dead leaves of *Acer campestre* (Sapindaceae). Cooke, IV 1874, distributed in Fungi Britannici no. 687 (K, holotype), also distributed in Fungi Britannici, ed. 2 no. 271 (K, isotype).

Included in the synonymy of *Apioplagiostoma acerinum* (Starbäck) Monod by Eriksson (1992), with which the types studied agree well.

Mycosphaerella acerina (Wallr.) Johanson ex Oudem., Rév. Champ. Pays-Bas 2: 212. 1897 = *Sphaeria acerina* Wallr., Fl. Cryptog. Germ. 2: 770. 1833 = *Sphaerella acerina* (Wallr.) Fuckel, Jahrb. Nassauischen Vereins Naturk. 23-24: 99. 1870.

Type — Germany: On lower surface of dead leaves of *Acer pseudoplatanus* (Sapindaceae).

The type was not found in B or elsewhere. Two specimens studied (Netherlands, Naaldwijk, van Trappen, XI 1865, L; also Germany, Oestrich, on *A. campestre* ["*pseudoplatanus*"], Fuckel, Fungi Rhenani Exsiccati no. 845, L) were both immature with *Asteromella* spermatial state, suggesting *M. punctiformis*. It was cited as synonymous with *M. punctiformis* by Tomilin (1979).

Mycosphaerella aceris Woron., Vestn. Tiflissk. Bot. Sada 35: 7. 1914 = *Sphaerella aceris* (Woron.) Sandu, Ciuperici Pyrenomycetes-Sphaeriales din România: 118. 1971, lacking reference to basionym, not validly published (Article 33.2).

Type — Georgia: *Acer laetum* (Sapindaceae).

No material was studied, as it was not sent on loan from LE or LEP.

Mycosphaerella acerna (Fautrey) Tomilin, Novosti Sist. Nizsh. Rast. 6: 118. 1970 ["1969", as "*acerina*"] = *Sphaerella acerna* Fautrey, in Roum., Rev. Mycol. (Paris) 13: 166. 1891.— Fig. 3.

Type — France: Côte-d'Or. On upper and lower surface of dead leaves of *Acer campestre* (Sapindaceae). Fautrey, distributed in Roumeguère, Fungi Selecti Exsiccati no. 5830, IV 1891 (PC, holotype).

Already cited as synonymous with *M. punctiformis* by Tomilin (1979), with which the holotype studied agrees well, with asci cylindrical, ascospores 7-10 × 2-2.5 µm.

Mycosphaerella acervata (Ellis & Everh.) M.E. Barr, Contr. Univ. Michigan Herb. 9: 596. 1972 = *Diatrype acervata* Ellis & Everh., J. Mycol. 4: 75. 1888 = *Diaporthe acervata* (Ellis & Everh.) Ellis & Everh., N. Amer. Pyrenomyc.: 738. 1892 = *Valsa acervata* (Ellis & Everh.) Kuntze, Revis. Gen. Pl. 3(2): 539. 1898 = *Planistromella acervata* (Ellis & Everh.) M.E. Barr, Mycotaxon 60: 434. 1996.— Fig. 4.

Type — USA: New Jersey, Newfield. On white spots with brown margins on upper and lower surface of living leaves of *Yucca filamentosa* (Asparagaceae). Ellis, VII 1888 (NY, holotype), also distributed in North American Fungi no. 2124 (NY, 5 isotypes).

Anamorph: given as *Coniothyrium* on the label, but presents a species of *Microsphaeropsis* (pers. obs.).

Accepted as *Planistromella acervata* (Ellis & Everh.) M.E. Barr by Barr (op. cit.), with ascomata aggregated on erumpent stromata, asci cylindrical, surrounded by large parenchymatal interascal cells, ascospores 22-30 (24-29 in the type) × 3.5-5 µm.

Sphaerella acetosellae G.H. Otth, nomen herbariorum (not validly published, Article 32) [as "*acetosella*"].

Authentic material — Germany: Brunngartenwald, on brown spots on upper and lower surface of living leaves of *Oxalis acetosella* (Oxalidaceae). Otth no. 142, VIII 1867 (B).

The material contains only a coelomycete.

Mycosphaerella achilleae Novoss., Bot. Mater. Otd. Sporov. Rast. Bot. Inst. Akad. Nauk SSSR 4(10-12): 35. 1938.

Type — France: *Achillea macrophylla* (Asteraceae).

No material was studied, as it was not sent on loan from LE or LEP.

Mycosphaerella acicola (Cooke & Harkn.) Lindau, in Engl. & Prantl, Natürlichen Pflanzenf. 1(1): 425. 1897 = *Sphaerella acicola* Cooke & Harkn., Grevillea 13: 20. 1884, non *Oligostroma acicola* Dearn., non *Eruptio acicola* (Dearn.) M.E. Barr, non *Mycosphaerella dearnessii* M.E. Barr.— Fig. 5.

Type — USA: California, San Francisco, Cemetery. On upper and lower surface of dead needles of *Pinus insignis* (Pinaceae). Harkness no. 2303, IV 1881 (K, holotype; BPI, K, isotypes); IV 1882 (BPI, topotype).

This is morphologically indistinguishable from *Mycosphaerella juniperina*, of which the heterotypic *Eruptio acicola* is also a synonym, with asci pyriform, ascospores 7-9 × 2.5-3 µm.

Mycosphaerella acilegna M. Morelet, Bull. Soc. Sci. Nat. Archéol. Toulon & Var 196: 7. 1971, nomen novum (Article 58) for *Mycosphaerella angelicae* (Fr.) Petr., Ann. Mycol. 19: 203. 1921, later homonym (illegitimate, Article 53), as Potebnia (Ann. Mycol. 8: 46. 1910) did not make the new combination = *Dothidea angelicae* Fr., Systema Mycol. 2: 561. 1823 = *Asteroma angelicae* (Fr.) Fr., Summa Veget. Scand. 2: 425. 1849 = *Phyllachora angelicae* (Fr.) Fuckel, Jahrb. Nassauischen Vereins Naturk. 23-24: 219. 1870 = *Oligostroma angelicae* (Fr.) Höhn. ex Petr., Ann. Mycol. 19: 203. 1921 = *Carlia angelicae* (Fr.) Höhn., Centralbl. Bakteriol., 2. Abth., 60: 2. 1923 = *Mycosphaerella friesii* Tomilin, Opredeletel' gribov roda *Mycosphaerella* Johans.: 145. 1979, superfluous nomen novum (illegitimate, Article 52).— Fig. 6.

Type — Kamchatka: On dead stems of *Angelica sylvestris* (Apiaceae).

According to Eriksson (1992), the type contains no *Mycosphaerella*. Material studied (Czech Republic, Mähren, Neudeck, Petrak, VI 1937, distributed in Mycotheca Generalis no. 1849, B) belongs to section *Caterva* and is morphologically indistinguishable from *M. subradians*, with asci cylindrical, ascospores 11-13 × 2.5-3.5 µm.

Mycosphaerella aconitorum Petr., Sydowia 10: 282. 1957 [“1956”].

Type — Austria: *Aconitum* (Ranunculaceae).

Cited as synonymous with *Mycosphaerella hieraciophila* by Tomilin (1979). The type nor any other material was found in W or elsewhere.

Mycosphaerella acori Höhn., in Strasser, Verh. Zool.-Bot. Ges. Wien 69: 360. 1919 = *Sphaerella acori* (Höhn.) Trotter, Syll. Fung. 24: 850. 1928.

Type — Austria: *Acorus calamus* (Acoraceae).

No type material was preserved in FH.

Mycosphaerella acrocomiicola Bat., Anais 5 Reun. Anu. Soc. Bot. Brasil 1954: 116. 1958.— Fig. 7.

Type — Brazil: On white leaf sections on upper surface of leaves of *Acrocomia intumescens* (Arecaceae). Batista exs. no. 14819 (URM 5194, holotype).

Anamorph: associated with *Gloeosporium cocophilum* Wakef. in the same spots.

This belongs to *Davidiella*, of which it represents a parasitic species, with asci pyriform, ascospores $9.5-11 \times 3-3.5 \mu\text{m}$. Therefore the following new combination is made:

Davidiella acrocomiicola (Bat.) Aptroot comb. nov., **MB 500338**. **Basionym:** *Mycosphaerella acrocomiicola* Bat., Anais 5 Reun. Anu. Soc. Bot. Brasil 1954: 116. 1958. It is morphologically indistinguishable from the Chagos material (not the type) of *Mycosphaerella cocophylla*.

Mycosphaerella actaeae (Rostr.) Tomilin, Novosti Sist. Nizsh. Rast. 1968: 163. 1968 = *Sphaerella actaeae* Rostr., Skr. Vidensk.-Selsk. Christiana, Math.-Naturvidensk. Kl. 1904: 21. 1904.

Type — Denmark: *Actaea spicata* (Ranunculaceae).

The type nor any other material was found in C or elsewhere.

Mycosphaerella actinidiae Syd. & Hara, in Syd. & P. Syd., Ann. Mycol. 11: 59. 1913 = *Sphaerella actinidiae* (Syd. & Hara) Trotter, Syll. Fung. 24: 860. 1928.

Type — Japan: Mino, Kawanye-mura. On upper and lower surface of dead leaves of *Actinidia* (Actinidiaceae). Hara, IV 1912 (S, holotype).

This is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, but ascospores immature.

Mycosphaerella adenophorae Sawada, Bull. Gov. Forest Exp. Sta. 53: 164. 1952.

Type — Taiwan: *Adenophora verticillata* (Campanulaceae). No material was found in BPI, where most Sawada types are.

Mycosphaerella adhatodae S. Ahmad, Biologia (Lahore) 15: 1. 1969.

Type — Pakistan: *Adhatoda vesca* (Acanthaceae).

No type material was preserved in ZT.

Mycosphaerella adonidina Petr., Sydowia 13: 76. 1959.

Type — Austria: Baden. On dead stems of *Adonis vernalis* (Ranunculaceae). Petr., V 1940 (W, holotype).

Cited as synonymous with *Mycosphaerella hieraciophila* by Tomilin (1979). The type contains an immature *Didymella*, with asci surrounded by pseudoparaphyses.

Mycosphaerella adonis (Sacc.) Lindau, Hilfsb. Sammeln Ascomyceten: 6. 1903 = *Sphaerella adonis* Sacc., Michelia 1: 122. 1878.— Fig. 8.

Type — Germany: Brandenburg. On upper and lower surface of dead leaves of *Adonis vernalis* (Ranunculaceae). Reinhardt (PAD, holotype).

The type shows that this belongs to section *Longispora*, and is morphologically indistinguishable from *M. latebrosa*, with asci cylindrical, ascospores $26-31 \times 4-5 \mu\text{m}$. Additional material studied (Unterfranken, Gerolzhofen, Sulzheim, Vill, III 1907, distributed in Fungi Bavarici no. 821, B, PAD) is immature.

Sphaerella adoxae Fuckel, Jahrb. Nassauischen Vereins Naturk. 27-28: 21. 1873.— Fig. 9.

Type — Germany: Rennrod. On upper and lower surface of dead tips of living leaves of *Adoxa moschatellina* (Adoxaceae). Fuckel, Fungi Rhenani Exsiccati no. 2645 (L, isotype).

The type material studied is immature.

Sphaerella adunca Niessl. See *Mycosphaerella adusta* Lindau.

Mycosphaerella adusta Lindau, Hilfsb. Sammeln Ascomyceten: 31. 1903, nomen novum (Article 58) for *Sphaerella adusta* Niessl, Oesterr. Bot. Z. 25: 86. 1875, later homonym (illegitimate, Article 53) = *Sphaerella adunca* Niessl, nomen herbariorum (not validly published, Article 32).

Type — Czech Republic: Karthaus. On dead stems of *Convolvulus arvensis* (Convolvulaceae). Niessl, VI 1874 (M, holotype [sub “*adunca*”]).

This belongs to section *Caterva* and is morphologically indistinguishable from *M. superflua*, with asci cylindrical, ascospores $14-17 \times 3-4.5 \mu\text{m}$.

Mycosphaerella adusta (Fuckel) Jacz., Opredelitel' gribov 2: 613. 1917, later homonym (illegitimate, Article 53) = *Sphaerella adusta* Fuckel, in Heuglin, Reisen nach dem Nordpolarmeer 3: 320. 1874 = *Venturia adusta* (Fuckel) E. Müll., Sydowia 11: 87. 1958 = *Sphaerella effusa* Sacc. & P. Syd., Syll. Fung. 14: 333. 1899, superfluous (illegitimate, Article 52) nomen novum (Article 58).

Type — Russia: *Epilobium latifolium* (Onagraceae).

Accepted as *Venturia adusta* (Fuckel) E. Müll. by Sivanesan (1977).

Mycosphaerella advena Syd., Ann. Mycol. 28: 82. 1930.— Fig. 10, 969.

Type — Venezuela: Puerto La Cruz. On upper surface of dead leaves of cf. *Maximilliana* (Arecaceae). Sydow, XII 1927 (B, BPI, isotypes).

This is morphologically indistinguishable from *Arecophila chamaeropsis* (Bertault & Malençon) K.D. Hyde, with ascomata immersed, asci bitunicate, paraphyses 2-3 μm wide, simple, septate, ascospores golden brown, $15-16 \times$

4.5-5 µm, ornamented with warts. Therefore the following new combination is proposed: ***Arecophila advena*** (Syd.) Y.Z. Wang, Aptroot & K.D. Hyde comb. nov., **MB 500338**. **Basionym:** *Mycosphaerella advena* Syd., Ann. Mycol. 28: 82. 1930.

Mycosphaerella aegopodii Potebnia, K istorii razvitiya nekotorykh askomitsetov. 1. *Mycosphaerella*. 2. *Gnomonia*, *Glomerella* i *Pseudopeziza*: 84. 1908 ≡ *Sphaerella aegopodii* (Potebnia) Sacc. & Trotter, in Sacc. & Traverso, Syll. Fung. 20: 817. 1911.

Type — Russia: Prov. Wiatka, near Wiatka. On small white spots on upper and lower surface of living leaves of *Aegopodium podagraria* (Apiaceae). Cardacova, VIII 1915 (BPI, topotype).

Anamorphs: *Phyllosticta aegopodii* Fuckel and *Septoria podagrariae* Lasch fide Potebnia (op. cit.).

Cited as synonymous with *Mycosphaerella podagrariae* (Roth : Fr.) Petr. by Eriksson (1992). The topotype studied contains only the two anamorphs mentioned by Potebnia.

Mycosphaerella aeluropodis Lobik, Materialy po floristicheskim i faunisticheskim obsledovaniyam Terskogo okruga: 25. 1928 [as “*aeluropodi*”].

Type — Russia: *Aeluropus litoralis* (Poaceae).

No material was studied, as it was not sent on loan from LE or LEP.

Sphaerella aequalis (Cooke) Auersw., in Gonn. & Rabenh., Mycol. Europaea 5-6: 4. 1869 ≡ *Sphaerella maculiformis* var. *aequalis* Cooke, J. Bot. 4: 246. 1866 [as “*maculaeformis* var.”].

Type — United Kingdom.

Cited as synonymous with *M. punctiformis* by Tomilin (1979). No material was found in K or elsewhere.

Mycosphaerella aequatoriensis Petr., Sydowia 2: 342. 1948.— Fig. 11.

Type — Ecuador: Prov. Tungurahua, Baños. On white spots on the upper surface of living leaves of *Eugenia* (Myrtaceae). Sydow no. 622 (BPI, isotype), also distributed in Reliquiae Petrakianae no. 1244, XII 1937 (H, L, isotypes).

Anamorph: *Septoria pichinchica* Petr. fide Petrak (1948).

This is a parasitic species, with asci clavate, ascospores 16-19 × 3-4 µm.

Mycosphaerella aesculi (Cocc. & Morini) Tomilin, Novosti Sist. Nizsh. Rast. 1968: 163. 1968 ≡ *Sphaerella aesculi* Cocc. & Morini, Mem. Reale Accad. Sci. Ist. Bologna, ser. 4, 8: 23. 1887.

Type — Italy: *Quercus* (Fagaceae).

Anamorph: *Septoria aesculicola* (Fr.) Fuckel fide Tomilin (1979).

No material has been found in any of the herbaria consulted.

Mycosphaerella aethiops (Fuckel) Lindau, Hilfsb. Sammeln Ascomyceten: 93. 1903 ≡ *Sphaeria aethiops* Fuckel, Hedwigia 3: 161. 1864 ≡ *Sphaerella aethiops* (Fuckel) Auersw., in Gonn. & Rabenh., Mycol. Europaea 5-6: 5. 1869.— Fig. 12.

Type — Germany: Oestrich. On lower surface of dead leaves of *Quercus robur* (Fagaceae). Fuckel, Fungi Rhenani Exsiccati no. 818, (BPI, L, isotypes).

The type material studied in L is immature, but the type in BPI is morphologically indistinguishable from additional material studied (Brandenburg, Bellinchen, on *Quercus pedunculata*, Sydow, Mycotheca Germanica no. 2686, VI 1933, L) which is *M. punctiformis*, with ascospores 9-11 × 3-3.5 µm. The type in BPI has asci cylindrical, ascospores 8.5-10 × 2.5-3 µm. Therefore, this species is morphologically indistinguishable from *M. punctiformis*.

Mycosphaerella aethiops var. *aroniae* Rehm, Ascomyceten no. 1344. 1900.— Fig. 13.

Type — Italy: Lago di Garda, Torbole, Monte Baldo. On upper and lower surface of dead leaves of *Aronia rotundifolia* (Rosaceae). Rehm, V 1900, distributed in Ascomyceten no. 1344 (B, isotype).

This is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores 7-9 × 2-2.5 µm.

Mycosphaerella affinis (G. Winter) Starbäck, Bih. Kongl. Svenska Vetensk.-Akad. Handl. 15(3, 2): 9. 1899 ≡ *Sphaerella affinis* G. Winter, in Thüm., Mycoth. Univ., cent. 1 no. 59. 1875.— Fig. 14.

Type — Germany: Bayreuth. On upper and lower surface of dead leaves of *Carlina vulgaris* (Asteraceae). Thümen, 1874, distributed in Mycotheca Universalis no. 59 (B, BPI, NY, isotypes).

Anamorph: *Cercospora carlinae* Sacc. fide Eriksson (1992) [= *Passalora carlinae* (Sacc.) U. Braun & Crous].

The type and additional material studied (Hungary, Eger, Jahn, VII 1916, distributed in Petrak, Flora Bohemiae et Moraviae Exsiccata no. 1234, L) belong to section *Longispora*, and show that this is morphologically indistinguishable from *M. millegrana*, with asci cylindrical, ascospores 17-22 × 3-4 µm.

Mycosphaerella afghanica Petr., Sydowia 7: 88. 1953.— Fig. 15.

Type — Afghanistan: Nuristan, Kamdesch. On upper and lower surface of dead spiny leaves of *Astragalus* (Fabaceae). Zilli, VI 1950 (W, holotype).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradicans*, with ascospores partly in dense groups, asci cylindrical, ascospores 10-13 × 3-3.5 µm.

Mycosphaerella africana Crous & M.J. Wingf., Mycologia 88: 450. 1996.

Type — South Africa: Western Cape, Stellenbosch, Stellenbosch Mountain. On spots on leaves of *Eucalyptus viminalis* (Myrtaceae). Crous, X 1994 (PREM 51917, holotype, not seen).

No material was studied of this recently described species.

Mycosphaerella agapanthi (Kalchbr. & Cooke) Lindau, in Engl. & Prantl, Natürlichen Pflanzenf. 1(1): 426. 1897 ≡ *Sphaerella agapanthi* Kalchbr. & Cooke, Grevillea 9: 31. 1880.— Fig. 16.

Type — South Africa: Cape. On upper surface of dead leaves of *Agapanthus umbellatus* (Alliaceae). Kalchbrenner no. 1342 (K, holotype; K, 2 isotypes).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. ammophilae*, with asci pyriform, ascospores 10-12 × 4-5 µm. Additional materials studied (Natal, Kentani, Pegler, 1916, B, 2×; also Tanzania, Lyamungu, Moshi no. 280/51, X 1951, IMI 150330; also Kenya, Nairobi, Gatumbi no. 3638, V 1925, IMI 196249; also Zimbabwe, Salisbury, Rothwell no. 18197, I 1963, IMI 98929; also Uganda, Kawanda, Leighton no. 00482, I 1962, IMI 93552) belongs also to *Davidiella*, but are *D. allicina*, with asci pyriform, ascospores 16-19 × 5-6.5 µm; the KwaZulu-Natal material contains only a coelomycete.

Mycosphaerella agapanthi T.S. Ramakr. & K. Ramakr. See *Mycosphaerella agapanthi-umbellati* T.S. Ramakr. & Sundaram.

Mycosphaerella agapanthi-umbellati T.S. Ramakr. & Sundaram, Proc. Indian Acad. Sci., Sect. B, 41: 190. 1955, nomen novum (article 58) for *Mycosphaerella agapanthi* T.S. Ramakr. & K. Ramakr., Proc. Indian Acad. Sci., Sect. B, 32: 98. 1951 ["1950"], later homonym (illegitimate, Article 53).

Type — India: *Agapanthus umbellatus* (Alliaceae).

No material was found in BPI, where other types from Ramakrishnan were preserved.

Mycosphaerella agaves (C. Massal.) Tomilin, Novosti Sist. Nizsh. Rast. 1967: 188. 1967 [as "*agavis*"] = *Sphaerella agaves* C. Massal., Atti Reale Ist. Veneto Sci. 74: 258. 1914 [as "*agavis*"].

Type — Italy: Treguago. On upper surface of dead leaves of *Agave americana* (Asparagaceae). Massalongo (PAD, isotype).

The type contains mostly coelomycetes, probably also a immature *Dothidea*.

Mycosphaerella aggregata (Schwein.) Stev., J. Agric. Univ. Puerto Rico 2(3): 155. 1918 = *Sphaeria aggregata* Schwein., Trans. Amer. Philos. Soc., ser. 2, 4: 213. 1832.

Type — USA: Philadelphia, Bethlehem. On wood. Schweinitz (PH, holotype; PH, isotype).

The type material contains an empty ascomycete, possibly *Melanomma pulvis-pyrius* (Pers. : Fr.) Fuckel.

Mycosphaerella aggregata Earle, in Seaver & Chardón, Sci. Surv. Porto Rico & Virg. Isl., New York Acad. Sci., 8: 61. 1926, later homonym (illegitimate, Article 53).— Fig. 18.

Type — Puerto Rico: Yauco. On upper surface of dead leaves of *Hemerocallis* ["*Hymerocallis*"] (Xanthorrhoeaceae). Heller no. 70a, 1899 (NY, holotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores 14-16 × 5-6.5 µm.

Mycosphaerella aggregata Carnegie & Keane see *Mycosphaerella gregaria* Carnegie & Keane.

Mycosphaerella agostinii (Nann.) M. Morelet, Bull. Soc. Sci. Nat. Archéol. Toulon & Var 196: 7. 1971 = *Sphaerella agostinii* Nann., Atti Reale Accad. Fisiocrit. Siena, ser. 10, 2: 437. 1927 = *Sphaerella crithmi* Nann., nomen herbariorum (not validly published, Article 32).— Fig. 17.

Type — Italy: Isoletto di S. Nicolicevlio. On upper and lower bracts and leaves of *Crithmum maritimum* (Apiaceae). Nannizzi, IX 1927 (SIENA, holotype, sub. "*crithmi* sp. nov.").

This belongs to *Davidiella* and is morphologically indistinguishable from *D. ammophilae*, with asci pyriform, ascospores 10-13 × 3-4.5 µm.

Mycosphaerella agrimoniae Syd. & P. Syd., Ann. Mycol. 40: 200. 1942.— Fig. 19.

Type — Germany: Brandenburg, Oberbarnim. On upper and lower surface of dead leaves of *Agrimonia eupatoria* (Rosaceae). Sydow, VI 1941, distributed in Mycotheca Germanica no. 3501 (L, isotype).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. superflua*, with asci clavate, ascospores 16-20 × 3-4 µm.

Mycosphaerella agrostidis (Castagne) Lindau, Hilfsb. Sammeln Ascomyceten: 6. 1903 = *Sphaeria agrostidis* Castagne, Cat. Plantes Marseille, Aix. 1845 = *Phyllachora agrostidis* (Castagne) Fuckel, Jahrb. Nassauischen Vereins Naturk. 23-24: 217. 1870 [as "*agrostis*"] = *Sphaerella agrostidis* (Castagne) Auersw., in Gonn. & Rabenh., Mycol. Europaea 5-6: 17. 1869.

Type — France: *Agrostis stolonifera* (Poaceae).

Cited as synonymous with *Davidiella allicina* (as *Mycosphaerella allicina*) by Tomilin (1979). The type was not received from PC.

Mycosphaerella agrostistachydis Anahosur, Sydowia 24: 287. 1970 ["1971, as *agrostistachidis*"].

Type — India: *Agrostistachys indica* (Poaceae).

No material has been found from this taxon in ZT or elsewhere.

Mycosphaerella aiacu (Speg.) Aptroot comb. nov., **MB 500508**. **Basionym:** *Sphaerella aiacu* Speg., Bol. Acad. Nac. Ci. 11: 204. 1888 ["1887"].— Fig. 20.

Type — Argentina: Isla Picton. On spots on upper and lower surface of living leaves of *Maytenus* (Celastraceae). Spegazzini no. 6179, V 1882 (LPS, holotype).

This is a parasitic species, with asci cylindrical, ascospores 10-13 × 1.5-2 µm.

Sphaerella ailanthi Cooke, Grevillea 6: 146. 1878.

Type — USA: South Carolina, Aiken. On lower surface of dead leaves of *Ailanthus* (Simaroubaceae). Ravenel, Fungi Americani Exsiccati no. 377 (BPI, K (3×), NY (3×), isotypes).

The type materials are postmature, but show that this is morphologically indistinguishable from *M. punctiformis*.

Mycosphaerella ailanthi House, New York State Mus. Bull. 233-234: 25. 1921, nomen novum (Article 58) for *Sphaerella ailanthi* Ellis & Barthol., New York State Mus.

Bull. 188: 40. 1916, nomen nudum (not validly published, Article 32) & later homonym (illegitimate, Article 53).

Type — USA: *Ailanthus* (Simaroubaceae).

No material was preserved in NY (where the Ellis herbarium is kept) or BPI.

Mycosphaerella ailanthina Sacc., nomen herbariorum (not validly published, Article 32).— Fig. 21. Authentic material — Portugal: Coimbra. On upper and lower surface of dead leaves of *Ailanthus glandulosa* (Simaroubaceae). Trotter, II 1891 (PAD).

The material belongs to *Davidiella*, and is morphologically indistinguishable from *D. clandestina*, with asci pyriform, ascospores 21-25 × 5.5-7.5 µm.

Mycosphaerella airicola Petr., Ann. Mycol. 34: 445. 1936.— Fig. 22.

Type — Finland: Lapponia, Laanik. On dead leaves and culms of *Aira caespitosa* (Poaceae). Kari, VI 1931 (W, holotype).

Anamorph: *Selenophoma fide* Eriksson (1992).

Cited as synonymous with *Guignardia graminicola* (Rostr.) Vasyag. by Eriksson (1992). However, examination of the type shows that this belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores 16-19 × 5-6 µm. On the type no *Guignardia* could be found, but *Pleospora herbarum* (Pers.: Fr.) Rabenh. was present as well.

Mycosphaerella alarum (Ellis & Halst.) Hara, A list of Japanese fungi hitherto known, ed. 3: 229. 1927 ≡ *Sphaerella alarum* Ellis & Halst., in Ellis & Everh., N. Am. Pyrenomyc.: 279. 1892.

Type — USA: New Jersey, New Brunswick. On dead fruits ["samarae"] of *Acer* (Sapindaceae). Halsted, IV 1891 (NY, holotype).

This is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores immature, ca 10-12 × 2.5-3 µm.

Mycosphaerella alba (Pass.) Tomilin, Novosti Sist. Nizsh. Rast. 1967: 183. 1967 ≡ *Sphaerella alba* Pass., Atti Reale Accad. Lincei, Mem. Cl. Sci. Fis., ser. 4, 7(2, 2): 44. 1891.— Fig. 23.

Type — Italy: *Populus alba* (Salicaceae).

No type material was found in any of the herbaria consulted. Material studied (Germany, Bremen, on bark, herb. Treviranus, B) is *Lophiostoma corticolum* (Fuckel) E.C.Y. Liew, Aptroot & K.D. Hyde, with asci thick-walled, pseudoparaphyses 2 µm wide, ascospores 3-septate, 25-27 × 7-8 µm.

Mycosphaerella albescens (Rabenh.) Lind ex Rehm, Ann. Mycol. 9: 3. 1911 ≡ *Sphaeria albescens* Rabenh., Deutsch. Kryptog.-Fl. 1: 192. 1844.— Fig. 24.

Type — Switzerland: On dead stems of *Vincetoxicum officinale* (Apocynaceae).

No type material was found in any of the herbaria consulted. Material studied (Czech Republic, Mähren, Sternberg, Piskoř, V 1928, distributed in Petrak, Mycotheca Generalis no. 262, B) belongs to section *Caterva*, and is

morphologically indistinguishable from *M. subradicans*, with asci cylindrical, ascospores 9-11 × 2.5-3 µm.

Mycosphaerella albizziae U.P. Singh, Mycologia 68: 175. 1976.— Fig. 25.

Type — India: On brown spots on upper and lower surface on living leaves of *Albizzia lebbek* (Fabaceae).

The location of the type is unknown and its preservation uncertain. Material studied (Uttar Pradesh, Seorahi, Deoria, Verma, X 1988, IMI 333084; also Gorakhpur University, on *Albizzia* sp., Verma, XII 1980, IMI 254019; also Rai, I 1979, IMI 234816) is all identical and is a parasitic species, with asci cylindrical, ascospores 14-16 × 3.5-4.5 µm.

Sphaerella albocrustata (Schwein.) Cooke, J. Bot. 21: 68. 1883 ≡ *Sphaeria albocrustata* Schwein., Trans. Amer. Philos. Soc., ser. 2, 4: 224. 1832 ≡ *Mycosphaerella albocrustata* (Schwein.) Crous & Corlett, Canad. J. Bot. 76: 1525. 1998.

Type — USA: Philadelphia. On lower surfaces of dead leaves of *Platanus* (Platanaceae). Schweinitz (PH, holotype; PH, isotype).

The type material is sterile, but it probably represents morphologically indistinguishable from *M. punctiformis*. Crous and Corlett (op. cit.) studied fertile possible isotype material in BPI and accepted the species, although they do not discuss the (probably non-existent) differences with *M. punctiformis*.

Sphaerella alboi Unamuno, Anales Jard. Bot. Madrid 1: 26. 1941.

Type — Spain: *Astragalus narbonensis* (Fabaceae).

No material could be studied, as the type was not found in MA.

Sphaerella alchemillae Kalchbr., Math. Természettud. Közlem. 7: 257. 1868.

Type — Hungary: *Alchemilla vulgaris* (Rosaceae).

No material could be studied, as the type was probably destroyed in B.

Mycosphaerella alchemillicola Vassiljevsky, Bolezni Rast. 14: 25. 1926 [as "alchimillicola"].

Type — Russia: *Alchemilla cymatophylla* (Rosaceae).

Anamorph: *Ovularia schroeteri* (Kühn) Sacc. fide Tomilin (1979) (= *Ramularia aplospora* Speg.).

No material of this species was sent on loan from LE or LEP.

Mycosphaerella alchemillicola Petr., Kryptog. Forsch. 2: 166. 1931, later homonym (illegitimate, Article 53).— Fig. 26.

Type — Germany: Allgau, Luckerskopf. On upper and lower surface of dead leaves of *Alchemilla* (Rosaceae). Ade, VII 1909 (W, holotype).

Cited as synonymous with *Mycosphaerella alchemillicola* Vassiljevsky by Tomilin (1979). The type shows that it belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradicans*, with asci cylindrical, ascospores 12-14 × 3-3.5 µm.

Mycosphaerella aleuritidicola Khokhr., Bot. Mater. Otd. Sporov. Rast. Bot. Inst. Komarova Akad. Nauk SSSR 7: 143. 1951 = *Mycosphaerella aleuritidis* Gutner, Opređ.: 536. 1966, later homonym (illegitimate, Article 53).

Type — Georgia: *Aleurites cordata* (Euphorbiaceae). Anamorph: *Ascochyta aleuritis* Khokhr. *fide* Tomilin (1979).

No material of this species was sent on loan from LE or LEP.

Mycosphaerella aleuritidis (I. Miyake) S.H. Ou, Sinensia 11: 183. 1940, based on an anamorph (illegitimate, Article 59) = *Cercospora aleuritidis* I. Miyake, Bot. Mag. (Tokyo) 26: 66. 1912 [as “*aleuritis*”] = *Pseudocercospora aleuritidis* (I. Miyake) Deighton, Mycol. Pap. 140: 138. 1976 [as “*aleuritis*”].

Type — Japan: *Aleurites* (Euphorbiaceae).

Anamorph: *Pseudocercospora aleuritidis* (I. Miyake) Deighton *fide* Deighton (op. cit.).

Mycosphaerella aleuritidis Gutner. See *Mycosphaerella aleuritidicola* Khokhr.

Mycosphaerella algarbiensis Dias, Mem. Soc. Brot. 21: 49. 1971.

Type — Portugal: Algarve, Albufeira. On dead stems of *Ruta chalepensis* (Rutaceae). Dias & Lucas 2289b, IV 1968 (LISE 91371, holotype).

The material contains *Pleospora herbarum* (Pers. : Fr.) Rabenh., a *Phoma*, and an immature *Mycosphaerella* with small ascospores belonging to *Davidiella*, and it is therefore probably an additional synonym of *D. ammophilae*.

Mycosphaerella algida Lar.N. Vassiljeva, Pirenomitsety i Lokuloaskomitsety Severa Dal'nego Vostoka: 80. 1987.

Type — Russia: *Neuroloma nudicaule* (Brassicaceae).

No material of this species was sent on loan from LE or LEP.

Mycosphaerella aliena (Pass.) Cash, Syll. Fung. 26: 358. 1972 = *Sphaerella aliena* Pass., Erbario Crittogamico Italiano, ser. 2 no. 1278. 1882.

Type — Italy: Parma, Collecchio. On dead stems of *Centaurea nigrescens* (Asteraceae). Passerini, distributed in Erbario Crittogamico Italiano no. 1278 (NY, isotype).

The isotype is overmature, but shows that it belongs to section *Caterva*, and is morphologically indistinguishable from *M. superflua*, with asci cylindrical.

Sphaerella aliena f. *clementei* Gonz. Frag., Broteria 22: 57. 1926.— Fig. 27.

Type — Spain: Cádiz, Benarem. On dead stems of *Centaurea clementei* (Asteraceae). Zuer no. 7598, VI 1921 (MA, holotype).

This is a *Guignardia*, with asci 50-70 × 12-15 μm, ascospores simple, 19-22 × 5-7 μm, surrounded by a gelatinous sheath.

Sphaerella aliena f. *hieracii* Gonz. Frag., Bol. Soc. Esp. Hist. Nat. 18: 366. 1918.— Fig. 28.

Type — Spain: Madrid. On dead stems of *Hieracium carpetanum* (Asteraceae). Frago no. 2403, VII 1916 (MA, holotype).

This is a *Guignardia*, with asci thick-walled, with ocular chamber, 35-45 × 12-15 μm, ascospores simple, clavate, 14-16 × 3.5-4.5 μm; the septa illustrated in the protologue are artifacts.

Mycosphaerella alismatis (Hollós) M. Morelet, Ann. Soc. Sci. Nat. Archéol. Toulon & Var 20: 105. 1968 = *Sphaerella alismatis* Hollós, Math. Természettud. Közlem. 35: 61. 1926.

Type — Hungary: *Alisma* (Alismataceae).

The type is not in BP and may have been destroyed during the war.

Mycosphaerella allescheri (Sacc.) Lindau, Hilfsb. Sammeln Ascomyceten: 119. 1903 [as “*Allesch.ii*”] = *Sphaerella allescheri* Sacc., Syll. Fung. 9: 612. 1891 [as “*allescherii*”], nomen novum for *Sphaerella thalictri* Allesch., Ber. Bot. Vereines Landshut 11: 60. 1889, later homonym (illegitimate, Article 53).

Type — Germany: *Thalictrum aquilegifolium* (Ranunculaceae).

No material could be studied; the type was probably destroyed in B.

Sphaerella allescheri f. *arctica* Allesch., in Allesch. & Henn., Biblioth. Bot. 49: 6. 1897.

Type — Greenland: *Thalictrum alpinum* (Ranunculaceae).

No material could be studied; the type was probably destroyed in B.

Mycosphaerella allicina (Fr. : Fr.) Vestergr., Bih. Kongl. Svenska Vetensk.-Akad. Handl. 22, Afd. 3, 6: 15. 1896 = *Sphaeria allicina* Fr., Kongl. Vetensk. Acad. Handl. 38: 247. 1817, sanctioned by Fr., Systema Mycol. 2: 437. 1823 = *Sphaerella allicina* (Fr. : Fr.) Auersw., in Gonn. & Rabenh., Mycol. Europaea 5-6: 19. 1869.— Fig. 29.

Type — Sweden: Skåne. On dead parts of living leaves of *Allium* (Alliaceae). Fr. no. F-09810 (UPS-FRIES, holotype).

Already synonymised with *Davidiella tassiana* (as *Mycosphaerella tassiana*) by von Arx (1949), with which the holotype and additional material (e.g. The Netherlands, Nunspeet, on *Allium porrum*, Beins, II 1899, L) agrees well, with asci pyriform, ascospores 22-27 × 6-8 μm.

This synonymy has been questioned, e.g. by Barr (1972), not on the basis of the indeed indistinguishable morphology (ascospores 18-22(-24) × 4.5-5.5 μm), but on the basis of the different host. It was declared that *Mycosphaerella allicina* grows on living leaves, whereas *D. tassiana* is saprophytic. However, in the specimens studied, the species is always growing on the rapidly dead leaf tips and margins of living leaves, thus truly saprophytic, possibly originating as endophyte (as is probably the rule in *Mycosphaerella*). As already indicated by von Arx (1949), *Mycosphaerella allicina* has priority over *Davidiella tassiana* if both are synonymised. Therefore the following combination is proposed: **Davidiella allicina** (Fr. : Fr.) Crous & Aptroot comb. nov., **MB 500339**. **Basionym:**

Sphaeria allicina Fr., Kongl. Vetensk. Acad. Handl. 38: 247. 1817, sanctioned by Fr., Systema Mycol. 2: 437. 1823.

Sphaerella allicina f. *allii-fallacis* Thüm., Mycotheca Universalis, fasc. 19 no. 1872. 1881.— Fig. 30.

Type — Austria. On dead stems of *Allium fallax* (Alliaceae). Thümen, Mycotheca Universalis no. 1872 (L, 2 isotypes).

This is morphologically indistinguishable from *D. allicina*, with ascospores 14-17 × 4-6 µm.

Sphaerella allicina f. *allii-porri* Thüm., Mycotheca Universalis, fasc. 20 no. 1946. 1881.

Type — Belgium: Malmedy. On upper and lower surface of dead leaves of *Allium porrum* (Alliaceae). Libert, distributed in Thümen, Mycotheca Universalis no. 1946 (BPI, isotype).

This is morphologically indistinguishable from *D. allicina*, although the isotype is immature, with asci pyriform.

Mycosphaerella allicina var. *arctica* (Rostr.) M. Morelet. See *Mycosphaerella tassiana* var. *arctica* (Rostr.) M.E. Barr.

Mycosphaerella allicina var. *ferlinii* M. Morelet, Ann. Soc. Sci. Nat. Archéol. Toulon & Var 20: 103. 1968.

Type — France: *Tagetes erecta* (Asteraceae).

No material could be studied, but there is no doubt that it is morphologically indistinguishable from *D. allicina*.

Mycosphaerella allii-cepae M.M. Jord., Maude & Burchill, Trans. Brit. Mycol. Soc. 86: 392. 1986 ≡ *Davidiella allii-cepae* (M.M. Jord., Maude & Burchill) Crous & U. Braun, in U. Braun *et al.*, Mycol. Progress 2: 10. 2003.

Type — United Kingdom. On upper and lower surface of dead leaves of *Allium porrum* ["*cepa*"] (Alliaceae). Jordan, III 1985 (IMI 296791, holotype).

Anamorph: *Cladosporium allii-cepae* (Ranoj.) M.B. Ellis, *vide* Jordan, Maude & Burchill (op. cit.).

Nothing identifiable could be found any more on the holotype material.

Sphaerella allioniae Speg., Anales Mus. Nac. Hist. Nat. Buenos Aires 23: 51. 1912.— Fig. 31.

Type — Argentina: Mendoza, Potrerillos, Rio Blanco. On stems of *Allionia incarnata* (Nyctaginaceae). Spegazzini no. 6131, III 1910 (LPS, holotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores 17-22 × 6-9 µm.

Sphaerella alnea (Fr. : Fr.) Auersw., in Gonn. & Rabenh., Mycol. Europaea 5-6: 3. 1869 ≡ *Sphaeria alnea* Fr., Observ. Mycol. 1: 185. 1818, sanctioned by Fr., Systema Mycol. 2: 520. 1823 ≡ *Laestadia alnea* (Fr. : Fr.) Auersw., Hedwigia 8: 177. 1869 ≡ *Guignardia alnea* (Fr. : Fr.) J. Schröt., in Cohn, Kryptog.-Fl. Schlesien 3(2): 330. 1894 ["1893"] ≡ *Gnomonina alnea* (Fr. : Fr.) Höhn., Ann. Mycol. 16: 47. 1918 ≡ *Plagiostoma alneum* (Fr. : Fr.) Arx, Antonie van Leeuwenhoek Ned. Tijdschr. Hyg. 17: 264. 1951 [as "*alnea*"].— Fig. 32.

Type — Sweden. On upper and lower surface of dead leaves of *Alnus* (Betulaceae). Fr., Scleromyceti Suecicae no. 59 (BPI, isotype).

Anamorph: *Cylindrosporella fide* Eriksson (1992).

Accepted as *Plagiostoma alneum* (Fr. : Fr.) Arx by Barr (1987). The isotype agrees, with ascomata skewed, asci clavate, but has relatively short, yellowish, simple to 1-septate ascospores of 11-14 × 3.5-4 µm.

Mycosphaerella alni (Fuckel) Tomilin, Opredelitel' grivob roda *Mycosphaerella* Johans.: 72. 1979 ≡ *Stigmata alni* Fuckel, Jahrb. Nassauischen Vereins Naturk. 23-24: 97. 1870 ≡ *Sphaerella alni* (Fuckel) Sacc., Michelia 1: 245. 1878 ≡ *Coleroa alni* (Fuckel) Petr. Ann. Mycol. 23: 63. 1925.— Fig. 33.

Type — Germany: Oestrich. Superficial in mycelium on grey spots on upper surface of living leaves of *Alnus glutinosa* (Betulaceae). Fuckel, Fungi Rhenani Exsiccati no. 1703 (L, isotype).

Already included in the synonymy of *Venturia alnea* (Fr.) E. Müll. by Sivanesan (1977), with which the type agrees, with grey, unequally septate ascospores of 14-17 × 4-6.5 µm.

Mycosphaerella alnicola (Peck) Jaap, Ann. Mycol. 15: 105. 1917 ≡ *Sphaerella alnicola* Peck, Annual Rep. New York State Mus. 40: 68. 1887.— Fig. 34.

Type — USA: On upper and lower surface of dead leaves of *Alnus viridis* (Betulaceae).

No type material was found in NY or BPI. Material studied (Switzerland, Glarus, Alp Platten, Müller, V 1949, CBS) belongs to section *Longispora*, and is morphologically indistinguishable from *M. latebroso*, with asci cylindrical, ascospores 25-30 × 2-3 µm.

Mycosphaerella alniviridis (De Not.) Tomilin, Novosti Sist. Nizsh. Rast. 1967: 188. 1967 ≡ *Sphaerella alniviridis* De Not., Comment. Soc. Crittog. Ital. 2(3): 487. 1867.— Fig. 35.

Type — Italy: Novara, Riva, Valdobbia. On upper and lower surface of dead leaves of *Alnus viridis* (Betulaceae). Carestia 615, 1869 (RO, lectotype); also Carestia 612, 1869 (RO-CESATI, topotype), also Carestia, X 1900, distributed in Sacc., Mycotheca Italica no. 633 (B, topotype).

The lectotype and topotype materials are identical, belong to *Davidiella*, and show that this is morphologically indistinguishable from *D. ammophilae*, with asci pyriform, ascospores 9-11 × 2.5-3.5 µm.

Mycosphaerella alnobetulae Jaap, Ann. Mycol. 15: 105. 1917 ≡ *Sphaerella alnobetulae* (Jaap) Trotter, Syll. Fung. 24: 853. 1928.

Type — Switzerland: *Alnus viridis* (Betulaceae).

No material could be studied; the type was probably destroyed in B.

Mycosphaerella alocasiae Syd. & P. Syd., Philipp. J. Sci., sect. C, Bot. 8: 195. 1913 ≡ *Sphaerella alocasiae* (Syd. & P. Syd.) Trotter, Syll. Fung. 24: 850. 1928.— Fig. 36.

Type — Philippines: Luzon, Manila. On white spots on upper surface of living leaves of *Alocasia indica* (Araceae).

Graff, XII 1911, distributed in Sydow, Fungi Exotici Exsiccati no. 30 (L, 2 isotypes).

Anamorphs: *Cercospora colocasiae* (Höhn.) Chupp [= *Passalora colocasiae* (Höhn.) U. Braun] (observed on spots on the lower surface in the type) and *Phyllosticta colocasiae* Höhn. *vide* Sivanesan (1984).

This is a parasitic species, with asci narrowly pyriform, ascospores $17-22 \times 4.5-5.5 \mu\text{m}$.

Mycosphaerella aloës Syd., Ann. Mycol. 37: 181. 1939.

Type — South Africa: *Aloë lineata* (Xanthorrhoeaceae).

No material was found in S, BPI or elsewhere.

Sphaerella aloysiae Pass., Atti Reale Accad. Lincei, Mem. Cl. Sci. Fis., ser. 4, 3: 8. 1887.

Type — Italy: *Lippia* ["*Aloysia*"] (Verbenaceae).

No material was found in any of the herbaria consulted.

Mycosphaerella alpina Arx, Sydowia 3: 68. 1949.— Fig. 37.

Type — Switzerland: Wallis, Zermatt, Riffelalp. On upper and lower surface of dead leaves of *Carex curvula* (Cyperaceae) [on label erroneously identified as "*Festuca*" (Poaceae)]. Von Arx, IX 1945 (ZT, holotype; ZT, isotype). Cited as synonym of *Mycosphaerella bulgarica* Petr. by Eriksson (1992). This belongs to section *Fusispora*, with asci pyriform, ascospores $24-32 \times 3.5-5 \mu\text{m}$.

Mycosphaerella alpiniae S.Q. Chen & P.K. Chi, in P.K. Chi, Fungal Diseases of Cultivated Medicinal Plants in Guangdong Province: 159. 1994.

Type — China: *Alpinia katsumadai* (Zingiberaceae).

No material could be studied as its preservation is uncertain.

Mycosphaerella alpiniicola S.Q. Chen & P.K. Chi, in P.K. Chi, Fungal Diseases of Cultivated Medicinal Plants in Guangdong Province: 35. 1994 [as "*alpinicola*"].

Type — China: *Alpinia oxyphylla* (Zingiberaceae).

No material could be studied as its preservation is uncertain.

Mycosphaerella alsinearum P. Karst., nomen herbariorum (not validly published, Article 32).

Authentic material — Finland. On upper and lower surface of dead leaves and calyces of *Spergularia rubra* (Caryophyllaceae). (L).

The material studied is immature.

Mycosphaerella alsines (Pass.) Magnus, Bull. Herb. Boissier, ser. 2, 3: 581. 1903 \equiv *Sphaerella alsines* Pass., Atti Reale Accad. Lincei, Mem. Cl. Sci. Fis., ser. 4, 3: 5. 1887.

Type — Italy: *Stellaria alsine* ["*Alsine*"] (Caryophyllaceae). Synonymised with *Davidiella allicina* (as *Mycosphaerella tassiana*) by von Arx (1949). No material was found in any of the herbaria consulted, but a slide made by von Arx from the type was studied and found to agree.

Mycosphaerella alsophila (Kirschst.) Tomilin, Novosti Sist. Nizsh. Rast. 14: 123. 1977 \equiv *Sphaerella alsophila* Kirschst., Kryptog.-Fl. Mark Brandenburg 7: 400. 1938.— Fig. 38.

Type — Germany: Niederbayern, Eisenstein. On dead stems of *Hieracium boreale* (Asteraceae). Kirschstein, VII 1937 (B, holotype).

Type and additional materials (On dead stems of *Arnica montana*, Kirschstein, VII 1933, B; also on dead stems of *Silene inflata*, Kirschstein, VII 1933, B; also Siegen, on dead stems of *Hieracium boreale*, Ludwig, V 1947, B) all agree and belong to *Davidiella* and this is morphologically indistinguishable from *D. ammophilae*, with asci pyriform, ascospores $6-9 \times 2.5-3.5 \mu\text{m}$.

Sphaerella alstroemeriae Speg., Revista Fac. Agron. Univ. Nac. La Plata, ep. 2, 6: 200. 1910.

Type — Chile: Valparaiso, Cerro Alegre. On dead stems of *Alstroemeria* (Alstroemeriaceae). Speg., I 1909 (LPS, holotype).

The type contains only a coelomycete.

Mycosphaerella altera (Pass.) House, New York State Mus. Bull. 233-234: 25. 1921 \equiv *Sphaerella altera* Pass., Atti Reale Accad. Lincei, Mem. Cl. Sci. Fis., ser. 4, 3: 10. 1887.— Fig. 39.

Type — Italy: On dead stems and branches of *Equisetum ramosum* (Equisetaceae).

Material studied (USA, California, Marion Co., Tomales State Park, on *Equisetum telmateia*, Bonar no. 1381, V 1970, L) belongs to *Davidiella* and this is morphologically indistinguishable from *D. allicina*, with asci narrowly pyriform, ascospores $16-19 \times 4-5 \mu\text{m}$, often 2-septate.

Mycosphaerella althaeina Lobik, Bolezni Rast. 17: 163. 1928.

Type — Russia: *Althaea rosea* (Malvaceae).

The type was not among the specimens sent on loan from LE or LEP.

Mycosphaerella alyssi (Hollós) Moesz, Balkán-Kutat. Tud. Eredm. 3: 132. 1926 \equiv *Sphaerella alyssi* Hollós, Ann. Hist.-Nat. Mus. Natl. Hung. 5: 44. 1907.

Type — Hungary: *Alyssum tortuosum* (Brassicaceae).

The type is not in BP and may have been destroyed during the war.

Mycosphaerella alyxiae Petr., Sydowia 8: 199. 1954.— Fig. 40.

Type — Australia: New South Wales, Twofold Bay. On lower surface of dead leaves of *Alyxia buxifolia* (Apocynaceae). Gauba, IV 1951, distributed in Reliquiae Petrakianae no. 1641 (H, L, isotypes).

Anamorph: *Stenella alyxiae* Yip and *Asteromella fide* Yip (1989).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. superflua*, with asci cylindrical, ascospores $15-17 \times 3.5-4 \mu\text{m}$.

Sphaerella alyxiae Cooke & Masee, Grevillea 16: 5. 1887.

Type — Australia: Victoria, Melbourne. On lower surface of dead leaves of *Alyxia* (Apocynaceae). Masee (NY, holotype; NY, isotype).

In the type only various coelomycetes are present, one of which may have led to the original description.

Mycosphaerella ambiens Starbäck, Bih. Kongl. Svenska Vetensk.-Akad. Handl. 14, Afd. 3, 5: 17. 1889 ≡ *Sphaerella ambiens* (Starbäck) Sacc., Syll. Fung. 9: 635. 1891.
Type — Sweden: *Berberis* (Berberidaceae).
No material was found in S or elsewhere.

Mycosphaerella ambigua (Fautrey & Lambotte) Tomilin, Opredelitel' gribov roda *Mycosphaerella* Johans.: 61. 1979 ≡ *Sphaerella ambigua* Fautrey & Lambotte, Rev. Mycol. (Toulouse) 17: 170. 1895.— Fig. 41.
Type — France: On dead stems of *Cichorium intybus* (Asteraceae). Fautrey, distributed in Roumeguère, Fungi Selecti Exsiccati no. 6882, VII 1895 (PC, holotype).
This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradians*, with asci broadly cylindrical but rather small, ascospores 10-12 × 3-3.5 µm.

Mycosphaerella ambiphylla A. Maxwell, Mycol. Res. 107: 355. 2003.
Type — Australia: *Eucalyptus globulus* (Myrtaceae).
Anamorph: *Phaeophleospora fide* Maxwell *et al.* (op. cit.). (= *Colletogloeopsis fide* Crous)
No material was studied of this recently described species.

Sphaerella ammophilae (Durieu & Mont.) Höhn., Sitzungsber. Kaiserl. Akad. Wiss., Math.-Naturwiss. Kl., Abt. 1, 119: 417. 1910 ≡ *Asterina ammophilae* Durieu & Mont., in Durieu, Fl. Algérie, Cryptog., 1: 548. 1849.— Fig. 42, 965.
Type — Algeria. On dead leaves of *Ammophila arenaria* (Poaceae). Durieu (PC, holotype).
The type material was also studied and annotated by von Höhnelt in 1909. This is the oldest epithet for the small-spored *Davidiella*. Therefore the following new combination is proposed here: **Davidiella ammophilae** (Durieu & Mont.) Aptroot comb. nov., **MB 500340**.
Basionym: *Asterina ammophilae* Durieu & Mont., in Durieu, Fl. Algérie, Cryptog., 1: 548. 1849. Material of this common species was partly annotated as *Mycosphaerella minor* and partly as *Mycosphaerella longissima*.

Mycosphaerella amomi P.K. Chi, Fungal Diseases of Cultivated Medicinal Plants in Guangdong Province: 44. 1994.
Type — China: *Amomum compactum* (Zingiberaceae).
No material was studied, as its preservation is uncertain.

Sphaerella amphigena Auersw., in Gonn. & Rabenh., Mycol. Europaea 5-6: 4. 1869, nomen nudum (not validly published, Article 32).
Authentic material — Germany: *Rubus fruticosus* (Rosaceae).
Cited as synonym of *Sphaerella punctiformis* [as “*sparsa*”] by Auerswald (op. cit.), cited as synonym with *M. punctiformis* by Tomilin (1979).

Mycosphaerella anacardiicola Bat., Anais 5 Reun. Anu. Soc. Bot. Brasil, 1954: 113. 1956 [as “*anacardiicola*”].
Type — Brazil: *Anacardium occidentale* (Anacardiaceae).
No material was studied, as it was not included in material received on loan from URM.

Sphaerella analoga Sacc. & Paol., Atti Reale Ist. Veneto Sci., ser. 6, 6: 408. 1888.
Type — Indonesia: On upper surface of white spots on living leaves of Melastomataceae. Paoletti (PAD, holotype).
The type contains only a coelomycete.

Mycosphaerella anarithma (Berk. & Broome) Lindau, Hilfsb. Sammeln Ascomyceten: 7. 1903 ≡ *Sphaeria anarithma* Berk. & Broome, Ann. Mag. Nat. Hist., ser. 3, 3: 375. 1859 (non Sacc. ≡ *Metasphaeria anarithma* (Sacc.) Sacc.)— Fig. 43.
Type — United Kingdom: Batheaston. On upper and lower surface of dead leaves of *Aira cespitosa* (Poaceae). Berkeley & Broome no. 210, 1854 (K, holotype).
The type and additional topotype material studied (Broome 1867, distributed in Rabenhorst, Fungi Europaei Exsiccati, ser. 2 no. 1136, CBS) show that this is morphologically indistinguishable from *Paraphaeosphaeria michotii* (Westend.) O.E. Erikss., with ascospores brown, ornamented, 1-2-septate, 17-24 × 5-7 µm, the epithet of which was published in the same year.

Mycosphaerella andersonii (Sacc.) Lindau, in Engl. & Prantl, Natürlichem Pflanzenf. 1(1): 425. 1897 ≡ *Sphaerella andersonii* Sacc., Syll. Fung. 9: 650. 1891, nomen novum for *Sphaerella conigena* Ellis & Everh., Proc. Acad. Nat. Sci. Philadelphia 1890: 230. 1890, later homonym (illegitimate, Article 53) ≡ *Keissleriella andersonii* (Sacc.) M.E. Barr, N. Amer. Fl., ser. 2, 13: 10. 1990.— Fig. 44.
Type — USA: Montana, Belt Mts. On scales of dead cones of *Pseudotsuga menziesii* [as “*Abies douglasii*”] (Pinaceae). Anderson no. 612, IX 1889 (NY, holotype; NY, isotype).
Mentioned to be a species of *Massarina* by Barr (1972), accepted as *Keissleriella andersonii* (Sacc.) M.E. Barr by Barr (op. cit.). The type, with ascomata ca. 500 µm, mostly immersed in the substratum, wall only covering the upper half, asci cylindrical, pseudoparaphyses numerous, mostly simple, ca. 1 µm wide, ascospores uniseriate, ca. 10 × 4 µm, agrees with the latter disposition, although no ascomatal setae (a character of the genus *Keissleriella*) are present.

Mycosphaerella andicola Speg., Anales Mus. Nac. Hist. Nat. Buenos Aires 23: 52. 1912.— Fig. 45.
Type — Argentina: Mendoza, Potrerillos. On bark of *Lycium longiflorum* (Solanaceae). Spegazzini no. 5780, III 1910 (LPS, holotype).
This is morphologically indistinguishable from *Arthopyrenia punctiformis*, with parenchyma between the asci, asci pyriform, ascospores 14-18 × 5-6 µm. It was already reported that it was no *Mycosphaerella* by Tomilin (1979).

Sphaerella andina Speg., Anales Mus. Nac. Hist. Nat. Buenos Aires 19: 350. 1909.— Fig. 46.
Type — Argentina: Chubut, Lago Blanco. On dead stems of *Thlaspi andicola* (Brassicaceae). Spegazzini, I 1903 (LPS, holotype).
This belongs to *Davidiella*, and is morphologically indistinguishable from *D. clandestina*, with asci pyriform, ascospores 22-25 × 7-8 µm.

Mycosphaerella andirae (Gonz. Frag. & Cif.) Cif., Quaderno 19: 230. 1961 = *Sphaerella andirae* Gonz. Frag. & Cif., in Cif. & Gonz. Frag., Bol. Soc. Esp. Hist. Nat. 26: 473. 1926.

Type — Dominican Republic: *Andira jamaicensis* (Fabaceae).

No material could be found in either the Ciferri herbarium in BPI, nor in MA, where the material of Gonz. Frag. is kept.

Mycosphaerella andrewsii (Sacc.) Davis, Trans. Wisconsin Acad. Sci. 24: 280. 1929 = *Sphaerella andrewsii* Sacc., Atti Mem. Reale Accad. Sci. 33: 165. 1917.

Type — USA: North Dakota, Anselm, Sandhills. On pale spots on upper and lower surface of living leaves of *Gentiana andrewsii* (Gentianaceae). Brenckle & Stevens, Fungi Dakotensis no. 385, VIII 1916 (B, IMI 16726, isotypes).

Spermatial state: *Asteromella fide* Petrak, in Davis (op. cit.). Only a coelomycete could be found on the isotypes studied.

Sphaerella andromedae Auersw., in Gonn. & Rabenh., Mycol. Europaea 5-6: 12. 1869 [as “*Sphaerella*”] = *Leptosphaeria andromedae* (Auersw.) Sacc., Syll. Fung. 2: 49. 1883 = *Wettsteinina andromedae* (Auersw.) M.E. Barr, Contr. Inst. Bot. Univ. Montréal 73: 8. 1959 = *Extrawettsteinina andromedae* (Auersw.) M.E. Barr, Mycotaxon 29: 502. 1987.

Type — Germany: *Cassiope* [“*Andromeda*”] *tetragona* (Ericaceae).

Accepted as *Extrawettsteinina andromedae* (Auersw.) M.E. Barr by Barr (op. cit.).

Mycosphaerella andromedae Miles, Pl. Dis. Reporter 19(5): 55. 1935, nomen novum (Article 58) for *Sphaerella andromedae* Tracy & Earle, Bull. Torrey Bot. Club 22: 176. 1895, later homonym (illegitimate, Article 53).— Fig. 47.

Type — USA: Mississippi, Ocean Springs. On lower surface of dead leaves of *Lyonia* [“*Andromeda*”] *nitida* [“*lucida*”] (Ericaceae). Earle, III 1888 (NY, 4 isotypes), also distributed in Reliquiae Petrakianae no. 476 (B, isotype).

Anamorph: *Phyllosticta elongata* Weid. *fide* Sivanesan (1984).

Synonymised with *Botryosphaeria vaccinii* (Shear) M.E. Barr by Barr (1970). However, the isotypes show that this is morphologically indistinguishable from *Glomerella cingulata* (Stoneman) Spauld. & H. Schrenk, with asci clavate, paraphyses simple, ascospores simple, 11-13 × 4-5.5 µm. The types also contain coelomycetes. Additional material studied (Netherlands, Aalsmeer, on white spots on upper surface of living leaves of *Andromeda axiliaris*, Kruseman & Schwarz, VI 1921, CBS) is a *Pseudomassaria*, with ascospores very asymmetrically septate, 16-20 × 5.5-7 µm.

Sphaerella andryalae Gonz. Frag., Trab. Mus. Nac. Ci. Nat., Ser. Bot. 4: 11. 1914.— Fig. 48.

Type — Spain: Navacerrada. On stems of *Andryala ragusina* (Asteraceae). Fragoso no. 863, VIII 1913 (MA, holotype).

This belongs to *Davidiella* and is morphologically indistinguishable from *D. ammophilae*, with asci pyriform, ascospores 11-13 × 3.5-4.5 µm. Additional material (La Manche, Acerros, Albo, VI 1935, MA) also belongs to *Davidiella*, but is *D. allicina*.

Mycosphaerella [“*Mycosphaerium*”] *anemones* Clem., Cryptogamae Formationum Coloradensium no. 214. 1906, nomen herbariorum (not validly published, Article 32).

Authentic material — USA: Colorado, Palsgrove Canyon. On dead stems of *Anemone globosa* (Ranunculaceae). Clements, VII 1906, Cryptogamae Formationum Coloradensium no. 214 (BPI).

The material studied contains only *Pleospora herbarum* (Pers. : Fr.) Rabenh. and a *Septoria* coelomycete.

Mycosphaerella anethi (Pers. : Fr.) Petr., Ann. Mycol. 25: 229. 1927 = *Sphaeria anethi* Pers., Observ. Mycol. 1: 67. 1796, sanctioned by Fr., Systema Mycol. 2: 429. 1823 = *Dothidea anethi* (Pers. : Fr.) Fr., Summa Veg. Scand.: 387. 1849 = *Phoma anethi* (Pers. : Fr.) Sacc., Michelia 2: 336. 1881.— Fig. 49.

Type — The Netherlands: On dead stems of *Anethum graveolens* (Apiaceae).

The type could not be found in L. Material studied, which agrees with the description (Czech Republic: Mährisch-Weißkirchen, Hranice, Petrak, V 1926, distributed in Reliquiae Petrakianae no. 1463, L) belongs to *Davidiella*, and is morphologically indistinguishable from *D. clandestina*, with asci nearly cylindrical, ascospores 19-23 × 4.5-5.5 µm.

Mycosphaerella angelicae Woron., Vestn. Tiflissk. Bot. Sada 28: 17. 1913 = *Sphaerella angelicae* (Woron.) Trotter, Syll. Fung. 24: 890. 1928, later homonym (illegitimate, Article 53).

Type — Georgia: On dead stems of *Angelica sylvestris* (Apiaceae).

Anamorph: *Passalora depressa* (Berk. & M.A. Curtis) Sacc. *fide* Von Arx (1983).

Cited as synonymous with *Mycosphaerella menthae* by Tomilin (1979). No material was included in the material received on loan from LE or LEP. Material studied (Latvia, Vidzeme, Vestiena, Starcs no. 0373, VII 1932, B) is postmature.

Mycosphaerella angelicae (Fr.) Petr. See *Mycosphaerella acilegna* M. Morelet.

Sphaerella angelicae Ellis & Everh., Proc. Acad. Nat. Sci. Philadelphia 44: 231. 1890 = *Didymella angelicae* (Ellis & Everh.) Sacc., Syll. Fung. 9: 663. 1891 = *Cercidospora angelicae* (Ellis & Everh.) Kuntze, Revis. Gen. Pl. 3(2): 453. 1898.— Fig. 50.

Type — Canada: London. On dead stems of *Angelica atropurpurea* (Apiaceae). Dearness no. 1715, VI 1890 (NY, holotype); also on *Angelica archangelica*, V 1890 (NY, topotype).

Synonymised with *Phomatospora angelicae* (Fuckel) Mouton by Von Arx & Müller (1954), with which the holotype agrees well, with asci cylindrical, paraphyses

simple, 2-3 µm wide, ascospores simple, ca. 9-11 × 3.5-4.5 µm.

Mycosphaerella angophorae Hansf., Proc. Linn. Soc. New South Wales 82: 217. 1957.— Fig. 51.

Type — Australia: New South Wales, Londonderry. On brown spots with purple margins on lower surface of living leaves of *Angophora bakeri* (Myrtaceae). Fraser, XII 1946 (K, holotype).

This is a parasitic species, with asci clavate, ascospores 12-13 × 2.5-3 µm.

Mycosphaerella angulata W.A. Jenkins, Phytopathology 32: 77. 1944.— Fig. 53.

Type — USA: Georgia, Experiment. On upper and lower surface of dead leaves of *Vitis rotundifolia* (Vitaceae). Jenkins, II 1941 (BPI, isotype).

Anamorph: *Cercospora brachypus* Ellis & Everh. *vide* Tomilin (1979) (= *Pseudocercospora brachypus* (Ellis & Everh.) X.J. Liu & Y.L. Guo).

This is a species of *Guignardia (sensu von Arx)*, with asci clavate, without hamathecium, ascospores simple, hyaline, 11-13 × 5-7 µm, without gelatinous sheath.

Sphaerella angulata Fuckel, Jahrb. Nassauischen Vereins Naturk. 27-28: 20. 1873 = *Laestadia angulata* (Fuckel) Sacc., Syll. Fung. 1: 424. 1882.— Fig. 52.

Type — Germany: Oestrich. On lower surface of dead leaves of *Berberis vulgaris* (Berberidaceae). Fuckel, Fungi Rhenani Exsiccati no. 2520 (G, holotype).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradians*, with asci cylindrical, ascospores 11-13 × 2.5-3.5 µm.

Mycosphaerella angustifoliorum A.W. Ramaley, Mycotaxon 40: 17. 1991.— Fig. 54.

Type — USA: Colorado, La Plata Co., Durango. On upper and lower surface of dead leaves of *Populus angustifolia* (Salicaceae). Ramaley, X 1988 (BPI, holotype).

Anamorph and spermatial state: *Clypeispora angustifoliorum* A.W. Ramaley and *Asteromella angustifoliorum* A.W. Ramaley *vide* Ramaley (op. cit.).

This is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores 7-8.5 × 2-2.5 µm.

Mycosphaerella annulata (Cooke) Miles, Pl. Dis. Reporter 19(5): 55. 1935 = *Sphaerella annulata* Cooke, Grevillea 6: 146. 1878.

Type — USA: Florida, Gainesville. On upper surface of dead leaves of *Magnolia glauca* (Magnoliaceae). Ravenel (K, holotype), also distributed in Fungi Americani Exsiccati no. 378 (K, 4 isotypes).

Synonymised with *Hyponectria magnoliae* (Schwein.) M.E. Barr by Barr (1977), with which the type studied agrees, although it is immature.

Mycosphaerella antarctica (Speg.) Tomilin, Novosti Sist. Nizsh. Rast. 1968: 163. 1968 = *Sphaerella antarctica* Speg., Bol. Acad. Nac. Ci. 11: 204. 1888 [“1887”].

Type — Argentina: *Fagus antarctica* (Fagaceae).

No material has been studied, as it was not included in a loan received from LPS.

Mycosphaerella anthemidina Petr., Ann. Mycol. 42: 52. 1944.— Fig. 55.

Type — Germany: Hessen-Nassau, Dillkreis, Donsbach. On dead stems and upper and lower surface of dead leaves of *Anthemis arvensis* (Asteraceae). Ludwig, VIII 1927, distributed in Petr., Mycotheca Generalis no. 271 (W 8039, holotype; B, W, isotypes).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradians*, with asci cylindrical, ascospores 12-15 × 3-4 µm.

Sphaerella anthistiriae McAlpine, Proc. Linn. Soc. New South Wales 28: 102. 1903.

Type — Australia: *Anthistiria australis* (Poaceae).

No material was studied; the preservation of the type is uncertain.

Mycosphaerella anthurii Miles, Trans. Illinois State Acad. Sci. 10: 252. 1917 = *Sphaerella anthurii* (Miles) Trotter, Syll. Fung. 24: 850. 1928.

Type — Puerto Rico: *Anthurium acaule* (Araceae).

No material was found in any of the herbaria consulted.

Sphaerella antivarensis Bubák, Bull. Herb. Boissier, ser. 2, 6: 399. 1906 = *Microthyrium antivarensis* (Bubák) Crous, Mycol. Res. 103: 618. 1999.— Fig. 56.

Type — Montenegro: Bar. On bark of branches of *Myrtus communis* (Myrtaceae). Bubák, IV 1903 (BPI, holotype).

This is not a *Microthyrium*, but morphologically indistinguishable from *Mycoporum hippocastani* (DC.) Coppins, with ascomata aggregated below a clypeus, in section greenish brown (darker in KOH) with meandering cells, without interascal filaments but with periphysoids in the ostiole, ascospores ca. 11 × 3.5 µm. It represents an extension of both the host range and the geographical range for this species.

Mycosphaerella antoniana (Unamuno) Cash, Syll. Fung. 26: 326. 1972 = *Sphaerella antoniana* Unamuno, Bol. Soc. Esp. Hist. Nat. 30: 210. 1930.— Fig. 57.

Type — Spain: León, San Roman de los Caballeros. On upper and lower surface of dead leaves of *Corrigiola telephifolia* (Caryophyllaceae). Alvarez no. 9017, IX 1929 (MA, holotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. ammophilae*, with asci pyriform, ascospores 11-13 × 3.5-5 µm. Additional material seen (León, Ponferrada, Rothmaler, IV 1933, MA) contains only a hyphomycete.

Mycosphaerella antonovii Petr., Hedwigia 68: 206. 1929 [“1928”].— Fig. 58.

Type — Russia: Siberia, Tsherepanovo, Tshumysh. On upper and lower surface of dead leaves of *Aconitum excelsum* (Ranunculaceae). Antonov no. 57, VI 1925 (W, holotype).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradicans*, with asci cylindrical, ascospores 12-15(-17) × 3-4 µm.

Sphaerella apertiuscula (Schwein.) Cooke, J. Bot. 21: 139. 1883 = *Sphaeria apertiuscula* Schwein., Trans. Amer. Philos. Soc., ser. 2, 4: 224. 1832.

Type — USA: New York. On lower surface of dead leaves of *Ulmus fulva* (Ulmaceae). Schweinitz. (PH, holotype; PH, isotype).

Both holo- and isotype are immature, but it appears morphologically indistinguishable from *M. punctiformis*.

Mycosphaerella aphyllanthis Bernaux, Rev. Pathol. Vég. Entomol. Agric. France 28: 144. 1949.

Type — France: Hérault, St. Clement. On dead parts of living stems of *Aphyllanthes monspeliensis* (Asparagaceae). Bernaux, X 1954 (BPI, 2 topotypes).

The topotypes are immature, but belong to *Davidiella*, and show that this is morphologically indistinguishable from *D. ammophilae*, with asci pyriform, ascospores small.

Mycosphaerella apiahyana (Speg.) Tomilin, Novosti Sist. Nizsh. Rast. 7: 202. 1970 = *Sphaerella apiahyana* Speg., Bol. Acad. Nac. Ci. 11: 518. 1889.— Fig. 59.

Type — Brazil: Apiahy. On lower surface of living leaves of Ericaceae. Puiggari, 1888 (LPS, holotype).

This is morphologically indistinguishable from *Microthyrium microscopicum* Desm., with ascomata applanate, superficial, paraphyses 1-2 µm wide, and ascospores 9-12 × 1.5-2 µm.

Mycosphaerella apocynica Petr., Ann. Mycol. 29: 366. 1931.— Fig. 60.

Type — Bulgaria: Anchralt am Schwarzem Murl. On dead stems of *Apocynum venetum* (Apocynaceae). Hruby, VI 1928 (NY, lectotype, here designated).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradicans*, with asci cylindrical, ascospores 9-12 × 5-6 µm. Also present on the material are *Glomerella cingulata* (Stoneman) Spauld. & H. Schrenk and *Pleospora herbarum* (Pers. : Fr.) Rabenh. The lectotype is selected in NY because no material was found in W.

Mycosphaerella apophlaeae Kohlm., in Kohlm. & Demoulin, Bot. Mar. 24: 13. 1981 = *Mycosphycias apophlaeae* (Kohlm.) Kohlm. & Volkm.-Kohlm., Systema Ascomycetum 16: 3. 1998.

Type — New Zealand: Snares Islands. In thallus of the red algae *Apophlaea lyallii* (Rhodophyta, Hypneaceae). Horning, Kohlmeyer no. 4028, II 1971 (NY, holotype).

Accepted as *Mycosphycias apophlaeae* (Kohlm.) Kohlm. & Volkm.-Kohlm. by Kohlmeyer & Volkmann-Kohlmeyer (1998). The type shows that this is a species of *Stigmidium*, with totally immersed ascomata which are only carbonized at the tip and a long (immersed, not protruding) ostiolar neck filled with periphyses. The genus *Mycosphycias* has been erected to accommodate the algal parasites excluded from *Mycosphaerella*, and the species is currently recognized as one of the two known taxa in it. However, it is scientifically preferably not to let the host relation

dominate the classification. As there are no morphological characters distinguishing this species from species from the much older genus *Stigmidium*, the following new combination is proposed: ***Stigmidium apophlaeae*** (Kohlm.) Aptroot comb. nov., **MB 500376**. **Basionym:** *Mycosphaerella apophlaeae* Kohlm., in Kohlm. & Demoulin, Bot. Mar. 24: 13. 1981.

Mycosphaerella applanata (Ellis & Everh.) Lind, Danish Fungi: 207. 1913 = *Sphaerella applanata* Ellis & Everh., J. Mycol. 4: 98. 1888.— Fig. 61.

Type — USA: Colorado, Sand Coulee. On dead stems of *Clematis ligusticifolia* (Ranunculaceae). Anderson no. 134, II 1888 (NY, holotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with asci not observed, ascospores 19-21 × 5.5-6.5 µm.

Mycosphaerella apula (Sacc. & D. Sacc.) Tomilin, Novosti Sist. Nizsh. Rast. 1968: 163. 1968 = *Sphaerella elatior* var. *apula* Sacc. & D. Sacc., Syll. Fung. 17: 636. 1905.

Type — Italy: Figgia. On white spots with brown margin on the upper surface of living leaves of *Magnolia grandiflora* (Magnoliaceae). Sacc., IV 1903 (PAD, holotype).

Cited as synonymous with *Mycosphaerella elatior* by Tomilin (1979). The type was found to be immature.

Mycosphaerella aquatica (Cooke) J.H. Miller, Mycologia 33: 79. 1941 = *Sphaerella aquatica* Cooke, J. Bot. 21: 106. 1883.— Fig. 62.

Type — USA: South Carolina, Seaboard. On lower surface of dead leaves of *Quercus aquatica* (Fagaceae). Ravenel no. 3262, 1881 (K, holotype), also distributed in Fungi Americani Exsiccati no. 690 (K (9x), NY, isotypes).

This belongs to section *Longispora*, and is morphologically indistinguishable from *M. millegrana*, with asci cylindrical, ascospores 14-17 × 2-2.5 µm.

Mycosphaerella aquatica Udaiyan & V.S. Hosag., J. Econ. Taxon. Bot. 15: 661. 1991, later homonym (illegitimate, Article 53), also not validly published (Article 37).

Type — India: On wood.

No material was studied and the preservation of a type is uncertain.

Mycosphaerella aquilegiae Murashk., in Murashk. & Ziling, Trudy Omsk. Selskokh. Inst. 9: 229. 1928.

Type — Russia: Siberia, Sajany. On *Aquilegia glandulosa* (Ranunculaceae).

No material was studied as the type was not included in loans from LE and LEP.

Mycosphaerella aquilegiae Sawada. See *Mycosphaerella sawadae* Tomilin.

Sphaerella aquilegiae Ellis & Galloway. See *Mycosphaerella aquilegiae-jonesii* Tomilin.

Mycosphaerella aquilegiae-jonesii Tomilin, Novosti Sist. Nizsh. Rast. 1968: 163. 1968, nomen novum (Article 58) for *Sphaerella aquilegiae* Ellis & Galloway, J. Mycol. 5: 66.

1889 (because the combination in *Mycosphaerella* already occupied in 1928 with another basionym).— Fig. 63.

Type — USA: Montana, Helena, Belt Mts., Yogo. On dead stems of *Aquilegia jonesii* (Ranunculaceae). Williamson, VII 1888 (NY, holotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores 16-19 × 5-6.5 μm, much narrower than mentioned in the description.

Mycosphaerella aquilina (Fr.) J. Schröt., in Cohn, Kryptog.-Fl. Schlesien 3(2): 341. 1894 ["1893"] = *Xyloma aquilina* Fr., Observ. Mycol. 2: 362. 1818 = *Sphaeria aquilina* (Fr.) Fr., Systema Mycol. 2: 522. 1823 = *Sphaerella aquilina* (Fr.) Auersw., in Unio Itin. Cryptog. no. 60. 1866 = *Leptosphaeria aquilina* (Fr.) Pass., Atti Soc. Crittog. Ital. 2: 47. 1879.— Fig. 64.

Type — Sweden: *Pteridium aquilinum* (Polypodiaceae).

According to Eriksson (1992) the type is a *Leptostroma*.

Material studied: India, Mt. Herson. On upper and lower surface of dead fronds of *Gleichenia linearis*. Ellis, V 1943 (IMI 152515). The material studied belongs to *M. punctiformis*, with asci cylindrical, ascospores 6-7 × 2-2.5 μm.

Mycosphaerella aquilina f. *aspidiurum* (Sacc.) Jaap, Verh. Bot. Vereins Prov. Brandenburg 57: 13. 1915 = *Sphaerella aquilina* f. *aspidiurum* Sacc., Ann. Mycol. 7: 435. 1909.

Type — Germany: On fronds of *Dryopteris filix-mas* ["*Aspidium filix mas*"] (Polypodiaceae).

Material studied: Switzerland: Lugano. Jaap, Fungi Selecti Exsiccati no. 615, IV 1913 (B).

Mycosphaerella aquilina f. *polypodii-filix-mas* Fautrey, in Roum., Rev. Mycol. (Toulouse) 13: 8. 1891.— Fig. 65.

Type — France: Noidan. On upper and lower surface of dead fronds of *Dryopteris filix-mas* ["*Polypodium*"] (Polypodiaceae). Fautrey, distributed in Roumeguère, Fungi Selecti Exsiccati no. 5543, V 1890 (PC, holotype).

This is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores 8-10 × 2-2.5 μm.

Mycosphaerella aquilina var. *infera* P. Karst., Hedwigia 23: 4. 1884.

Type — Finland: *Pteridium aquilinum* (Polypodiaceae).

No material was studied, as the type could not be found in H.

Mycosphaerella arachidicola Khokhr., Bolezni i vrediteli maslichnykh kul'tur 1(2): 29. 1934 = *Didymosphaeria arachidicola* (Khokhr.) Alcorn, Punith. & G.J.P. McCarthy, Trans. Brit. Mycol. Soc. 66: 354. 1976 = *Didymella arachidicola* (Khokhr.) Tomilin, Opredelitel' gribov roda *Mycosphaerella* Johans.: 285. 1979.

Type — Georgia: Abkhazia. On living leaves of *Arachis hypogaea* (Fabaceae). (LEP, holotype).

Anamorph: *Ascochyta adzameithica* Shoschiaschvili fide Alcorn, Punithalingam & McCarthy (op. cit.).

The type shows that it should indeed be accepted as *Didymella arachidicola* (Khokhr.) Tomilin, with copious hamathecium filaments.

Mycosphaerella arachidicola W.A. Jenkins. See *Mycosphaerella arachidis* Deighton.

Mycosphaerella arachidis Deighton, Trans. Brit. Mycol. Soc. 50: 328. 1938, nomen novum (Article 58) for *Mycosphaerella arachidicola* W.A. Jenkins, J. Agric. Res. 56: 325. 1938, later homonym (illegitimate, Article 53) = *Mycosphaerella jenkinsii* Tomilin, Novosti Sist. Nizsh. Rast. 1968: 165. 1968, superfluous (illegitimate, Article 52) nomen novum (Article 58).

Type — USA: Georgia, Experiment. On upper and lower surface of dead leaves of *Arachis hypogaea* (Fabaceae). Jenkins, VI 1937 (BPI, holotype).

Anamorph: *Cercospora arachidicola* Hori (= *Passalora arachidicola* (Hori) U. Braun) fide Sivanesan (1984).

The holotype is very much decayed and only the anamorph could be found on it, or on any of the other specimens studied.

Mycosphaerella arachnoidea F.A. Wolf, Mycologia 28: 275. 1936.— Fig. 66, 984.

Type — USA: North Carolina, Durham. On upper surface of dead leaves of *Morus rubra* (Moraceae). Wolf, V 1935 (BPI, holotype; IMI 227775, isotype, slide only).

Anamorph: *Cercospora arachnoidea* F.A. Wolf fide Sivanesan (1984).

This belongs to section *Caterva*, with asci cylindrical, ascospores 7.5-10 × 1.5-2.5 μm (in the holotype; the isotype slide contains ascospores of 16-19 × 3-3.5 μm, which are more in accordance with the protologue).

Mycosphaerella araliae Bunkina & Koval, Bot. Zhurn. (Kiev) 20: 94. 1963.

Type — Russia: *Kalopanax septemlobus* (Araliaceae).

Anamorph: *Phyllosticta araliae* Ellis & Everh. fide Bunkina & Koval (op. cit.).

No material was studied as the type was not included in loans from LE and LEP.

Sphaerella araliae Cooke & Harkn., Grevillea 9: 9. 1880.— Fig. 67.

Type — USA: California. On dead stems of *Aralia californica* (Araliaceae). Harkness no. 1246 (K, holotype).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradians*, with asci cylindrical, ascospores 11-13 × 2-2.5 μm.

Mycosphaerella araneosa (Rehm) Lindau, Hilfsb. Sammeln Ascomyceten: 125. 1903 = *Sphaerella araneosa* Rehm, in Arnold, Verh. K.K. Zool.-Bot. Ges. Wien 23: 115. 1873 [as "*Sphärella*"] = *Epicymatia araneosa* (Rehm) Sacc., Syll. Fung. 1: 572. 1882 = *Sphaerellothecium araneosum* (Rehm) Zopf, Nova Acta Acad. Caes. Leop.-Carol. German. Nat. Cur. 70(2): 178. 1897 = *Phaeosphaerella araneosa* (Rehm) Sacc. & D. Sacc., Syll. Fung. 17: 676. 1905 = *Endococcus araneosus* (Rehm) H. Olivier, Bull. Acad. Int. Géogr. Bot.

17: 127. 1907 = *Discothecium araneosum* (Rehm) Vouaux, Bull. Soc. Mycol. France 29: 55. 1913.

Type — Austria: Tirol, Waldrast. On thallus and apothecia of *Ochrolechia* ["*pallescens* var."] *upsaliensis* ["*tartarea*"] (Ascomycota, Ochrolechiaceae). Arnold, VIII 1872, distributed in Rehm, Ascomyceten no. 133 (NY, isotype).

Accepted as *Endococcus araneosus* (Rehm) H. Olivier by Clauzade, Diederich & Roux (1989). Now generally accepted as *Sphaerellothecium araneosum* (Rehm) Zopf, with which the type agrees well with the typical superficial, creeping subiculum.

Mycosphaerella araucariae (Rehm) Arx, Acta Bot. Neerl. 7: 512. 1958 = *Lizonia araucariae* Rehm, Hedwigia 40: 104. 1901.

Type — Brazil: *Araucaria brasiliensis* (Araucariaceae). No material studied, as the type could not be found in S.

Sphaerella arbuti (Fr.) Sacc., Syll. Fung. 1: 536. 1882 = *Xyloma arbuti* Fr., Kungl. Svenska Vetensk.-Akad. Handl. 1818: 361. 1818 = *Sphaeria arbuti* (Fr.) Fr., Systema Mycol. 2: 523. 1823.

Type — Sweden: *Arctostaphylos alpina* (Ericaceae).

No material studied, as there is no type specimen in UPS or elsewhere in any of the herbaria consulted.

Mycosphaerella arbuticola (Peck) Jaap, Ann. Mycol. 14: 13. 1916 = *Sphaerella arbuticola* Peck, Bull. Torrey Bot. Club 10: 75. 1883.— Fig. 68.

Type — USA: California, Santa Cruz Mts. On black spots on upper surface of living leaves of *Arbutus menziesii* (Ericaceae). Pringle no. 124, VII 1882 (NY, isotype), also distributed in Ellis & Everhart, North American Fungi no. 1682 (L, isotype); also distributed in Rabenhorst, Fungi Europaei Exsiccati no. 3761 (L, isotype).

The isotype in NY shows that this is a parasitic species, with asci cylindrical, ascospores 13-15 × 3.5-4 µm. The isotypes in L contain only empty ascomata; additional material studied (California, on white spots on lower surface of living leaves of *Umbellularia californica*, Hansen, Flora of the Sequoia Gigantea Region no. 1014, VII 1894, L) is a parasitic species, with asci cylindrical, ascospores 11-14 × 3.5-4 µm.

Sphaerella arcana Cooke, J. Bot. 4: 246. 1866 = *Sphaerella maculiformis* var. *arcana* (Cooke) Auersw., in Gonn. & Rabenh., Mycol. Europaea 5-6: 5. 1869 [as "*maculaeformis* var."].— Fig. 69.

Type — United Kingdom: Kent, Darenth. On lower surface of dead leaves of *Castanea vesca* (Fagaceae). Cooke (K, holotype).

Cited as synonymous with *M. maculiformis*, which is morphologically indistinguishable from *M. punctiformis*, by Tomilin (1979), with which the holotype agrees, with ascomata in groups, asci cylindrical, ascospores 8-11 × 2.5-3 µm.

Mycosphaerella arctica (Fuckel) Lavrov, Trudy Tomsk. Gosud. Univ. 110: 174. 1951 = *Sphaeria arctica* Fuckel, in Heuglin, Reisen nach dem Nordpolarmeer 3: 94. 1874 =

Didymella arctica (Fuckel) Berl. & Voglino, Syll. Fung., Addit. ad vol. 1-4: 88. 1886.— Fig. 70.

Type — Novaya Zemlya: Sabine Island. On upper and lower surface of dead leaves of *Poa alpina* (Poaceae). Heuglin (G, holotype).

This is a species of *Davidiella*. It was previously synonymised with *D. allicina* (as *Mycosphaerella tassiana*) by von Arx (1949), but asci are pyriform, ascospores 20-28 × 6-8 µm. Therefore it is morphologically indistinguishable from *D. clandestina*.

Mycosphaerella ardisiae (Cif. & Gonz. Frag.) Cif., Quaderno 19: 230. 1961 = *Sphaerella ardisiae* Cif. & Gonz. Frag., in Gonz. Frag. & Cif., Estac. Agron. Haina, ser. D, Bot., 5: 5. 1926.

Type — Dominican Republic: *Ardisia obovata* (Myrsinaceae).

No material could be found in either the Ciferri herbarium in BPI, nor in MA, where the material of Gonz. Frag. is kept.

Sphaerella arenariae Ellis & Everh., nomen herbariorum (not validly published, Article 32).

Authentic material — USA: Colorado, Larimer Co., Cameron Pass. On upper and lower surface of dead leaves of *Arenaria fendleri* (Caryophyllaceae). Ellis, VII 1894 (NY).

The material is immature, and the species was therefore never published.

Mycosphaerella arenariicola Bubák, in Hand.-Mazz., Ann. K.K. Naturhist. Hofmus. 23: 103. 1909 = *Sphaerella arenariicola* (Bubák) Sacc. & Trotter, Syll. Fung. 22: 123. 1913.— Fig. 71.

Type — Turkey: Trapezunti, Ulugoba Mt., Fol. On upper surface of dead leaves of *Arenaria rotundifolia* (Caryophyllaceae). Handel-Mazzetti no. 665, VII 1907 (BPI, holotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with asci pyriform and thick-walled, ascospores 13-16 × 3-4 µm, a bit immature and therefore much narrower than usual.

Mycosphaerella areola Ehrlich & F.A. Wolf, Phytopathology 22: 238. 1932.— Fig. 72.

Type — USA: North Carolina, Durham. On leaves of *Gossypium herbaceum* (Malvaceae). Wolf, X 1934 (IMI 227778, isotype, slide only).

Anamorph: *Ramularia gossypii* (Speg.) Cif. *fide* Sivanesan (1984). Additional material: USA, Alabama, Lee Co., Auburn. Atkinson, X 1890 (IMI no. 227779, slide only).

This is a parasitic species, with asci cylindrical, ascospores 12-15 × 2.5-3.5 µm.

Sphaerella areola (Fuckel) Auersw., in Gonn. & Rabenh., Mycol. Europaea 5-6: 2. 1869 = *Sphaeria areola* Fuckel, Fungi Rhenani Exsiccati, suppl. fasc. 7 no. 2160 = *Laestadia areola* (Fuckel) Sacc., Syll. Fung. 1: 422. 1882 = *Anisostomula areola* (Fuckel) Höhn., Ann. Mycol. 16: 49. 1918.

Type — Germany: Oestrich. On upper surface of dead leaves of *Quercus* (Fagaceae). Fuckel, Fungi Rhenani Exsiccati no. 2160.

Accepted as *Anisostomula areola* (Fuckel) Höhn. by von Arx & Müller (1954).

Mycosphaerella aretiae Höhn., Sitzungsber. Kaiserl. Akad. Wiss., Math.-Naturwiss. Kl., Abt. 1, 116: 110. 1907 = *Sphaerella aretiae* (Höhn.) Sacc. & Trotter, Syll. Fung. 22: 132. 1913.— Fig. 73.

Type — Austria: *Aretia alpina* (Myrsinaceae).

The type was not found in FH. Material studied (France, Dept. Vaucluse, Ventoux, on upper surface of dead leaves of *Douglasia vitaliana*, Reverchon, distributed in Reliquiae Petrakianae no. 1642, V 1877, L) belongs to *Davidiella allicina*, with asci pyriform, ascospores 16-19 × 4.5-5.5 µm.

Mycosphaerella argentinensis Frezzi, Revista Invest. Agropecu., ser. 5, Patol. Veg. 6: 149. 1969.

Type — Argentina: *Arachis hypogaea* (Fabaceae).

Synonymised with *Didymella arachidicola* (Khokhr.) Tomilin by Aptroot (1995a).

Sphaerella argyrophylli Bubák, Ann. K.K. Naturhist. Hofmus. 28: 197. 1914.— Fig. 74.

Type — Iraq: Gharra, Abd el Asis. On dead petioles of *Astragalus argyrophyllus* (Fabaceae). Handel-Mazzetti no. 1732, VI 1910 (BPI, holotype).

Already synonymised with *Davidiella allicina* (as *Mycosphaerella tassiana*) by Von Arx (1949), with which the holotype agrees well, with asci pyriform, ascospores 14-17 × 5-6.5 µm.

Mycosphaerella ariadna (Sacc.) Lindau, in Engl. & Prantl, Natürlichen Pflanzenf. 1(1): 424. 1897 = *Sphaerella ariadna* Sacc., Michelia 1: 34. 1877.— Fig. 75.

Type — Italy: On upper and lower surface of white spots with brown margins on living leaves of *Coronilla emerus* (Fabaceae). Saccardo, X 1876 (PAD, holotype).

This belongs to *Davidiella*, of which it represents a parasitic species, with ascomata pale (not much carbonized), asci pyriform, ascospores 9-11 × 3-4.5 µm. Therefore the following new combination is made: **Davidiella ariadna** (Sacc.) Aptroot comb. nov., **MB 500341**. **Basionym:** *Sphaerella ariadna* Sacc., Michelia 1: 34. 1877. Additional material studied (Vittario, Trotter, IX 1905, PAD) agrees.

Sphaerella ariadna subsp. *tormentillae* Sacc. See *Mycosphaerella tormentillae* (Sacc.) Tomilin.

Mycosphaerella aristolochiae Syd. & P. Syd., Ann. Mycol. 12: 555. 1914 = *Sphaerella aristolochiae* (Syd. & P. Syd.) Trotter, Syll. Fung. 24: 852. 1928.

Type — Philippines: Luzon, Cariti Prov. On white spots with black margins on lower surface of living leaves of *Aristolochia tagala* (Aristolochiaceae). Robinson, Bureau of Science no. 18265, II 1913 (BPI, isotype).

The type and additional material studied (Prov. Sorsogon, Irosin, Elmer, Phillipine Islands Plants no. 14415, III-VIII 1916, L, 2×) belong to section *Plaga*, with asci cylindrical,

ascospores immature (also not reported by the original authors).

Sphaerella aristolochiae Roum. See *Mycosphaerella clematitidis* (Oudem.) Johanson ex Oudem.

Mycosphaerella aristoteliae (Cooke) Aptroot comb. nov., **MB 500507**. **Basionym:** *Sphaerella aristoteliae* Cooke, Grevillea 15: 16. 1886.— Fig. 76.

Type — New Zealand. On white spots with brown margins on upper surface of living leaves of *Aristolochia racemosa* (Elaeocarpaceae). Colenso (K, holotype).

This is a parasitic species, with asci clavate, ascospores 16-19 × 3-4 µm.

Sphaerella armeriae Kirschst., nomen herbariorum (not validly published, Article 32).

Authentic material — Germany: Westhavelland, Grossluhnitz. On dead stems of *Armeria vulgaris* (Plumbaginaceae). Kirschstein (B).

This belongs to *Davidiella* and is morphologically indistinguishable from *D. ammophilae*, with asci pyriform, ascospores postmature.

Mycosphaerella armoraciae Johanson ex Oudem., Rév. Champ. Pays-Bas. 2: 211. 1897, nomen novum (Article 58) for *Sphaerella armoraciae* (Fuckel) Oudem., Fungi Neerl. Exsiccati no. 176, based on an anamorph (illegitimate, Article 59) = *Ascochyta armoraciae* Fuckel, Jahrb. Nassauischen Vereins Naturk. 23-24: 388. 1870.

Type — Netherlands: Culemborg. On white spots on upper surface of living leaves of *Armoracia rusticana* (Brassicaceae). Hulsebosch, X 1875, distributed in Oudemans, Fungi Neerlandici Exsiccati no. 176 (L, holotype).

Anamorph: *Ascochyta armoraciae* Fuckel *fide* Oudemans (op. cit.).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores 13-16 × 5-6 µm.

Sphaerella arnicae Speg., Atti Soc. Crittog. Ital. 3: 51. 1881.

Type — Italy: Mantelao. On upper surface of dead leaves of *Arnica montana* (Asteraceae). Spegazzini, IX 1879 (PAD, isotype).

This material is immature and, according to the original drawing on the label, ascospores have never been seen in it.

Mycosphaerella aronici Volkart, Ber. Deutsch. Bot. Ges. 21: 480. 1903 = *Sphaerella aronici* (Volkart) Sacc. & Traverso, in Sacc. & Trotter, Syll. Fung. 22: 134. 1913.

Type — Germany: *Aronicum* (Asteraceae).

Anamorphs: *Fusicladiella aronici* (Sacc.) Höhn. *fide* von Arx (1983) and *Phyllosticta aronici* (Fuckel) Sacc. *fide* Saccardo & Trotter (1913).

No type material was preserved in ZT. Material studied (Switzerland, Pilatus, on upper surface of dead leaves, Jaap, Fungi Selecti Exsiccati no. 469, VIII 1910, L and Graubünden, Albulapass, on stems of *Doronicum*

grandiflorum, Müller, VIII 1949, CBS) contains only coelomycetes, including *Ascochyta* and *Septoria*.

Mycosphaerella ["*Mycosphaerium*"] *artemisiae* Clem., Cryptogamae Formationum Coloradensium no. 215. 1906, nomen herbariorum (not validly published, Article 32).

Authentic material — USA: Colorado, Ruxton Dell. On dead stems of *Artemisia canadensis* (Asteraceae). Clements, VII 1906, Cryptogamae Formationum Coloradensium no. 215 (BPI).

The material studied contains only immature ascomycetes and a coelomycete.

Mycosphaerella artemisiae Tilak, Mycopathol. Mycol. Appl. 10: 218. 1959.

Type — India: *Artemisia nilgirica* (Asteraceae).

Anamorph: *Septoria nilgiriensis* Tilak *vide* Tilak (op. cit.).

No material was studied as the preservation of the type is uncertain.

Sphaerella arthoniae Arnold, Verh. K.K. Zool.-Bot. Ges. Wien 22: 304. 1872 = *Pharcidia arthoniae* (Arnold) Arnold, Flora 57: 152. 1874 = *Sphaerulina arthoniae* (Arnold) Sacc., Syll. Fung. 17: 695. 1905 = *Sphaeria arthoniae* (Arnold) H. Olivier, Bull. Acad. Int. Géogr. Bot. 17: 165. 1907.

Type — Austria: *Arthonia radiata* (Ascomycotina, Arthoniaceae).

Accepted as *Pharcidia arthoniae* (Arnold) Arnold by Clauzade, Diederich & Roux (1989).

Mycosphaerella arthopyrenioides (Auersw.) Lindau, Hilfsb. Sammeln Ascomyceten: 72. 1903 = *Sphaerella arthopyrenioides* Auersw., in Gonn. & Rabenh., Mycol. Europaea 5-6: 15. 1869 = *Mycosphaerella tassiana* var. *arthopyrenioides* (Auersw.) M.E. Barr, Contr. Inst. Bot. Univ. Montréal 73: 26. 1959 = *Mycosphaerella allicina* var. *arthopyrenioides* (Auersw.) M. Morelet, Bull. Soc. Sci. Nat. Archéol. Toulon & Var 195: 7. 1971.

Type — Austria: Steiermark, Aducaut. On dead stems of *Papaver bursari* (Papaveraceae). Niessl (PAD, isotype).

Synonymised with *Davidiella allicina* (as *Mycosphaerella tassiana*) by von Arx (1949). The isotype contains only a coelomycete. The holotype was probably destroyed in B.

Mycosphaerella arthraconicola Naumov, Bull. Soc. Mycol. France 30: 75. 1914 = *Sphaerella arthraconicola* (Naumov) Trotter, Syll. Fung. 24: 865. 1928.

Type — Russia: *Arthraxon ciliaris* (Poaceae).

No material was studied, as the type was not included in loans received from LE or LEP.

Mycosphaerella artocarpi F. Stevens & K. Young, in F. Stevens, Bernice P. Bishop Mus. Bull. 19: 101. 1925.— Fig. 77.

Type — Hawaii: Oahu. On upper and lower surface of dead leaves of *Artocarpus communis* (Moraceae). Degener & Iwasaki no. 3872, III 1930 (B, BPI, topotypes).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradians*, with ascomata large,

mostly immersed in the soft host tissue, asci cylindrical, ascospores $9-11 \times 2.5-3 \mu\text{m}$.

Mycosphaerella artocarpi Bat., Anais 5 Reun. Anu. Soc. Bot. Brasil 1954: 114. 1956, later homonym (illegitimate, Article 53).

Type — Brazil: *Artocarpus integrifolia* (Moraceae).

No material was studied, as the type was not included in a loan received from URM.

Sphaerella artocreas Rabenh., nomen herbariorum (not validly published, Article 32).

Authentic material — Germany: Rabenhorst, 1855, distributed in Klotzsch, Herbarium Vivum Mycologicum no. 360 (B).

Cited as morphologically indistinguishable from *M. punctiformis* by Tomilin (1979).

Mycosphaerella arundinariae (G.F. Atk.) Earle, in Mohr, Contr. U.S. Natl. Herb. 6: 174. 1901 = *Sphaerella arundinariae* G.F. Atk., Cornell Univ. Sci. Bull. 3: 9. 1897.

Type — USA: Alabama. On leaves of *Arundinaria tecta* (Poaceae). Atkinson.

Cited as synonymous with *M. recutita*, which is morphologically indistinguishable from *Davidiella disseminata*, by Tomilin (1979). No material was found in the herbaria consulted.

Sphaerella asarifolia Cooke, J. Bot. 21: 138. 1883 = *Laestadia asarifolia* (Cooke) Sacc., Syll. Fung. 2: XXXIII. 1883.

Type — USA: *Asarum arifolium* (Aristolochiaceae). (K, holotype).

Not studied in detail, as the species was already redispensed in *Laestadia*.

Mycosphaerella asclepiadis Siemaszko, Acta Soc. Bot. Pol. 1: 19. 1923.

Type — Georgia: Adzharskaya, Batumi. On lower surface of dead leaves of *Asclepias incarnata* (Apocynaceae). Woronow, IX 1917 (BPI, isotype).

Cited as synonymous with *Mycosphaerella galatea* by Tomilin (1979). The isotype studied contains only a coelomycete.

Mycosphaerella ascophylli Cotton, Trans. Brit. Mycol. Soc. 3: 96. 1909 = *Sphaerella ascophylli* (Cotton) Sacc. & Trotter, Syll. Fung. 22: 147. 1913 = *Mycosphyrcias ascophylli* (Cotton) Kohlm. & Volkm.-Kohlm., Systema Ascomycetum 16: 3. 1998.— Fig. 78.

Type — United Kingdom: Dorset, Studland. In thallus of the brown algae *Ascophyllum nodosum* (Phaeophyta, Fucaceae). Cotton, V 1915 (K, topotype).

Anamorph: *Septoria ascophylli* Melnik & Yu. Petrov *vide* Eriksson (1992).

The holotype and other material studied (Ireland, Kerry, Dingle Peninsula, Baile, on thallus of *Pelvetia canaliculata*, Poelt, Plantae Graecenses no. 196, VIII 1978, L; also United Kingdom, Dorsetshire, Lulworth Cove, Rhodes no. 4738, V 1931, B) shows that this is a species of *Stigmidium*, with ostioles with sparsely septate, $15-20 \times 2 \mu\text{m}$ large

paraphysoids, asci pyriform, ascospores 16-20 × 5-6 µm. The genus *Mycophycias* was erected by Kohlmeyer & Volkman-Kohlmeier (1998) to accommodate the algal parasites (called mycobionts in their paper) excluded from *Mycosphaerella*, and the species is currently recognised as *Mycophycias ascophylli* (Cotton) Kohlm. & Volkm.-Kohlm. However it is scientifically preferable not to let the host relation dominate the classification. As there are no morphological characters distinguishing it from the much older genus *Stigidium*, the following new combination is proposed: ***Stigidium ascophylli*** (Cotton) Aptroot comb. nov., **MB 500377**. **Basionym:** *Mycosphaerella ascophylli* Cotton, Trans. Brit. Mycol. Soc. 3: 96. 1909.

Sphaerella ascoscypha Tassi, Bull. Lab. Orto Bot. Reale Univ. Siena 3: 94. 1900.

Type — Australia: On green leaves of *Casuarina* (Casuarinaceae). Tassi, 1900 (SIENA, holotype). The holotype is immature and the identity of this fungus remains unknown; it is probably no *Mycosphaerella*.

Mycosphaerella asensioi (Unamuno) Cash, Syll. Fung. 26: 328. 1972 ≡ *Sphaerella asensioi* Unamuno, Revista Mauritania Tanger, 153-154: 4. 1940 [as “*Sphaerelle asensioi*”].— Fig. 79.

Type — Morocco: Playa de Berritez Centa. On upper surface of dead leaves of *Phagnalon saxatile* (Asteraceae). Unamuno no. 10984, III 1940 (MA, holotype).

This belongs to section *Caterva*, with ascomata tiny, asci cylindrical to clavate, ascospores 11-13 × 2.5-3.5 µm.

Mycosphaerella asiminae (Ellis & Kellerm.) Tomilin, Opredelitel' gribov roda *Mycosphaerella* Johans.: 46. 1979 ≡ *Sphaerella asiminae* Ellis & Kellerm., in Ellis & Everh., N. Amer. Pyrenomyc.: 268. 1892.

Type — USA: Ohio. On white spots with black margins on upper surface of living leaves of *Asimina triloba* (Annonaceae). Kellerman (NY, holotype).

In the very sparse material only a coelomycete could be found.

Mycosphaerella asparagi (Bres.) Magnus, in Dalla Torre & Sarnth., Fl. Tirol, Vorarlberg, Liechtenstein, III, Pilze: 468. 1905 ≡ *Sphaerella asparagi* Bres., Fungi Tridentini II: 43, 1892.— Fig. 80.

Type — Italy: On dead stems of *Asparagus officinalis* (Asparagaceae). Bresadola, II 1889 (S, holotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. clandestina*, with asci pyriform, ascospores 20-25 × 6-8 µm.

Sphaerella asparagi var. *minor* Unamuno, Revista Mauritania Tanger 153-154: 5. 1940.

Type — Morocco: Ceuta, Parque de San Amaro. On dead stems of *Asparagus plumosus* (Asparagaceae). Unamuno no. 10982, XI 1940 (MA, holotype).

The holotype contains only an immature ascomycete.

Mycosphaerella asperifolii (E. Bommer, M. Rousseau & Sacc.) Tomilin, Novosti Sist. Nizsh. Rast. 7: 202. 1970 ≡

Sphaerella asperifolii E. Bommer, M. Rousseau & Sacc., Ann. Mycol. 3: 508. 1905.

Type — Belgium: De Panne. On upper and lower surface of dead leaves of *Cynoglossum vulgare* (Boraginaceae). Bommer & Rousseau, III 1904 (BR, holotype).

This is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical and ascospores immature.

Mycosphaerella asperulae (Roum. & Fautrey) Vestergr., Hedwigia 42: 103. 1903 ≡ *Sphaerella asperulae* Roum. & Fautrey, Rev. Mycol. (Toulouse) 16: 171. 1894.

Type — France: On white spots on upper surface of living leaves of *Asperula odorata* (Rubiaceae). Fautrey, distributed in Roumeguère, Fungi Selecti Exsiccati no. 6686, XII 1891 (PC, holotype).

The holotype contains only coelomycetes.

Mycosphaerella asperulata L. Holm & K. Holm, Bot. Not. 132: 212. 1979.— Fig. 81.

Type — Sweden: Uppland, Jerusalem. On upper and lower surface of dead fronds of *Polypodium vulgare* (Polypodiaceae). Holm no. 1469a, VII 1978 (UPS, holotype).

The holotype and additional material studied (Finland, Kulanko and Mustiana, Alanko no. 6836a, VI 1968 and no. 16889, VI 1971, H, paratypes; also Austria, Steiermark, Koralmpe, Poelt, Plantae Graecenses no. 450, V 1983, L) belong to section *Caterva*, and show that this is morphologically indistinguishable from *M. filicum*, with asci broadly cylindrical, ascospores 13-18 × 3-4.5 µm. The ascomata appear mostly at the ends of the nervation and at hydrotodes, strongly suggesting that the species is endophytic and has already infected the whole frond before dead. The species therefore does not have to colonize the fronds after secession.

Mycosphaerella asphodelina (Alcalde) M. Morelet, Ann. Soc. Sci. Nat. Archéol. Toulon & Var 20: 105. 1968 ≡ *Sphaerella asphodelina* Alcalde, Anales Jard. Bot. Madrid 7: 464. 1947.

Type — Spain: Salamanca, La Alberca. On dead stems of *Asphodelus* (Xanthorrhoeaceae). Alcalde no. 12700, VI 1946 (MA, holotype).

The holotype is overmature, but shows that it belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*.

Mycosphaerella aspidii (Höhn.) L. Holm & K. Holm, Bot. Not. 132: 213. 1979 ≡ *Sphaerella asplenii* var. *aspidii* Höhn., in Rehm, Ann. Mycol. 7: 136. 1909 ≡ *Carlia aspidii* (Höhn.) Höhn., Ann. Mycol. 16: 62. 1918.— Fig. 82.

Type — Austria: Steiermark, Schladming. *Dryopteris* [“*Aspidium*”] *filix-mas* (Polypodiaceae). Höhnel (FH-Höhnel, holotype), also distributed in Rehm, Ascomyceten no. 1809, VIII 1908 (FH-Höhnel isotype).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. filicum*, with asci cylindrical, ascospores 10-12 × 2-2.5 µm.

Mycosphaerella asplenii (Auersw.) Lindau, in Engl. & Prantl, Natürlichen Pflanzenf. 1(1): 426. 1897 ≡ *Sphaerella*

asplenii Rabenh. ex Auersw., in Gonn. & Rabenh., Mycol. Europaea 5-6: 20. 1869.— Fig. 83.

Type — Czech Republic: Moravia, Naniest. On upper surface of dead leaves of *Asplenium septentrionale* (Polypodiaceae). Niessl (B, possible holotype: material seen by Auerswald).

The type and additional materials studied (Italy, Lugano, Jaap no. 616, L; also Botzen, Kühbacher Wald, Hausmann, distributed in Rabenhorst, Fungi Europaei Exsiccati no. 2438, B) belong to section *Caterva*, and show that this is morphologically indistinguishable from *M. filicum*, with asci cylindrical, ascospores $12-18 \times 3-4 \mu\text{m}$.

Mycosphaerella asplenii (Jaap) Thirum. & Govindu, Sydowia 8: 345. 1954, based on an anamorph (illegitimate, Article 59) and also a later homonym (illegitimate, Article 53) = *Cercospora asplenii* Jaap, Ann. Mycol. 14: 43. 1916.

Type — Germany: *Asplenium* (Polypodiaceae).

Not studied, to be accepted as *Cercospora asplenii* Jaap.

Sphaerella asplenii f. *trichomanis* Thüm., Fungi Austriaci no. 243. 1871.— Fig. 84.

Type — Austria: Niederösterreich, Krems. On upper surface of dead leaves of *Asplenium trichomanes* (Polypodiaceae). Thümen, 1871, distributed in Fungi Austriaci no. 243 (B, isotype).

The very small ascomata show that this belongs to section *Caterva*, and is morphologically indistinguishable from *M. filicum*, with asci cylindrical, ascospores $12-14 \times 4 \mu\text{m}$.

Sphaerella asplenii var. *aspidii* Höhn. See *Mycosphaerella aspidii* (Höhn.) L. Holm & K. Holm.

Mycosphaerella assimilata (J. Kunze) Lindau, Hilfsb. Sammeln Ascomyceten: 97. 1903 = *Sphaerella assimilata* J. Kunze, in Sacc., Syll. Fung. 1: 486. 1882 = *Sphaerella assimilata* f. *ribis-alpinae* J. Kunze, Fungi Selecti Exsiccati no. 246.— Fig. 85.

Type — Germany: Eisleben, Wolferode. On upper and lower surface of dead leaves of *Ribes alpinum* (Grossulariaceae). Kunze, Fungi Selecti Exsiccati no. 246, V 1879. (L, holotype).

This is no *Mycosphaerella* according to Tomilin (1979). However, the holotype contains typical *M. punctiformis*, with asci cylindrical, ascospores $8-10 \times 2-2.5$ and also a *Venturia* species, with ascospores greenish brown, asymmetrically septate, $11-13 \times 3-4 \mu\text{m}$. Therefore this is morphologically indistinguishable from *M. punctiformis*.

Sphaerella assimilata f. *ribis-alpinae* J. Kunze. See *Mycosphaerella assimilata* (J. Kunze) Lindau.

Sphaerella assurgens Cooke, Grevillea 15: 16. 1886 [as “*asurgens*”] = *Trichothelium assurgens* (Cooke) Aptroot & Lücking, Mycol. Res. 105: 511. 2001.— Fig. 86.

Type — New Zealand. On upper and lower surface of living fronds of *Trichomanes venosum* (Hymenophyllaceae). Colenso no. 200, received XII 1885 (K, holotype).

This is an older epithet for the lichenized ascomycetes *Trichothelium marianense* Harada and *T. nanum* Malcolm

& Vězda. Therefore, it has recently been combined into *Trichothelium* (Aptroot & Lücking 2001).

Mycosphaerella astericola Hara, J. Pl. Protect. 5: 536. 1918.

Type — Japan: *Aster* (Asteraceae).

No material studied, as the type was not found in TNS or BPI.

Sphaerella asteriniformis Sacc. & P. Syd. See *Mycosphaerella asterinoides* Lindau.

Mycosphaerella asterinoides Lindau, in Engl. & Prantl, Natürlichen Pflanzenf. 1(1): 424. 1897, nomen novum (Article 58) for *Sphaerella asterinoides* Pat., Bull. Soc. Mycol. France 8: 132. 1892, later homonym (illegitimate, Article 53) = *Sphaerella asteriniformis* Sacc. & P. Syd., Syll. Fung. 14: 533. 1899 [as “*asterinaeformis*”], superfluous (illegitimate, Article 52) nomen novum (Article 58).— Fig. 87.

Type — Ecuador: Pinchincha. On upper surface of green, but dead leaves of a Solanaceae. VII 1891 (FH-Patouillard, holotype).

This is probably a species of *Rehmiomycella*, with ascomata cupular, arranged in groups on immersed stroma, ascospores $15-18 \times 3.5-4.5 \mu\text{m}$.

Mycosphaerella asterinoides (Ellis & Everh.) Fairm., Proc. Rochester Acad. Sci. 4: 176. 1905, later homonym (illegitimate, Article 53) = *Sphaerella asterinoides* Ellis & Everh., J. Mycol. 4: 98. 1888.

Type — USA: New York, Clyde. On dead stems of *Dipsacus* (Dipsacaceae). Cook no. 539, IV 1888 (NY, holotype).

The type and additional material studied (On dead stems and involucra of *Dipsacus silvestris*, Saccardo no. 1479, L) contain immature material of *Pleospora herbarum* (Pers. : Fr.) Rabenh., with asci bitunicate, with ocular chamber, with parenchymatous tissue between them, ascospores thick-walled, hyaline, 1-septate, $22-26 \times 8-12 \mu\text{m}$.

Mycosphaerella asteroma (Fr.) Johanson ex Oudem., Rév. Champ. Pays-Bas 2: 208. 1897 = *Dothidea asteroma* Fr., Systema Mycol. 2: 560. 1823 = *Ascospora asteroma* (Fr.) Fuckel, Jahrb. Nassauischen Vereins Naturk. 23-24: 94. 1870 = *Sphaerella asteroma* (Fr.) P. Karst., Bidrag Kännedom Finlands Natur Folk 23: 181. 1873.— Fig. 88.

Type — Sweden: Småland, Femsjö. On upper and lower surface of dead leaves of *Convallaria majalis* (Asparagaceae). Fries no. F-09809 (UPS-FRIES, holotype). Spermatial states and anamorphs: *Asteroma reticulata* (DC.) Chevallier ex Fr. *vide* Saccardo (1882), *Asteromella convallariae* (Cavara) Petr. and *Septoria brunneola* (Fr.) Niessl *vide* Eriksson (1992).

The holotype is overmature and in bad shape. Additional material studied (Germany, Brandenburg, Tamsel, Vogel, distributed in Sydow, Mycotheca Germanica no. 1899, IV 1922, L) belongs to section *Caterva*, and confirms that this is morphologically indistinguishable from *M. subradians*, with asci cylindrical, ascospores $10-13 \times 2.5-3.5 \mu\text{m}$.

Sphaerella asteroma var. *asphodeli* Pat. ex Pit., Contr. Fl. Maroc: 63. 1931.

Type — Morocco: *Asphodelus microcarpus* (Xanthorrhoeaceae).

No specimens were studied, as no type material was preserved in FH.

Sphaeria asteroma var. *caryophyllacearum* Klotzsch, Herbarium Vivum Mycologicum no. 1453. 1851 = *Sphaeria asteroma* var. *caryophyllacearum* f. *saponariae* Klotzsch, Herbarium Vivum Mycologicum no. 1453. 1851.

Type — Italy: Vercellis. On upper and lower surface of dead leaves of *Saponaria officinalis* (Caryophyllaceae). Cesati, 1848, distributed in Klotzsch, Herbarium Vivum Mycologicum no. 1453 (B, isotype, 3×, sub f. *saponariae*).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradians*, like the nominal variety, but the studied material is immature.

Sphaeria asteroma var. *caryophyllacearum* f. *saponariae* Klotzsch. See *Sphaeria asteroma* var. *caryophyllacearum* Klotzsch.

Sphaeria asteroma var. *polygonati* DC., in Klotzsch, Herbarium Vivum Mycologicum no. 1454. 1851.

Type — Germany: Driesen. On upper and lower surface of dead leaves of *Polygonatum multiflorum* ["*vulgaris*"] (Asparagaceae). Lasch, 1848, distributed in Klotzsch, Herbarium Vivum Mycologicum no. 1454 (B, 2 isotypes).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradians*, like the nominal variety, but the studied material is immature.

Sphaeria asteroma var. *violae* DC., in Klotzsch, Herbarium Vivum Mycologicum no. 846. 1846.

Type — Germany: Sonnewalde. On upper and lower surface of dead leaves of *Viola odorata* (Violaceae). Kretzschmar, 1843, distributed in Klotzsch, Herbarium Vivum Mycologicum no. 846 (B, isotype).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradians*, like the nominal variety, but the studied material is immature.

Mycosphaerella astragali (Cooke) Tomilin, Novosti Sist. Nizsh. Rast. 1968: 163. 1968 = *Sphaerella astragali* Cooke, J. Bot. 21: 136. 1883, nomen novum (Article 58) for *Sphaeria astragali* Curr., Trans. Linn. Soc. London 22: 331. 1859, later homonym (illegitimate, Article 53) of *Sphaeria astragali* Rabenh. (1844).— Fig. 89.

Type — Canada: Arctic. On dead petioles of *Astragalus* (Fabaceae). Richardson, 1827 (K, holotype).

The type shows that this belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores 17-22 × 4.5-6 µm. Additional material studied (Germany, Hessen, Dillkreis, Donsbachtal, on *Astragalus glycyphyllos*, Ludwig, V 1939, B) belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradians*, with asci cylindrical, ascospores 10-12 × 2.5-3 µm.

Mycosphaerella astragalina I.E. Brezhnev, Bot. Mater. Otd. Sporov. Rast. Bot. Inst. Komarova Akad. Nauk SSSR 7: 185. 1951 = *Phaeosphaerella astragalina* (I.E. Brezhnev)

Tomilin, Opredelitel' gribov roda *Mycosphaerella* Johans.: 285. 1979.

Type — Russia: *Astragalus glycyphyllos* (Fabaceae).

No material was studied, as the type was not included in loans received from LE or LEP.

Mycosphaerella asunciensis Starbäck, Ark. Bot. 5(7): 21. 1905 = *Sphaerella asunciensis* (Starbäck) Sacc. & Traverso, Syll. Fung. 20: 818. 1911.

Type — Paraguay: Asuncion, villa Morra. On upper surface (not in spots) of living leaves of *Trichilia* (Meliaceae). Malme, Regnellian Expedition, Fungi no. 376, VII 1893 (S, holotype).

The holotype contains only a coelomycete and lichens, none of which match the protologue.

Mycosphaerella athamantae (Parisi) M. Morelet, Ann. Soc. Sci. Nat. Archéol. Toulon & Var 20: 105. 1968 = *Sphaerella athamantae* Parisi, Bull. Orto Bot. Regia Univ. Napoli 9: 58. 1928.

Type — Libya: *Athamanta* (Apiaceae).

No material was studied, as the type was not included in a loan received from PC.

Mycosphaerella atichae Hansf., nomen herbariorum (not validly published, Article 32).— Fig. 90. Authentic material — New Zealand: Auckland, Little Barrier Island. On upper surface of dead leaves, with or on stromata of *Atichia* on *Myrsine* ["*Suttonia*"] *australis* (Myrsinaceae). Dingley no. 15907, X 1945 (K, IMI no. 86120).

The material contains various fungi, e.g. Phyllachoraceae with simple ascospores of 15-17 × 9-10 µm and an ascomycete with strongly apiosporous ascospores of 7-9 × 1.5-2 µm.

Mycosphaerella atomus (Desm.) Johanson ex Oudem., Rév. Champ. Pays-Bas 2: 209. 1897 = *Sphaeria atomus* Desm., Ann. Sci. Nat. Bot., ser. 2, 15: 143. 1841 = *Sphaerella atomus* (Desm.) Fuckel, Jahrb. Nassauischen Vereins Naturk. 23-24: 99. 1870.— Fig. 91.

Type — France. On upper surface of dead leaves of *Fagus sylvatica* (Fagaceae). Desmazières, Plantes Cryptogames de France no. 1293 (BPI, isotype).

The type contains only a coelomycete with such tiny ("*atomus*") conidiomata that it could never have been a *Mycosphaerella*. Additional material studied (Germany, Brandenburg, on upper surface of dead leaves of *Fagus sylvatica* var. *atropupurea*, Vogel, distributed in Sydow, Mycotheca Germanica no. 3307, V 1939, L; also The Netherlands, Zuid-Beveland, van den Bosch, L) is *M. punctiformis*, with asci cylindrical, ascospores 8-9 × 2-2.5 µm. It was cited as synonymous with *M. fagi*, which is morphologically indistinguishable from *M. punctiformis*, by Tomilin (1979).

Sphaerella atra (Lév.) Sacc., Syll. Fung. 1: 498. 1882 = *Sphaeria atra* Lév., Ann. Sci. Nat. Bot., ser. 3, 3: 50. 1845.

Type — Australia: *Grevillea* (Proteaceae).

No material was studied, as the type was not included in a loan received from PC.

Mycosphaerella atractylodis Koval., Bot. Mater. Otd. Sporov. Rast. Bot. Inst. Komarova Akad. Nauk SSSR 16: 99. 1963.

Type — Russia: *Atractylodes ovata* (Asteraceae). Cited as synonymous with *Davidiella allicina* (as *Mycosphaerella allicina*) by Tomilin (1979). No material was studied, as the type was not included in loans received from LE or LEP.

Mycosphaerella atropae Syd. & P. Syd., Ann. Mycol. 14: 245. 1916 ≡ *Sphaerella atropae* (Syd. & P. Syd.) Trotter, Syll. Fung. 24: 887. 1928.— Fig. 92.

Type — Germany: Saarbrücken. On dead stems of *Atropa belladonna* (Solanaceae). Ludwig, IV 1914, distributed in Sydow, Mycotheca Germanica no. 1335 (L, isotype).

Anamorphs: *Phoma atropae* Roum. and *Ramularia atropae* Allesch. *vide* Tomilin (1979).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradians*, with asci cylindrical, ascospores 10-12 × 3-3.5 μm. It was cited as synonymous with *M. montellica*, which is however morphologically indistinguishable from *Davidiella ammophilae*, by Tomilin (1979).

Mycosphaerella aucubae Mark.-Let., Bolezni Rast. 16: 195. 1928 [“1927”].

Type — Russia: *Aucuba japonica* (Cornaceae). Anamorphs: associated with *Phyllosticta aucubicola* Sacc. and *Ascochyta aucubicola* G. Winter *vide* Markova-Letova (op. cit.).

No material was studied, as the type was not included in loans received from LE or LEP.

Mycosphaerella aucupariae (Lasch) Johanson ex Oudem., Rév. Champ. Pays-Bas 2: 215. 1897 ≡ *Sphaeria aucupariae* Lasch, in Klotzsch, Herb. Vivum Mycol., ed. 1 no. 459. 1843 ≡ *Sphaerella aucupariae* (Lasch) Plowr., Sphaeriacei Britannici, ed. 2 no. 92. 1875 ≡ *Venturia aucupariae* (Lasch) O. Rostr., in Lind, Danish Fungi: 213. 1913.

Type — Germany. On lower surface of dead leaves of *Sorbus aucuparia* (Rosaceae). Klotzsch, Herbarium Vivum Mycologicum no. 459 (B, holotype, sub *Septoria sorbi*).

Anamorph: *Septoria sorbi* Lasch *vide* Lasch (op. cit.).

This contains only the anamorph, *Septoria sorbi*.

Mycosphaerella audibertiae Rehm, Ann. Mycol. 7: 533. 1909 ≡ *Sphaerella audibertiae* (Rehm) Sacc. & Trotter, Syll. Fung. 22: 131. 1913.

Type — USA: California, Los Angeles Co., Claremont. On dead stems of *Audibertia polystachya* (Lamiaceae). Baker no. 5221, III 1909 (S, holotype).

Already cited as synonymous with *Davidiella allicina* (as *Mycosphaerella allicina*) by Tomilin (1979), with which the holotype agrees well.

Mycosphaerella auerswaldii (Fleischh.) Lindau, Hilfsb. Sammeln Ascomyceten: 32. 1903 ≡ *Sphaerella auerswaldii* Fleischh., in Auersw., in Gonn. & Rabenh., Mycol. Europaea 5-6: 11. 1869.

Type — Germany: *Cornus sanguinea* (Cornaceae).

No material was studied; the type was probably destroyed in B.

Mycosphaerella aurantia A. Maxwell, Mycol. Res. 107: 353. 2003.

Type — Australia: *Eucalyptus globulus* (Myrtaceae).

No material was studied of this recently described species.

Mycosphaerella aurantiorum Ruggieri, Boll. Staz. Patol. Veg. Roma 15: 343. 1935.

Type — Italy: *Citrus sinensis* (Rutaceae).

Anamorph: *Septoria aurantiorum* Ruggieri *vide* Ruggieri (op. cit.).

No material was studied, as the type was not included in a loan received from RO and its preservation is uncertain.

Mycosphaerella aurea R. Stone, Phytopathology 6: 425. 1916 ≡ *Sphaerella aurea* (R. Stone) Trotter, Syll. Fung. 24: 885. 1928.— Fig. 93.

Type — Canada: Ontario, Guelph. On lower surface of dead leaves of *Ribes aureum* (Grossulariaceae). Stone, V 1915 (NY, holotype; NY, isotype).

Anamorph: *Septoria aurea* Ellis & Everh. and *Septoria ribis* *vide* Eriksson (1992).

Cited as synonym: *Mycosphaerella ribis* (Sacc.) Feltgen by Eriksson (1992), but the name *ribis* has priority and it is moreover morphologically indistinguishable from *M. latebrosa*. The type contains mostly the *Septoria* anamorph, but also ascomata which show that it belongs to section *Longispora*, and is morphologically indistinguishable from *M. millegrana*, with asci cylindrical, ascospores immature, 14-16 × 2-2.5 μm.

Mycosphaerella australis (Speg.) Lindau, in Engl. & Prantl, Natürlichen Pflanzenf. 1(1): 425. 1897 ≡ *Sphaerella australis* Speg., Bol. Acad. Nac. Ci. 11: 204. 1888 [“1887”].

Type — Argentina: Isla de Los Estados. On lower surface of dead leaves of *Berberis ilicifolia* (Berberidaceae). Spegazzini, III 1882 (LPS, holotype).

The type is overmature.

Mycosphaerella babajani Negru & R. Sandor, Izv. Akad. Naul Arm. SSR, Biol. Nauki 18: 58. 1965.

Type — Romania: *Betula nana* (Betulaceae).

No material was studied, as the location of the type is unknown.

Mycosphaerella baccharidiphila (Speg.) Cash, Syll. Fung. 26: 329. 1972 ≡ *Sphaerella baccharidiphila* Speg., Bol. Acad. Nac. Ci. 25: 57. 1921.— Fig. 94.

Type — Chile: Los Perales. On dead stems of *Baccharis* (Asteraceae). Spegazzini no. 6175, 1917 (LPS, holotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores 16-20 × 6-8 μm.

Sphaerella baccharidis Pat., in Pat. & Lagerh., Bull. Herb. Boissier 3: 70. 1895.— Fig. 95.

Type — Ecuador: San Jorge. On upper and lower surface of dead, but still green leaves of *Baccharis* (Asteraceae). Lagerheim (FH-Patouillard, holotype).

This is probably a species of *Rehmiomycella*, with ascomata cupular, arranged in groups on immersed stroma, asci clavate, ascospores 30-40 × 10-16 µm, with 2 µm thick walls.

Mycosphaerella bacillifera (P. Karst.) Lind, Danish Fungi: 205. 1913 = *Sphaerella bacillifera* P. Karst., Hedwigia 22: 179. 1883 = *Leptosphaeria bacillifera* (P. Karst.) Munk, Dansk Bot. Ark. 17(1): 378. 1957 = *Monascostroma bacilliferum* (P. Karst.) O.E. Erikss., Mycotaxon 15: 196. 1982 [as “*bacillifera*”].

Type — Finland: *Scheuchzeria palustris* (Scheuchzeriaceae).

Accepted as *Monascostroma bacilliferum* (P. Karst.) O.E. Erikss. by Eriksson (1992).

Mycosphaerella badensis (Niessl) Lindau, Hilfsb. Sammeln Ascomyceten: 82. 1903 = *Sphaerella badensis* Niessl, Oesterr. Bot. Z. 25: 129. 1875.— Fig. 96.

Type — Austria: Baden near Wien. On dead leaves of *Poa badensis* (Poaceae). Niessl (M, lectotype, here designated). Additional material studied: Germany: Tolau. Niessl (M, paratype). Already synonymised with *Davidiella allicina* (as *Mycosphaerella tassiana*) by von Arx (1949), with which both lecto- and paratype agree well, with asci pyriform, ascospores 15-19 × 5.5-7 µm.

Mycosphaerella bakeri Rehm, Ann. Mycol. 7: 533. 1909 = *Sphaerella bakeri* (Rehm) Sacc. & Trotter, Syll. Fung. 22: 134. 1913.

Type — USA: *Gnaphalium* (Asteraceae).

Synonymised with *Davidiella allicina* (as *Mycosphaerella tassiana*) by von Arx (1949). No material was found in S (where Rehm's material is) or BPI (where Baker's material is).

Mycosphaerella balansae (Speg.) Tomilin, Novosti Sist. Nizsh. Rast. 6: 118. 1970 [“1969”] = *Sphaerella balansae* Speg., Anales Soc. Ci. Argent. 26: 33. 1888.— Fig. 97.

Type — Brazil: Guarapi. On lower surface of dead leaves of a Sapindaceae. Balansa no. 4081 [“4087”], IX 1883 (LPS, holotype).

This belongs to section *Longispora*, and is morphologically indistinguishable from *M. millegrana*, with ascomata seemingly arranged in stromata, asci pyriform, ascospores 17-23 × 3.5-4 µm.

Sphaerella balcanica Bubák, Bot. Közlem. 1915: 56. 1915.— Fig. 98.

Type — Montenegro: Pim, Božur planina. In brown spots (often leaf sectors surrounded by veins) with dark margins on upper and lower surface of living leaves of *Trifolium medium* subsp. *balcanicum* (Fabaceae). Rohlern, VII 1906 (BPI, holotype).

This is a parasitic species, with ascomata with pale walls, asci cylindrical, ascospores 8-12 × 2-3 µm. The species seems to deserve recognition, but the combination in *Mycosphaerella* cannot be made because the combination already exists for a different taxon. Therefore the following new name is proposed here: ***Mycosphaerella bubakii*** Aptroot nom. nov. **MB 500332**. pro *Sphaerella balcanica*

Bubák, Bot. Közlem. 1915: 56. 1915. Additional material studied (Germany, Westfalen, Siegen, on upper and lower surface of dead leaves of *Trifolium medium*, Ludwig, VIII 1927, B, NY) is *Davidiella allicina*, with asci pyriform, ascospores 15-18 × 4-5.5 µm.

Mycosphaerella balcanica Bubák & Picb., in Picb., Ann. Mycol. 35: 138. 1937.— Fig. 99.

Type — Bulgaria: *Silene inflata* var. *balcanica* (Caryophyllaceae).

No material was found in BPI, where the Bubák herbarium is kept.

Mycosphaerella baldensis (C. Massal.) Tomilin, Novosti Sist. Nizsh. Rast. 1967: 188. 1967 = *Sphaerella baldensis* C. Massal., in Sacc., Ann. Mycol. 9: 250. 1911.

Type — Italy: *Salix arbuscula* (Salicaceae).

No material was included in a loan from RO.

Mycosphaerella balsamopopuli Nevod., Griby SSSR, fasc. 1 no. 7. 1952.— Fig. 100.

Type — Russia: prov. Moskovskaja, distr. Podolskij, Klenovo. On upper and lower surface of dead leaves of *Populus balsamifera* (Salicaceae). Nevodovsky, Moscow Community for Investigators of Nature, Fungi USSR no. 7, V 1951 (LE 34957, holotype; L, isotype).

As already reported by Tomilin (1979), this belongs to section *Longispora*, and is morphologically indistinguishable from *M. populi*, with asci cylindrical, ascospores 25-28(-35) × 2.5-3 µm.

Mycosphaerella balsamorhizae Earle, Bull. New York Bot. Gard. 3: 292. 1904 [as “*balsamorrhizae*”] = *Sphaerella balsamorhizae* (Earle) Sacc. & Trotter, Syll. Fung. 22: 135. 1913 [as “*balsamorrhizae*”].

Type — USA: Nevada, Ormsby Co., Carson, King's Canyon. On dead stems and petioles of *Balsamorhiza* (Asteraceae). Baker no. 1230, VII 1904 (NY, holotype; BPI, isotype).

The types contain only *Pleospora herbarum* (Pers. : Fr.) Rabenh., with ascospores densely muriform, brown, 48-55 × 24-30, immature specimens of which (with hyaline, 1-septate ascospores) may have led to the description of this species.

Mycosphaerella bambusae (Pat. & Gaillard) Hara, Bot. Mag. (Tokyo) 27: 249. 1913 = *Sphaerella bambusae* Pat. & Gaillard, Bull. Soc. Mycol. France 4: 112. 1888.— Fig. 101.

Type — Venezuela: Caracas. On dead culms of *Bambusa* (Poaceae). Ernst no. 7b, 1888 (FH-Patouillard, holotype).

This belongs to *Davidiella* and is morphologically indistinguishable from *D. ammophilae*, with asci pyriform (nearly globose), ascospores 9-11 × 3-3.5 µm.

Sphaerella bambusae (I. Miyake & Hara) Sacc. See *Mycosphaerella phyllostachydicola* Tomilin.

Sphaerella bambusae var. *cocoas* Rehm. See *Mycosphaerella cocoas* (Rehm) J. Fröhl. & K.D. Hyde.

Mycosphaerella bambusae var. *phyllostachydis* Hara. See *Mycosphaerella phyllostachydis* (Hara) Hara.

Mycosphaerella bambusifolia I. Miyake & Hara, Bot. Mag. (Tokyo) 24: 338. 1910 = *Sphaerella bambusifolia* (I. Miyake & Hara) Sacc. & Trotter, Syll. Fung. 22: 145. 1913 [as “*bambusicola*”].

Type — Japan: prov. Mino, Kawauye-mura. On spots on upper surface of living leaves of *Phyllostachys reticulata* (Poaceae). Hara, VI 1912, distributed in Sydow, Fungi Exotici Exsiccati no. 127 (L, topotype). Topotype material studied is immature.

Mycosphaerella bambusina (Syd. & E.J. Butler) Tomilin, Novosti Sist. Nizsh. Rast. 1967: 188. 1967 = *Sphaerella bambusina* Syd. & E.J. Butler, Ann. Mycol. 9: 407. 1911.— Fig. 103.

Type — India: Assam, Wahjarin. On white spots on upper surface of living leaves of *Bambusa* (Poaceae). Butler 1375 (S, holotype).

This is morphologically indistinguishable from *Paraphaeosphaeria michotii* (Westend.) O.E. Erikss., with asci cylindrical, paraphyses septate, ca. 2 µm wide, ascospores becoming brown, 2-3-septate, 15-19 × 4-5 µm; mostly immature material, with paler and smaller ascospores.

The species was synonymised with *Mycosphaerella bambusifolia* by Tomilin (1979).

Mycosphaerella banksiae (Cooke & Masee) Tomilin, Novosti Sist. Nizsh. Rast. 1968: 163. 1968 = *Sphaerella banksiae* Cooke & Masee, in Cooke, Grevillea 16: 114. 1888 = *Dothiorella banksiae* (Cooke & Masee) Hansf., Proc. Linn. Soc. New South Wales 81: 43. 1956.

Type — Australia: Victoria, Melbourne. On upper surface of dead leaves of *Banksia integrifolia* (Proteaceae). Campbell no. 403 (NY, holotype; K, 2 isotypes).

The holo- and isotypes contain only the coelomycete *Colletotrichum* sp.

Mycosphaerella baptisiicola (Cooke) Earle, in Mohr, Contr. U.S. Natl. Herb. 6: 174. 1901 = *Sphaeria baptisiicola* Cooke, Grevillea 11: 110. 1883 [as “*baptisiaecola*”] = *Sphaerella baptisiicola* (Cooke) Berl. & Voglino, Syll. Fung., Addit. ad vol. 1-4: 81. 1886 [as “*baptisiaecola*”].— Fig. 104.

Type — USA: South Carolina, Aiken. On dead stems of *Baptisia leucantha* (Fabaceae). Ravenel, distributed in Fungi Americani Exsiccati no. 680 (BPI, isotype).

The isotype and additional material studied (USA, New Jersey, Newfield, on dead stems of *Baptisia tinctoria*, Ellis & Everh., distributed in North American Fungi no. 1799, V 1886, L) belong to *Phyalospora*, with ascospores non-septate, 17-19 × 5-5.5 µm, surrounded by a 1-2 µm thick gelatinous sheath.

Mycosphaerella bardanae (Hazsl.) Lindau, Hilfsb. Sammeln Ascomyceten: 60. 1903 = *Sphaerella bardanae* Hazsl., Math. Természettud. Közlem. 25: 108. 1893 [“1892”].

Type — Hungary: *Arctium* [“*Lappa*”] (Asteraceae).

Cited as synonymous with *Davidiella allicina* (as *Mycosphaerella allicina*) by Tomilin (1979). The type is not in BP and may have been destroyed during the war.

Mycosphaerella barnadesiae Petr., Sydowia 4: 502. 1950.— Fig. 106.

Type — Ecuador: Prov. Tungurahua, Baños. On lower surface of dead leaves of *Barnadesia parviflora* (Asteraceae). Sydow, distributed in Reliquiae Petrakianae no. 648, VIII 1948 (H, L, isotypes).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. superflua*, with asci cylindrical, ascospores 15-19 × 4.5-5.5 µm.

Mycosphaerella basicola (A.B. Frank) Lindau, Hilfsb. Sammeln Ascomyceten: 111. 1903 = *Sphaerella basicola* A.B. Frank, Ber. Deutsch. Bot. Ges. 13: 62. 1895.— Fig. 105.

Type — Germany: Berlin, Dahlem, Botanical Garden. *Secale cereale* (Poaceae). VII 1894 (B, isotype).

The material is not well preserved, but shows that this is probably a species of *Didymella*.

Mycosphaerella bataticola Khokhr. & Dyur., in Dunin & Yakimovich, Bolezni batata i mery borby s nimi: 90. 1934.

Type — Georgia: *Ipomoea batatas* (Convolvulaceae).

No material was included in loans from LE or LEP.

Mycosphaerella baudysiana Picb., in Baudyš & Picb., Práce Morav. Přír. Společn. 2: 155. 1925.

Type — Montenegro: *Inula crithmoides* (Asteraceae).

No material was found in BPI, where the Bubák herbarium is kept, and the location of this type is unknown.

Mycosphaerella bauhiniae Starbäck, Bih. Kongl. Svenska Vetensk.-Akad. Handl. 25(3): 56. 1899 = *Sphaerella bauhiniae* (Starbäck) Sacc. & Trotter, Syll. Fung. 16: 469. 1902.— Fig. 107.

Type — Brazil: Matto Grosso, Macoco. On white to brown (in isotype) spots on lower surface of living leaves of *Bauhinia* (Fabaceae). Lindman, 1st Regnellian Expedition, Fungi no. B 522, IV 1894 (S, holotype), also distributed in Vestergren, Micromycetes Rariores Selecti no. 827 (BPI, isotype).

The holotype contains unidentifiable, old, superficial ascomata that have not belonged to a *Mycosphaerella*, filled with an actinomycete. The isotype shows that they belong to a species of *Microthyriaceae*, probably a species of *Muyocopron*, with ascomata conical, asci cylindrical, ascospores simple, 5.5-7 × 3-3.5 µm.

Mycosphaerella beaglensis (Speg.) Cash, Syll. Fung. 26: 330. 1972 = *Sphaerella beaglensis* Speg., Bol. Acad. Nac. Ci. 27: 359. 1924.— Fig. 108.

Type — Argentina: Fuegia, Ushuaia. On dead stems of *Senecio longipes* (Asteraceae). Spegazzini, I 1924 (LPS, holotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. clandestina*, with asci pyriform, ascospores 20-22 × 5-7 µm.

Sphaerella beckhausii Fuckel, nomen herbariorum (not validly published, Article 32).— Fig. 109.

Authentic material — Germany: Östrich. On upper surface of dead leaves of *Euphorbia cyparissias* (Euphorbiaceae). Fuckel no. 248, 1870 (B).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores 17-21 × 4.5-7 µm.

Sphaerella begoniae Pat., Bull. Herb. Boissier 3: 70. 1895.

Type — Ecuador: San Jorge. On white spots on lower surface of dead leaves of *Begonia* (Begoniaceae). 1895 (FH-Patouillard, holotype).

The type contains only old ascomata and its identity remains unknown.

Mycosphaerella belladonnae (Briard & Har.) Tomilin, Novosti Sist. Nizsh. Rast. 8: 149. 1971 = *Sphaerella belladonnae* Briard & Har., Rev. Mycol. (Toulouse) 13: 15. 1891 [as “belladonnae”].

Type — France: Aube, Pont du Seine. On living leaves of *Atropa belladonna* (Solanaceae). Hariot, IX 1890 (PC, isotype).

Cited as synonymous with *Davidiella allicina* (as *Mycosphaerella allicina*) by Tomilin (1979). No fungus was present on the isotype studied, but the reported synonymy is unlikely because of the host (living leaves).

Mycosphaerella bellona (Sacc.) Spagorow, Trudy Bot. Inst. Kharkovsk. Univ. 28: 11. 1915 = *Sphaerella bellona* Sacc., Michelia 1: 244. 1878.

Type — Italy: *Pyrus communis* (Rosaceae).

Anamorph: *Phyllosticta pyrina* Sacc. fide Saccardo (op. cit.), which is morphologically indistinguishable from *Phoma pomorum* Thüm. fide Boerema, Dorenbosch & van Kesteren (1977).

No material was studied as the type is not held in PAD and not found in any of the other herbaria consulted; its anamorph does not suggest a *Mycosphaerella*.

Mycosphaerella bellula Crous & M.J. Wingf., Mycotaxon 46: 20. 1993.

Type — South Africa: *Protea repens* (Proteaceae).

No material was studied of this recently described species.

Mycosphaerella benguetensis Syd., in Syd. & Petr., Ann. Mycol. 29: 196. 1931.

Type — Philippines: *Coriaria intermedia* (Coriariaceae).

No material was found in S or any of the herbaria consulted.

Mycosphaerella berberidiphilum (Speg.) N. Lundq., nomen herbariorum (not validly published), based on anamorph (illegitimate, Article 59) [as “berberidicola”] = *Coniothyrium berberidiphilum* Speg., Bol. Acad. Ci. Córdoba 27: 394. 1924 [as “berberidiphila”].

Type — Argentina: On dead stems of *Berberis* (Berberidaceae).

Anamorphs: *Coniothyrium berberidicola* Speg. or *Coniothyrium bergii* Speg. according to label.

The type was not included in a loan from LPS. Material studied (Neuquén, Lago Triful, Gamundi, X 1957, CBS) contains only a *Microsphaeropsis*.

Mycosphaerella berberidis (Auersw.) Lindau, in Engl. & Prantl, Natürlichen Pflanzenf. 1(1): 424. 1897 = *Sphaerella berberidis* Auersw., in Gonn. & Rabenh., Mycol. Europaea 5-6: 3. 1869, nomen novum (article 58) for *Sphaeria berberidis* Nitschke ex Fuckel, Fungi Rhenani Exsiccati no. 1771. 1868, nomen nudum (not validly published, Article 32).— Fig. 110.

Type — Germany: On lower surface of dead leaves of *Berberis vulgaris* (Berberidaceae). Fuckel, Fungi Rhenani Exsiccati no. 1771 (L, isotype).

Anamorph and spermatial state: *Septoria berberidis* Niessl fide Sivanesan (1984) and *Asteromella* fide von Arx (1949).

The isotype material studied is immature. Additional material studied (Germany, Brandenburg, Dahmsdorf, Syd., Mycotheca Germanica no. 3502, IV 1941, L) belongs to section *Longispora*, and shows that this is morphologically indistinguishable from *M. latebrosa*, with asci cylindrical, ascospores 20-26 × 2.5-3.5 µm.

Mycosphaerella berkeleyi W.A. Jenkins, J. Agr. Res. 56: 331. 1938.

Type — USA: Georgia, Experiment. On upper and lower surface of dead leaves of *Arachis hypogaea* (Fabaceae). Jenkins, VI 1937 (BPI, holotype).

Anamorph: *Phaeoisariopsis personata* (Berk. & M.A. Curtis) von Arx = *Cercosporidium personatum* (Berk. & M.A. Curtis) Deighton, fide von Arx (1983) (= *Passalora personata* (Berk. & M.A. Curtis) S.A. Khan & M. Kama).

The holotype contains of totally decayed material with an overmature ascomycete.

Mycosphaerella berlesiana (Traverso) Tomilin, Novosti Sist. Nizsh. Rast. 1968: 164. 1968 = *Sphaerella berlesiana* Traverso, Fl. Ital. Cryptog. 1(11): 613. 1913, nomen novum (Article 58) for *Sphaerella simulans* Berl. & Bres., Annuario Soc. Alpinisti Tridentini 14: 27. 1889, later homonym (illegitimate, Article 53).

Type — Italy: *Quercus* (Fagaceae).

Cited as synonymous with *Mycosphaerella quercina* by Tomilin (1979).

Sphaerella betulae Auersw., in Gonn. & Rabenh., Mycol. Europaea 5-6: tab. 3, f. 36. 1869 = *Guignardia betulae* (Auersw.) Sacc. & Trotter, Syll. Fung. 22: 77. 1913.

Type — Germany: *Betula* (Betulaceae).

The type was probably destroyed in B.

Mycosphaerella bhandardarensis Patw., Mycopathol. Mycol. Appl. 29: 368. 1966.

Type — India: Bhandardara, Pune. On white spots on upper surface of living leaves of *Arisaema murrayi* (Araceae). Patwardhan no. 213 (LWG, holotype).

Anamorph: *Colletotrichum bhandardarensis* Patw. fide Patwardhan (op. cit.).

The type material studied contains only the anamorph, suggesting that it might have been morphologically

indistinguishable from *Glomerella cingulata* (Stoneman) Spauld. & H. Schrenk.

Mycosphaerella bhauria (Cooke) Lindau, in Engl. & Prantl, *Natürlichen Pflanzenf.* 1(1): 425. 1897 ≡ *Sphaerella bhauria* Cooke, *Grevillea* 6: 118. 1878.— Fig. 111.

Type — India: Dinagepore. On lower surface of dead, but still green leaves of *Symplocos spicata* (Symplocaceae). (K, 7 isotypes).

This is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores 7-9 × 2.5-3 µm.

Mycosphaerella biberwierensis (Auersw.) Lindau, *Hilfsb. Sammeln Ascomyceten*: 38. 1903 ≡ *Sphaerella biberwierensis* Auersw., in Gonn. & Rabenh., *Mycol. Europaea* 5-6: 9. 1869.— Fig. 112.

Type — Austria: Biberwier. On upper and lower surface of dead leaves of *Dryas octopetala* (Rosaceae). Auerswald, VI 1869 (B, holotype; L, isotype).

The isotype material studied contains only *Pleospora herbarum* (Pers. : Fr.) Rabenh., with ascospores densely muriform, ca. 40 × 15 µm.

Sphaerella bicalcarata Ces., in Rabenh., *Fungi Europaei Exsiccati*, ed. nov., ser. 2 no. 1561. 1872 ≡ *Diaporthe bicalcarata* (Ces.) Niessl, *Verh. Naturf. Vereins Brünn* 14: 169. 1876 ≡ *Ceriospora bicalcarata* (Ces.) Sacc., *Syll. Fung.* 2: 186. 1883 ≡ *Ceriosporella bicalcarata* (Ces.) Berl., *Icones Fung.* 1: 121. 1894 ≡ *Frondispora bicalcarata* (Ces.) K.D. Hyde, *Sydowia* 45: 208. 1993.— Fig. 113.

Type — Italy: On upper surface of dead petioles of *Chamaerops humilis* (Arecaceae). Cesati, 1872, distributed in Rabenhorst, *Fungi Europaei Exsiccati*, ed. nov., ser. 2, no. 1561 (CBS, isotype).

This is the type species of *Frondispora*, with ascospores 25-30 × 5-7 µm, with two polar appendages of 6-9 µm long. It is accepted here as *Frondispora bicalcarata* (Ces.) K.D. Hyde.

Sphaerella bidwellii (Ellis) Ellis, in Britton, *Cat. Plants New Jersey*: 522. 1890 [“1888”] ≡ *Sphaeria bidwellii* Ellis, *Bull. Torrey Bot. Club* 7: 90. 1880 ≡ *Physalospora bidwellii* (Ellis) Sacc., *Syll. Fung.* 1: 441. 1882 ≡ *Laestadia bidwellii* (Ellis) Viala & Ravaz, *Progrès Agricole*: 492. 1888 ≡ *Guignardia bidwellii* (Ellis) Viala & Ravaz, *Bull. Soc. Mycol. France* 8: 63. 1892 ≡ *Carlia bidwellii* (Ellis) Magnus, *Bull. Soc. Mycol. France* 8: 63. 1892 ≡ *Phyllachorella bidwellii* (Ellis) Theiss., *Verh. Zool.-Bot. Vereins Wien* 69: 11. 1919 ≡ *Botryosphaeria bidwellii* (Ellis) Petr., *Sydowia* 11: 440. 1958 [“1957”].

Type — USA: New Jersey, Vineland. On twigs of *Vitis* (Vitaceae). Bidwell, IV 1890 (NY, holotype); also distributed in Ellis & Everh., *North American Fungi* no. 596 (NY, 2 isotypes).

Anamorphs: *Phyllostictina uvicola* (Berk. & M.A. Curtis) Höhn. *vide* Barr (1972), *Phyllosticta ampelicida* (Engelman) van der Aa and *Leptodothiorella* *vide* Sivanesan (1984).

Accepted as *Botryosphaeria bidwellii* (Ellis) Petr. by Barr (1972), with which the types agree well.

Mycosphaerella biguttulata Rieuf, *Cah. Rech. Agron.* 15: 36. 1962.

Type — Morocco: *Argania spinosa* (Sapotaceae).

The type was not found in PC and its location is unknown.

Mycosphaerella bixae Crous & R.L. Benchimol, in Crous *et al.*, *Sydowia* 52: 79. 2000.

Type — Brazil: *Bixa orellana* (Bixaceae).

Anamorph: *Passalora bixae* Crous & R.L. Benchimol (Crous & Braun, 2003)

No material was studied of this recently described species.

Sphaerella boehmeriae Rabenh., *Fungi Europaei Exsiccati*, ed. nov., ser. 2, no. 2341. 1876 ≡ *Metasphaeria boehmeriae* (Rabenh.) Sacc., *Syll. Fung.* 2: 156. 1883.

Type — India: Calcutta. On dead stems of *Boehmeria nivea* (Urticaceae). Kurz, I 1874, distributed in Rabenhorst, *Fungi Europaei Exsiccati*, ed. nov., ser. 2, no. 2341 (L, isotype).

In the isotype material studied only a *Phoma* coelomycete could be found, with conidia fitting the original description of the spores.

Mycosphaerella bohémica (Bubák & Kabát) Sacc. ex Trotter, *Syll. Fung.* 24: 893. 1928 ≡ *Rehmiellopsis bohémica* Bubák & Kabát, in Bubák, *Naturwiss. Z. Forst-Landw.* 6: 320. 1910.

Type — Czech Republic: *Abies alba* (Pinaceae).

No material was found in BPI, where the Bubák herbarium is kept, and the location of this type is unknown.

Sphaerella bolivari Caball., *Mem. Real Soc. Esp. Hist. Nat.* 15: 312. 1929.

Type — Spain: Madrid, Casa de Campo. On upper and lower surface of dead leaves of *Phlomis herba-venti* (Lamiaceae). Caballero no. 9582, IV 1929 (MA, holotype).

The holotype contains nothing identifiable.

Mycosphaerella bolleana B.B. Higgins, *Amer. J. Bot.* 7: 443. 1920 ≡ *Sphaerella bolleana* (B.B. Higgins) Trotter, *Syll. Fung.* 24: 879. 1928.

Type — USA: On upper and lower surface of leaf of *Ficus carica* (Moraceae).

Anamorph: *Pseudocercospora bolleana* (Thüm.) Sivan. *vide* Sivanesan (1984).

No material was found in BPI, where the Higgins herbarium is kept. Material studied (Romania, Constanța, Hagieni, Constantinescu & Negru, *Herbarium Mycologicum Romanicum* no. 2046, X 1970, L) contains only the anamorph *Pseudocercospora bolleana* (Thüm.) Sivan. and an *Asteromella* spermatial state.

Mycosphaerella bombycina T.S. Viswan., in T.S. Viswan. & Tilak, *Sydowia* 14: 309. 1960.— Fig. 114.

Type — India: Karnataka, Coorg. On upper and lower surface of dead leaves of *Callistemon lanceolata* [as “*Syzygium cumini*”] (Myrtaceae). Anahosur no. 545, X 1966 (LWG, holotype).

This is morphologically indistinguishable from *M. punctiformis*, with ascospores somewhat aggregated, asci cylindrical, ascospores 7-9 × 2-2.5 µm.

Mycosphaerella bonae-noctis (Sacc.) Syd. & P. Syd., Ann. Mycol. 15: 205. 1917 = *Sphaerella bonae-noctis* Sacc., Bol. Soc. Brot. 11: 61. 1893.— Fig. 115.

Type — São Tomé: Benfica. On pale spots with brown margins on upper surface of living leaves of *Ipomoea bonanox* (Convolvulaceae). Trotter, VI 1885 (PAD, holotype).

Cited as synonymous with *Davidiella allicina* (as *Mycosphaerella allicina*) by Tomilin (1979). However, the holotype shows that this is a species of *Didymella*, with paraphyses sparse, 2 µm wide, asci clavate, ascospores 20-23 × 7-8.5 µm.

Sphaerella bonaerensis Speg., Anales Soc. Ci. Argent. 10: 139. 1880.

Type — Argentina: Buenos Aires, Barracas. On upper and lower surface of dead leaves of *Celtis tala* (Cannabaceae). Spegazzini, V 1880 (LPS, holotype).

Anamorph: *Cercospora dubia* Spegazzini fide Spegazzini (op. cit.) (= *Passalora dubia* (Riess) U. Braun).

The type was immature, and according to annotations of the original author, ascospores have never been observed in the specimen.

Sphaerella boquilae Speg., Revista Fac. Agron. Vet., ep. 2, 6: 53. 1910.— Fig. 116.

Type — Chile: Concepción, Cerro Caracol. On lower surface of dead leaves of *Boquila trifoliata* (Lardizabalaceae). Spegazzini, I 1909 (LPS, holotype).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradians*, with asci cylindrical, ascospores 8-11 × 2-2.5 µm.

Mycosphaerella borealis (Bubák & Vleugel) M.E. Barr, Contr. Univ. Michigan Herb. 9: 602. 1972 = *Sphaerella borealis* Bubák & Vleugel, in Vleugel, Svensk Bot. Tidskr. 11: 309. 1917 = *Phaeosphaerella borealis* (Bubák & Vleugel) Tomilin, Novosti Sist. Nizsh. Rast. 9: 119. 1972.— Fig. 970.

Type — Sweden: Luleå, Björkin. On lower surface of dead leaves of *Alnus incana* var. *borealis* (Betulaceae). Vleugel, VI 1914 (BPI, holotype).

This is best accepted as *Phaeosphaerella borealis* (Bubák & Vleugel) Tomilin, with asci cylindrical, without hamathecial filaments or cells, ascospores greenish black, 11-12 × 4-4.5 µm. Also present in the type is isotype material of *Planistromella conglomeratiformis* (Bubák & Vleugel) Aptroot (see below).

Mycosphaerella borrieriae Kranz, Nova Hedwigia 18: 236. 1969.— Fig. 117.

Type — Guinea: Kindia. On upper and lower surface of dead leaves of *Borreria verticillata* (Rubiaceae). Kranz no. 264, V 1962 (IMI no. 95629a, holotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores 15-18 × 4.5-4.5 µm.

Mycosphaerella botrychii (Rostr.) Savile, Mycologia 51: 297. 1959 = *Sphaerella botrychii* Rostr., Bot. Tidsskr. 22: 265. 1899.— Fig. 118.

Type — Denmark: Pileå Stadion. On upper and lower surface of dead fronds of *Botrychium lunaria* (Botrychiaceae). Lindberg, VIII 1891 (C, holotype).

This belongs to section *Longispora*, and is morphologically indistinguishable from *M. millegrana*, with ascomata small, in dense clusters, asci cylindrical, ascospores 13-15 × 1.5-1.7 µm.

Mycosphaerella boureae Syd., nomen herbariorum (not validly published, Article 32).— Fig. 119.

Authentic material — Philippines: Luzon, Prov. Bataan, between Limay and Lamao. On white spots with brown margins on upper surface of living leaves of *Boerhavia* ["*Bourea*"] *erecta* (Nyctaginaceae). Graft, XI 1912, distributed in Sydow, Fungi Exotici Exsiccati no. 128 (BPI). This is a parasitic species, with asci cylindrical, ascospores 14-19 × 3-4 µm.

Mycosphaerella brachypodii Schief., nomen herbariorum (not validly published, Article 32).— Fig. 120.

Authentic material — Germany: Hannover, Hildesheim, Knebel. On dead leaf tips of *Brachypodium sylvaticum* (Poaceae). Schieferdecker, IX 1949 (B).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. disseminata*, with asci pyriform, ascospores 12-14 × 2.5-3.5 µm.

Mycosphaerella brachyscomes Petr., Sydowia 9: 562. 1955 [as "*brachycomes*"].— Fig. 122.

Type — Australia: Australian Capital Territory, Mt. Coree. On dead stems and involucra of *Brachyscome diversifolia* (Asteraceae). Gauba, XII 1954 (BPI, isotype), also distributed in Reliquiae Petrakianae no. 1246, XII 1955 (H, L, topotypes or possibly isotypes if the date is wrong).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. ammophilae*, with asci pyriform, ascospores 8-10 × 3-4 µm.

Mycosphaerella brachythecha (Cooke) Magnus, in Dalla Torre & Sarnth., Fl. Tirol, Vorarlberg, Liechtenstein, III, Pilze: 467. 1905 = *Sphaerella brachythecha* Cooke, Grevillea 7: 88. 1879.— Fig. 121.

Type — United Kingdom: Scotland, Grampian, Forres. On grey spots on upper surface of living leaves of *Vaccinium vitis-idaea* (Ericaceae). Keith (K, holotype).

Cited as synonymous with *Mycosphaerella stemmatea* by Tomilin (1979), with which the holotype agrees well. It is a parasitic species, with asci cylindrical, ascospores 14-17 × 3-4 µm.

Sphaerella brachythecha Cooke & Harkn. See *Mycosphaerella harknessii* (Sacc.) Tomilin.

Mycosphaerella bracteophila (Pass.) Tomilin, Novosti Sist. Nizsh. Rast. 6: 119. 1969 = *Sphaerella bracteophila* Pass., Atti Reale Accad. Lincei, Mem. Cl. Sci. Fis., ser. 4, 3: 5. 1887.— Fig. 123.

Type — Italy: Parma. On upper and lower surface of dead but still attached fruit bracts of *Tilia argentea* (Malvaceae). Passerini, I 1887, distributed in Roumeguère, Fungi Gallici Exsiccati no. 4055 (NY, isotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores $16-22 \times 5.5-7 \mu\text{m}$.

Mycosphaerella braheae Siemaszko, Acta Soc. Bot. Pol. 1: 19. 1923.— Fig. 124.

Type — Georgia: Adzharskaya, Batumi. On upper surface of dead leaves of *Brahea edulis* (Arecaceae). Siemaszko, IX 1917 (BPI, isotype).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradians*, with asci cylindrical, ascospores $10-12 \times 2.5-3 \mu\text{m}$.

Mycosphaerella brassicicola (Duby) Lindau, in Engl. & Prantl, Natürlichen Pflanzenf. 1(1): 424. 1897 = *Sphaeria brassicicola* Duby, in DC., Botan. Gallicum, ed. 2, 2: 712. 1830 [as “*brassicaecola*”] non *Sphaeria brassicicola* Berk. & Broome (1860) = *Dothidea brassicae* Desm., Ann. Sci. Nat. Bot., ser. 2, 17. 1842 = *Sphaerella brassicicola* (Duby) Ces. & De Not., Comment. Soc. Crittog. Ital. 1(4): 238. 1863 [as “*brassicaecola*”].— Fig. 125.

Type — France: On upper and lower surface of dead leaves of *Brassica* (Brassicaceae). Desmazières, Plantes Cryptogames de France no. 95 (B-Desmazières, isotype).

Spermatial state: *Asteromella brassicae* (F. Chevallier) Boerema & van Kesteren *vide* Corlett (1988).

The isotype studied is empty. Additional material studied (Netherlands, Utrecht, Vereeniging voor de Flora v. Nederland enz. no. 318, L) belongs to section *Caterva*, and is morphologically indistinguishable from *M. superflua*, with asci cylindrical, ascospores $13-16 \times 4.5-5.5 \mu\text{m}$.

Sphaerella briardi Sacc., in Briard & Har., Rev. Mycol. (Toulouse) 12: 177. 1890 = *Sphaerella patouillardii* subsp. *briardi* (Sacc.) Sacc., Syll. Fung. 9: 635. 1891.

Type — France: *Buxus sempervirens* (Buxaceae).

Cited as synonymous with *M. buxicola* by Tomilin (1979), which is however based on an anamorph. Teleomorph material called thus belongs to section *Longispora*, and is morphologically indistinguishable from *M. latebrosa*.

Mycosphaerella briedeliae Syd. & P. Syd., Ann. Mycol. 12: 199. 1914 [as “*brideliae*”] = *Sphaerella briedeliae* (Syd. & P. Syd.) Trotter, Syll. Fung. 24: 862. 1928 [as “*brideliae*”].— Fig. 126.

Type — Philippines: Luzon, Prov. Laguna, Los Baños. On white spots on upper surface of living leaves of *Briedelia stipularis* (Euphorbiaceae). Baker no. 2577, I 1914 (S, holotype).

This is a parasitic species, with asci cylindrical, ascospores $11-13 \times 3-3.5 \mu\text{m}$. Additional material studied (prov. Rizal, Antipolo, Ramos, distributed in Cryptogamae Vindobonenses no. 3267, VIII, L) is immature.

Mycosphaerella brionnensis (Sacc. & Malbr.) Tomilin, Novosti Sist. Nizsh. Rast. 7: 202. 1970 = *Sphaerella brionnensis* Sacc. & Malbr., Atti Reale Ist. Veneto Sci., ser. 6, 1: 1275. 1883.— Fig. 127.

Type — France: Eure, Brionne. On upper and lower surface of dead leaves of *Angelica sylvestris* (Apiaceae).

Malbranche, distributed in Roumeuguère, Fungi Selecti Exsiccati no. 4933 (NY, isotype).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. superflua*, with asci cylindrical, ascospores $13-16 \times 3-3.5 \mu\text{m}$.

Sphaerella bromi Auersw. See *Mycosphaerella longissima* (Fuckel) Lindau.

Mycosphaerella bromi-albidis Nevod. See *Mycosphaerella nevodovskii* Tomilin.

Sphaerella broteriana Alcalde, Anales Jard. Bot. Madrid 6: 402. 1946 [“1945”].— Fig. 128.

Type — Spain: La Bujeda. On dead stems, calyces and upper and lower surface of dead leaves of *Potentilla rupestris* (Rosaceae). Alcalde & Caball. no. 12593, VI 1935 (MA, holotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. ammophilae*, with asci pyriform, ascospores $8-10 \times 3-4 \mu\text{m}$.

Mycosphaerella brunnea Hansf., Proc. Linn. Soc. New South Wales 81: 35. 1956.— Fig. 129.

Type — Australia: Queensland, Brisbane. On living leaves of *Tristania* (Myrtaceae). Bailey no. 506 (IMI no. 59149, isotype, slide only).

This is a parasitic species, with asci cylindrical, ascospores $15-18 \times 3-4 \mu\text{m}$.

Sphaerella brunnea (Berk. & M.A. Curtis) Cooke, Grevillea 6: 146. 1878 = *Depazea brunnea* Berk. & M.A. Curtis, in Berk., Grevillea 4: 155. 1876 = *Laestadia brunnea* (Berk. & M.A. Curtis) Sacc., Syll. Fung. 1: 429. 1882.

Type — USA: *Acer rubrum* (Sapindaceae). Curtis (K, holotype).

The type contains an unidentifiable ascomycete.

Mycosphaerella brunneola (Fr. : Fr.) Johanson ex Oudem., Rév. Champ. Pays-Bas 2: 207. 1897 = *Sphaeria brunneola* Fr., Systema Mycol. 2: 526. 1823, sanctioned by Fr., Systema Mycol. 2: 526. 1823 = *Ascospora brunneola* (Fr. : Fr.) Fuckel, Jahrb. Nassauischen Vereins Naturk. 23-24: 94. 1870 = *Sphaerella brunneola* (Fr. : Fr.) Cooke, Handb. Brit. Fungi 2: 922. 1871.— Fig. 130.

Type — Sweden: On upper and lower surface of dead leaves of *Convallaria majalis* (Asparagaceae). Fr., Scleromyceti Suecici no. 284 (UPS, holotype; B, isotype).

Spermatial states and anamorphs: *Asteroma subradians* Fr. *vide* Saccardo (1882), *Asteromella convallariae* (Cavara) Petr. and *Septoria brunneola* (Fr.) Niessl *vide* Eriksson (1992).

Cited as probably synonymous with *M. subradians* by Eriksson (1992). However, both type specimens studied are in bad condition, although they suggest that this synonymy is correct. Additional material studied (Germany, Oberfranken, Langheim, Rohnfelder, distributed in Allesch. & Schnabl, Fungi Bavarici no. 537, IV 1896, L) belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradians*, with asci cylindrical, ascospores $10-12 \times 2.5-3.5 \mu\text{m}$.

Sphaerella brunneola f. *convallariae* Rabenh., Fungi Europaei Exsiccati no. 1854. 1874.

Type — Germany: Schwaben, Dounstetten. On upper and lower surface of dead leaves of *Convallaria majalis* (Asparagaceae). Kemmler, 1873, distributed in Rabenhorst, Fungi Europaei Exsiccati no. 1854 (L, isotype).

As the nominal form, this belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradians*.

Mycosphaerella brunneomaculans T. Kobay., Trans. Mycol. Soc. Japan 25: 255. 1984.

Type — Paraguay: *Tabebuia flavescens* (Bignoniaceae).

The type was not found in TNS and its location is unknown.

Sphaerella bryi Gonz. Frag., Trab. Mus. Nac. Ci. Nat., Ser. Bot. 10: 78. 1916 [as “*bryii*”].— Fig. 131.

Type — Spain: Sevilla, Pedrozo de la Sierra. On setae and thecae of *Bryum provinciale* [“*capillare* var. *meridionale*”] (Musci, Bryaceae). Frago no. 1851, V 1916 (MA, holotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. ammophilae*, with asci pyriform, ascospores 9-11 × 3-4 µm.

Sphaerella bryoniae Auersw., in Gonn. & Rabenh., Mycol. Europaea 5-6: 15. 1869 ≡ *Sphaeria bryoniae* (Auersw.)

Fuckel, Jahrb. Nassauischen Vereins Naturk. 23-24: 112. 1870 ≡ *Didymosphaeria bryoniae* (Auersw.) Niessl, Oesterr. Bot. Z. 25: 130. 1875 ≡ *Didymella bryoniae* (Auersw.) Rehm, Ber. Naturhist. Vereins Augsburg 26: 27. 1881.

Type — Germany: On upper and lower surface of dead leaves of *Bryonia cretica* [“*dioica*”] (Cucurbitaceae). Rabenh., Fungi Europaei Exsiccati no. 746 (B, isotype).

Anamorph: *Ascochyta cucumis* Fautrey & Roum. *vide* Corlett *et al.* (1986).

Material studied (Czech Republic, Brno [“Brünn”], Niessl, distributed in Rabenhorst, Fungi Europaei Exsiccati no. 1852, L) contains only the anamorph, *Ascochyta cucumis* Fautrey & Roum.

Mycosphaerella buckinghamiae Crous & Summerell, in Crous *et al.*, Australasian Plant Pathology 29: 272. 2000.

Type — Australia: *Buckinghamia* (Proteaceae).

No material was studied of this recently described species.

Mycosphaerella bulgarica Petr., Ann. Mycol. 29: 367. 1931.

Type — Bulgaria: *Carex* (Cyperaceae).

The type was not found in W nor in any other herbarium consulted.

Mycosphaerella bumeliae (Cooke) J.H. Miller, Mycologia 33: 79. 1941 ≡ *Sphaerella bumeliae* Cooke, Grevillea 7: 54. 1878.— Fig. 132.

Type — USA: Georgia, Darien. On upper surface of dead leaves of *Bumelia* (Sapotaceae). Ravenel no. 2434 (K, holotype).

Cited as synonymous with *Mycosphaerella hermione* by Tomilin (1979), which is morphologically indistinguishable from *M. punctiformis*. This indeed belongs to section *Mycosphaerella* s.s., but is characterised by ascomata which

are piercing the epidermis in groups, cylindrical asci and very slender ascospores of 8-9 × 1.5-2 µm.

Mycosphaerella buna R. Kaneko & Kakishima, Mycoscience 42: 60. 2001.

Type — Japan: *Fagus crenata* (Fagaceae).

Anamorph: *Pseudocercospora fide* Kaneko & Kakishima (op. cit.).

No material was studied of this recently described species.

Mycosphaerella bupleuri (Rota-Rossi) Tomilin, Novosti Sist. Nizsh. Rast. 7: 202. 1970 ≡ *Sphaerella bupleuri* Rota-Rossi, Atti Ist. Bot. Univ. Pavia ser. 2, 13: 207. 1914.

Type — Italy: *Bupleurum graminifolium* (Apiaceae).

Cited as synonymous with *Davidiella allicina* (as *Mycosphaerella allicina*) by Tomilin (1979). No material was found in any of the herbaria consulted.

Mycosphaerella bupleurina Petr., Hedwigia 68: 207. 1928.— Fig. 133.

Type — Russia: Altay. On dead stems of *Bupleurum multinerve* (Apiaceae). Antonov no. 444, VIII 1925 (W, holotype).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradians*, with asci cylindrical, ascospores 11-13 × 2.5-3.5 µm.

Mycosphaerella burnati Cruchet, Bull. Soc. Vaud. Sci. Nat. 44: 473. 1909 ≡ *Sphaerella burnati* (Cruchet) Sacc. & Traverso, Syll. Fung. 20: 818. 1911.

Type — Switzerland: *Lloydia serotina* (Asparagaceae).

Spermatial state: Associated with *Asteroma lloydiae* Cruchet *vide* Cruchet (op. cit.).

Synonymised with *Davidiella allicina* (as *Mycosphaerella tassiana*) by von Arx (1949). No material was found in any of the herbaria consulted.

Sphaerella buxi (DC. ex Desm.) Ces. & De Not., Comment. Soc. Crittog. Ital. 1(4): 237. 1863 ≡ *Sphaeria buxi* DC. ex

Desm., Fl. Fr. 6: 146. 1815 ≡ *Hyponectria buxi* (DC. ex Desm.) Sacc., Michelia 1: 250. 1878 ≡ *Laestadia buxi* (DC. ex Desm.) Sacc., Syll. Fung. 2: XXXI. 1883 ≡ *Guignardia buxi* (DC. ex Desm.) Lindau, Hilfsb. Sammeln Ascomyceten: 21. 1903.— Fig. 134.

Type — France: On lower surface (not in spots) of living leaves of *Buxus sempervirens* (Buxaceae). Desmazières, Plantes Cryptogames de France no. 1280 (BPI, isotype).

Accepted as *Hyponectria buxi* (DC. ex Desm.) Sacc. by von Arx & Müller (1954), with which the isotype agrees well, with asci cylindrical, ascospores simple, 13-15 × 4-5 µm.

Mycosphaerella buxicola (DC.) Tomilin, Novosti Sist. Nizsh. Rast. 6: 119. 1970 [“1969”], based on an anamorph

(illegitimate, Article 59) ≡ *Sphaeropsis lichenoides* var. *buxicola* DC., Fl. Franç. 5: 147. 1815, based on an anamorph (illegitimate, Article 59) ≡ *Sphaeria buxicola* (DC.) Fr., Systema Mycol. 2: 528. 1823, based on an anamorph (illegitimate, Article 59).

Type — France: *Buxus sempervirens* (Buxaceae).

The species is incorrectly based on an anamorph. Teleomorph material called thus belongs to section

Longispora, and is morphologically indistinguishable from *M. latebrosa*.

Sphaerella buxifolia Cooke, J. Bot. 21: 69. 1883 = *Laestadia buxifolia* (Cooke) Sacc., Syll. Fung. 2: XXXI. 1883.

Type — India: *Buxus sempervirens* (Buxaceae).

No material was studied as the species was already placed in *Laestadia* by Saccardo (op. cit.).

Mycosphaerella byliana Syd., Ann. Mycol. 22: 423. 1924.

Type — South Africa: *Olea* (Oleaceae).

The type was not found in S or any of the herbaria consulted.

Mycosphaerella byrsonimae Bat. & Peres, in Bat., Peres, Cavalcanti & Heringer, Atas Inst. Micol. 3: 221. 1966.

Type — Brazil: On white spots on upper surface of living leaves of *Byrsonima basiloba* (Malpighiaceae). Batista exs. no. 15286 (URM 21791, holotype).

On the type specimen, which at the same time is the presumed type of several other fungi, only a *Phoma* is present.

Mycosphaerella cacaliae Ziling, Trudy Bot. Inst. Akad. Nauk SSSR, ser. 2, Sporov. Rast., fasc. 3: 680. 1936.

Type — Russia: Far East, Pos'tet. On pale spots with brown margins on upper surface of living leaves of *Cacalia aconitifolia* (Asteraceae). Ziling, VII 1928 (BPI, isotype).

Anamorph: *Septoria cacaliae-aconitifoliae* Ziling, fide Ziling (op. cit.).

The isotype contains the anamorph, but also an *Ascochyta* coelomycete with 1-septate conidia that might have been the basis for the description of the *Mycosphaerella*.

Mycosphaerella caespitosa (Ellis & Everh.) Diehl, in W.B. Cooke, Mycobiota N. Amer., fasc. 4 no. 93. 1940 = *Sphaerella caespitosa* Ellis & Everh., J. Mycol. 9: 166. 1903.— Fig. 135.

Type — USA: Texas, Meridan. On lower surface of dead leaves of *Quercus virginiana* (Fagaceae). Long no. 957, IV 1901 (BPI, isotype).

No type material is preserved in NY; the isotype in BPI might be the only existing type material left. It shows that this is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores 8-10 × 2-3 μm. Additional material studied (California, Del Norte Co., on *Quercus vaccinifolia*, Parks no. 3398, IV 1933 & no. 6014, V 1937, both B) are *Planistromella operculata* (see below).

Sphaerella caladii (Schwein.) Sacc., in P. Syd., Syll. Fung. 12: 712. 1897 = *Depazea caladii* Schwein., in Berk. & M.A. Curtis, J. Acad. Nat. Sci. Philadelphia, ser. 2, 2: 291. 1853 = *Sphaeria caladii* (Schwein.) Sacc., Syll. Fung. 1: 524. 1882.

Type — Surinam. On white spots with brown margins on upper and lower surface of living leaves of *Caladium* (Araceae). (K, isotype).

No type material was preserved in PH, but an isotype ex herbarium Schweinitz was found in K. It is a parasitic species, with asci cylindrical, ascospores 13-15 × 3-4 μm.

Mycosphaerella calamagrostidis Petr., Hedwigia 65(1): 226. 1925 = *Mycosphaerella petrakii* Tomilin, Novosti Sist. Nizsh. Rast. 1970: 200. 1970, superfluous (illegitimate, Article 52) nomen novum (Article 58).— Fig. 136.

Type — Poland: Zentrale Waldkarpaten, Chomiak. On dead culms of *Calamagrostis montana* (Poaceae). Petr., Fungi Polonici Exsiccati no. 641, VII 1918 (W 16123, lectotype; W 11069, isolectotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. ammophilae*, with asci pyriform, ascospores 7.5-9 × 2.5-3 μm.

Mycosphaerella calamagrostidis Volkart. See *Mycosphaerella volkartii* Tomilin.

Mycosphaerella calamagrostidis H.C. Greene. See *Mycosphaerella greenei* Tomilin.

Mycosphaerella calceoli Kirschst. & Kirulis, in Kirschst., Ann. Mycol. 33: 210. 1935.— Fig. 138.

Type — Latvia: Prov. Vidzeme, Pededze. On *Cypripedium calceolus* (Orchidaceae). Kirulis no. 922 & no. A.1008, VI 1934 (B, syntypes).

Mycosphaerella californica (Cooke & Harkn.) Lindau, in Engl. & Prantl, Natürlichen Pflanzenf. 1(1): 426. 1897 = *Sphaerella californica* Cooke & Harkn., in Cooke, J. Bot. 21: 136. 1883.— Fig. 139.

Type — USA: California. On dead leaves of Poaceae. Harkness no. 1242 (K, 2 isotypes).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. disseminata*, with asci clavate, ascospores 11-13 × 4-5 μm, surrounded by a gelatinous sheath.

Mycosphaerella callistea (Syd. & P. Syd.) Rehm, Ann. Mycol. 8: 300. 1910 [as "callista"] = *Sphaerella callistea* Syd. & P. Syd., Ann. Mycol. 7: 439. 1909.— Fig. 140.

Type — Germany: Mecklenburg, Müritz. On lower surface of dead sectors of living leaves of *Osmunda regalis* (Osmundaceae). Sydow, Mycotheca Germanica no. 785, VIII 1909 (L, isotype); also distributed in Kryptogamae exsiccatae no. 2622 (L, isotype).

The isotype materials studied, which agree well with the description, contains immature material of *Pleospora herbarum* (Pers.: Fr.) Rabenh., with asci bitunicate, with paraplectenchymatous tissue between them, ascospores still hyaline and 1-septate, 22-28 × 10-13 μm. Therefore this is morphologically indistinguishable from *Pleospora herbarum* (Pers.: Fr.) Rabenh.

Sphaerella callunae De Not., Sferiacei Italici 1: 88. 1863 = *Physalospora callunae* (De Not.) Sacc., Syll. Fung. 1: 447. 1882.— Fig. 141.

Type — Italy: Monsinu. On corticate branches of *Calluna vulgaris* (Ericaceae). De Notaris, X 1862 (RO, holotype).

This is a species of *Arthopyrenia*, close to *A. punctiformis* Pers., but differing e.g. by the minute, ca. 0.1 mm wide ascomata with ascomal wall KOH negative, ostiole thickened, pseudoparaphyses 1.5 μm wide, asci pyriform to clavate, ascospores 13-15 × 4.5-6 μm, surrounded by a

gelatinous sheath. Therefore it is transferred here to the genus as *Arthopyrenia callunae* (De Not.) Aptroot comb. nov., **MB 500335**. **Basionym:** *Sphaerella callunae* De Not., *Sferiacei Italici* 1: 88. 1863. According to the additional material studied, it is apparently not host-restricted. Additional materials seen (The Netherlands, Friesland, Wadden Sea Island Vlieland, Lange Paal, on corticate branches of *Calluna vulgaris* (Ericaceae), Aptroot no. 54843, IX 2002, ABL; also: Same locality, on bark of *Pinus pinaster* (Pinaceae), Aptroot no. 54836, IX 2002, ABL) agree. Von Arx & Müller (1954), without studying the type, expressed the view that this was morphologically indistinguishable from *Pseudophacidium ledi* (Alb. & Schwein.) P. Karst.

Sphaerella callunae Roum., *Fungi Gallici Exsiccati* no. 795. 1880, nomen herbariorum (not validly published, Article 32, later homonym (illegitimate, Article 53).

Authentic material — France: Morvan. On upper surface of dead leaves of *Calluna vulgaris* (Ericaceae). Grognot, distributed in Roumeguère, *Fungi Gallici Exsiccati* no. 795 (L).

The authentic material studied contains only empty ascomata.

Mycosphaerella calopogonii (Gonz. Frag. & Cif.) Cif., *Ist. Bot. Reale Univ. Reale Lab. Crittog. Pavia Atti*, ser. 5, 19: 135. 1962 = *Sphaerella calopogonii* Gonz. Frag. & Cif., *Estac. Agron. Moca*, ser. B, Bot., 11: 16. 1928.

Type — Dominican Republic: *Calopogon orthocarpi* (Orchidaceae).

No material could be studied as it was not present in MA or BPI, where respectively the Frago and Ciferri herbaria are held.

Mycosphaerella calotropidis T.S. Viswan., in T.S. Viswan. & Tilak, *Sydowia* 14: 308. 1960.

Type — India: *Calotropis gigantea* (Apocynaceae).

No material could be studied as the location of the type is unknown.

Mycosphaerella [*“Mycosphaerium”*] *calthae* Clem., *Cryptogamae Formationum Coloradensium* no. 216. 1906, nomen herbariorum (not validly published, Article 32).— Fig. 142.

Authentic material — USA: Colorado, Cabin Canyon. On upper and lower surface of dead leaves and petioles of *Caltha leptosepala* (Ranunculaceae). Clements, VII 1906, *Cryptogamae Formationum Coloradensium* no. 216 (BPI).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradians*.

Mycosphaerella calycanthi Koshk. & Frolov, *Mikoflora Podgornoi Ravniny Kopetdaga i Tsentral'nykh Karakumov*: 83. 1973, nomen nudum (not validly published, Article 32).

Type — Turkmenia: *Calycanthus laevigatus* (Calycanthaceae).

No material could be studied as the type was not included in loans from LE or LEP.

Mycosphaerella calycicola (Pass.) Jacz., *Gribnye i Bakterialnye Bolezni Klevera*: 59. 1916 = *Sphaerella calycicola* Pass., *Erbario Crittogamico Italiano*, ser. 2, no. 1462. 1885.— Fig. 143.

Type — Italy: Parma, Vigheffio. On dead, but still attached, calyces of *Trifolium angustifolium* (Fabaceae). Passerini, IX 1884, distributed in *Erbario Crittogamico Italiano*, ser.2, no. 1462 (NY, isotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores 15-19 × 5-6.5 µm.

Mycosphaerella camarae Dias, *Mem. Soc. Brot.* 21: 49. 1970.— Fig. 144.

Type — Portugal: Algarve, Monohique. On dead stems of *Tuberaria guttata* (Cistaceae). Dias & Lucas 2268, IV 1968 (LISE 91372, holotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. ammophilae*, with asci pyriform, ascospores 9-10 × 2.5-3.5 µm.

Mycosphaerella camelliae Petch, *Ann. Roy. Bot. Gard. (Peradeniya)* 9: 319. 1925.— Fig. 145.

Type — Sri Lanka: Concordia. On dead spots on upper and lower surface of living leaves of *Camellia theifera* (Theaceae). Petch no. 6554, X 1922 (K, holotype).

Anamorph: Associated with *Phyllosticta theae* Speschnev *vide* Petch (op. cit.).

This is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores 7-8 × 2-2.5 µm.

Sphaerella camelliae Cooke, *Grevillea* 13: 4. 1884 [as “*camilleae*”] = *Laestadia camelliae* (Cooke) Berl. & Voglino, *Syll. Fung.*, *Addit. ad vol.* 1-4: 62. 1886 [as “*camilleae*”].

Type — India: *Camellia theifera* (Theaceae). (K, holotype). Already synonymised with *Glomerella cingulata* (Stoneman) Spauld. & H. Schrenk by von Arx & Müller (1954), for which it is an older name. The name *Glomerella cingulata* has been widely used for one of the most common ascomycetes, which, for instance, is present with the highest number of specimens in the IMI herbarium. However, several older names are already reported, e.g. by von Arx & Müller (1954). Therefore, the name *Gnomoniopsis cingulata* Stoneman will have to be proposed for conservation over *Sphaerella camelliae* and other names.

Mycosphaerella campanulae (Ellis & Kellerm.) Naumov, *Mater. Mikol. Fitopatol. Rossii* 1: 41. 1916 = *Sphaerella campanulae* Ellis & Kellerm., *Amer. Naturalist* 17: 1166. 1883.— Fig. 146.

Type — USA: Ohio, Fairfield Co. On dead stems of *Campanula americana* (Campanulaceae). Kellerman no. 386, VII 1883 (NY, holotype; NY, isotype); also distributed in Ellis & Everhart, *North American Fungi* no. 1673, VII 1883 (L, NY (2×), isotypes).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. superflua*, with asci cylindrical, ascospores 14-16 × 3.5-4.5 µm. It was cited as synonymous with *M. isariphora* by Tomilin (1979).

Mycosphaerella camphorosmae Lobik, Materialy po floristicheskim i faunisticheskim obsledovaniyam Terskogo okruga: 25. 1928.

Type — Russia: *Camphorosma monspeliaca* (Amaranthaceae).

No material could be studied as the type was not included in loans from LE or LEP.

Sphaerella campoi Speg., Bol. Acad. Nac. Ci. 25: 58. 1921.— Fig. 147.

Type — Chile: Victoria, Mariluán. On lower surface of dead leaves of *Azara* (Salicaceae). Campo, V 1918 (LPS, holotype).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradians*, with asci cylindrical, ascospores $9-11 \times 2-2.5 \mu\text{m}$.

Sphaerella canadensis Ellis & Everh., N. Amer. Pyrenomyc.: 280. 1892.— Fig. 148.

Type — Canada: Ontario, London. On scales of dead cones of *Thuja occidentalis* (Cupressaceae).

Already cited as synonymous with *Scirrhia conigena* (Peck) M.E. Barr by Barr (1972) with which material studied: (USA: California, Trinidad, Humboldt County, on living leaves (scales) of *T. plicata*, Parks, X 1932, B) agrees, even though it grows on living leaves, with asci cylindrical, surrounded by copious pseudoparenchymatic cells of ca. $4-8 \mu\text{m}$, ascospores $18-20 \times 3.5-5 \mu\text{m}$.

Mycosphaerella canariensis Petr., Bot. Jahrb. Syst. 62, Beibl. 142: 117. 1928.— Fig. 149.

Type — Canary Islands: Tenerife. On upper and lower surface of dead leaves of *Plantago arborescens* (Plantaginaceae). Ade, V 1926 (W 10054, lectotype; W 26900, isolectotype).

Cited as synonymous with *M. plantaginis* (Sollm.) Vesterg. by Tomilin (1979), which is morphologically indistinguishable from *M. punctiformis*.

However, the type and additional material (Madeira: Funchal, Ade, VI 1926, W, 2 \times) shows that this belongs to *Davidiella*, and is morphologically indistinguishable from *D. ammophilae*, with asci pyriform, ascospores $8-11 \times 2.5-4 \mu\text{m}$. In the type materials, also a *Phaeosphaeria* is present.

Sphaerella canariensis Tassi. See *Mycosphaerella euphorbiae-canariensis* Tomilin.

Mycosphaerella canavaliae Syd., Ann. Mycol. 21: 100. 1923.

Type — Philippines: Luzon, Bataan. On brown spots on lower surface of living leaf of *Canavalia gladiata* (Fabaceae). Reyes, Bureau of Science no. 39283, IX 1920 (S, holotype).

The type contains only a coelomycete.

Mycosphaerella canephora Saccas, I.F.C.C. Bull. 16: 287. 1981, type not designated (not validly published, Article 37).

Authentic material — Central African Republic: *Coffea robusta* (Rubiaceae).

No material could be studied as the type was not designated and no material was included in a loan from PC.

Sphaerella canescens P. Karst., Fungi Fenniae Exsiccati no. 895. 1869.

Type — Finland: *Salix acutifolia* (Salicaceae).

Cited as morphologically indistinguishable from *Venturia chlorospora* (Ces.) P. Karst. by Müller & von Arx (1962).

Sphaerella canifaciens (Fuckel) Auersw., in Gonn. & Rabenh., Mycol. Europaea 5-6: 16. 1869 [as “*canificans*”] = *Sphaeria canifaciens* Fuckel, Fungi Rhenani Exsiccati, no. 1774. 1868 [as “*canofaciens*”].— Fig. 150.

Type — Germany: Oestrich, Hallgarten. On upper and lower surface of dead leaves of *Triticum repens* (Poaceae). Fuckel, Fungi Rhenani Exsiccati no. 1774 (G, holotype; B, isotype).

The holotype is somewhat immature, but shows that it belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradians*, with asci cylindrical, ascospores ca. $12 \times 3 \mu\text{m}$.

Mycosphaerella cannabis (G. Winter) Magnus, in Dalla Torre & Sarnth., Fl. Tirol, Vorarlberg, Liechtenstein, III, Pilze: 465. 1905 = *Sphaerella cannabis* G. Winter, Hedwigia 11: 145. 1872 = *Didymella cannabis* (G. Winter) Arx, in E. Müll. & Arx, Beitr. Kryptogamenfl. Schweiz 11(2): 365. 1962.— Fig. 151.

Type — Austria: on dead stems of *Cannabis sativa* (Cannabaceae).

Anamorph: *Phyllosticta cannabis* (Lasch) Speg. fide Müller & von Arx (1962).

Material studied (Latvia, Vidzeme, Vestiena, Starcs no. D149, IX 1932, B), belongs to section *Caterva*, and suggests that this is morphologically indistinguishable from *M. superflua*, with asci cylindrical, ascospores $11-13 \times 4-5 \mu\text{m}$. Additional material studied (Germany, Osthavelland, Nauen, Röder, IX 1936, B) contains only a coelomycete.

Sphaerella cannae Speg., Anales Mus. Nac. Hist. Nat. Buenos Aires 19 (ser. 3, 12): 350. 1909.— Fig. 152.

Type — Argentina: Buenos Aires, Santa Catalina. On dead stems of *Canna indica* (Cannaceae). Spegazzini, XI 1905 (LPS, holotype).

This is a *Didymella*, with paraphyses copious, up to $1.5 \mu\text{m}$ wide, asci clavate, ascospores $10-12 \times 4-5 \mu\text{m}$.

Mycosphaerella capparidis Vasyag., in Vasyag., Byzova & Tartenova, Flora Sporovykh Rastenii Kazakhstana 12. Sumchatye Griby 2. Lokuloaskomitsety: 44. 1987.

Type — Russia: *Capparis spinosa* (Brassicaceae).

No material could be studied as the type was not included in loans from LE or LEP.

Mycosphaerella capreolatae (Pass.) Miles, Trans. Illinois Acad. Sci. 10: 249. 1917 = *Sphaerella capreolatae* Pass.,

Atti Reale Accad. Lincei, Mem. Cl. Sci. Fis., ser. 4, 6: 458. 1890 [as "*capreolata*"].

Type — Italy: *Bignonia capreolata* (Bignoniaceae).

Anamorph: Associated with *Gloeosporium bignoniae* Pass. *vide* Passerini (op. cit.).

No material could be studied as the type was not found in any of the herbaria consulted.

Sphaerella caprifoliorum (Desm.) Sacc., *Michelia* 2: 315. 1881 = *Sphaeria caprifoliorum* Desm., *Ann. Sci. Nat. Bot.*, ser. 2, 13: 188. 1840.

Type — France: On upper and lower surface of dead leaves of *Lonicera caprifolium* (Adoxaceae). Desmazières, *Fungi Exsiccati* no. 799 (L, isotype); also distributed in *Plantes Cryptogames de France* no. 1299 (BPI, isotype).

The isotype materials studied contain an overmature ascomycete, probably a *Pleospora*, certainly no *Mycosphaerella*.

Mycosphaerella capronii (Sacc.) Lind, *Skr. Vidensk.-Selsk. Christiana, Math.-Naturvidensk. Kl.* 1909(9): 16. 1910 ["1909"] = *Sphaerella capronii* Sacc., *Syll. Fung.* 1: 487. 1882.— Fig. 153.

Type — United Kingdom. On upper and lower surface of dead leaves of *Salix* (Salicaceae). (IMI no. 182085, isotype, slide only).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradians*, with asci cylindrical, ascospores 12-15 × 3-4 µm. Additional material studied (on *Salix triandra*, Klotzsch, *Herbarium Vivum Mycologicum* no. 366, L, sub *Sphaeria salicicola*) contains only a coelomycete.

Mycosphaerella capsellae A.J. Inman & Sivan., in A.J. Inman, Sivan., Fitt & R.L. Evans, *Mycol. Res.* 95: 1339. 1991.— Fig. 154.

Type — United Kingdom: Hertfordshire, Harpenden, Rothamsted Experimental Station. On dead pods (fruits) of *Brassica napus* ssp. *oleifera* (Brassicaceae). Inman, XI 1990 (IMI no. 344668, holotype).

Anamorph: *Pseudocercospora capsellae* (Ellis & Everh.) Deighton *vide* Inman, Sivanesan, Fitt & Evans (op. cit.).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. superflua*, with asci cylindrical, ascospores 16-19 × 3-4.5 µm.

Sphaerella carectorum (Berk. & M.A. Curtis) Cooke, *J. Bot.* 21: 137. 1883 = *Sphaeria carectorum* Berk. & M.A. Curtis, *Grevillea* 4: 153. 1876.

Type — USA: *Carex folliculata* (Cyperaceae).

No material could be studied as the type was not found in any of the herbaria consulted.

Mycosphaerella caricae Syd. & P. Syd., *Ann. Mycol.* 11: 403. 1913.— Fig. 155.

Type — Philippines: Luzon, Prov. Laguna, Los Baños. On upper and lower surface of white spots on living leaves of *Carica papaya* (Caricaceae). Reyes no. 1512, VII 1913 (S, holotype).

Anamorph: *Phoma caricae-papayae* (Tarr) Punith. *vide* Sivanesan (1984).

This is a parasitic species, with asci broadly cylindrical, ascospores 15-19 × 4.5-6 µm.

Mycosphaerella caricae (Maubl.) Hansf., *Proc. Linn. Soc. London* 153: 22. 1941, later homonym (illegitimate, Article 53) = *Sphaerella caricae* Maubl., *Bull. Soc. Mycol. France* 29: 358. 1913.

Type — Brazil: *Carica papaya* (Caricaceae).

Anamorph: *Asperisporium caricae* (Speg.) Maubl. *vide* Maublanc (op. cit.).

No material could be studied as the type was not included in a loan from PC.

Mycosphaerella caricicola (Fuckel) Lindau, *Hilfsb. Sammeln Ascomyceten*: 24. 1903 = *Sphaeria caricicola* Fuckel, *Fungi Rhenani Exsiccati* no. 1772. 1868 = *Sphaerella caricicola* (Fuckel) Fuckel, *Jahrb. Nassauischen Vereins Naturk.* 23-24: 101. 1870.— Fig. 156.

Type — Germany: Rauenthal. On upper and lower surface of dead leaves of *Carex riparia* ["*paludosa*" on label] (Cyperaceae). Fuckel, *Fungi Rhenani Exsiccati* no. 1772 (G, holotype; B, isotype).

This belongs to section *Fusispora*, with ascomata immersed, asci pyriform, ascospores 15-18 × 2.5-3.5 µm. Additional material studied (Brandenburg, on upper and lower surface of dead leaves of *Scirpus sylvaticus*, Vogel, distributed in Sydow, *Mycoteca Germanica* no. 3104, V 1936, L) belongs to section *Caterva*, and is morphologically indistinguishable from *M. superflua*, with asci cylindrical, ascospores 14-16 × 3.5-4 µm.

Mycosphaerella caricis (Dearn. & House) Petr. & Syd., *Ann. Mycol.* 22: 383. 1924 = *Laestadia caricis* Dearn. & House, *New York State Mus. Bull.* 205-206: 52. 1919 ["1918"] = *Guignardia caricis* (Dearn. & House) Dearn. & House, *New York State Mus. Bull.* 266: 73. 1925.

Type — USA: *Carex stricta* (Cyperaceae).

No material was studied, as the species was already redisposed in *Guignardia*.

Mycosphaerella carinthiaca Jaap, *Ann. Mycol.* 6: 210. 1908 = *Sphaerella carinthiaca* (Jaap) Sacc. & Trotter, *Syll. Fung.* 22: 128. 1913.— Fig. 157.

Type — Austria: On spots on lower surface of living leaves of *Trifolium medium* (Fabaceae).

Anamorph: *Ramularia trifolii* Jaap *vide* Jaap (op. cit.).

Authentic material studied (Germany, Eifel, Daun, Jaap, *Fungi Selecti Exsiccati* no. 374, VIII 1909, L) is a parasitic species of *Davidiella*, with asci pyriform, ascospores 11-14 × 3-4 µm. Therefore the following new combination is made: **Davidiella carinthiaca** (Jaap) Aptroot comb. nov., **MB 500342**. **Basionym:** *Mycosphaerella carinthiaca* Jaap, *Ann. Mycol.* 6: 210. 1908.

Sphaerella carlii Fuckel. See *Mycosphaerella oxalidis* (Rabenh.) Magnus.

Mycosphaerella carlinae (G. Winter) Feltgen, *Recueil Mém. Trav. Soc. Bot. Grand-Duché Luxembourg* 14: 286. 1899 = *Sphaerella carlinae* G. Winter, *Hedwigia* 10: 162. 1871.

Type — Germany: On dead stems and leaves of *Carlina vulgaris* (Asteraceae).

Anamorph and spermatial state: *Asteromella carlinae* Petr. and *Cercospora carlinae* Sacc. *vide* Tomilin (1979) (= *Passalora carlinae* (Sacc.) U. Braun & Crous).

Topotype material studied (Berlin, Gr̄unewald, Sydow no. 2829, XI 1889, B) is badly preserved. Additional material (Czech Republic, Weiβkirchen, Petrak, 1924, B) belongs to *Davidiella*, and is morphologically indistinguishable from *D. ammophilae*, with asci pyriform, ascospores 10-11 × 2.5-3 μm.

Mycosphaerella carniolica (Niessl) Lindau, Hilfsb. Sammeln Ascomyceten: 38. 1903 = *Sphaerella carniolica* Niessl, Oesterr. Bot. Z. 25: 85. 1875.— Fig. 158.

Type — Yugoslavia: On upper and lower surface of dead leaves of *Draba ciliata* (Brassicaceae). Niessl (M, holotype).

Already synonymised with *Davidiella allicina* (as *Mycosphaerella tassiana*) by von Arx (1949), with which the holotype agrees well, with asci pyriform, ascospores 16-19 × 6-7 μm.

Sphaerella carniolica var. *major* Niessl, Oesterr. Bot. Z. 25: 86. 1875.— Fig. 159.

Type — Austria: Dazeu. On upper and lower surface of dead leaves of *Draba aizoides* (Brassicaceae). Niessl (M, holotype).

The type and additional material studied (Steiermark, Lietzen, Niessl, M) belong to *Davidiella*, and shows that this is morphologically indistinguishable from *D. clandestina*, with asci pyriform, ascospores 21-24 × 6-8 μm.

Mycosphaerella caroliniana (F.A. Wolf) J.H. Miller, Mycologia 33: 79. 1941 = *Sphaerella caroliniana* F.A. Wolf, J. Elisha Mitchell Sci. Soc. 41: 95. 1925 = *Sphaerella oxydendri* F.A. Wolf, nomen herbariorum (not validly published, Article 32).— Fig. 160.

Type — USA: North Carolina, Raleigh. On upper and lower surface of dead leaves of *Oxydendrum arboreum* (Ericaceae). Wolf, III 1924 (BPI, holotype; NY, isotype, sub “*S. oxydendri*”).

Anamorph: *Phyllosticta* *vide* Wolf (op. cit.).

This is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores 7-10 × 2-2.5 μm

Mycosphaerella carphae (Speg.) Cash, Syll. Fung. 26: 332. 1972 = *Sphaerella carphae* Speg., Bol. Acad. Nac. Ci. 27: 360. 1924.— Fig. 161.

Type — Argentina: Tierra del Fuego, Bahia Sholl Bay. On upper and lower surface of dead leaves of *Carpha schoenoides* (Cyperaceae). Spegazzini no. 6190, I 1924 (LPS, holotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores 19-23 × 7-9 μm; the measurements in the protologue are based on immature parts of the type.

Sphaerella carpinea (Fr.) Cooke, J. Bot. 4: 248. 1866 = *Xyloma carpinea* Fr., Observ. Mycol. Fl. Suecicam 2: 363.

1818 [as “*carpini*”] = *Sphaeria carpinea* (Fr.) Fr., Systema Mycol. 2: 523. 1823 = *Ascospora carpinea* (Fr.) Fr., Summa Veg. Scand.: 425. 1849 = *Laestadia carpinea* (Fr.) Sacc., Syll. Fung. 1: 426. 1882 = *Guignardia carpinea* (Fr.) J. Schröt., in Cohn, Kryptog.-Fl. Schlesien 3(2): 330. 1894 [“1893”] = *Carlina carpinea* (Fr.) Kuntze & Bonord. in Kuntze, Revis. Gen. Pl. 2(3): 846. 1898 = *Apiosporopsis carpinea* (Fr.) Mariani, Atti Soc. Ital. Sci. Nat. 50: 165. 1911 = *Sphaerognomonia carpinea* (Fr.) Potebnia ex Höhn., Ber. Deutsch. Bot. Ges. 35: 634. 1917 = *Gnomonia carpinea* (Fr.) Kleb., Haupt- und Nebenfruchtformen der Askomyzeten: 274. 1918.

Type — Sweden: On upper and lower surface of dead leaves of *Carpinus betulus* (Betulaceae).

Accepted as *Sphaerognomonia carpinea* (Fr.) Potebnia ex Höhn. by Barr (1978). Material examined (Netherlands, on *Carpinus betulus*, Van de Sande Lacoste no. 159b, L) is in agreement with this disposition.

Mycosphaerella carpogena (Pass.) Tomilin, Novosti Sist. Nizsh. Rast. 1968: 164. 1968 = *Sphaerella carpogena* Pass., Atti Reale Accad. Lincei, Mem. Cl. Sci. Fis., ser. 4, 3: 5. 1887.

Type — Italy: *Liriodendron tulipifera* (Magnoliaceae).

No material could be studied as the type was not found in any of the herbaria consulted.

Mycosphaerella caryigena (Ellis & Everh.) Demaree & Cole, J. Agric. Res. 44: 145. 1932, based on an anamorph (illegitimate, Article 59) = *Cylindrosporium caryigenum* Ellis & Everh., Proc. Acad. Nat. Sci. Philadelphia 45: 168. II 1894 [“1893”, as “*caryogenum*”] = *Pseudocercospora caryigena* (Ellis & Everh.) Sivan., Bitunicate Ascomycetes and their anamorphs: 201. 1984.

Type — USA: *Hicorius amarus*. [“*Carya*”] (Juglandaceae). Material seen: USA: California, Albany. On *Hicorius pecan*. Demaree, VII 1938 (IMI no. 227618).

The material seen indeed contains only the anamorph, and suggests that the species is correctly classified as *Pseudocercospora caryigena*.

Mycosphaerella caryophyllata Boriquet & Heim, in Heim & Boriquet, Rev. Pathol. Vég. Entomol. Agric. France 26: 16. 1939, lacking Latin description (not validly published, Article 36).

Type — Madagascar: *Dianthus* [“*Caryophyllus*”] (Caryophyllaceae).

No material could be studied as the type was not included in a loan from PC.

Sphaerella caryophyllea Cooke & Harkn., in Cooke, Grevillea 14: 9. 1885 = *Laestadia caryophyllea* (Cooke & Harkn.) Berl. & Voglino, Syll. Fung., Addit. ad vol. 1-4: 62. 1886.— Fig. 162.

Type — USA: California, Placer Co., Roseville. On dead stems of *Dianthus* (Caryophyllaceae). Harkness no. 2183, II 1881 (BPI, holotype; BPI, isotype [“no. 2183a”]).

This is a species of *Botryosphaeria*, with asci pyriform, thick-walled, ascospores hyaline, simple, 14-17 × 4.5-5.5 μm.

Mycosphaerella caryophylli (Pass.) Cruchet, Bull. Soc. Vaud. Sci. Nat. 55: 44. 1923 ≡ *Sphaerella caryophylli* Pass., Atti Reale Accad. Lincei, Mem. Cl. Sci. Fis., ser. 4, 3: 5. 1887.

Type — Italy. On dead stems of *Dianthus caryophyllus* (Caryophyllaceae).

The type was not found in any of the herbaria consulted. Material studied (Germany, Hessen, Dillkreis, Donsbach, on *Petrorhagia saxifraga*, Ludwig, V 1934, B, 2×) is overmature.

Mycosphaerella casiozumi Hara, J. Pl. Protect. 5: 536. 1918. Type — Japan.

No material could be studied as the type was not included in a loan from TNS, and no material was found in BPI.

Mycosphaerella cassiae Syd., Leaflet Philipp. Bot. 9: 3120. 1925.

Type — Philippines: Luzon, Irosin, Sorsogon. On brown spots on upper surface of living leaves of *Cassia alata* (Fabaceae). Elmer, Philippine Islands Plants no. 15193, IX 1915 (BPI, isotype); also 1916 (L, topotype).

The isotype and topotype contain only a *Phoma*, on the lower surface of the spots.

Mycosphaerella cassiae F. Stevens. See *Mycosphaerella frauxii* M. Morelet.

Mycosphaerella cassiae Tilak, in T.S. Viswan. & Tilak, Sydowia 14: 311. 1960, later homonym (illegitimate, Article 53).

Type — India: *Cassia tora* (Fabaceae).

No material could be studied as the the location of the type is unknown and no material was found in BPI.

Mycosphaerella cassinopsidis (Kalchbr. & Cooke) Tomilin, Opredelitel' gribov roda *Mycosphaerella* Johans.: 139. 1979 ≡ *Sphaerella cassinopsidis* Kalchbr. & Cooke, Grevillea 9: 31. 1880 [as "*cassinopsis*"].— Fig. 163.

Type — South Africa: Cape of Good Hope, Somerset-East. On spots on upper surface of living leaves of *Cassinopsis ilicifolia* ["*capensis*"] (Icacinaceae). Owan, distributed in Rabenhorst-Winter, Fungi Europaei Exsiccati no. 3445 (L, isotype).

This is a parasitic species, with asci broadly cylindrical, ascospores 14-16 × 3-4 μm.

Mycosphaerella cassiopes M.E. Barr. See *Mycosphaerella inconspicua* Vesterg.

Sphaerella cassythae McAlpine, Proc. Linn. Soc. New South Wales 28: 102. 1903.

Type — Australia: *Cassytha glabella* (Lauraceae).

No material could be studied as the the location of the type is unknown and no material was found in any of the herbaria consulted.

Mycosphaerella castagnei (Har. & Briard) Jaap, Ann. Mycol. 14: 13. 1916 ≡ *Sphaerella castagnei* Har. & Briard, in Briard, Rev. Mycol. (Toulouse) 13: 18. 1891, nomen novum (Article 58) for *Sphaerella celtidis* Briard & Har., in

Briard, Rev. Mycol. (Toulouse) 12(48): 177. 1890, later homonym (illegitimate, Article 53).

Type — France: Montaud-les-Miramas. On lower surface of dead leaves of *Celtis australis* (Cannabaceae). Castagne, distributed in Vestergren, Micromycetes Rariores Selecti no. 1230 (BPI, isotype).

This is immature, but is morphologically indistinguishable from *M. punctiformis*. Additional material (Botanical Garden, XII 1888, Pass., distributed in Roumeguère, Fungi Selecti Exsiccati no. 5043, PC) agrees.

Sphaerella castaneae Tognini, Atti Ist. Bot. Univ. Pavia, ser. 2, 3: 52. 1893.

Type — Italy: *Castanea vesca* (Fagaceae).

No material could be studied as the the location of the type is unknown and no material was found in any of the herbaria consulted.

Mycosphaerella castanicola Kleb., Z. Pflanzenkrankh. Pflanzenschutz 44: 2. 1934.

Type — Germany: On spots on lower surface of living leaves of *Castanea vesca* (Fagaceae).

Anamorph: *Septoria castanicola* Desm. fide Tomilin (1979). No type material was found. Material examined (Italy, Toscane, Lizzano, van der Aa no. 8095, IX 1981, CBS) contains only an *Asteromella* anamorph.

Mycosphaerella castanopsidis (Dearn.) Petr., Ann. Mycol. 39: 311. 1941 ≡ *Dothidella castanopsidis* Dearn., Mycologia 16: 155. 1924.

Type — USA: Oregon, Lane Co., Star. Breaking through the leaf cuticle on brown spots on the upper surface of living leaves of *Castanopsis chrysophylla* (Fagaceae). Boyce, VI 1921 (IMI no. 215780, isotype); also VI 1923 (IMI no. 26789, topotype).

Cited as synonymous with *Mycosphaerella janus* by Barr (1972). This is indeed a *Dothidella* species, morphologically indistinguishable from *Dothidella janus* (Berk. & M.A. Curtis) Höhn., with hamathecium consisting of up to 4 μm wide pseudoparaphyses, asci cylindrical, ascospores 20-25 × 3.5-4.5 μm.

Mycosphaerella castillae Stev. & A.M.J. Watson, in Stev., Mycologia 35: 634. 1943 [as "*castilloae*"].— Fig. 164.

Type — Costa Rica. On white spots with brown margins on upper surface of living leaves of *Castilla* ["*Castilloa*"] *costaricana* (Moraceae). Lorenz no. 3071, IX 1940 (NY, isotype).

This is a parasitic species, with asci cylindrical, thick-walled, ascospores 20-26 × 5-7 μm.

Mycosphaerella castillejae Murashk., Trudy Omsk. Selskokh. Inst. 3: 125. 1924 [as "*castilleya*"].— Fig. 165.

Type — Russia: *Castilleja pallida* (Orobanchaceae).

No material could be studied as the type was not included in loans from LE and LEP.

Mycosphaerella catesbeyi (Cooke) J.H. Miller, Mycologia 33: 80. 1941 ≡ *Sphaerella catesbeyi* Cooke, Grevillea 7: 53. 1878.

Type — USA: South Carolina, Aiken. On upper and lower surface of dead leaves of *Quercus catesbeysi* (Fagaceae). Ravenel no. 2046 (K, holotype); also distributed in Fungi Americani Exsiccati no. 383 (K, 3 isotypes).

This belongs to section *Mycosphaerella* s.s., with asci cylindrical, ascospores $12-14 \times 3.5-5 \mu\text{m}$.

Mycosphaerella cattleyae Cash & A.M.J. Watson, Mycologia 47: 732. 1955.— Fig. 166.

Type — Panama: Canal Zone, Balboa; intercepted in Texas, Laredo. On white spots with brown margins on upper surface of living leaves of *Cattleya* (Orchidaceae). Leary, VI 1945 (BPI, holotype).

This is a species of *Planistromella*, with asci cylindrical, surrounded by a parenchymatous tissue of 4-10 μm wide cells, ascospores $13.5-15 \times 3-3.5 \mu\text{m}$. Therefore the following new combination is proposed here:

Planistromella cattleyae (Cash & A.M.J. Watson) Aptroot comb. nov., **MB 500371**. **Basionym:** *Mycosphaerella cattleyae* Cash & A.M.J. Watson, Mycologia 47: 732. 1955.

Mycosphaerella caulicola (P. Karst.) Lindau, Hilfsb. Sammeln Ascomyceten: 39. 1903 \equiv *Sphaerella caulicola* P. Karst., Bidrag Kännedom Finlands Natur Folk 23: 169. 1873.— Fig. 167.

Type — Russia: Kola. On *Galeopsis versicolor* (Lamiaceae). Karsten no. 3672, VII 1861 (H, holotype).

The type belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradians*, with asci cylindrical, ascospores $10-12 \times 3-3.5 \mu\text{m}$. Additional material, which is cited in the original publication as probably belonging to this species (Finland, on *Chamaenerion* ["*Epilobium*"] *angustifolium* (Onagraceae), Karsten, H) and other material examined (Czech Republic, Hranice, Podhorn, Petrak, on dead stems of *Galeopsis*, distributed in Reliquiae Petrakianae no. 1851, VI 1932, L) belong to *Davidiella*, and are morphologically indistinguishable from *D. ammophilae*, with asci small, pyriform, ascospores $10-11 \times 3-3.5 \mu\text{m}$.

Mycosphaerella cecropiae Bat., J.L. Bezerra & Matta, Publ. Inst. Micol. Univ. Fed. Pernambuco 309: 7. 1961.— Fig. 168.

Type — Brazil: On brown spots with black margins on upper surface of living leaves of *Cecropia peltata* (Urticaceae). Batista exs. no. 14818 (URM 20640, holotype).

Spermatial state: *Asteromella fide* Batista, Bezerra & Matta (op. cit.).

This belongs to *Davidiella*, of which it represents a parasitic species, with asci pyriform, ascospores $15-19 \times 3-4.5 \mu\text{m}$. Therefore the following new combination is made:

Davidiella cecropiae (Bat., J.L. Bezerra & Matta) Aptroot comb. nov., **MB 500343**. **Basionym:** *Mycosphaerella cecropiae* Bat., J.L. Bezerra & Matta, Publ. Inst. Micol. Univ. Fed. Pernambuco 309: 7. 1961.

Mycosphaerella cecropiae var. *macrocarpa* Bat., J.L. Bezerra & Matta, Publ. Inst. Micol. Univ. Fed. Pernambuco 309: 10. 1961.— Fig. 169.

Type — Brazil: On lower surface of the leaves, below white spots on the upper surface of living leaves of *Mouriri pusa*

(Melastomataceae). Batista exs. no. 14819 (URM 20641, holotype).

This belongs to *Davidiella*, of which it represents the parasitic species *D. cecropiae*, with asci pyriform, ascospores $15-19 \times 3-4.5 \mu\text{m}$; it is morphologically indistinguishable from the nominal variety (see above).

Mycosphaerella cedrelae (Speg.) M. Morelet, Ann. Soc. Sci. Nat. Archéol. Toulon & Var 20: 105. 1968 \equiv *Sphaerella cedrelae* Speg., Anales Mus. Nac. Hist. Nat. Buenos Aires 31: 409. 1922.— Fig. 170.

Type — Paraguay: Asunción. On white spots with brown rims on upper surface of dead leaves of *Cedrela fissilis* (Meliaceae). Spegazzini no. 6297, IX 1919 (LPS, holotype).

This is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores $9-11 \times 2.5-3 \mu\text{m}$.

Mycosphaerella celtidis (Pass.) Tomilin, Novosti Sist. Nizsh. Rast. 1968: 164. 1968 \equiv *Sphaerella celtidis* Pass., in Roum., Rev. Mycol. (Toulouse) 11: 196. 1889.

Type — Italy: On lower surface of dead leaves of *Celtis australis* (Cannabaceae). Passerini, distributed in Roumeguère, Fungi Selecti Exsiccati (PC, holotype).

The holotype contains only empty ascomata.

Sphaerella celtidis Briard & Har. See *Mycosphaerella castagnei* (Har. & Briard) Jaap.

Mycosphaerella centellae Petr., Ann. Mycol. 22: 39. 1924.— Fig. 171.

Type — Indonesia: Celebes, Madjune, Polenati. On brown spots on lower surface of living leaves of *Hydrocotyle* ["*Centella*"] *asiatica* (Araliaceae). Gäumann, III 1920 (W, holotype).

This is a parasitic species, with asci cylindrical, ascospores $13-16 \times 3-4.5 \mu\text{m}$. It is morphologically indistinguishable from *Mycosphaerella hydrocotyles-asiaticae*.

Mycosphaerella centellae-asiaticae (Pat.) Petr. See *Mycosphaerella hydrocotyles-asiaticae* (Pat.) Petr.

Sphaerella cephalariae Auersw., in Gonn. & Rabenh., Mycol. Europaea 5-6: 14. 1869 \equiv *Venturia cephalariae* (Auersw.) Kalchbr. & Cooke, Grevillea 9: 31. 1880 \equiv *Laestadia cephalariae* (Auersw.) Sacc., Syll. Fung. 1: 425. 1882.

Type — Germany: *Cephalaria alpina* (Dipsacaceae). Accepted as *Venturia cephalariae* (Auersw.) Kalchbr. & Cooke by Sivanesan (1977).

Mycosphaerella cepharanthae Sawada, Taiwan Univ. Coll. Agric. Special Publ. 8: 61. 1959.— Fig. 172.

Type — Taiwan: Taipeh. On brown spots on upper and lower surface of living leaves of *Stephania cepharantha* (Menispermaceae). Sawada, X 1949 (BPI, isotype).

This is a parasitic species of *Davidiella*, with asci pyriform, ascospores $8.5-10.5 \times 2.5-3 \mu\text{m}$. Therefore the following new combination is made: **Davidiella cepharanthae** (Sawada) Aptroot comb. nov., **MB 500344**. **Basionym:**

Mycosphaerella cepharanthae Sawada, Taiwan Univ. Coll. Agric. Special Publ. 8: 61. 1959.

Mycosphaerella cerasella Aderh., Ber. Deutsch. Bot. Ges. 18: 246. 1900 = *Sphaerella cerasella* (Aderh.) Sacc. & P. Syd., Syll. Fung. 16: 469. 1902.— Fig. 173.

Type — Germany: *Prunus cerasus* (Rosaceae).

Associated with *Cercospora cerasella* Sacc. *vide* Aderhold (op. cit.) (= *Passalora circumscissa* (Sacc.) U. Braun).

The location of the type is unknown; it may have been destroyed in B. Material examined (Netherlands, Eemnes-Buiten, on upper and lower surface of dead leaves of *Prunus virginica*, von Arx, V 1949, CBS) belongs to section *Longispora*, and is morphologically indistinguishable from *M. millegrana*, with asci cylindrical, ascospores 16-20 × 2.5-3.5 µm.

Mycosphaerella cerasicola (Pass.) Koshk., Mikromitsety yuzhnogo Turkmenistana: 67. 1977 = *Sphaerella cerasicola* Pass., Atti Reale Accad. Lincei, Mem. Cl. Sci. Fis., ser. 4, 3: 6. 1887.

Type — Italy: *Prunus cerasus* (Rosaceae).

Anamorph: Associated with *Coniothyrium cerasi* Pass. *vide* Passerini (op. cit.).

No material was studied as the type was not found in any of the herbaria consulted.

Mycosphaerella cerasina (Cooke) Feltgen, Recueil Mém. Trav. Soc. Bot. Grand-Duché Luxembourg 15: 257. 1903 = *Sphaerella cerasina* Cooke, Grevillea 7: 54. 1878.— Fig. 174.

Type — USA: South Carolina, Aiken. On upper and lower surface of dead leaves of *Prunus* ["*Cerasus*"] *laurocerasus* (Rosaceae). Ravenel, Fungi Americani Exsiccati no. 380 (K, holotype; K, 4 isotypes), also distributed in Ellis, North American Fungi no. 794 (K, isotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with ascospores 14-18 × 4-5.5 µm, but mature asci have not been seen. Additional material examined (Germany, Brandenburg, Strausberg, Sydow, Mycotheca Germanica no. 3503, V 1941, L) belongs to *M. punctiformis*, with asci cylindrical, ascospores 9-10 × 3-3.5 µm.

Sphaerella cerasina subsp. *padina* P. Karst. See *Mycosphaerella padina* (P. Karst.) Jørst.

Mycosphaerella cerastii (Rabenh.) Magnus. See *Mycosphaerella stellarinearum* (Rabenh.) Johanson.

Sphaerella ceratoniae Pass., Funghi Parmensis III: 28. 1882.— Fig. 176.

Type — Italy: Emergent through the epidermis on the upper surface of dead leaves of *Ceratonia siliqua* (Fabaceae).

The type was not found in any of the herbaria consulted. Material studied (Spain, Mallorca, El Arenal, van der Aa no. 1701, V 1969, CBS) belongs to section *Longispora*, and is morphologically indistinguishable from *M. millegrana*, with asci cylindrical to narrowly pyriform, ascospores 12-15 × 2-3.5 µm.

Mycosphaerella cercidicola (Ellis & Kellerm.) F.A. Wolf, Mycologia 32: 135. 1940 = *Sphaerella cercidicola* Ellis & Kellerm., Bull. Torrey Bot. Club 11: 123. 1884.— Fig. 178.

Type — USA: Kansas, Manhattan. On upper and lower surface of dead leaves of *Cercis canadensis* (Fabaceae). Kellerman no. 550, VI 1884 (NY, holotype; NY, isotype).

Anamorph: *Cercospora cercidicola* Ellis *vide* Sivanesan (1984) (= *Passalora cercidicola* (Ellis) U. Braun).

This belongs to section *Caterva* and is morphologically indistinguishable from *M. subradicans*, with asci cylindrical, ascospores 10-12 × 2.5-3.5 µm. Additional material studied (USA, North Carolina, Durham, Wolf no. 71296, V 1939, B, IMI no. 127139) probably belongs to *M. punctiformis*, with asci cylindrical, ascospores small.

Mycosphaerella cercidis (Pass.) Tomilin, Novosti Sist. Nizsh. Rast. 1968: 164. 1968 = *Sphaerella cercidis* Pass., Erbario Crittogamico Italiano, ser. 2 no. 1463. 1885.— Fig. 179.

Type — Italy: Parma, Botanical Garden. On upper and lower surface of dead leaves of *Cercis japonica* (Fabaceae). Passerini, IV 1884, distributed in Erbario Crittogamico Italiano, ser. 2 no. 1463 (NY, isotype).

Anamorph: *Septoria cercidis* Fr. *vide* Tomilin (1979).

This is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores 8-10 × 2-3 µm.

Mycosphaerella cerei Henn., Hedwigia 43: 85. 1904 = *Sphaerella cerei* (Henn.) Sacc. & D. Sacc., Syll. Fung. 17: 639. 1905.

Type — Brazil: *Cereus macrogonus* (Cactaceae).

No material could be studied as the type may have been destroyed in B, and no material was found in SP.

Mycosphaerella ceres (Sacc.) Siemaszko, Mater. Mikol. Fitopatol. Rossii 1: 25. 1915 = *Sphaerella ceres* Sacc., Nuovo Giorn. Bot. Ital. 7: 302. 1875.— Fig. 177.

Type — Italy: *Sorghum vulgare* (Poaceae). Saccardo (IMI no. 143008, isotype, slide only).

Anamorph: *Ascochyta sorghi* Sacc. *vide* Saccardo (1882).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. disseminata*, with asci pyriform, ascospores 13-16 × 5-7 µm. Additional material (Herb. Mycol. Cooke, 1885, IMI no. 146996, slide only) agrees.

Sphaerella ceriospora (Duby) Ces. & De Not., Comment. Soc. Crittog. Ital. 1(4): 237. 1863 = *Sphaeria ceriospora* Duby, in Rabenh., Herb. Vivum Mycol., cent. 20 no. 1937.

1855 = *Ceriospora dubyi* Niessl, Verh. Naturf. Vereins Brünn 14: 169. 1876 = *Hindersonia ceriospora* (Duby) J. Schröt., in Cohn, Kryptog.-Fl. Schlesien 3(2): 393. 1894 ["1893"].

Type — Switzerland: On dead stems of *Humulus lupulus* (Urticaceae).

Accepted as *Ceriospora dubyi* Niessl by Müller & von Arx (1962). Material studied (Germany, Voitsberg, Niessl, distributed in Rabenhorst, Fungi Europaei Exsiccati no. 1560, L) contains only a coelomycete.

Mycosphaerella cesatiana (Speg.) Tomilin, Novosti Sist. Nizsh. Rast. 1967: 188. 1967 = *Sphaerella cesatiana* Speg., in Traverso, Fl. Ital. Cryptog. 1(11): 613. 1913.— Fig. 180. Type — Italy. On lower surface of dead leaves of *Alnus glutinosa* (Betulaceae). Spegazzini, V 1877 (PAD, holotype).

Already cited as synonymous with *Mycosphaerella alnicola* by Tomilin (1979). The type shows that this belongs, just like *M. alnicola*, to section *Longissima*, and is morphologically indistinguishable from *M. latebrosa*, with asci cylindrical, ascospores 20-25 × 4-4.5 µm, although most material is overmature.

Sphaerella chalcographa Kalchbr., Math. Természettud. Közlem. 7: 258. 1868.

Type — Hungary: *Glyceria spectabilis* (Poaceae).

No material could be studied as the type may have been destroyed in B.

Mycosphaerella chamaemori (P. Karst.) Lindau, in Engl. & Prantl, Natürlichem Pflanzenf. 1(1): 424. 1897 = *Sphaerella chamaemori* P. Karst., Fungi Fennici Exsiccati no. 899. 1869 = *Venturia chamaemori* (P. Karst.) Arx, Acta Bot. Neerl. 6: 340. 1957.

Type — Finland: *Rubus chamaemorus* (Rosaceae).

Accepted as *Venturia chamaemori* (P. Karst.) Arx by Sivanesan (1977).

Mycosphaerella chamaemori L. Holm & K. Holm. See *Mycosphaerella holmii* O.E. Erikss.

Mycosphaerella chamaenerii Savile, Canad. J. Bot. 40: 1388. 1962.— Fig. 181.

Type — Canada: Hazard Inlet, Somerset Island, Franklin. On dead leaves of *Chamaenerion* ["*Epilobium*"] *latifolium* (Onagraceae). Savile no. 3590B, VII 1958, ex. Herb. DAOM no. 84792 (IMI no. 226862, isotype, slide only).

Anamorph: *Ramularia chamaenerii* Rostr. *fide* Savile (op. cit.).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores 18-21 × 6-8 µm.

Mycosphaerella chamaeropsis (Traverso) Tomilin, Opredelitel' gribov roda *Mycosphaerella* Johans.: 56. 1979 = *Sphaerella chamaeropsis* Traverso, Malpighia 14: 464. 1900.— Fig. 182.

Type — Italy: Candanabbia. On brown spots with black margins on upper and lower surface of living leaves of *Chamaerops humilis* (Arecaceae). Traverso, distributed in Pollacci, Fungi Longobardiae Exsiccati no. 320 (BPI, isotype, as "*Sphaerella*"); also Ciferri (BPI, possible isotype).

Anamorph: Associated with *Diplodia passeriniana* Thüm. *fide* Saccardo & Sydow (1901).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores 19-23 × 4-6 µm. The *Diplodia* is also present.

Mycosphaerella chardonii (Gonz. Frag. & Cif.) Cif., Quaderno 19: 231. 1961 = *Sphaerella chardonii* Gonz. Frag. & Cif., Estac. Agron. Moca, ser. Bot. no. 11: 17. 1928.

Type — Dominican Republic: Diego de Ocampo. On white spots with black margins on upper surface of living leaves of *Guarea trichilioides* (Meliaceae). Ciferri no. 2490, VII 1929 (BPI, 2 topotypes).

All materials studied are immature.

Mycosphaerella chaubattiensis S.K. Bose & A.J. Roy, Progr. Hort. 1: 59. 1970.

Type — India: *Pyrus communis* (Rosaceae).

No material could be studied as the location of the type is unknown.

Mycosphaerella chelidonii (Fautrey & Lambotte) Guyot, Bull. Soc. Mycol. France 62: 82. 1946 = *Sphaerella chelidonii* Fautrey & Lambotte, Rev. Mycol. (Toulouse) 17: 170. 1895.— Fig. 183.

Type — France: On stems of *Chelidonium majus* (Papaveraceae). Fautrey, distributed in Roumeguère, Fungi Selecti Exsiccati no. 6883, V 1895 (PC, holotype).

This belongs to section *Caterva* and is morphologically indistinguishable from *M. subradicans*, with asci cylindrical, ascospores 10-12 × 3-3.5 µm. Additional material studied (Sweden, Södermanland, Nacka, Nackanäs, Romell, VII 1890, B) agrees.

Mycosphaerella chenopodii Dearn. & Barthol., in Dearn., Mycologia 16: 157. 1924.

Type — USA: *Chenopodium leptophyllum* (Amaranthaceae).

No material was found in any of the herbaria consulted.

Mycosphaerella chenopodiicola (Speg.) Tomilin, Novosti Sist. Nizsh. Rast. 1968: 164. 1968 = *Sphaerella chenopodiicola* Speg., Anales Mus. Nac. Hist. Nat. Buenos Aires 19 (ser. 3, 12): 351. 1909 [as "*chaenopodiicola*"] = *Paradidymella chenopodiicola* (Speg.) Tomilin, Novosti Sist. Nizsh. Rast. 9: 119. 1972.

Type — Argentina: *Chenopodium hircinum* (Amaranthaceae).

The type was not included in a loan from LPS.

Mycosphaerella chimaphilae (Ellis & Everh.) Höhn., Sitzungsber. Kaiserl. Akad. Wiss., Math.-Naturwiss. Kl., Abt. 1, 126: 355. 1917 = *Sphaerella chimaphilae* Ellis & Everh., Proc. Acad. Nat. Sci. Philadelphia 45: 447. II 1894 ["1893"].— Fig. 184.

Type — USA: Delaware, Faulkland. On upper and lower surface of dead, but still attached leaves of *Chimaphila umbellata* (Ericaceae). Commons no. 479, V 1887 (NY, holotype).

This belongs to section *Caterva* and is morphologically indistinguishable from *M. subradicans*, with asci cylindrical, ascospores 11-13 × 2.5-3.5 µm.

Sphaerella chimaphilae Peck. See *Mycosphaerella chimaphilina* (Peck) House.

Mycosphaerella chimaphilina (Peck) House, New York State Mus. Bull. 233-234: 26. 1921 = *Sphaerella chimaphilina* Peck, in Sacc., Syll. Fung. 11: 297. 1895, nomen novum (Article 58) for *Sphaerella chimaphilae* Peck, Annual Rep. New York State Mus. 47: 150. XII 1894, later homonym (illegitimate, Article 53).— Fig. 185.

Type — USA: *Chimaphila umbellata* (Ericaceae).

Cited as synonymous with *Mycosphaerella pyrolae* by Tomilin (1979).

The type was not found in any of the herbaria consulted. Additional material seen (Canada, Ontario, Temagami Island, on dead, blackened spots on upper surface of living leaves of *Chimaphila umbellata* (Ericaceae), Jackson, University of Toronto, Cryptogamic Herbarium no. 4250, VI 1932, IMI no. 26958) is *M. punctiformis*, with asci cylindrical, ascospores $10-11 \times 2.5-3 \mu\text{m}$.

Sphaerella chionanthi (Berk. & Broome) Cooke, J. Bot. 21: 107. 1883 = *Sphaeria chionanthi* Berk. & Broome, J. Linn. Soc., Bot. 14: 128. 1875.

Type — Sri Lanka: *Chionanthus zeylanica* (Oleaceae).

No material was studied as the type was not found in K nor in any of the other herbaria consulted.

Sphaerella chlorina Sacc. & Trab., in Trab., Étude sur l'Halifa: 48. 1889 = *Mycosphaerella stipicola* Tomilin, Novosti Sist. Nizsh. Rast. 1966: 173. 1966, superfluous (illegitimate, Article 52) nomen novum (Article 58).

Type — Algeria: *Stipa tenacissima* (Poaceae). Muheria (PAD, holotype).

Spermatial state: *Asteroma fide* Saccardo & Trabut (op. cit.). Cited as synonymous with *M. recutita*, which is morphologically indistinguishable from *Davidiella disseminata*, by Tomilin (1979). The type contains only a coelomycete, which is in accordance with an illustration by Saccardo on the original material.

Mycosphaerella chlorogali Fairm., in Millsp. & L. Nutt., Publ. 212. Field Columbian Mus., Bot. Ser. 5: 348. 1923 [as "*chlorogalli*"].— Fig. 186.

Type — USA: On dead stems of *Chlorogalum pomeridianum* (Alliaceae).

The type was not found in any of the herbaria consulted. Material seen (USA, California, Los Angeles, Plunkett, Southern Californian Fungi, IMI no. 154430) belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores $13-15 \times 4.5-6 \mu\text{m}$.

Sphaerella chlorospila Sacc., Malpighia 13: 438. 1900 ["1899"].

Type — Italy: Riva-Valdobbia. On upper and lower surface of dead leaves of *Gentiana purpurea* (Gentianaceae). Carestia, IX 1898 (PAD, holotype).

The type contains only immature ascomata of an unidentifiable fungus.

Sphaerella chlorospora (Ces.) Ces. & De Not., Comment. Soc. Crittog. Ital. 1(4): 237. 1863 = *Sphaeria chlorospora* Ces., in Rabenh., Fungi Europaei Exsiccati, ed. nov., ser. 2 no. 48. 1859 = *Venturia chlorospora* (Ces.) P. Karst.,

Bidrag Kännedom Finlands Natur Folk 23: 189. 1873 = *Endostigme chlorospora* (Ces.) Syd., Ann. Mycol. 21: 173. 1923.

Type — Italy: *Salix* (Salicaceae).

Anamorph: *Fusicladium fide* Nüesch (1960).

Accepted as *Venturia chlorospora* (Ces.) P. Karst. by Sivanesan (1977).

Mycosphaerella chlouna (Cooke) Lindau, in Engl. & Prantl, Natürlichen Pflanzenf. 1(1): 426. 1897 [as "*chlorina*"] = *Sphaerella chlouna* Cooke, Grevillea 5: 121. 1877.

Type — United Kingdom: *Phalaris arundinacea* (Poaceae). (PAD, isotype).

Cited as synonymous with *M. recutita*, which is morphologically indistinguishable from *Davidiella disseminata*, by Tomilin (1979). The isotype contains a coelomycete with 1-septate, hyaline conidia, which could well have been confused with an ascomycete.

Sphaerella chondri H.L. Jones, Oberlin College Lab. Bull. 9: 3. 1898.

Type — USA: Massachusetts, Nahauf. In thallus of *Chondrus crispus* (Rhodophyta, Gigartinales). Jones, II 1894 (BPI, isotype).

This is morphologically indistinguishable from *Lautitia danica* (Berl.) S. Schatz according to J. & E. Kohlmeyer (1979). The isotype agrees, with ascomata fully immersed, partly in groups, asci bitunicate, with paraphysoids, no ascospores were observed.

Sphaerella chondrospora (Ces.) Ces. & De Not., Comment. Soc. Crittog. Ital. 1(4): 237. 1863 = *Sphaeria chondrospora* Ces., in Rabenh., Herb. Vivum Mycol. no. 1932b. 1855 = *Cryptospora chondrospora* (Ces.) Rehm, Ber. Naturhist. Vereins Augsburg 26: 83. 1881 = *Cryptosporella chondrospora* (Ces.) Sacc., Syll. Fung. 1: 467. 1882 = *Pseudomassaria chondrospora* (Ces.) Jacz., Bull. Herb. Boissier 2: 663. 1894 = *Aplacodina chondrospora* (Ces.) Ruhland, Hedwigia 39: 38. 1900 = *Spegazzinula chondrospora* (Ces.) Höhn., Ann. Mycol. 2: 42. 1904 = *Apiospora chondrospora* (Ces.) Sacc. & D. Sacc., Syll. Fung. 17: 658. 1905.

Type — Germany: *Tilia* (Malvaceae).

Accepted as *Pseudomassaria chondrospora* (Ces.) Jacz. by Müller & von Arx (1962).

Mycosphaerella chorinensis (Kirschst.) Tomilin, Novosti Sist. Nizsh. Rast. 1968: 164. 1968 = *Sphaerella chorinensis* Kirschst., Kryptog.-Fl. Mark Brandenburg 7(3): 382. 1938.— Fig. 187.

Type — Germany: Oberbarnim, Choriner Forst. *Rumex acetosa* (Polygonaceae). Kirschstein, VI 1917 (B, holotype). Cited as synonymous with *Mycosphaerella leptasca* by Tomilin (1979).

Mycosphaerella chrysanthemi (Tassi) Tomilin, Opredelitel' gribov roda *Mycosphaerella* Johans.: 60. 1979 = *Sphaerella chrysanthemi* Tassi, Bull. Lab. Orto Bot. Siena 3: 117. 1900 = *Didymella chrysanthemi* (Tassi) Garib. & Gullino, L'Agricoltura Ital. 71: 286. 1971.

Type — Italy: Siena, Botanical Garden. On upper surface of dead leaves of *Chrysanthemum marginatum* (Asteraceae). Tassi, 1901 (SIENA, holotype).

Anamorph: *Ascochyta chrysanthemi* F. Stevens *vide* Sivanesan (1984).

This is morphologically indistinguishable from *Pleospora herbarum* (Pers. : Fr.) Rabenh., with ascospores brown, muriform, 20-25 × 10-14 µm; the material is partly immature, and that part may have led to the description in (*Myco*-)*Sphaerella*.

Mycosphaerella chrysobalani Miles, Trans. Illinois State Acad. Sci. 10: 252. 1917 = *Sphaerella chrysobalani* (Miles) Trotter, Syll. Fung. 24: 882. 1928.— Fig. 188.

Type — Puerto Rico: Rio Piedras. On white spots with black margins on upper surface of living leaves of *Chrysobalanus icaco* (Chrysobalanaceae). Stevens no. 5699, XI 1913 (BPI, 2 isotypes).

This is a parasitic species of *Davidiella*, with asci pyriform, ascospores thick-walled, ends rounded, 18-23 × 3.5-4.5 µm. Therefore the following new combination is made: **Davidiella chrysobalani** (Miles) Aptroot comb. nov., **MB 500345**. **Basionym:** *Mycosphaerella chrysobalani* Miles, Trans. Illinois State Acad. Sci. 10: 252. 1917. Additional material (Aibonito, Fink no. 1735, I 1916, B, 2×) agrees.

Mycosphaerella chrysobalanicola Petr. & Cif., Ann. Mycol. 8: 389-390. 1930.

Type — Dominican Republic: *Chrysobalanus icaco* (Chrysobalanaceae).

No material could be studied, as the type was not present in W or BPI, or in any of the other herbaria consulted.

Sphaerella chusqueicola Speg., Revista Fac. Agron. Vet. Univ. La Plata, ep. 2, 6: 57. 1910.

Type — Chile: Cerro Caracol de Concepción. On living leaves of *Chusquea cumingii* (Poaceae). Spegazzini, I 1909 (LPS, holotype).

The type contains only a stromatic coelomycete, suggesting a Phyllachoraceae anamorph.

Sphaerella ciconii Losa, Collect. Bot. 2: 217. 1949.

Type — Spain: *Erodium ciconium* (Geraniaceae).

No material could be studied, as the type was not present in MA or in any of the herbaria consulted.

Sphaerella cicutae Kirchn., in Thüm., Fungi Austr., cent. 10 no. 964. 1872 = *Laestadia cicutae* (Kirchn.) Sacc., Syll. Fung. 1: 430. 1882.

Type — Czech Republic: Bohemia, Plöckensteiner See. On dead stems of *Cicuta virosa* (Apiaceae). Kirchner, 1872, distributed in Thümen, Fungi Austriaca no. 964 (NY, isotype).

The isotype contains a *Septoria* coelomycete and a postmature ascomycete; not likely to be a *Mycosphaerella*, as the ascospores are simple according to the protologue.

Mycosphaerella ciliata (Ellis & Everh.) House, New York State Mus. Bull. 233-234: 26. 1921 = *Sphaerella ciliata* Ellis & Everh., Proc. Acad. Nat. Sci. Philadelphia 42: 231. 1890.— Fig. 189.

Type — Canada: London. On dead stems of *Steironema ciliatum* (Myrsinaceae). Dearness no. 1640 [“1650”], IV 1890 (NY, holotype).

This belongs to section *Caterva* and is morphologically indistinguishable from *M. subradians*, with asci cylindrical, ascospores 10-12 × 3-3.5 µm.

Mycosphaerella cinerascens (Fleischh.) Vesterg., Bot. Not. 1897: 267. 1897 = *Sphaerella cinerascens* Fleischh., in Rabenh., Fungi Europaei Exsiccati, ed. nov., ser. 2 no. 845. 1865, nomen novum (Article 58) for *Sphaeria cinerascens* Fuckel, Fungi Rhenani Exsiccati no. 824. 1863, later homonym (illegitimate, Article 53) of *Sphaeria cinerascens* Schwein., Trans. Amer. Philos. Soc. (Philadelphia), ser. 2, 4: 225. 1832 = *Endostigme cinerascens* (Fleischh.) Jørst., Nytt Mag. Naturvidensk. 84: 252. 1944 = *Spilosticta cinerascens* (Fleischh.) Petr., Sydowia 1: 197. 1947.— Fig. 190.

Type — Germany: Zange. On lower surface of dead leaves of *Sorbus aria* (Rosaceae). Fuckel, Fungi Rhenani Exsiccati no. 824 (L, isotype).

Anamorph: *Cercospora ariae* Fuckel *vide* Fuckel (op. cit.).

Cited as synonymous with *Venturia inaequalis* (Cooke) G. Winter by Sivanesan (1977), with which the type agrees well, with asci cylindrical, ascospores greenish brown, asymmetrically septate, 12-15 × 5-6 µm.

Sphaerella cinereo-nebulosa (Desm.) Sacc., Syll. Fung. 1: 538. 1882 = *Sphaeria cinereo-nebulosa* Desm., Bull. Soc. Bot. France 4: 816. 1857.

Type — France: On upper and lower surface of dead leaves of *Phalaris arundinacea* (Poaceae). Desmazières, Plantes Cryptogames de France no. 370 (B-Desmazières, isotype).

The isotype contains only the coelomycete *Phaeoseptoria*, with conidia 3-septate, brown, 16-22 × 3.5-4.5 µm.

Mycosphaerella cinnafolia R. Sprague, Mycologia 50: 814. 1959 [“1958”].

Type — USA: *Cinna latifolia* (Poaceae).

No material was studied as the type was not preserved in FH, nor found in any of the herbaria consulted.

Mycosphaerella cinnamomicola (Gonz. Frag. & Cif.) Cif., Quaderno 19: 231. 1961 = *Sphaerella cinnamomicola* Gonz. Frag. & Cif., in Cif. & Gonz. Frag., Bol. Soc. Esp. Hist. Nat. 27: 271. 1927.

Type — Dominican Republic: Moca. On pale spots with brown margins on upper surface of living leaf tips of *Cinnamomum zeylanicum* (Lauraceae). Ciferri, X 1926 (B, BPI, isotypes).

The isotypes contain only a coelomycete.

Mycosphaerella cinxia (Sacc.) Lindau, in Engl. & Prantl, Natürlichen Pflanzenf. 1(1): 425. 1897 = *Sphaerella cinxia* Sacc., Nuovo Giorn. Bot. Ital. 7: 303. 1875.

Type — Italy: *Lilium candidum* (Liliaceae).

No material was studied as the type was not found in any of the herbaria consulted.

Mycosphaerella circe (Sacc.) Tomilin, Novosti Sist. Nizsh. Rast. 1968: 164. 1968 = *Sphaerella circe* Sacc., Nuovo Giorn. Bot. Ital. 7: 301. 1875.

Type — Italy. On white spots on upper surface of living leaves of *Phytolacca decandra* (Phytolaccaceae). Saccardo, IX (PAD, holotype).

The type contains only a coelomycete.

Mycosphaerella circumdans (Pass.) Tomilin, Novosti Sist. Nizsh. Rast. 1968: 164. 1968 ≡ *Sphaerella circumdans* Pass., Nuovo Giorn. Bot. Ital. 7: 256. 1875.— Fig. 191.

Type — Italy: Parma. On upper surface of dead leaves of *Platanus occidentalis* (Platanaceae). Passerini (PAD, holotype).

This belongs to *Davidiella*, and shows that this is morphologically indistinguishable from *D. clandestina*, with asci pyriform, ascospores 21-25 × 7-8 μm. The original description was based on immature spores, which are abundantly present in the type as well.

Sphaerella circumscissa G. Winter, Bol. Soc. Brot. 4: 13. 1886.

Type — South Africa: *Aristolochia* (Aristolochiaceae).

No material was studied as the type was probably destroyed in B and no material was found in any of the herbaria consulted.

Mycosphaerella circumvaga (Desm.) Vesterg., Bot. Not. 1902: 125. 1902 ≡ *Dothidea circumvaga* Desm., Ann. Sci. Nat. Bot., ser. 3, 8: 175. 1847 ≡ *Sphaerella circumvaga* (Desm.) Sacc., Michelia 2: 56. 1880.— Fig. 192.

Type — France: On on dead stems of *Medicago falcata* (Fabaceae). Desmazières, Plantes Cryptogames de France no. 88 (B-Desmazières, isotype).

The isotype studied is immature. Additional material examined (Germany, Thüringen, Sondershausen, Oertel, distributed in Sydow, Mycotheca Germanica no. 235, III 1904, L) belongs to *Davidiella*, and shows that this is morphologically indistinguishable from *D. ammophilae*, with asci small, pyriform to globose, ascospores 9-10 × 3-3.5 μm.

Dothidea circumvaga var. *hyperici* Desm., Ann. Sci. Nat. Bot., ser. 3, 8: 175. 1847.

Type — France: On dead stems of *Hypericum perforatum* ["vulgare"] (Hypericaceae). Desmazières, Plantes Cryptogames de France no. 89 (B-Desmazières, isotype).

The isotype studied is immature. It seems morphologically indistinguishable from the nominal form, and thus belongs to *Davidiella*, and is morphologically indistinguishable from *D. ammophilae*.

Mycosphaerella cirsii Cruchet, Bull. Soc. Vaud. Sci. Nat. 55: 47. 1923.

Type — France: *Cirsium lanceolatum* (Asteraceae).

No material could be studied, as the type was not included in a loan from PC.

Mycosphaerella cirsii-arvensis Petr., Hedwigia 65(1): 227. 1925.— Fig. 193.

Type — Poland: Stryj, Podhorce. On dead stems of *Cirsium arvense* (Asteraceae). Petrak, VI 1917, distributed in Fungi Polonici Exsiccati no. 642 (BPI, isotype).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradians*, with asci broadly cylindrical, ascospores 9.5-11.5 × 3-3.5 μm. Additional materials (Weißkirchen, Cernotin. On upper and lower surface of dead leaves of *Cirsium arvense* (Asteraceae). Petrak, V 1924, distributed in Mycotheca Generalis no. 1440, B and Germany, Dillenburg, Ludwig, distributed in Reliquiae Petrakianae no. 1247, V 1949, L) agree.

Mycosphaerella citri Whiteside, Phytopathology 62: 263. 1972.— Fig. 194.

Type — USA: Florida, University of Florida, Citrus Experimental Station. On upper and lower surface of dead leaves of *Citrus paradisi* (Rutaceae). Whiteside no. 108 (K, isotype); also on lower surface of dried leaves, VII 1996 (IMI no. 141543, topotype, also dried cultures).

Anamorph: *Stenella citri-grisea* (F.E. Fisher) Sivan. *fade* Sivanesan (1984).

The isotype is morphologically indistinguishable from *Davidiella ammophilae*, with asci pyriform, ascospores 6-7 × 2.5-3 μm. The topotype belongs to *M. punctiformis*, with asci cylindrical, ascospores 7-9 × 2-2.5 μm. The taxonomic position of this species is as yet unresolved, as cultures of *Stenella citri-grisea* do not cluster with *Davidiella* in phylogenetical analyses. Additional material examined (Surinam, van Brussel, VI 1975, CBS) contains only the anamorph, *Stenella citri-grisea*.

Mycosphaerella citricola Tilak, in T.S. Viswan. & Tilak, Sydowia 14: 310. 1960 ≡ *Mycosphaerella tilakii* Tomilin, Opredelitel' gribov roda *Mycosphaerella* Johans.: 240. 1979, superfluous (illegitimate, Article 52) nomen novum (Article 58).— Fig. 195.

Type — India: On dead, partly white, spots on upper surface of living leaves of *Citrus medica* (Rutaceae).

Material studied: India, Mahdya Pradesh, Jabalpur. Rajak, H 504, 22, 412, VI 1977 (IMI no. 214018); also Nepal, Khairnitar, Jujubhai, III 1976 (IMI no. 212502); also Bhutan, on *C. jamblin*, Pereguine, IX 1984, H 1045/84/4814 (IMI no. 290378). The location of the type is unknown. All materials studied is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores 9-10 × 2.5-3 μm.

Sphaerella citricola McAlpine. See *Mycosphaerella limonis* Tomilin.

Mycosphaerella citrullina Grossenb., New York Agric. Exp. Sta. Techn. Bull. 9: 226. 1909 [as "(Sm.) Grossenb. comb. nov.", but with description of the ascospores and therefore considered valid, see Article 59] ≡ *Sphaerella citrullina* (Chester) C.O. Sm., Delaware Coll. Agric. Exp. Sta. Bull. 70: 3. 1905, based on an anamorph (illegitimate, Article 59) ≡ *Phyllosticta citrullina* Chester, Bull. Torrey Bot. Club 1891: 374. 1891.

Type — USA: New York. On dead plant material of *Citrullus* (Cucurbitaceae). Grossenbacher (BPI, syntypes).

Anamorphs: *Diplodina citrullina* (Chester) Grossenb. *fade* Grossenbacher (op. cit.) ≡ *Ascochyta cucumis* Fautrey & Roum. *fade* Eriksson (1992).

Cited as synonymous with *Didymella bryoniae* (Auersw.) Rehm by Eriksson (1992). The many syntypes in BPI agree well.

Mycosphaerella cladii Cruchet, Bull. Soc. Vaud. Sci. Nat. 55: 41. 1923.

Type — Switzerland: *Cladium mariscus* (Cyperaceae). No material could be studied as the type was not included in a loan from PC.

Mycosphaerella clallamensis R. Sprague, Mycologia 49: 837. 1958 [“1957”].

Type — USA: *Bromus laevipes* (Poaceae). No material was studied as the type was not preserved in FH, nor found in any of the herbaria consulted.

Mycosphaerella clandestina (Niessl) Lind, Biol. Meddel. Kongel. Danske Vidensk. Selsk. 11(2): 71. 1934 ≡ *Sphaerella clandestina* Niessl, Verh. Naturf. Vereins Brünn 10: 18. 1872.— Fig. 196.

Type — Austria: Brenner. On upper and lower surface of dead leaves of *Primula minima* × *glutinosa* (Primulaceae). Niessl (M, holotype).

Although it was synonymised with *M. primulae* by von Arx (1949), examination of the holotype shows that it belongs to *Davidiella*, with asci pyriform, ascospores 23-27 × 6-8 µm. Therefore the following new combination is made: **Davidiella clandestina** (Niessl) Aptroot comb. nov., **MB 500346**. **Basionym:** *Sphaerella clandestina* Niessl, Verh. Naturf. Vereins Brünn 10: 18. 1872. Additional material studied (Bosenstein, Niessl, M, 2×) agrees.

Mycosphaerella cleidionii (Berk. & Broome) Aptroot comb. nov., **MB 500333**. **Basionym:** *Sphaeria cleidionii* Berk. & Broome, J. Linn. Soc., Bot. 14: 129. 1875 ≡ *Sphaerella cleidionii* (Berk. & Broome) Sacc., Syll. Fung. 1: 498. 1882 [as “*cleidii*”].— Fig. 197.

Type — Sri Lanka: On pale spots on upper and lower surface of living leaves of *Cleidion javanicum* (Euphorbiaceae), (K, holotype; K, 4 isotypes; IMI 164504, isotype).

This is a parasitic species, with asci cylindrical, ascospores 14-16 × 3-3.5 µm.

Mycosphaerella clematidina Petr., Sydowia 1: 144. 1947.— Fig. 198.

Type — Austria: Niederösterreich, Baden, Pfaffstätten. On dead stems of *Clematis recta* (Ranunculaceae). Petrak, V 1939 (NY, isotype), also V 1940, distributed in Mycotheca Generalis no. 1848 (B, topotype).

The types and authentic material examined (Vienna, Patzak, distributed in Reliquiae Petrakianae no. 1248, VI 1955, L) belong to section *Caterva*, and is morphologically indistinguishable from *M. subradians*, with asci cylindrical, ascospores 8-10 × 2-2.5 µm.

Sphaerella clematidis (Jaap) Kirschst., Kryptog.-Fl. Mark Brandenburg 7(13): 377. 1938 ≡ *Mycosphaerella punctiformis* var. *clematidis* Jaap, Verh. Bot. Vereins Prov. Brandenburg 59: 30. 1917.— Fig. 740.

Type — Germany: Brandenburg, Triglitz in der Prignitz. On upper and lower surface of dead leaves of *Clematis jackmannii* (Ranunculaceae). Jaap, Fungi Selecti Exsiccati no. 770, VI 1916 (L, isotype).

Cited as synonymous with *Mycosphaerella hermione* by Tomilin (1979), which is morphologically indistinguishable from *M. punctiformis*, with which the isotype agrees well, with asci cylindrical, ascospores 8-10 × 2-2.5 µm.

Mycosphaerella clematidis (Oudem.) Johanson ex Oudem., Rév. Champ. Pays-Bas 2: 217. 1897 ≡ *Sphaerella clematidis* Oudem., Ned. Kruidk. Arch., ser. 2, 4: 521. 1886, nomen novum (Article 58) for *Sphaerella aristolochiae* Roum., Fungi Selecti Gallici Exsiccati no. 1601. 1881, nomen nudum (not validly published, Article 32).— Fig. 199.

Type — Netherlands: Vogelenzang. On dead stems of *Aristolochia clematidis* (Aristolochiaceae). Oudemans, VI 1876 (L, holotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores 15-20 × 5.5-7.5 µm. Additional material (France, on spots on living leaves, Roumeguère, Fungi Selecti Gallici Exsiccati no. 1601, L) contains only empty ascomata.

Mycosphaerella clethrae Hara, J. Pl. Protect. 5: 461. 1918.

Type — Japan: *Clethra* (Clethraceae). No material could be studied as the type was not included in a loan from TNS.

Mycosphaerella cleyerae T. Kobay., Mem. Nat. Sci. Mus., Tokyo 9: 87. 1976.

Type — Japan: *Cleyera japonica* (Theaceae). No material could be studied as the type was not included in a loan from TNS.

Mycosphaerella clidemiae Bat. & Peres, in Bat., Peres & Poroca, Atas Inst. Micol. 5: 89. 1967.

Type — Brazil: *Clidemia hirta* (Melastomataceae). No material could be studied as the type was not present in URM.

Mycosphaerella clintoniana (House) House. See *Mycosphaerella rhododendri* Feltgen.

Mycosphaerella clusiae F. Stevens, Trans. Illinois State Acad. Sci. 10: 181. 1917 ≡ *Sphaerella clusiae* (F. Stevens) Trotter, Syll. Fung. 24: 869. 1928 ≡ *Bertia clusiae* (F. Stevens) Gonz. Frag. & Cif., Bol. Real Soc. Esp. Hist. Nat. 24: 494. 1936.— Fig. 201.

Type — Puerto Rico: Maricao, Monte Alegreillo. On lower surface of dead leaves of *Clusia rosea* (Clusiaceae). Stevens no. 1374, V 1913 (BPI, isotype).

This is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores 8-9.5 × 2-2.5 µm. Additional material studied (Venezuela, Caracas, Catuche, Syd., Fungi exotici exsiccati no. 810, XII 1927, L) belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradians*, with asci cylindrical, ascospores 11-13 × 2.5-3.5 µm.

Mycosphaerella clymenia (Sacc.) Johanson ex Oudem., Rév. Champ. Pays-Bas 2: 217. 1897 ≡ *Sphaerella clymenia* Sacc., Michelia 1: 35. 1877.— Fig. 202.

Type — Italy: Treviso, Montello. On pale brown spots with brown margins on upper and lower surface of living leaves of *Lonicera caprifolium* (Adoxaceae). Saccardo, IX 1876, distributed in Mycotheca Veneta no. 908 (BPI, 2 isotypes), also IX 1902, distributed in Mycotheca Italica no. 1031 (B, toptype).

Anamorph: *Phyllosticta vulgare* f. *loniceriae* Desm. fide Saccardo (1882).

Isotypes and additional material studied (Parma, Collecchio, Passerini, distributed in Thümen, Mycotheca Universalis no. 852, 1874, B, L, also in Herbarium Mycologicum Oeconomicum no. 580, B, L; also Netherlands, Putten, Schoonderbeek, VIII 1885, L; also Germany, Westfalen, Siegen, on *Lonicera periclymenum*, Sydow, Mycotheca Germanica no. 2526, IX 1929, L) belong to section *Plaga*, with asci cylindrical, ascospores 10-12 × 3-3.5 µm (in the isotype immature, 7.5-9.5 × 2-2.5 µm). One of the isotypes also contains a coelomycete.

Sphaerella clypeata Sousa da Câmara & A.T. Vascon., Agron. Lusit. 15: 186. 1953.

Type — Spain: *Jasminum* (Oleaceae).

No material could be studied as the type was not present in MA or LISE.

Mycosphaerella coacervata Syd., Ann. Mycol. 22: 296. 1924.

Type — New Zealand: Wellington, Palmerston North. On pale brown spots with brown margins on lower surface of living leaves of *Coprosma robusta* (Rubiaceae). Cunningham no. 1841, VI 1919 (BPI, isotype).

The isotype contains only various coelomycetes.

Mycosphaerella cocoës (Rehm) J. Fröhl. & K.D. Hyde, Sydowia 50: 173. 1999 [nom. illeg., Art. 52: superfluous name for *Mycosphaerella frenumbensis*] ≡ *Sphaerella bambusae* var. *cocoës* Rehm, Hedwigia 40: 110. 1901.— Fig. 102.

Type — Brazil: On upper surface of dead leaves of *Cocos nucifera* (Arecaceae). Ule no. 1185 (S, holotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. ammophilae*, with ascomata often in groups or rows of 2-4, asci pyriform, ascospores 9.5-11.5 × 3-3.5 µm. Reported by Fröhlich & Hyde (1998) as morphologically indistinguishable from *M. frenumbensis*, of which it is a later synonym at species level.

Sphaerella cocoïna Sacc., nomen herbariorum (not validly published, Article 32).

Authentic material — Tahiti. On upper surface of dead leaves of *Cocos nucifera* (Arecaceae). Brunaud (PAD).

Nothing identifiable is present on the material preserved under this name, which has rightly never been published.

Sphaerella cocophylla Cooke, Grevillea 5: 102. 1877 ≡ *Laestadia cocophylla* (Cooke) Sacc., Syll. Fung. 1: 431. 1882 [as “*cocophila*”].— Fig. 203.

Type — Guyana: Demerara. On upper surface of dead leaves of *Cocos nucifera* (Arecaceae). Cooke, distributed in Thümen, Mycotheca Universalis no. 1452 (B, L, isotypes).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. ammophilae*, with asci pyriform, ascospores 9-11 × 3-4 µm. Additional material studied (Chagos, on spots on upper surface of living leaves, Seaward, 1996, CBS) belongs to *Davidiella acrocomiicola*, with asci pyriform, ascospores 9-10 × 2-2.5 µm.

Mycosphaerella codonopsidis Jacz., nomen herbariorum (not validly published, Article 32).

Authentic material — Russia: Primorskij, Ovchinnikovaja. On white spots on upper surface of living leaves of *Codonopsis silvestris* (Campanulaceae). Dyukina, VII 1913 (LEP, L).

The materials studied contain only a *Septoria*, probably the anamorph.

Mycosphaerella coerulea (Ellis & Everh.) Tracy & Earle, in Greene, Plantae Bakerianae 1(1): 33. 1901 ≡ *Sphaerella coerulea* Ellis & Everh., Proc. Acad. Nat. Sci. Philadelphia 46: 334. 1894.— Fig. 204.

Type — USA: Colorado, junction of Big South and Cache la Poudre rivers. On dead stems of *Aquilegia coerulea* (Ranunculaceae). Crandall no. 11, VII 1894 (NY, holotype).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. superflua*, with asci cylindrical, ascospores 13-16 × 3.5-4.5 µm, not reaching 20 µm as was mentioned in the protologue. Additional material studied (Colorado, San Juan Mts., Bethel, distributed in Ellis & Everhart, North American Fungi no. 3522, VII 1894, L) contains only *Pleospora herbarum* (Pers. : Fr.) Rabenh., both immature and mature material.

Mycosphaerella coffeae F. Noack, Z. Pflanzenkrankh. 11: 200. 1901 ≡ *Sphaerella coffeae* (F. Noack) Sacc. & D. Sacc., Syll. Fung. 17: 640.— Fig. 205.

Type — Brazil: On upper surface of dead, often marginal, leaf sections of living leaves of *Coffea arabica* (Rubiaceae). Materials seen: Western Samoa, Uholn. Johnston no. 148, VI 1961 (IMI no. 88819); also Uganda, Kabanyolo University Farm. On *C. dybowskii*. Hansford, C.L. 00642, VIII 1962 (IMI no. 98815); also Gizo, Solomon Islands. On *C. robusta*. Brown, II 1971 (IMI no. 155387); also Kenya, Jacaranda. XI 1971 (IMI 164617, dried culture); also Cuba, Bayamo. Urtiaga, III 1967 (IMI 126566). The type may have been destroyed in B. All materials seen belongs to section *Caterva*, suggesting that this is morphologically indistinguishable from *M. subradicans*, with asci cylindrical, ascospores 11-15 × 3-5 µm.

Mycosphaerella coffeae Sawada, Taiwan Univ. Coll. Agric. Special Publ. 8: 62. 1959, later homonym (illegitimate, Article 53).

Type — Taiwan: *Coffea* (Rubiaceae).

No material was studied as the type was not present in BPI, where many Sawada isotypes are kept.

Mycosphaerella coffeicola (Cooke) Stev. & Wellman, J. Wash. Acad. Sci. 34: 262. 1944 ≡ *Sphaerella coffeicola* Cooke, Grevillea 9: 11. 1880.— Fig. 206.

Type — Venezuela: On leaves of *Coffea arabica* (Rubiaceae). Ernst, 1880 (K, holotype; K, isotype; IMI 268439, isotype, slide only).

This is morphologically indistinguishable from *Lophiostoma rubi* (Fuckel) E.C.Y. Liew, Aptroot & K.D. Hyde, with asci with ocular chamber, cellular pseudoparaphyses, and fusiform ascospores of 22-25 × 5-7 μm.

Mycosphaerella coggygriae Zerova, Bot. Zhurn. (Kiev) 8: 76. 1951.

Type — Russia: *Cotinus coggygria* (Anacardiaceae).

Anamorph: *Septoria rhoïna* Berk. & M.A. Curtis fide Tomilin (1979).

No material could be studied as the type was not included in loans from LE and LEP.

Mycosphaerella collina (Sacc. & Speg.) Tomilin, Novosti Sist. Nizsh. Rast. 8: 149. 1971 ≡ *Sphaerella collina* Sacc. & Speg., in Sacc., Michelia 1: 379. 1878.

Type — Italy: Colpono. On upper and lower surface of dead leaves of *Lonicera xylosteum* (Adoxaceae). Spegazzini, 1 1878 (PAD, holotype).

The type was badly preserved, and most probably contains immature material of *Pleospora herbarum* (Pers. : Fr.) Rabenh.

Sphaerella collina f. *caulicola* Berl., Sacc. & Roum., Rev. Mycol. (Toulouse) 11: 120: 1889.

Type — Spain: *Lonicera etrusca* (Adoxaceae).

No material could be studied as the type was not included in a loan from PC and not found in any of the herbaria consulted.

Mycosphaerella colocasiae Hara, J. Pl. Protect. 4: 355. 1917 ≡ *Sphaerella colocasiae* (Hara) Trotter, Syll. Fung. 24: 851. 1928.

Type — Japan. On white spots on upper surface of living leaves of *Colocasia antiquorum* (Araceae). Hara, 1916 (TNS 209262, holotype).

Type and additional material seen (Hara, X 1925, TNS 209886) contain only coelomycetes.

Mycosphaerella colombiensis Crous & M.J. Wingf., Mycologia Memoir 21: 41. 1998.

Type — Colombia: Pinal Farm. On leaves of *Eucalyptus urophylla* (Myrtaceae). Wingfield, V 1995 (PREM 54396, holotype, not seen).

No material was studied of this recently described species.

Mycosphaerella colorata (Peck) Earle, in Mohr, Contr. U.S. Natl. Herb. 6: 174. 1901 ≡ *Sphaerella colorata* Peck, Annual Rep. New York State Mus. 29: 62. 1878.— Fig. 207.

Type — USA: New York, Center. On white to brown spots on upper surface of living leaves of *Kalmia angustifolia* (Ericaceae). Peck, distributed in Ellis, North American Fungi no. 899 (L, 2 isotypes).

Isotypes and additional material studied (Canada, Newfoundland, Port aux Basques, Cain no. 40745, VIII 1963, L; also Chocarua, Farlow, VII 1908, B) belong to section *Plaga*, with asci cylindrical, ascospores 15-17 × 2.5-3 μm.

Mycosphaerella columbariae Feltgen, Recueil Mém. Trav. Soc. Bot. Grand-Duché Luxembourg 1901: 182. 1901 ≡ *Sphaerella columbariae* (Feltgen) Sacc. & P. Syd., Syll. Fung. 16: 1133. 1902.

Type — Luxembourg: *Scabiosa columbaria* (Dipsacaceae). No material was studied as the type was not found in any of the herbaria consulted.

Mycosphaerella columbi Rehm, Ann. Mycol. 6: 120. 1908 ≡ *Sphaerella columbi* (Rehm) Sacc. & Trotter, Syll. Fung. 22: 133. 1913.— Fig. 208.

Type — USA: Ohio, Columbus. On upper and lower surface of brown parts of (possibly still living) leaves of *Plantago rugelii* (Plantaginaceae). Kellerman no. 6750, 1907 (S, holotype; B, isotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. ammophilae*, with asci pyriform, ascospores 11-13 × 3-3.5 μm.

Mycosphaerella columbiae Syd. & P. Syd., Ann. Mycol. 14: 360. 1916 ≡ *Sphaerella columbiae* (Syd. & P. Syd.) Trotter, Syll. Fung. 24: 889. 1928.— Fig. 209.

Type — Philippines: Luzon, Prov. Laguna, Los Baños. On white spots on upper surface of living leaves of *Colona* ["*Columbia*"] *serratifolia* (Malvaceae). Baker no. 3043, I 1914 (S, holotype).

This is a parasitic species, with asci broadly cylindrical, ascospores 16-19 × 4.5-5.5 μm.

Mycosphaerella comari (J. Schröt.) Tomilin, Opredelitel' gribov roda *Mycosphaerella* Johans.: 221. 1979 ≡ *Stigmatea comari* J. Schröt., in Cohn, Kryptog.-Fl. Schlesien 3(2): 331. 1894 ["1893"] ≡ *Sphaerella schroeteriana* Kirschst., Kryptog.-Fl. Mark Brandenburg 7(3): 415. 1938, superfluous (illegitimate, Article 52) nomen novum (Article 58).

Type — Poland: *Comarum palustre* (Rosaceae).

Synonymised with *Venturia palustris* Sacc., E. Bommer & M. Rousseau by Sivanesan (1977).

Sphaerella comedens Pass., in Rabenh., Fungi Europaei Exsiccati, ed. nov., ser. 2 no. 2439. 1876 ≡ *Laestadia comedens* (Pass.) Sacc., Syll. Fung. 1: 430. 1882.

Type — Italy: On white spots on upper surface of living leaves of *Ulmus campestris* (Ulmaceae). Passerini, 1876, distributed in Rabenhorst, Fungi Europaei Exsiccati, ed. nov., ser. 2 no. 2439 (CBS, isotype).

The isotype studied contains only a *Microsphaeropsis* anamorph.

Sphaerella comedens (Schwein.) Cooke, J. Bot. 21: 69. 1883, later homonym (illegitimate, Article 53) ≡ *Sphaeria comedens* Schwein., in Berk. & M.A. Curtis, J. Acad. Nat. Sci. Philadelphia, ser. 2, 2: 210. 1853.

Type — Surinam: On lower surface of dead leaf (PH, holotype).

The holotype contains only a coelomycete.

Mycosphaerella communis Crous & J.P. Mansilla in Crous *et al.*, Stud. Mycol. 50: 203. 2004.

Type — Spain: Pontevedra, Lourizán, Areeiro, on leaves of *Eucalyptus globulus* (Myrtaceae). Mansilla, XII 2002 (CBS 9900, holotype); culture ex-type CBS 114238 = CPC 10440. Anamorph: *Dissoconium commune* Crous & J.P. Mansilla *vide* Crous *et al.* (op. cit.).

No material was studied of this recently described species.

Mycosphaerella compositarum (Auersw.) J. Schröt., in Cohn, Kryptog.-Fl. Schlesien 3(2): 338. 1894 [“1893”] = *Sphaerella compositarum* Auersw., in Gonn. & Rabenh., Mycol. Europaea 5-6: 15. 1869.— Fig. 210.

Type — Germany: On dead stems of *Cichorium intybus* [and *Carlina acaulis* according to the protologue] (Asteraceae). Auerswald (B, holotype and isotype).

Synonymised with *Davidiella allicina* (as *Mycosphaerella tassiana*) by von Arx (1949), with which the types agree. Additional material studied (Leipzig, on stems of *Cichorium intybus*, Winter, distributed in Rabenhorst, Fungi Europaei Exsiccati no. 1558, V 1871, L) contains only an immature ascomycete, possibly *Pleospora herbarum* (Pers. : Fr.) Rabenh., with ascospores hyaline, 1-septate, 20-24 × 7-9 µm, and an anamorph.

Mycosphaerella concentrica (Racib.) Joa.E. Taylor & Crous, Mycol. Res. 107: 657. 2003 = *Gibellina concentrica* Racib., Parasitische Algen und Pilze Javas 2: 11. 1900 = *Teratosphaeria concentrica* (Racib.) E. Müll., Beitr. Kryptogamenfl. Schweiz 11, 2: 316. 1962.

Type — Indonesia: *Lasianthus* (Rubiaceae).

No material was studied of this recently described species.

Sphaerella coneglanensis Speg., Michelia 1: 455. 1879.

Type — Italy: *Calycanthus floridus* (Calycanthaceae).

No material could be studied as the type was not included in a loan from LPS and not found in any of the herbaria consulted.

Mycosphaerella conferta (Speg.) Lindau, in Engl. & Prantl, Natürlichen Pflanzenf. 1(1): 425. 1897 = *Sphaerella conferta* Speg., Anales Soc. Ci. Argent. 18: 279. 1884.— Fig. 211.

Type — Brazil: Guarapi. On upper surface of dead leaves of a Sapindaceae. Balansa no. 3891, VII 1882, distributed in Plantas de Paraguay no. 216 (NY, isotype); also distributed in Roumeguère, Fungi Sel. Exsiccati no. 4135 (NY, isotype).

This belongs to section *Longispora*, and is morphologically indistinguishable from *M. millegrana*, with ascomata in dense groups, asci cylindrical, ascospores 14-18 × 2.5-3 µm.

Mycosphaerella confinis (P. Karst.) Dearn., Rep. Canad. Arctic Exped. 1913-1918 4, C: 6. 1923 = *Sphaerella confinis* P. Karst., Öfvers. Förh. Kongl. Svenska Vetensk.-Akad. 1872(2): 106. 1872.

Type — Svalbard: *Draba* (Brassicaceae).

Synonymised with *Davidiella allicina* (as *Mycosphaerella tassiana*) by von Arx (1949). Material studied (Sweden, Gutland, on upper and lower surface of dead leaves of *Arabis gerardii* (Brassicaceae), Hamberg, VII 1891, distributed in Romell, Fungi Exsiccati Preasertim Scandinaviae no. 181, PAD, 2×) seems to agree, with asci pyriform, ascospores immature.

Sphaerella confinis subsp. *hambergii* Romell & Sacc. See *Mycosphaerella hambergii* (Romell & Sacc.) Petr.

Mycosphaerella confusa F.A. Wolf, Mycologia 28: 85. 1936, nomen novum (Article 58) for *Mycosphaerella dubia* F.A. Wolf, Mycologia 27: 355. 1935, later homonym (illegitimate, Article 53).— Fig. 212.

Type — USA: North Carolina, Durham. On upper surface of dead, overwintered leaves of *Rubus* (Rosaceae). Wolf, VIII 1933 (IMI no. 227780, isotype, slide only); also VII 1934 (IMI no. 227774, topotype, slide only).

Anamorph: *Pseudocercospora rubi* (Sacc.) Deighton *vide* Deighton (1976).

Types and additional material (Brazil, São Paulo, Araraquara, Viegas, White Herbarium no. 1496, VI 1933, IMI 227773, slide only) show that this is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores 7-9 × 2-2.5 µm.

Mycosphaerella conglomerata (Wallr.) Lindau, in Engl. & Prantl, Natürlichen Pflanzenf. 1(1): 424. 1897 = *Sphaeria conglomerata* Wallr., Fl. Cryptog. Germ. 2: 814. 1833 = *Sphaerella conglomerata* (Wallr.) Rabenh., Fungi Europaei Exsiccati, ed. nov., ser. 2 no. 150. 1860.— Fig. 213.

Type — Germany: On lower surface of dead leaves of *Alnus incana* (Betulaceae). (B, 2 possible isotypes).

Material studied (Germany, Brandenburg, Prenden, on *Alnus glutinosa*, Sydow, Mycotheca Germanica no. 1537, IV 1916, L) is *M. punctiformis*, with asci cylindrical, ascospores 8-9 × 2-3 µm.

Sphaerella conglomerata f. *alnicola* Rabenh., Fungi Europaei Exsiccati, ed. nov., ser. 2 no. 150. 1860.

Type — Germany: Oestrich. On lower surface of dead leaves of *Alnus incana* (Betulaceae). Fückel, distributed in Rabenhorst, Fungi Europaei Exsiccati, ed. nov., ser. 2 no. 150 (L, 2 isotypes).

This is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores 8-9 × 2-3 µm.

Sphaerella conglomerata f. *siliquastris* Desm., in Roum., Fungi Gallici Exsiccati no. 1944. 1881.

Type — France: On veins on lower surface of dead leaves of *Cercis siliquastrum* (Fabaceae). Roumeguère, X 1881, Fungi Gallici Exsiccati no. 1944 (L, isotype).

The isotype studied contains only empty ascomata.

Sphaerella conglomeratiformis Bubák & Vleugel, in Vleugel, Svensk Bot. Tidskr. 11: 309. 1917.— Fig. 214, 988.

Type — Sweden: Luleå, Björkin. On lower surface of dead leaves of *Alnus incana* var. *borealis* (Betulaceae). Vleugel, VI 1914 (BPI, holotype).

This is a species of *Planistromella*, with ascomata in stromata, asci cylindrical, surrounded by a few hamathecial cells, ascospores $9.5-10 \times 2-2.5 \mu\text{m}$. Therefore the following new combination is proposed: ***Planistromella conglomeratiformis*** (Bubák & Vleugel) Aptroot comb. nov., **MB 500362**. **Basionym:** *Sphaerella conglomeratiformis* Bubák & Vleugel, in Vleugel, Svensk Bot. Tidskr. 11: 309. 1917. Also present in the type is isotype material of *Phaeosphaerella borealis* (Bubák & Vleugel) Tomilin (see above).

Sphaerella congregata (Lév.) Sacc., Syll. Fung. 1: 534. 1882 = *Sphaeria congregata* Lév., Ann. Sci. Nat. Bot., ser. 3, 5: 261. 1846.

Type — Indonesia: On leaves.

No material could be studied as the type was not included in a loan from PC and not found in any of the herbaria consulted.

Sphaerella conicola Peck, in Berl. & Voglino, Syll. Fung., Addit. ad vol. 1-4: 465. 1886.— Fig. 215.

Type — USA: New York, Schroon River. On scales of dead cones of *Thuja* [*“Arbor vitae”*] Cupressaceae). Peck (NY, isotype).

Cited as synonymous with *Scirrhia conigena* (Peck) M.E. Barr by Barr (1972), with which the isotype agrees well, with asci pyriform, surrounded by parenchymatous tissue, ascospores yellowish, $20-27 \times 5-6 \mu\text{m}$.

Mycosphaerella conigena (Peck) House, New York State Mus. Bull. 233-234: 26. 1921 = *Sphaerella conigena* Peck, Annual Rep. New York State Mus. 33: 34. 1883 [*“1880”*] = *Scirrhia conigena* (Peck) M.E. Barr, Contr. Univ. Michigan Herb. 9: 566. 1972.

Type — USA: New York, Albany Co., Helderberg Mts. On scales of dead cones of *Thuja occidentalis* (Cupressaceae). Peck, VII (NY, isotype).

Accepted as *Scirrhia conigena* (Peck) M.E. Barr by Barr (op. cit.), with which the isotype agrees.

Sphaerella conigena Peck. See *Mycosphaerella peckii* Lindau.

Sphaerella conigena Ellis & Everh. See *Mycosphaerella andersonii* (Sacc.) Lindau.

Mycosphaerella conigena (Bubák) Sacc., in Trotter, Syll. Fung. 24: 893. 1928, later homonym (illegitimate, Article 53) = *Rehmiellopsis conigena* Bubák, Ann. Mycol. 12: 206. 1914 = *Diplosphaerella conigena* (Bubák) Tomilin, Opredelitel' gribov roda *Mycosphaerella* Johans.: 285. 1979.

Type — Austria: *Pinus* (Pinaceae).

No material was studied; the type was not found in BPI. It is a *Delphinella*, most probably *D. peckii* (Lindau) M.E. Barr.

Mycosphaerella consociata (Rehm) Magnus, in Dalla Torre & Sarnth., Fl. Tirol, Vorarlberg, Liechtenstein, III, Pilze: 466. 1905 = *Sphaerella consociata* Rehm, Hedwigia 24: 238. 1885.

Type — Austria: *Trifolium pallescens* (Fabaceae).

Synonymised with *Davidiella allicina* (as *Mycosphaerella tassiana*) by von Arx (1949). The type was not found in S or in any of the herbaria consulted. Material studied (USA, Colorado, Mount Garfield, on dead petioles of *Trifolium nanum*, Clements, Cryptogamae Formationum Coloradensium no. 218, VIII 1906, S [sub *“Mycosphaerium consociatum”*]) contains only *Pleospora herbarum* (Pers. : Fr.) Rabenh.

Mycosphaerella conspicua (Syd. & P. Syd.) Bat. & Cavalcanti, in Bat., Peres, Cavalcanti & Heringer, Atas Inst. Micol. 3: 222. 1966 = *Sphaerella conspicua* Syd. & P. Syd., Ann. Mycol. 5: 356. 1907.— Fig. 216.

Type — Brazil: Minas Gerais, Confins. On upper and lower surface of dead leaves of *Myrsine* (Myrsinaceae). Noack no. 586, VII 1897 (S, holotype).

This belongs to section *Longispora*, and is morphologically indistinguishable from *M. latebrosa*, with asci cylindrical, ascospores $24-28 \times 3-5 \mu\text{m}$.

Mycosphaerella contraria Hansf., Proc. Linn. Soc. London 153: 22. 1941.

Type — Uganda: Kampala plantation. On white spots with black margins on upper surface of living leaves of *Dioscorea* (Dioscoreaceae). Chandler, VI 1937 (IMI no. 7956, isotype).

Anamorph: *Pseudocercospora contraria* (Syd. & P. Syd.) Deighton fide Sivanesan (1984).

The isotype studied contains only the anamorph, *Pseudocercospora contraria*. Additional material studied (Papua-New Guinea, Utabia, on dead stems and dead leaves of *D. bulbifera* and *D. albata*, IX 1984, IMI no. 296667; also Papua-New Guinea, Saramandi, on *D. alata*, Quin, VIII 1984, IMI no. 288622) contains *M. punctiformis*, with asci cylindrical and ascospores $9-10 \times 2-2.5 \mu\text{m}$. Further material (Solomon Islands, Guadalcanal, Honiara, on *D. alata*, Jackson, V 1983, IMI no. 277542; also Solomon Islands, Isabel, Honiara, on *D. bulbifera*, Mason no. 166, V 1959, IMI no. 77271) contains a *Didymella* with very variable ascospores.

Mycosphaerella convallariae McKeen & R.C. Zimmer, Canad. J. Bot. 42: 667. 1964 [as *convallaria*] = *Didymella convallariae* (McKeen & R.C. Zimmer) Arx, Beih. Nova Hedwigia 87: 288. 1987.

Type — Canada: *Convallaria majalis* (Asparagaceae).

Anamorph: *Ascochyta majalis* C. Massal. fide McKeen & Zimmer (op. cit.).

No material studied; already accepted as *Didymella convallariae* (McKeen & R.C. Zimmer) Arx by von Arx (op. cit.).

Mycosphaerella convexula (Schwein.) F.V. Rand, Phytopathology 1: 135. 1911 = *Sphaeria convexula* Schwein., Trans. Amer. Philos. Soc., ser. 2, 4: 224. 1832 = *Sphaerella convexula* (Schwein.) Thüm., Mycotheca Universalis no. 1149. 1878.— Fig. 217.

Type — USA: Philadelphia, Bethlehem. On lower surface of dead leaves of *Juglans* (Juglandaceae). Schweinitz (PH, holotype).

Anamorphs: *Cercospora halstedii* Ellis & Everh. (= *Passalora halstedii* (Ellis & Everh.) U. Braun & Crous) and *Phyllosticta convexula* Bubák *vide* Tomilin (1979).

The holotype is immature, but this is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical. Additional material studied (USA, Ohio, Amanda, on upper and lower surface of dead leaves of *Carya amara*, Kellerman, distributed in Rabenhorst-Winter, Fungi Europaei Exsiccati no. 3257, VI 1883, L) is *M. punctiformis*, with asci cylindrical, ascospores $7-9 \times 2.5-3.5$ μm . It was cited as synonymous with *Mycosphaerella dendroides* by Tomilin (1979).

Sphaerella convexula f. *caryae-tomentosae* Thüm., Mycotheca Universalis no. 1149. 1878.

Type — U.S.A.: Carolina, Aiken. On upper and lower surface of dead leaves of *Carya tomentosa* (Juglandaceae). Ravenel no. 2225 (NY, isotype), also distributed in Thümen, Mycotheca Universalis no. 1149 (L, NY, isotypes).

This is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores $7-9 \times 2.5-3.5$ μm .

Sphaerella cookeana Auersw., in Gonn. & Rabenh., Mycol. Europaea 5-6: 2. 1869, nomen novum (Article 58) for *Sphaerella punctiformis* Cooke, J. Bot. 4: 246. 1866, later homonym (illegitimate, Article 53), nomen novum (Article 58) for *Sphaeria punctiformis* Desm., Plant. Cryptog., ed. 1 no. 1794. 1845, later homonym (illegitimate, Article 53) = *Laestadia cookeana* (Auersw.) Sacc., Syll. Fung. 1: 421. 1882 = *Guignardia cookeana* (Auersw.) Lindau, Hilfsb. Sammeln Ascomyceten: 92. 1903 = *Anisostomula cookeana* (Auersw.) Höhn., Ann. Mycol. 16: 48. 1918 = *Hyponectria cookeana* (Auersw.) M.E. Barr, Mycologia 68: 618. 1976.

Type — France: On lower surface of dead leaves of *Quercus* (Fagaceae). Desmazières, Plantes Cryptogames de France no. 1794 (BPI, isotype).

Accepted as *Hyponectria cookeana* (Auersw.) M.E. Barr by Eriksson (1992). The isotype is immature, but agrees, as does additional material studied (Germany, Sachsen, Pirna, on *Q. pedunculata*, Krieger, VI 1909, B; also Schleswig-Holstein, Sachsenwald, on *Q. robur*, Jaap, Fungi Selecti Exsiccati no. 422, III 1910, B).

Mycosphaerella cookei (Linds.) Sacc. & D. Sacc., in Sacc., Syll. Fung. 17: 649. 1905 = *Microthelia cookei* Linds., Trans. Roy. Soc. Edinburgh 25: 537. 1869 = *Arthopyrenia cookei* (Linds.) Arnold, Flora 57: 139. 1874 = *Polycarpella cookei* (Linds.) Theiss. & Syd., Ann. Mycol. 16: 28. 1918 = *Delphinella cookei* (Linds.) E. Müll., in E. Müll. & Arx, Beitr. Kryptogamenfl. Schweiz 11(2): 28. 1962.

Type — United Kingdom. On thallus of *Trapelia coarctata* ["*Lecanora dispersa*"] (Ascomycota, Argyriales, Trapeliaceae). (K, holotype).

Already cited as synonymous with *Muellerella lichenicola* (Sommerf.) D. Hawksw. by Clauzade, Diederich & Roux (1989) and therefore not studied in detail.

Sphaerella cooperta (Desm.) Ces. & De Not., Comment. Soc. Crittog. Ital. 1(4): 237. 1863 = *Sphaeria cooperta* Desm., Ann. Sci. Nat. Bot., ser. 3, 11: 355. 1849 =

Laestadia cooperta (Desm.) Sacc., Syll. Fung. 1: 426. 1882 = *Phacidium cooperta* (Desm.) Höhn., Oesterr. Bot. Z. 57: 324. 1907 = *Anisostomula cooperta* (Desm.) Höhn., Ann. Mycol. 16: 49. 1918.

Type — France: *Quercus coccifera* (Fagaceae).

Accepted as *Anisostomula cooperta* (Desm.) Höhn. by von Arx & Müller (1954) and therefore not studied.

Mycosphaerella coptis (Schwein.) House, New York State Mus. Bull. 233-234: 27. 1921 = *Sphaeria coptis* Schwein., Trans. Amer. Philos. Soc., ser. 2, 4: 224. 1832 = *Sphaerella coptis* (Schwein.) Farlow, Appalachia 3: 247. 1884 = *Laestadia coptis* (Schwein.) Ellis & Everh., N. Amer. Pyrenomyc.: 261. 1892.

Type — USA: New York, Pocono. On spots on upper and lower surface of living leaves of *Coptis trifolia* (Ranunculaceae). Schweinitz (PH, holotype; PH, isotype).

Anamorph: *Septoria coptis* Berk. & M.A. Curtis *vide* Sivanesan (1984).

Both holo- and isotype are immature. Additional material studied (USA, Massachusetts, Chepacco Lake, Seymour, distributed in Ellis & Everh., North American Fungi no. 2358, VI 1888, L) contains only the anamorph.

Mycosphaerella cordatae Ananthan., Mycopathol. Mycol. Appl. 22: 97. 1964 [as "*cordata*"].

Type — India: Katraj, Pune. On white spots on upper surface of living leaves of *Aspidopterys cordata* (Malpighiaceae). Ananthanarayanan no. 169b, VIII 1962 (LWG, holotype).

Anamorph: *Phyllosticta aspidopteridis* Ananthan. *vide* Ananthanarayanan (op. cit.).

The type material studied contains only the anamorph.

Mycosphaerella cordylinicola (Speg.) Tomilin, Novosti Sist. Nizsh. Rast. 1967: 188. 1967 = *Sphaerella cordylinicola* Speg., Anales Mus. Nac. Hist. Nat. Buenos Aires 23: 51. 1912.

Type — Argentina: *Cordyline dracaenoides* (Asparagaceae).

No material was studied as the type was not included in a loan from LPS.

Mycosphaerella corispermii Lobik, Materialy po floristicheskim i faunisticheskim obsledovaniyam Terskogo okruga: 26. 1928.

Type — Russia: *Corispermum hyssopifolium* (Amaranthaceae).

No material was studied as the type was not included in loans from LE or LEP.

Mycosphaerella corni Kill. & Likhité, Bull. Soc. Mycol. France 42: 224. 1926.

Type — France: *Cornus sanguinea* (Cornaceae).

Anamorph: *Ramularia corni* (Desm.) Kill. & Likhité *vide* Killian & Likhité (op. cit.).

No material was studied as the type was not included in a loan from PC.

Mycosphaerella corni (Schwein.) J.H. Miller. See *Mycosphaerella cornifolia* Lindau.

Mycosphaerella corni Verma & Kamal, Indian Phytopathol. 40: 461. 1987, later homonym (illegitimate, Article 53) [as “*cornii*”].

Type — India: Uttar Pradesh, Naini Tal. On dead leaves of *Cornus macrophylla* (Cornaceae). Verma, XI 1985 (IMI no. 299707, isotype).

Anamorph: *Thyrostroma corni* Verma & Kamal *vide* Verma & Kamal (op. cit.).

The isotype studied contains only the anamorph, *Thyrostroma corni*.

Mycosphaerella cornicola Tehon & Daniels, Mycologia 17: 240. 1925.

Type — USA: Illinois, Jo Daviess Co., Apple River Canyon. On bark of living twigs of *Cornus stolonifera* (Cornaceae). Young no. 13596, VII 1924 (BPI, isotype).

Cited as a probable synonym of *Leiosphaerella vexata* (Sacc.) E. Müll. by Müller & von Arx (1962). The isotype contains only a *Phomopsis* coelomycete, with conidia simple, 30-35 × 12-15 µm, surrounded by paraphyses, which may have led to the original description of the species.

Mycosphaerella cornifolia (Cooke) Lindau, in Engl. & Prantl, Natürlichen Pflanzenf. 1(1): 425. 1897 = *Sphaerella cornifolia* Cooke, J. Bot. 21: 108. 1883, nomen novum (Article 58) for *Mycosphaerella corni* (Schwein.) J.H. Miller, Mycologia 33: 80. 1941, later homonym (illegitimate, Article 53) = *Sphaeria corni* Schwein., Trans. Amer. Philos. Soc., ser. 2, 4: 225. 1832.

Type — USA: New York, Pocono. On lower surface of dead leaves of *Cornus* (Cornaceae). Schweinitz (PH, holotype; PH, K, isotypes).

Both holo- and isotypes are immature, but this is morphologically indistinguishable from *M. punctiformis*. Additional material studied (USA, Londonville, New York, Albany, on *C. florida*, House, IV 1945, B) agrees.

Mycosphaerella coronillae-variae Petr., Ann. Mycol. 21: 1. 1923.— Fig. 218.

Type — Czech Republic: Weißkirchen, Svřčow. On dead stems of *Coronilla varia* (Fabaceae). Petrak, Mycotheca Generalis no. 1552, V 1936 (W, topotype; B, isotopotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. disseminata*, with asci pyriform, ascospores 12-14 × 3-4 µm.

Sphaeria corrigiolae H. Hoffm., Icon. Analyt. F., Heft 2, taf. 8. 1862 [as “*Sphaeria (Sphaerella) corrigiolae*”].

Type — Germany: *Corrigiola* (Caryophyllaceae).

No material was studied; the type may have been destroyed in B.

Mycosphaerella corylaria (Wallr.) Syd., Mycotheca Germanica no. 1538. 1921 = *Sphaeria corylaria* Wallr., Fl. Cryptog. Germ. 2: 770. 1833 = *Sphaerella corylaria* (Wallr.) Fuckel, Jahrb. Nassauischen Vereins Naturk. 23-24: 99. 1870.— Fig. 219.

Type — Germany: On lower surface of dead leaves of *Corylus avellana* (Betulaceae).

Cited as synonymous with *M. punctiformis* by Tomilin (1979).

The type was not found in B. Material studied (Germany, Brandenburg, Sophienstad, Sydow, Mycotheca Germanica, no. 1538, V 1916, L) is *M. punctiformis*, with asci cylindrical, ascospores 8-9 × 2.5-3.5 µm.

Mycosphaerella corylina (P. Karst.) Tomilin, Novosti Sist. Nizsh. Rast. 1967: 188. 1967 = *Sphaerella sparsa* subsp. *corylina* P. Karst., Hedwigia 23: 4. 1884.

Type — Finland: On lower surface of dead leaves of *Corylus avellana* (Betulaceae).

The type was not found in H. Material studied (Netherlands, Nunspeet, Beins no. 83, I 1903, L) is immature. Cited as synonymous with *M. maculiformis*, which is morphologically indistinguishable from *M. punctiformis*, by Tomilin (1979).

Mycosphaerella costii Saccas, I.F.C.C. Bull. 16: 294. 1981, type not designated (not validly published, Article 37).

Authentic material — Central African Republic: *Coffea* (Rubiaceae).

No material was studied as the type was not designated and no material was included in a loan from PC.

Mycosphaerella cotoneastri Dzhagal., Trudy Sukhumsk. Bot. Sada 15: 118. 1964.

Type — Georgia: *Cotoneaster salicifolia* (Rosaceae).

No material was studied as the type was not included in loans from LE or LEP.

Mycosphaerella coymiana Jaap, Ann. Mycol. 12: 11. 1914 = *Sphaerella coymiana* (Jaap) Trotter, Syll. Fung. 24: 875. 1928.— Fig. 220.

Type — Germany: On dead stems of *Vicia pisiformis* (Fabaceae).

The type was not found in B or in any of the other herbaria consulted. Material examined (Germany, Prov. Hessen-Nassau, Dillkreis, Donsbachtal, Ludwig, distributed in Sydow, Mycotheca Germanica no. 3105, V 1936, L) belongs to *Davidiella*, and suggests that this may be morphologically indistinguishable from *D. ammophilae*, with asci small, pyriform, ascospores 10-12 × 4-5 µm.

Mycosphaerella crassa (Auersw.) Lindau, Hilfsb. Sammeln Ascomyceten: 86. 1903 = *Sphaerella crassa* Auersw., in Gonn. & Rabenh., Mycol. Europaea 5-6: 7. 1869.— Fig. 221.

Type — Germany: on upper surface of dead leaves of *Populus tremula* (Salicaceae).

No material was studied; the type was not found in B or in any of the other herbaria consulted. Material studied (Italy, on *Populus alba*, PAD) belongs to *Davidiella*, and suggests that this may be morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores 15-22 × 6-8 µm.

Sphaerella crastophila Sacc., Atti Soc. Veneto-Trentina Sci. Nat. 2: 142. 1873 = *Leptosphaeria crastophila* (Sacc.) Sacc., Syll. Fung. 2: 61. 1883.

Type — Italy: *Festuca* (Poaceae).

No material was studied as the species was already disposed of as *Leptosphaeria crastophila* (Sacc.) Sacc. by Saccardo (op. cit.).

Mycosphaerella crataegi (Fuckel) Johanson ex Oudem., Rév. Champ. Pays-Bas 2: 215. 1897 = *Sphaeria crataegi* Fuckel, Fungi Rhenani Exsiccati no. 2162. 1867 = *Sphaerella crataegi* (Fuckel) Auersw., in Gonn. & Rabenh., Mycol. Europaea 5-6: 4. 1869 = *Carlia crataegi* (Fuckel) Höhn., Hedwigia 62: 57. 1920.— Fig. 222.

Type — Germany: On upper and lower surface of dead leaves of *Crataegus oxyacantha* (Rosaceae). Fuckel, Fungi Rhenani Exsiccati no. 2162 (L, isotype).

Anamorph: *Septoria oxyacanthae* Kunze *fide* Saccardo (1882).

The isotype studied is immature. Additional material studied (Czech Republic, Weißkirchen, Petrak, IV 1914, distributed in Flora Bohemiae et Moraviae Exsiccata no. 1020, L) is a species of *Sphaerulina*, probably *Sphaerulina tarda* (Harkn.) M.E. Barr, with asci cylindrical, ascospores 1-3-septate, in one bundle, sometimes curved, 40-50 × 2-3 µm.

Mycosphaerella crataegicola Bondartsev & Tranzschel, Bolezni Rast. 7: 49. 1913 = *Sphaerella crataegicola* (Bondartsev & Tranzschel) Trotter, Syll. Fung. 24: 883. 1928.

Type — Russia: *Crataegus sanguinea* (Rosaceae).

Anamorphs: *Phyllosticta michailovskoensis* Elenkin & Ohl *fide* Trotter (1928) and *Septoria crataegicola* Bondartsev & Tranzschel *fide* Bondartsev & Tranzschel (op. cit.).

Cited as synonymous with *Mycosphaerella crataegi* by Tomilin (1979). No material was studied as the type was not included in loans from LE or LEP.

Mycosphaerella craterispermi Hansf., Proc. Linn. Soc. London 156: 110. 1944.— Fig. 223.

Type — Uganda: Entebbe Road. On upper and lower surface of living leaves of *Craterispermum laurinum* (Rubiaceae). Hansford no. 3196, V 1943 (K, holotype).

This is a parasitic species of *Davidiella*, with asci pyriform to clavate, ascospores becoming pale brownish, 23-27(-35) × 5-6(-7) µm. Therefore the following new combination is made: **Davidiella craterispermi** (Hansf.) Aptroot comb. nov., **MB 500347**. **Basionym:** *Mycosphaerella craterispermi* Hansf., Proc. Linn. Soc. London 156: 110. 1944.

Mycosphaerella creberrima (Penz. & Sacc.) Syd. & P. Syd., Philipp. J. Sci., C, Bot. 8: 271. 1913 = *Sphaerella creberrima* Penz. & Sacc., Malpighia 11: 397. 1897.— Fig. 224.

Type — Indonesia: Java, Tjibodas. On grey spots with brown margins on upper surface of living leaves of *Erythralium* (Erythraliaceae). Penzig (PAD, holotype).

This is a parasitic species, with asci cylindrical, ascospores 12-14 × 3-3.5 µm. Additional material studied (Philippines, Luzon, Los Baños, on *E. scandens*, Foxworth, XII 1928, Bureau of Science no. 19654, NY) contains only a coelomycete.

Mycosphaerella crebra (Fautrey & Lambotte) Feltgen, Recueil Mém. Trav. Soc. Bot. Grand-Duché Luxembourg

15: 257. 1902 = *Sphaerella crebra* Fautrey & Lambotte, Rev. Mycol. (Toulouse) 18: 144. 1896 = *Carlia crebra* (Fautrey & Lambotte) Höhn., in Krieger, Fungi Saxonici Exsiccati no. 2460. 1919.

Type — France: On dead stems of *Linaria vulgaris* (Plantaginaceae). Fautrey, distributed in Roumeguère, Fungi Selecti Exsiccati no. 7278, V 1896 (PC, holotype).

The holotype is immature. The species was cited as synonymous with *Mycosphaerella linariae* by Tomilin (1979).

Sphaerella crenata Brunaud, Actes Soc. Linn. Bordeaux, ser. 5, 2: 96. 1889 [“1888”].

Type — France: *Spiraea crenata* (Rosaceae).

No material has been studied, as the location of the type is unknown and its preservation uncertain.

Mycosphaerella crepidophora (Mont.) Rehm, Ann. Mycol. 5: 210. 1907 = *Depazea crepidophora* Mont., in Berk., Some notes upon the cryptogamic portion of the plants collected in Portugal by F. Welwitsch. The Fungi: 10. 1853 = *Sphaerella crepidophora* (Mont.) Sacc. Syll. Fung. 1: 479. 1882.

Type — Portugal: On upper and lower surface of dead leaves of *Viburnum tinus* (Adoxaceae).

The type was not included in a loan from PC. Material studied (Italy, Genua, Jaap, Fungi Selecti Exsiccati no. 618, I 1913, BPI) is *M. punctiformis*, with asci cylindrical, ascospores immature, and also contains a coelomycete.

Sphaerella crepini (Westend.) Ces. & De Not., Comment. Soc. Crittog. Ital. 1(4): 237. 1863 = *Sphaeria crepini* Westend., Herbar Cryptogamique Belgique, fasc. 19 no. 911. 1854.

Type — Belgium: On dead scales of *Lycopodium* (Lycopodiaceae).

The type was not found in BR. Material studied (Germany, Oestrich, on *L. clavatum*, Fuckel, Fungi Rhenani no. 837, B) contains only a coelomycete.

Mycosphaerella crini Siemaszko, Acta Soc. Bot. Pol. 1: 19. 1923.

Type — Georgia: *Crinum* (Alliaceae).

Anamorph: Associated with *Phyllosticta crinicola* Siemaszko *fide* Siemaszko (op. cit.).

The type was not found in BPI, where some other Siemaszko types are preserved.

Sphaerella crithmi Nann. See *Mycosphaerella agostinii* (Nann.) M. Morelet.

Mycosphaerella crotalariae (Petch) Hansf., Proc. Linn. Soc. London 153: 21. 1942 = *Sphaerella crotalariae* Petch, Ann. Roy. Bot. Gard. (Peradeniya) 3: 2. 1906.

Type — Sri Lanka: Peradeniya. On pale spots on upper surface of living leaves of *Crotalaria striata* (Fabaceae). Petch no. 2256, VI 1905 (K, holotype; K, 2 isotypes); Same locality and host, III 1917 (IMI no. 59658, topotype, slides only).

Anamorphs: *Cercospora crotalariae* Petch (= *Cercospora apii* s.lat.) and *Phyllosticta crotalariae* Petch *vide* Petch (op. cit.).

The iso- and topotypes contain only an *Ascochyta*, with 1-septate conidia, that fits the protologue.

Mycosphaerella cruchetii M. Morelet, Bull. Soc. Sci. Nat. Archéol. Toulon & Var 176: 7. 1968, nomen novum (Article 58) for *Mycosphaerella syringae* Cruchet, Bull. Soc. Vaud. Sci. Nat. 55: 39. 1923.

Type — Switzerland: *Syringa vulgaris* (Oleaceae).

The type was not included in a loan from PC; its preservation is uncertain.

Mycosphaerella cruciatae (Lambotte & Fautrey) Tomilin, Novosti Sist. Nizsh. Rast. 8: 150. 1971 = *Sphaerella cruciatae* Lambotte & Fautrey in Fautrey & Lambotte, Rev. Mycol. (Toulouse) 17: 170. 1895.

Type — France: On dead stems of *Galium cruciata* (Rubiaceae). Fautrey, distributed in Roumeguère, Fungi Selecti Exsiccati no. 6884, IV 1895 (PC, holotype).

This is a *Didymella*, with paraphyses present, asci cylindrical, ascospores 18-22 × 4.5-5.5 µm.

Mycosphaerella cruciferarum (Fr.) Lindau, in Engl. & Prantl, Natürlichem Pflanzenf. 1(1): 424. 1897 = *Sphaeria cruciferarum* Fr., Systema Mycol. 2: 525. 1823 = *Sphaerella cruciferarum* (Fr.) Sacc., Michelia 2: 315. 1881 = *Carlia cruciferarum* (Fr.) Höhn., Ber. Deut. Bot. Ges. 36: 314. 1918.— Fig. 225.

Type — France: On dead stems of *Cheiranthus cheiri* (Brassicaceae). Mougeot (UPS-FRIES, holotype).

The holotype is immature, as already noticed by von Arx and Corlett, who both studied the holotype before. Additional material studied (Netherlands, Nunspeet, on dead stems of *Sisymbrium* ["*Erysimum*"] *officinale* (Brassicaceae), Beins no. 454, IX 1901, L) belongs to *Davidiella*, and represents a small-spored form of *D. allicina*, with asci pyriform, ascospores 12-15 × 4-5 µm.

Sphaerella cruciferarum f. *alliariae* Rabenh., Fungi Europaei Exsiccati, ed. nov., ser. 2 no. 1853. 1874.— Fig. 226.

Type — Czech Republic: Brno ["Brünn"]. On dead stems of *Alliaria petiolata* (Brassicaceae). Niessl, distributed in Rabenhorst, Fungi Europaei Exsiccati, ed. nov., ser. 2 no. 1853 (L, isotype).

Synonymised with *Mycosphaerella cruciferarum* by von Arx (1949). This belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores 15-18 × 3.5-4.5 µm.

Sphaerella cruciferarum f. *lepidii-graminifolii* Roum., Fungi Gallici Exsiccati no. 572.

Type — France: Pyrenées Orientales, Ille. On dead stems of *Lepidium graminifolium* (Brassicaceae). Roumeguère, VI 1879, distributed in Fungi Selecti Gallici Exsiccati no. 572 (L, isotype).

The isotype studied is immature.

Sphaerella cruenta Sacc., nomen herbariorum (not validly published, Article 32).

Authentic material — Italy: On pale spots with brown margins on upper surface of living leaves of *Polygonatum multiflorum* (Asparagaceae).

This is a coelomycete, probably an *Ascochyta*.

Mycosphaerella cruenta (Sacc.) Latham, Mycologia 26: 525. 1934, based on an anamorph (illegitimate, Article 59) = *Cercospora cruenta* Sacc., Michelia 2: 149. 1880 = *Pseudocercospora cruenta* (Sacc.) Deighton, Mycol. Pap. 140: 142. 1976.

Type — Italy: *Phaseolus* (Fabaceae).

Anamorph: *Pseudocercospora cruenta* (Sacc.) Deighton *vide* Tomilin (1979).

Mycosphaerella crus-galli (Ellis & Kellerm.) Lindau, in Engl. & Prantl, Natürlichem Pflanzenf. 1(1): 426. 1897 [as "*cruris-galli*?"] = *Sphaerella crus-galli* Ellis & Kellerm., J. Mycol. 3: 105. 1887.— Fig. 227.

Type — USA: Kansas, Manhattan. On upper and lower surface of dead leaves of *Panicum crus-galli* (Poaceae). Kellerman & Swingle no. 1067, VIII 1887 (NY, holotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores 17-20 × 4.5-6 µm.

Mycosphaerella cryptica (Cooke) Hansf., Proc. Linn. Soc. New South Wales 81: 35. 1956 = *Sphaerella cryptica* Cooke, Grevillea 20: 5. 1891.

Type — Australia: Victoria, Melbourne. On red-brown spots with purple margins on upper surface of dead leaves of *Eucalyptus* (Myrtaceae). Martin 753, IV 1977 (K, holotype; IMI 59211, NY, isotypes); also Martin 753 (IMI 59212, paratype).

Anamorph: *Colletogloeopsis nubilosum* (Ganapathi & Corbin) Crous & M.J. Wingf. *vide* Crous (1998).

This is a parasitic species, with asci cylindrical to clavate, ascospores 10-12 × 2.5-3.5 µm.

Mycosphaerella cryptomeriae Shiro & Hara, in Hara, J. Pl. Protect. 5: 462. 1918.— Fig. 228.

Type — Japan. On dead needles of *Cryptomeria japonica* (Cupressaceae). Hara, X 1911 (TNS 209263, holotype).

Type and additional material seen (Hara, TNS 209887) show that this is morphologically indistinguishable from *Mycosphaerella juniperina*, with ascomata thick-walled, breaking through the epidermis, ascospores 8-10 × 2.5-3.5 µm.

Sphaerella cryptotaeniae Henn., Bot. Jahrb. Syst. 37: 161. 1905.

Type — Japan: *Cryptotaenia japonica* (Apiaceae).

No material was studied; the type has probably been destroyed in B.

Mycosphaerella crystallina Crous & M.J. Wingf., Mycologia 88: 451. 1996.

Type — South Africa: Kwazulu-Natal Province, Umvoti. On leaves of *Eucalyptus bicostata* (Myrtaceae). Wingfield, X 1994 (PREM 51922, holotype, not seen).

Anamorph: *Pseudocercospora crystallina* Crous & M.J. Wingf. *vide* Crous (1998).

No material was studied of this recently described species.

Mycosphaerella cuboniana (D. Sacc.) Tomilin, Novosti Sist. Nizsh. Rast. 6: 119. 1970 [“1969”] = *Sphaerella cuboniana* D. Sacc., in Sacc. & D. Sacc., Syll. Fung. 17: 638. 1905.

Type — Italy: *Vitis vinifera* (Vitaceae).

No material was studied; it is no *Mycosphaerella* according to Tomilin (1979).

Mycosphaerella cucumis (Fautrey & Roum.) W.F. Chiu & J.C. Walker, J. Agric. Res. 78: 98. 1949, based on an anamorph (illegitimate, Article 59) = *Ascochyta cucumis* Fautrey & Roum., Rev. Mycol. (Toulouse) 13: 79. 1891.— Fig. 229.

Type — France: *Cucurbita* (Cucurbitaceae).

Anamorph: *Ascochyta cucumis* Fautrey & Roum. *vide* Chiu & Walker (op. cit.).

The type was not included in a loan form PC. Material studied (Surinam, Groningen, on upper surface of dead leaves of *Citrullus vulgaris*, van Hoof, X 1961, CBS) is *Didymella bryoniae* (Auersw.) Rehm, with asci cylindrical, surrounded by paraphyses, ascospores 12-15 × 4.5-5.5 µm, which is the current name for the teleomorph of *Ascochyta cucumis* Fautrey & Roum.

Mycosphaerella cucurbitae (Rostr.) Lind, Danish Fungi: 209. 1913 = *Sphaerella cucurbitae* Rostr., Plantepatologie: 456. 1902.— Fig. 230.

Type — Denmark: Lyngby. On white spots on upper surface of living leaves of *Cucurbita pepo* (Cucurbitaceae). Rostrup, IX 1897 (C, holotype).

Although on the type was annotated in 1945 by Munk that no ascomata are present, some overmature ascomata were found, showing that this is morphologically indistinguishable from *Didymella bryoniae* (Auersw.) Rehm, with ascospores 10-12 × 4.5-5 µm.

Sphaerella cucurbitacearum (Fr.) Cooke, J. Bot. 21: 71. 1883 = *Sphaeria cucurbitacearum* Fr., Systema Mycol. 2: 502. 1823 = *Laestadia cucurbitacearum* (Fr.) Sacc., Syll. Fung. 2: XXXIII. 1883.

Type — Sweden: Cucurbitaceae.

No material was studied as this species was already reported to have non-septate ascospores by Saccardo (op. cit.).

Mycosphaerella cunninghamiae Woron., Vestn. Tiflissk. Bot. Sada 28: 16. 1913 = *Sphaerella cunninghamiae* (Woron.) Trotter, Syll. Fung. 24: 858. 1928.

Type — Georgia: *Cunninghamia lanceolata* [“*sinensis*”] (Taxodiaceae).

Authentic material studied (Emergent through the epidermis of dead needles of *Cunninghamia japonica*, LEP 1764) belongs to section *Caterva*, with asci cylindrical, ascospores 18-21 × 3-3.5 µm.

Mycosphaerella cunninghamii Syd., Ann. Mycol. 22: 295. 1924.— Fig. 232.

Type — New Zealand: *Rubus australis* (Rosaceae).

No material was studied, as the type was not found in S or in any of the herbaria consulted.

Mycosphaerella cupaniae (Rehm) Arx, in E. Müll. & Arx, Beitr. Kryptogamenfl. Schweiz 11(2): 358. 1962 = *Lizonia cupaniae* Rehm, Ann. Mycol. 5: 527. 1907 = *Lizoniella cupaniae* (Rehm) Sacc. & Trotter, Syll. Fung. 22: 165. 1913 = *Pseudosphaerella cupaniae* (Rehm) Höhn., Sitzungsber. Kaiserl. Akad. Wiss., Math.-Naturwiss. Kl., Abt. 1, 120: 426. 1911.

Type — Brazil: *Cupania* (Sapindaceae).

No material was studied, as the type was not found in S or in any of the herbaria consulted.

Mycosphaerella cuprea (Sacc.) Siemaszko, in Woronow, Trudy Tiflissk. Bot. Sada, ser. 2, 3: 136. 1923 = *Sphaerella cuprea* Sacc., Ann. Mycol. 7: 432. 1909.— Fig. 233.

Type — Italy: Napoli, Botanical garden. Emergent through the epidermis on the upper surface of dead leaves of *Ceratonia siliqua* (Fabaceae). Cufino, V 1904 (PAD, holotype).

The type belongs to section *Caterva*, and this is morphologically indistinguishable from *M. subradians*, with asci cylindrical, ascospores 12-14 × 3-4 µm. Additional material studied (Spain, Ibiza, Cala Llonga, van der Aa no. 7827, IV 1981, CBS) belongs to section *Longispora*, and is morphologically indistinguishable from *M. millegrana*, with asci cylindrical to narrowly pyriform, ascospores 12-15 × 2-3 µm.

Sphaerella curva P. Karst., Hedwigia 22: 179. 1883.

Type — Finland: *Ribes alpinum* (Grossulariaceae).

No material was studied, as the type was not found in H or in any of the herbaria consulted.

Mycosphaerella curvulata (Pass.) Tomilin, Novosti Sist. Nizsh. Rast. 1967: 188. 1967 = *Sphaerella curvulata* Pass., in Rabenh., Hedwigia 15: 106. 1876.

Type — Italy: Parma, superficial on the lower surface of dead leaves of *Salix alba* (Salicaceae). Passerini, distributed in Rabenhorst, Fungi Europaei Exsiccata, ed. nov., ser. 2 no. 2052 (CBS, isotype).

The isotype studied is immature, but the loosely grouped, superficial ascomata agree macroscopically with *Venturia chlorospora* (Ces.) P. Karst.

Mycosphaerella cussoniae Crous & M.J. Wingf., Mycotaxon 46: 22. 1993.

Type — South Africa: *Cussonia thyrsoiflora* (Araliaceae).

No material was studied of this recently described species.

Mycosphaerella cuttsiae Sivan. & R.G. Shivas, Mycol. Res. 106: 355. 2002.

Type — Australia: *Cuttsia* (Grossulariaceae).

No material was studied of this recently described species.

Sphaerella cyananthi Pat., Rev. Mycol. (Toulouse) 8: 180. 1886.— Fig. 234.

Type — China: Yunnan. On stems of *Cyananthus* (Gesneriaceae). Delavay (FH-Patouillard, holotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores $15-18 \times 5.5-7 \mu\text{m}$.

Mycosphaerella cyanae F. Stevens & K. Young, in F. Stevens, Bernice P. Bishop Mus. Bull. 19: 101. 1925.

Type — USA: *Cyanea angustifolia* (Lobeliaceae).

No material was studied, as the type was not found in BPI or NY or in any of the other herbaria consulted.

Sphaerella cyclogena Sacc. & Paol., Atti Reale Ist. Veneto Sci., ser. 6, 6: 408. 1888.

Type — Indonesia. On pale spots with dark margins on upper surface of living leaves of an herbaceous plant with quadrangular stems (possibly Verbenaceae). Paoletti (PAD, holotype).

The type contains only old material of a coelomycete.

Mycosphaerella cydoniae Grove, J. Bot. 56: 285. 1918.— Fig. 235.

Type — United Kingdom: Herefordshire, Hereford. On upper and lower surface of dead leaves of *Cydonia vulgaris* (Rosaceae). Grove, 3 May 1918 (K, lectotype, here designated; 17 May, K, topotype).

Both lecto- and topotype show that this is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores $7-9 \times 2-2.5 \mu\text{m}$.

Mycosphaerella cydoniae (Vogolino) Kirchn., Die Krankheiten und Beschädigungen unserer landwirtschaftlichen Kulturpflanzen, ed. 3: 492. 1923, later homonym (illegitimate, Article 53) = *Sphaerella cydoniae* Vogolino, Ann. Reale Accad. Agric. Torino 48: 430. 1906 [“1905”].

Type — Italy: *Cydonia vulgaris* (Rosaceae).

Cited as synonymous with *Davidiella allicina* (as *Mycosphaerella allicina*) by Tomilin (1979). No material was studied, as the type was not found in any of the herbaria consulted.

Mycosphaerella cynodontis (Unamuno) M. Morelet, Ann. Soc. Sci. Nat. Archéol. Toulon & Var 20: 105. 1968 = *Sphaerella cynodontis* Unamuno, Mem. Real Soc. Esp. Hist. Nat. 15: 345. 1929.

Type — Spain: *Cynodon dactylon* (Poaceae).

Cited as synonymous with *Mycosphaerella jenensis* by Tomilin (1979). No material was studied, as the type was not found in MA or in any of the herbaria consulted.

Mycosphaerella cyparissiae (Pass.) Tomilin, Novosti Sist. Nizsh. Rast. 6: 119. 1970 [“1969”] = *Sphaerella cyparissiae* Pass., Atti Reale Accad. Lincei, Mem. Cl. Sci. Fis., ser. 4, 3: 8. 1887.

Type — Italy: *Euphorbia cyparissias* (Euphorbiaceae). No material was studied, as the type was not found in any of the herbaria consulted.

Mycosphaerella cyparissincola Petr., Sydowia 10: 287. 1957 [“1956”].— Fig. 236.

Type — Czech Republic: Weißkirchen, Hrabuvka. On dead stems of *Euphorbia cyparissias* (Euphorbiaceae). Petrak,

distributed in Reliquiae Petrakianae no. 2472, VI 1917 (H, L, isotypes).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradicans*, with asci cylindrical, ascospores $9-11 \times 2.5-3 \mu\text{m}$. It was cited as synonymous with *M. montellica* by Tomilin (1979), but that is morphologically indistinguishable from *D. ammophilae*.

Mycosphaerella cyperi Sivan. & R.G. Shivas, Mycol. Res. 106: 357. 2002.

Type — Australia: *Cyperus porta-tartarus* (Cyperaceae).

No material was studied of this recently described species.

Mycosphaerella cypripedii (Peck) House, New York State Mus. Bull. 233-234: 27. 1921 = *Sphaerella cypripedii* Peck, Annual Rep. New York State Mus. 51: 296. 1898 [“1897”].

Type — USA: *Cypripedium* (Orchidaceae).

Cited as synonymous with *Davidiella allicina* (as *Mycosphaerella allicina*) by Tomilin (1979). No material was studied, as the type was not found in BPI or NY or in any of the other herbaria consulted.

Mycosphaerella cytisi-sagittalis (Auersw.) Lindau, in Engl. & Prantl, Natürlichen Pflanzenf. 1(1): 425. 1897 = *Sphaerella cytisi-sagittalis* Auersw., in Gonn. & Rabenh., Mycol. Europaea 5-6: 10. 1869.

Type — Austria: *Cytisus sagittalis* (Fabaceae).

No material was studied, as the type has probably been destroyed in B.

Mycosphaerella dacrydii Butin, Sydowia 27: 274. 1975 [“1973-1974”].— Fig. 237.

Type — Chile: Cordillera pelada. On lower surface of green, still living scales/needles of *Dacrydium foncki* (Podocarpaceae). Butin, III 1970 (ZT, holotype).

This is a species of Parmulariaceae, probably a *Cycloshizon*, with ascomata with meandric cells, asci clavate, with $3 \mu\text{m}$ thick walls and thick-walled ascospores surrounded by a gelatinous sheath. The type also contains a *Mycosphaerella*, but with much smaller ascospores than mentioned in the protologue, which might therefore have been based on two widely different fungi.

Mycosphaerella dactylidis (Pass.) Kirchn., Die Krankheiten und Beschädigungen unserer landwirtschaftlichen Kulturpflanzen, ed. 3: 156. 1923 = *Sphaerella dactylidis* Pass., Atti Reale Accad. Lincei, Mem. Cl. Sci. Fis., ser. 4, 3: 9. 1887.

Type — Italy: *Dactylis glomerata* (Poaceae).

Cited as synonymous with *Davidiella allicina* (as *Mycosphaerella allicina*) by Tomilin (1979). No material was studied, as the type was not found in any of the herbaria consulted.

Sphaerella dactylidis f. *matritensis* Gonz. Frag., Bol. Soc. Esp. Hist. Nat. 18: 366. 1918.— Fig. 238.

Type — Spain: Madrid, Moriello. On dead leaves of *Koeleria* [wrongly identified as “*Dactylis glomerata*”] (Poaceae). Cuestal no. 1670, I 1916 (MA, holotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores 16-19 × 6-7.5 μm.

Mycosphaerella dahliae (Cooke & Ellis) Coons, Rep. Michigan Acad. Sci. 14: 273. 1912 = *Sphaerella dahliae* Cooke & Ellis, Grevillea 7: 42. 1878.

Type — USA: New Jersey, Newfield. On dead stems of *Dahlia* (Asteraceae). Ellis no. 2988 (K, holotype).

Anamorph: Associated with *Colletotrichum* ["*Vermicularia*"] *compactum* fide Saccardo (1882).

The holotype contains only coelomycetes.

Mycosphaerella dalbergiae E. Müll. & S. Ahmad, Sydowia 9: 243. 1955.— Fig. 239.

Type — Pakistan: Lahore, Changa Manga. On lower surface of dead leaves of *Dalbergia sissoo* (Fabaceae). Ahmad, distributed in Reliquae Petrakianae no. 2252, IV 1961, (H, L, isotypes).

Anamorph: *Cercospora sissoo* Syd. fide Tomilin (1979) (= *Colletogloeum sissoo* (Syd.) B. Sutton).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. superflua*, with ascomata in groups of 1-7, asci cylindrical, ascospores 14-16 × 3-4 μm.

Mycosphaerella dalmatica (Picb.) Petr., Sydowia 9: 484. 1955 = *Dothidella dalmatica* Picb., Práce Morav. Přír. Společn. 7(12): 2. 1932.

Type — Croatia: Dalmatia, Popovici. On upper surface of dead leaves of *Globularia allyssum* (Plantaginaceae). Prostmigg, III 1930 (IMI no. 24194, isotype).

The isotype contains only a coelomycete.

Mycosphaerella danaeae Petr. & Cif., Ann. Mycol. 28: 390. 1930.— Fig. 240.

Type — Dominican Republic: prov. Azua, San Juan de la Maguana. On upper surface of dead leaf sectors of *Danaea nodosa* (Polypodiaceae). Ciferri, VIII 1929, Mycoflora Domingensis Exsiccata no. 172 (L, NY, isotypes).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores 14-16 × 4.5-5.5 μm. The isotype in NY contains only foliicolous lichens.

Mycosphaerella danica Tomilin, Novosti Sist. Nizsh. Rast. 11: 252. 1974, nomen novum (Article 58) for *Mycosphaerella lindiana* Munk, Dansk Bot. Ark. 17(1): 328. 1957, later homonym (illegitimate, Article 53).— Fig. 241.

Type — Denmark: Søndernshovgaard. On dead stems of *Chamaenerion* [as "*Epilobium*"] *angustifolium* (Onagraceae). Lind, III 1925 (C, holotype).

The holotype contains mostly *Discostroma tostum* (Berk. & Broome) Brockmann, but the presence of some ascomata belonging to *Davidiella* shows that it is morphologically indistinguishable from *D. ammophilae*, with asci pyriform, ascospores 8-10 × 3-3.5 μm.

Mycosphaerella danubialis O. Săvul., Bull. Sect. Sci. Acad. Roumaine 22: 308. 1940 = *Sphaerella danubialis* (O.

Săvul.) Sandu, Ciuperci Pyrenomycetes-Sphaeriales din România: 124. 1971.

Type — Romania: *Oryza sativa* (Poaceae).

Cited as synonymous with *Mycosphaerella oryzae* by Tomilin (1979). The preservation of the type is unknown; it was not included in the widespread exsiccate series.

Mycosphaerella daphnes I.E. Brezhnev, Bot. Mater. Otd. Sporov. Rast. Bot. Inst. Komarova Akad. Nauk SSSR 7: 185. 1951.

Type — Russia: *Daphne sophia* (Thymelaeaceae).

No material was studied, as the type was not included in loans from LE and LEP.

Mycosphaerella daphniphylli Syd. & Hara, in Syd. & P. Syd., Ann. Mycol. 11: 57. 1913 = *Sphaerella daphniphylli* (Syd. & Hara) Trotter, Syll. Fung. 24: 862. 1928.— Fig. 242.

Type — Japan: Satsuma, Kagoshima. On white spots with red margins on upper and lower surface of living leaves of *Daphniphyllum glaucescens* (Daphniphyllaceae). Kawagoi, V 1912 (S, holotype).

This is a parasitic species, with asci broadly cylindrical, ascospores 16-20 × 4-5 μm.

Mycosphaerella dauci Nevod., Bot. Mater. Otd. Sporov. Rast. Bot. Inst. Komarova Akad. Nauk SSSR 14: 169. 1961.

Type — Russia: *Daucus carota* (Apiaceae).

No material was studied, as the type was not included in loans from LE and LEP.

Mycosphaerella daviesiae Petr., Sydowia 8: 200. 1954.

Type — Australia: *Daviesia latifolia* (Fabaceae).

No material was studied, as the type was not found in W, nor in any of the herbaria consulted.

Mycosphaerella daviesicola Beilharz & Pascoe, Mycotaxon 82: 364. 2002.

Type — Australia: *Daviesia mimosoides* (Fabaceae).

Anamorph: *Verrucisporota daviesiae* (Cooke & Massee) Beilharz & Pascoe fide Beilharz & Pascoe (op. cit.).

No material was studied of this recently described species.

Mycosphaerella davisii F.R. Jones, Mycologia 36: 523. 1944.— Fig. 243.

Type — USA: Wisconsin, Madison, Olin Park. On dead stems and upper and lower surface of dead leaves of *Melilotus alba* (Fabaceae). Jones, VII 1942 (IMI no. 201401, isotype, slides only).

Anamorph: *Cercospora davisii* Ellis & Everh. fide Jones (op. cit.).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradians*, with asci cylindrical, ascospores 11-13 × 2.5-3.5 μm.

Mycosphaerella dealbans Tomilin, Novosti Sist. Nizsh. Rast. 1967: 188. 1967, nomen novum (Article 58) for *Sphaerella dealbans* Speg., Bol. Acad. Nac. Ci. 23: 469. 1919, later homonym (illegitimate, Article 53).— Fig. 244.

Type — Brazil: Apiahy. On dead stems of *Agapanthus umbellatus* (Alliaceae). Puiggari no. 261, VII 1890 (LPS, holotype).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradians*, with asci cylindrical, ascospores $10-12 \times 2-2.5 \mu\text{m}$. It was cited as synonymous with *Mycosphaerella pittieri* by Tomilin (1979).

Sphaerella dealbans Müll. Arg., Flora 56: 507. 1872 = *Pharcidia dealbans* (Müll. Arg.) Sacc. & D. Sacc., Syll. Fung. 17: 647. 1905.

Type — Spain. On thallus of *Endocarpon loscosi* (Ascomycota, Verrucariaceae).

Accepted as *Pharcidia dealbans* (Müll. Arg.) Sacc. & D. Sacc. by Clauzade, Diederich & Roux (1989).

Mycosphaerella dearnessii M.E. Barr, Contr. Univ. Michigan Herb. 9: 587. 1972, nomen novum (Article 58) for *Oligostroma acicola* Dearn., Mycologia 18: 251. 1926 = *Scirrhia acicola* (Dearn.) Sigg., Phytopathology 29: 1076. 1939 = *Systemma acicola* (Dearn.) F.A. Wolf & Barbour, Phytopathology 31: 70. 1941 = *Eruptio acicola* (Dearn.) M.E. Barr, Mycotaxon 60: 438. 1996 non *Mycosphaerella acicola* (Cooke & Harkn.) Lindau (1897).

Type — USA: *Pinus* (Pinaceae).

Anamorph: *Lecanosticta acicola* (Thüm.) Syd. fide Barr (op. cit.).

The type was not studied. Material studied: China, Fujian Province, on *P. elliotii*, Zhu, VI 1985 (IMI no. 300262). According to the recent redescription by Barr (1996), this is morphologically indistinguishable from *Mycosphaerella juniperina*.

Mycosphaerella decidua (Ellis & Kellerm.) Tomilin, Novosti Sist. Nizsh. Rast. 8: 150. 1971 = *Sphaerella decidua* Ellis & Kellerm., Bull. Torrey Bot. Club 11: 122. 1884.— Fig. 245.

Type — USA: Kansas, Manhattan. On pale spots with dark margins on lower surface of living leaves of *Vernonia* (Asteraceae). Kellerman no. 556, VI 1884 (NY, holotype).

This is a parasitic species of *Davidiella*, with asci broadly cylindrical, ascospores thick-walled, $14-17 \times 6-7.5 \mu\text{m}$. Therefore the following new combination is made:

Davidiella decidua (Ellis & Kellerm.) Aptroot comb. nov., **MB 500348**. **Basionym:** *Sphaerella decidua* Ellis & Kellerm., Bull. Torrey Bot. Club 11: 122. 1884. The specimen cited is the holotype rather than a syntype as reported by Barr, Huhndorf & Rogerson (1996), because the other specimen is from a different host and represents the authentic material of a different (albeit unpublished) taxon, see below.

Sphaerella decidua var. *scrophulariae* Ellis & Kellerm., nomen herbariorum (not validly published, Article 32).

Authentic material — USA: Kansas, Manhattan. On pale spots with dark margins on upper surface of living leaves of *Scrophularia nodulosa* (Scrophulariaceae). Kellerman no. 563, VI 1884 (NY).

Mycosphaerella degeneri Petr., Sydowia 7: 389. 1953.— Fig. 246.

Type — Hawaii: Molokai, Mokomoko. On lower surface of living leaves, not in spots, but surrounded by a purplish stain, of *Euphorbia* (Euphorbiaceae). Degener & Wiebke, distributed in Reliquiae Petrakianae no. 1852, VI 1928 (H, L, isotypes).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores $16-21 \times 4-6.5 \mu\text{m}$.

Mycosphaerella deightonii M. Morelet, Bull. Soc. Sci. Nat. Archéol. Toulon & Var 29: 9. 1973.— Fig. 247.

Type — USA: New Jersey, Newfield. On dead parts of leaves of *Yucca filamentosa* (Asparagaceae). Ellis no. 2150, VI 1874 (NY, holotype, with *Cercospora concentrica*; NY isotype, slide only).

Anamorph: *Stigmina concentrica* (Cooke & Ellis) Deighton fide Morelet (op. cit.) (= *Pseudocercospora concentrica* (Cooke & Ellis) U. Braun & Crous).

This is morphologically indistinguishable from *Planistromella acervata* (Ellis & Everh.) M.E. Barr, with ascomata grouped in erumpent stromata, asci cylindrical, surrounded by large parenchymatic cells, ascospores $23-29 \times 4.5-6 \mu\text{m}$.

Mycosphaerella dejanira (Sacc.) Tomilin, Novosti Sist. Nizsh. Rast. 1968: 165. 1968 = *Sphaerella dejanira* Sacc., Nuovo Giorn. Bot. Ital. 7: 300. 1875.

Type — Italy: On pale spots with brown margins on upper surface of living leaves of *Aruncus dioicus* ["*Spiraea aruncus*"] (Rosaceae). VIII and IX (PAD, syntypes)

Anamorph: *Phyllosticta arunci* Sacc. fide Saccardo (1882).

Both type specimens contain only coelomycetes.

Mycosphaerella delegatensis R.F. Park & Keane, Trans. Brit. Mycol. Soc. 83: 95. 1984.— Fig. 248.

Type — Australia: Victoria, Narbethong. On spots on upper and lower surface of living leaves of *Eucalyptus delegatensis* (Myrtaceae). Park 1, VI 1983 (IMI no. 280471, isotype); also Park 2 (IMI 280472, isotype of the anamorph).

Anamorph: *Stagonospora delegatensis* R.F. Park & Keane fide Park & Keane (op. cit.), *Phaeophleospora delegatensis* (R.F. Park & Keane) Crous fide Crous (1998).

This is a parasitic species, with asci cylindrical, ascospores $17-25 \times 4.5-5.5 \mu\text{m}$.

Mycosphaerella delphinii Tomilin, Bot. Mater. Otd. Sporov. Rast. Bot. Inst. Komarova Akad. Nauk SSSR 12: 271. 1959.

Type — Russia: *Delphinium cuneatum* (Ranunculaceae).

Cited as synonymous with *Mycosphaerella delphiniicola* by Tomilin (1979). No material was studied, as the type was not included in loans from LE and LEP.

Mycosphaerella delphiniicola Earle, in Greene, Plantae Bakerianae 2(1): 19. 1901 = *Sphaerella delphiniicola* (Earle) Sacc. & D. Sacc., Syll. Fung. 16: 1133. 1902.

Type — USA: Colorado, Pagosa Peak. On dead stems of *Delphinium* (Ranunculaceae). Baker no. 37, VIII 1899 (K, NY (2×), isotypes).

The isotypes contain various postmature ascomycetes.

Mycosphaerella dendrobii-nobilis Katum., Trans. Mycol. Soc. Japan 24: 268. 1983.— Fig. 250.

Type — Japan: *Dendrobium nobile* (Orchidaceae).

The type was not included in a loan from TNS. Material seen (Thailand, intercepted at San Francisco, on white spots with black margins on upper and lower surface of living leaves of *Dendrobium* sp., Torbett, III 1986, BPI) is a parasitic species, with asci cylindrical, ascospores 9-12 × 2-3.5 µm.

Mycosphaerella dendroides (Schwein.) Demaree & Cole, J. Agric. Res. 40: 785. 1930 ≡ *Sphaeria dendroides* Schwein., Schriften Naturf. Ges. Leipzig 1: 47. 1822 ≡ *Sphaerella dendroides* (Schwein.) Cooke, J. Bot. 21: 108. 1883.— Fig. 251.

Type — USA: Philadelphia. On lower surface of dead leaves of *Carya* (Juglandaceae). Schweinitz (K, isotype).

No type material was preserved in PH, the isotype in K is postmature, but probably represents *M. punctiformis*. Additional material (North Carolina, Ravenel, Fungi Caroliniana no. 61, BPI, 2×) belongs to section *Longissima*, and is morphologically indistinguishable from *M. latebrosa*, with asci cylindrical, ascospores 23-26 × 3-4 µm.

Mycosphaerella dendromeconis (Cooke & Harkn.) Earle, in Baker, West Amer. Plants, West Coast Ser. 1, Fungi. Pacific Slope Fungi Exsiccati: 4 no. 657. 1902 ≡ *Sphaerella dendromeconis* Cooke & Harkn., Grevillea 9: 9. 1880.— Fig. 249.

Type — USA: California. Mt. Jamalpais. On dead stems of *Dendromecon rigidum* (Papaveraceae). Harkness no. 1386, V 1880 (BPI, holotype; BPI, isotype), also distributed in Ellis, North American Fungi no. 798 (BPI, L, isotypes).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores 14-15.5(-18) × 4-6 µm.

Mycosphaerella denigrans (Kirschst.) Tomilin, Opredeletel' gribov roda *Mycosphaerella* Johans.: 77. 1979 ≡ *Sphaerella denigrans* Kirschst., Ann. Mycol. 37: 103. 1939.— Fig. 252.

Type — Germany: Königstein. On dead stems of *Erysimum hieraciifolium* (Brassicaceae). Krieger, VII 1895 (B, holotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. ammophilae*, with asci pyriform, ascospores 10-12 × 2.5-3 µm.

Mycosphaerella dennettiae Sivan. & Okpala, Trans. Brit. Mycol. Soc. 72: 520. 1979.— Fig. 253.

Type — Nigeria. Breaking through the epidermis of black spots on lower surface of living leaves of *Dennettia tripetala* (Annonaceae). Okpala, VI 1977 (IMI no. 21688, isotype).

This is a parasitic species, with asci cylindrical, ascospores 17-21 × 3.5-4.5 µm.

Mycosphaerella densa (Rostr.) Lind, Rep. Sci. Results Norw. Exped. Novaya Zemlya 1921, 19: 12. 1924 ≡ *Sphaerella densa* Rostr., Bot. Tidsskr. 14: 225. 1885.— Fig. 254.

Type — Iceland: Reykjavik. On upper and lower surface of dead leaves of *Arenaria norvegica* (Caryophyllaceae). Graülen, VII 1868 (C, holotype).

Anamorph (presumed): *Ramularia fide* Corlett (1988).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores 14-18 × 3.5-5.5 µm.

Mycosphaerella depazeaeformis (Auersw.) Lindau, Hilfsb. Sammeln Ascomyceten: 72. 1903 ≡ *Sphaeria depazeaeformis* Auersw., in Rabenh., Herb. Vivum Mycol. no. 1641. 1852 ≡ *Sphaerella depazeaeformis* (Auersw.) Ces. & De Not., Comment. Soc. Crittog. Ital. 1(4): 238. 1863.— Fig. 255.

Type — Switzerland: Uetewalde. On spots on upper and lower surface of living leaves of *Oxalis acetosella* (Oxalidaceae). Auerswald, VII 1851, distributed in Klotzsch, Herbarium Vivum Mycologicum, ed. 1 no. 1641 (B (2×), L, isotypes).

Isotypes and additional material studied (Austria, Lunz, Petrak, distributed in Reliquiae Petrakianae no. 650, VII 1939, L), belong to section *Plaga*, with asci cylindrical, ascospores 12.5-14.5(-16) × 2.5-3.5 µm.

Mycosphaerella depressa (Berk.) Theiss., Verh. Zool.-Bot. Vereins Wien 66: 375. 1916 ≡ *Sphaeria depressa* Berk., in Hooker, Bot. Antarctic Voy. I, Fl. Antarctica: 172. 1880 ≡ *Physalospora depressa* (Berk.) Sacc., Syll. Fung. 1: 444. 1882 ≡ *Sphaerella depressa* (Berk.) Cooke, J. Bot. 21: 137. 1883, later homonym (illegitimate, Article 53).— Fig. 256.

Type — New Zealand: Campbell Island. On outer surface of dead leaves of *Luzula crinita* (Juncaceae). Hooker (K, holotype).

This is a *Phaeosphaeria*, with ascospores 3-septate, fusiform, pale brown, 35-40 × 5-7 µm.

Mycosphaerella depressa (Peck) House, New York State Mus. Bull. 233-234: 27. 1921, later homonym (illegitimate, Article 53) ≡ *Sphaerella depressa* Peck, Annual Rep. New York State Mus 33: 34. 1883 [“1880”] ≡ *Hyponectria depressa* (Peck) M.E. Barr, Mycologia 69: 961. 1977.

Type — USA: *Mulgedium* (Asteraceae).

No material studied, as it is accepted as *Hyponectria depressa* (Peck) M.E. Barr by Barr (op. cit.).

Mycosphaerella deschampsiae R. Sprague, Mycologia 46: 78. 1954.

Type — USA: *Deschampsia atropurpurea* (Poaceae).

Cited as synonymous with *M. recutita*, which is morphologically indistinguishable from *Davidiella disseminata*, by Tomilin (1979). No type material was preserved in FH, and no material was found in any of the herbaria consulted.

Mycosphaerella deschmannii (Voss) Lind, Danish Fungi: 209. 1913 ≡ *Sphaerella deschmannii* Voss, Mycol. Carniolica 3. Ascomycetes: 156. 1891.

Type — Yugoslavia: On upper and lower surface of dead (reddish) part of living leaves of *Gentiana pneumonanthe* (Gentianaceae).

The type was not found in M. Material examined (Denmark, Jutland, Frederikshavn, Lind, VII 1906, distributed in Vestergren, Micromycetes Rariores Selecti no. 1231, M) contains only a rust.

Mycosphaerella desmazieri (Mont.) Jaap, Verh. Bot. Vereins Prov. Brandenburg 64: 23. 1922 = *Sacidium desmazieri* Mont., in Desm., Plantes Cryptog. France, ed. 2 no. 351. 1856 = *Sphaerella desmazieri* (Mont.) Sacc., Syll. Fung. 11: 301. 1895.— Fig. 257.

Type — France: On upper and lower surface of dead leaves of *Iris pseudacorus* (Iridaceae). Desmazières, Plantes Cryptogames de France no. 351 (B-Desmazières, isotype). The isotype studied is immature. The species was synonymised with *Mycosphaerella iridis* by von Arx (1949). Additional materials studied (Germany, Ostprignitz, Triglitz, Jaap, IV 1905, B; also Westhavelland, Kirschstein, IV 1911, B) both belong to section *Caterva*, and are specimens of *M. subradicans*, with asci cylindrical, ascospores 9-10 × 2.5-3 µm.

Mycosphaerella desmodii (G. Winter) Tomilin, Novosti Sist. Nizsh. Rast. 1968: 165. 1968 = *Sphaerella desmodii* G. Winter, J. Mycol. 1: 121. 1885.

Type — USA: *Desmodium canescens* (Fabaceae). Cited as synonymous with *M. subradicans* by Tomilin (1979). No type material was preserved in B. Material studied (USA, Arkansas, Little Rock, on pale spots with brown margins on upper surface of living leaves of *D. viridiflorum*, Merrill no. 748, VIII 1938, BPI) is immature.

Mycosphaerella desmodiifolii Bat. & Peres, in Bat., Peres & Poroca, Atas Inst. Micol. 5: 90. 1967.— Fig. 258.

Type — Jamaica. On white spots on upper and lower surface of living leaves of *Desmodium* (Fabaceae). Batista exs. no. 20155 (URM 28224, holotype). Associated with *Phyllosticta fide* Batista & Peres (op. cit.). This is a parasitic species of *Davidiella*, morphologically indistinguishable from *D. ariadna*, with asci pyriform, ascospores 9.5-10 × 3-3.5 µm.

Sphaerella destructiva Berk. & Broome, Trans. Linn. Soc. London, ser. 2, 2: 71. 1882 = *Laestadia destructiva* (Berk. & Broome) Berl. & Voglino, Syll. Fung., Addit. ad vol. 1-4: 62. 1886.

Type — Australia: Queensland, Brisbane. On upper and lower surface (not in spots) of living leaves of *Medicago sativa* (Fabaceae). Bailey (BPI, isotype). The isotype is immature, but shows that this is morphologically indistinguishable from *Glomerella cingulata* (Stoneman) Spauld. & H. Schrenk, for which it would represent an older epithet.

Mycosphaerella deutziae Syd., Ann. Mycol. 19: 138. 1921 = *Sphaerella deutziae* (Syd.) Kirschst., Kryptog.-Fl. Mark Brandenburg 7(3): 340. 1938.— Fig. 259.

Type — Germany: Brandenburg, Tamsel, Arboretum. On upper and lower surface of dead leaves of *Deutzia lemoinae* (Saxifragaceae). Vogel, V 1915, distributed in Sydow, Mycotheca Germanica no. 1539 (L, isotype).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradicans*, with asci cylindrical, ascospores 8-11 × 2-2.5 µm. It was cited as synonymous with *Mycosphaerella parallelogramma* by Tomilin (1979).

Mycosphaerella devia Petr. & Cif., Ann. Mycol. 30: 210. 1932.— Fig. 260.

Type — Dominican Republic: Rio Ozame. On small brown spots with black margins on upper surface of living leaves of *Dalbergia ecastiphylla* (Fabaceae). Ciferri, 3485, XII 1929 (W, holotype).

Spermatial state: *Asteroma fide* Tomilin (1979).

This is a parasitic species, with asci cylindrical, ascospores 14-17 × 3-4.5 µm.

Mycosphaerella dianellae F. Stevens & Weddon, in F. Stevens, Bernice P. Bishop Mus. Bull. 19: 102. 1925.

Type — USA: *Dianella odorata* (Xanthorrhoeaceae). No material was studied, as the type was not found in BPI or NY or in any of the other herbaria consulted.

Mycosphaerella dianellincola Petr., Sydowia 9: 563. 1955.— Fig. 261.

Type — Australia: New South Wales, Wyalong. On upper surface of dead leaves of *Dianella revoluta* (Xanthorrhoeaceae). Gauba, V 1955, distributed in Reliquiae Petrakianae no. 2685 (H, L, B, isotypes).

The isotype in B shows that this belongs to *Davidiella*, and is morphologically indistinguishable from *D. ammophilae*, with asci pyriform, ascospores 9-11 × 2.5-3 µm. The isotypes in H and L contain only a *Diplodia* coelomycete. It was cited as synonymous with *M. petchii* (as *M. zeylanica*) by Tomilin (1979).

Mycosphaerella dianthi (C.C. Burt) Jørst., Meld. Statens Plantepatol. Inst. 1, C: 17. 1945 = *Didymellina dianthi* C.C. Burt, Trans. Brit. Mycol. Soc. 20: 214. 1936 = *Davidiella dianthi* (C.C. Burt) Crous & U. Braun, in U. Braun *et al.*, Mycol. Progress 2: 10. 2003.

Type — United Kingdom. On brown spots on upper surface of living leaves of *Dianthus barbatus* (Caryophyllaceae). Anamorph: *Cladosporium echinulatum* (Berk.) G.A. de Vries *fide* de Vries (1952).

Material studied: United Kingdom, Canterbury, University of Kent, on *D. caryophyllus*, III 1984 (IMI no. 284361 and no. 284362); also Jersey, Trinity, Howard Davis Farm, Bradshaw, IX 1980 (IMI no. 251383); also Isle of Man, Ramsey, on *Dianthus* "Sweet William", Hopkins, IV 1971 (IMI no. 169508). All materials studied contain only the anamorph, *Cladosporium echinulatum*.

Mycosphaerella dichrostachydis van der Byl, S. African J. Sci. 25: 186. 1928.

Type — Zimbabwe: *Dichrostachys nyassana* (Fabaceae). No material was studied, as the type was not found in B or in any of the other herbaria consulted.

Mycosphaerella dictamni Petr., Sydowia 1: 145. 1947.— Fig. 262.

Type — Austria: Hainburg, Hundsheimerkogel. On upper and lower surface of dead leaves of *Dictamnus alba*

(Rutaceae). Petrak, V 1940 (W, lectotype, here designated; W, isolectotype).

Spermatial state and anamorph: *Asteromella dictamni* Petr. and *Septoria dictamni* Fuckel *vide* Tomilin (1979).

This belongs to section *Longispora*, and is morphologically indistinguishable from *M. latebrosa*, with asci cylindrical, ascospores 22-24 × 2.5-4 µm.

Mycosphaerella didymelloides Petr., Bot. Jahrb. Syst. 62, Beibl. 142: 118. 1928.— Fig. 263.

Type — Madeira: Funchal. Mostly immersed in upper and lower surface of dead leaves of *Eucalyptus globulus* (Myrtaceae). Ade, VI 1926 (W, holotype; W, isotype).

Cited as synonymous with *M. molleriana* by Tomilin (1979), but excluded from *Mycosphaerella* by Crous (1998). This is indeed no *Mycosphaerella*, but probably a *Didymella*, with pseudoparaphyses, asci cylindrical, ascospores 9-11 × 3-4 µm.

Mycosphaerella didymopanacis Miles, Trans. Illinois Acad. Sci. 10: 249. 1917 = *Sphaerella didymopanacis* (Miles) Trotter, Syll. Fung. 24: 852. 1928.

Type — Puerto Rico: Anasco. On white spots with brown margins on upper and lower surface of living leaves of *Didymopanax mortonii* (Araliaceae). Stevens no. 3591, X 1913 (NY, isotype).

This is a parasitic species, but the material is badly preserved and overmature.

Mycosphaerella dieffenbachiae (Gonz. Frag. & Cif.) Cif., Quaderno 19: 232. 1961 = *Sphaerella dieffenbachiae* Gonz. Frag. & Cif., in Cif. & Gonz. Frag., Bol. Soc. Esp. Hist. Nat. 27: 326. 1927.— Fig. 264.

Type — Dominican Republic: *Dieffenbachia seguinea* (Araceae).

The type was not found in BPI or MA or in any of the other herbaria consulted. Material studied (Cuba, Bayamo, on upper and lower surface of dead leaf parts of living leaves of *D. picta*, Urtiaga no. F635, VI 1967, IMI no. 128250) belongs to *Davidiella*, and is morphologically indistinguishable from *D. ammophilae*, with asci pyriform, ascospores 7-9 × 2.5-3.5 µm.

Mycosphaerella digitalis (Ferraris) Tomilin, Novosti Sist. Nizsh. Rast. 8: 150. 1971 = *Sphaerella digitalis* Ferraris, Malpighia 16: 451. 1902.

Type — Italy: *Digitalis lutea* (Plantaginaceae).

No material was studied as the type was not found in any of the herbaria consulted.

Mycosphaerella digitalis-ambiguae Arx, Sydowia 3: 92. 1949.

Type — Switzerland: Sankt Gallen, Weesen. On upper and lower surface of dead leaves of *Digitalis ambigua* (Plantaginaceae). Müll., VII 1948 (CBS, holotype), also distributed in distributed in Reliquiae Petrakianae no. 1249 (H, isotype).

Anamorph and spermatial state: *Ramularia digitalis-ambiguae* von Arx and *Asteromella digitalis-ambiguae* von Arx *vide* von Arx (op. cit.).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradians*, with asci cylindrical, ascospores 12-14 × 2.5-3.5 µm. It was cited as synonymous with *M. mariae* by Tomilin (1979).

Mycosphaerella dioscoreae (Pass.) Tomilin, Novosti Sist. Nizsh. Rast. 1967: 188. 1967 = *Sphaerella dioscoreae* Pass., Atti Reale Accad. Lincei, Mem. Cl. Sci. Fis., ser. 4, 3: 8. 1887.— Fig. 265.

Type — Italy: On dead stems of *Dioscorea batatas* (Dioscoreaceae).

The type was not found in any of the herbaria consulted. Material studied (Spain, Madrid, Botanical garden, Caballero, IV 1923, BPI) belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores thick-walled, 17-18 × 5.5-6.5 µm.

Mycosphaerella dioscoreicola Syd. & P. Syd., Ann. Mycol. 14: 361. 1916 = *Sphaerella dioscoreicola* (Syd. & P. Syd.) Trotter, Syll. Fung. 24: 860. 1928.— Fig. 266.

Type — Philippines: Luzon, Prov. Laguna, Los Baños. On white spots with black margins on upper and lower surface of living leaves of *Dioscorea aculeata* (Dioscoreaceae). Baker no. 4121, XII 1915 (S, holotype).

This belongs to *Davidiella*, of which it represents a parasitic species, with asci pyriform, ascospores 12-14 × 5-6 µm. Therefore the following new combination is made:

Davidiella dioscoreicola (Syd. & P. Syd.) Aptroot comb. nov., **MB 500349**. **Basionym:** *Mycosphaerella dioscoreicola* Syd. & P. Syd., Ann. Mycol. 14: 361. 1916. Additional material studied (Indonesia, Java, Bogor, Botanical Garden, on upper and lower surface of dead leaves of *Dioscorea esculenta*, Boedijn no. 5173, L) contains only a coelomycete.

Mycosphaerella diospyri Syd. & P. Syd., Ann. Mycol. 11: 59. 1913 = *Sphaerella diospyri* (Syd. & P. Syd.) Trotter, Syll. Fung. 24: 860. 1928.

Type — Japan: Mino, Kawanye-mure. On upper and lower surface of dead leaves of *Diospyros kaki* (Ebenaceae). Hara, IV 1912 (S, holotype).

This is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores small but somewhat immature.

Sphaerella dircae Ellis & Everh., Proc. Acad. Nat. Sci. Philadelphia 45: 134. 1893.— Fig. 267.

Type — Canada: London. On white spots with black margins on upper surface of living leaves of *Dirca palustris* (Thymelaeaceae). Dearness no. 1941, VII 1892 (NY, holotype).

Anamorph: a *Macrosporium* is reported from the same spots by Ellis and Everhart (op. cit.).

This belongs to *Davidiella*, of which it represents a parasitic species, with asci pyriform, ascospores 15-19 × 5-7 µm. Therefore the following new combination is made:

Davidiella dircae (Ellis & Everh.) Aptroot comb. nov., **MB 500350**. **Basionym:** *Sphaerella dircae* Ellis & Everh., Proc. Acad. Nat. Sci. Philadelphia 45: 134. 1893.

Mycosphaerella discophora Syd., Ann. Mycol. 28: 81. 1930.— Fig. 268.

Type — Venezuela: Caracas, Cotiza. On spots on upper surface of living leaves of *Echites trifida* (Apocynaceae). Sydow, distributed in Reliquiae Petrakianae no. 67, II 1927 (H, L, isotypes).

This is a parasitic species, with asci cylindrical, ascospores $12-15 \times 3-3.5 \mu\text{m}$.

Mycosphaerella discophora var. *macrospora* Bat. & Peres, in Bat., Peres, Cavalcanti & Heringer, Atas Inst. Micol. 3: 223. 1960.

Type — Brazil: On upper surface of dead leaves of *Hancornia speciosa* (Apocynaceae). Batista exs. no. 19504 (URM 36241, holotype, as “*macrospora*”).

The type specimen contains only an immature ascomycete.

Sphaerella discrepans Kirschst., Ann. Mycol. 37: 103. 1939.— Fig. 269.

Type — Germany: Oberbayern, Bad Heilbrunn. On dead stems of *Chamaenerion* [as “*Epilobium*”] *angustifolium* (Onagraceae). Kirschstein, V 1938 (B, holotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. ammophilae*, with asci pyriform, ascospores $10-12 \times 2.5-3 \mu\text{m}$.

Mycosphaerella dispersa (Lahm ex Körb.) J. Schröt., in Cohn, Kryptog.-Fl. Schlesien 3(2): 342. 1894 [“1893”] = *Arthopyrenia dispersa* Lahm ex Körb., Parerga Lichenologica: 388. 1865 = *Pharcidia dispersa* (Lahm ex Körb.) G. Winter, in Rabenh., Krypt.-Fl. Deutschl., Österr. Schweiz 2, 1(2): 346. 1885 = *Stigmidium dispersum* (Lahm ex Körb.) D. Hawksw., Kew Bull. 30: 201. 1975.

Type — Germany: On thallus of *Lecanora* (Ascomycota, Lecanoraceae).

Accepted as *Stigmidium dispersum* (Lahm ex Körb.) D. Hawksw. by Clauzade, Diederich & Roux (1989).

Mycosphaerella disseminata (De Not. & Carestia) Tomilin, Novosti Sist. Nizsh. Rast. 1967: 188. 1967 = *Sphaerella disseminata* De Not. & Carestia, Erbario Crittogamico Italiano, ser. 2 no. 391. 1871 = *Leptosphaeria disseminata* (De Not. & Carestia) Rabenh., Fungi Europaei Exsiccati no. 1348. 1870, later homonym (illegitimate, Article 53), non De Not. (1867).— Fig. 270.

Type — Italy: Piedmonte, Valsesta. On dead leaves and culms of Poaceae. Carestia (RO, holotype; L, isotype), also distributed in Roumeguère, Fungi Gallici Exsiccati no. 2394 [“2891”] (L, isotype).

Anamorph: *Septoria fide* Saccardo (1882).

This belongs to *Davidiella*, and is morphologically indistinguishable from *Mycosphaerella recutita* sensu von Arx, with asci pyriform, ascospores $13-15 \times 4-5 \mu\text{m}$, for which it represents an older name than *M. wichuriana*, which was suggested as the oldest epithet for this taxon by Eriksson (1992). Therefore the following new combination is made: **Davidiella disseminata** (De Not. & Carestia) Aptroot comb. nov., **MB 500351**. **Basionym:** *Sphaerella disseminata* De Not. & Carestia, Erbario Crittogamico Italiano, ser. 2 no. 391. 1871.

Sphaerella distincta P. Karst., Öfvers Förh. Kongl. Svenska Vetensk.-Akad. 1872(2): 107. 1872 = *Didymella distincta* (P. Karst.) O.E. Erikss., Ark. Bot. 6: 448. 1967.

Type — Svalbard, Norway: *Glyceria vahliana* (Poaceae).

No material was studied as the species was accepted as *Didymella distincta* (P. Karst.) O.E. Erikss. by Eriksson (op. cit.).

Mycosphaerella ditissima Syd. & P. Syd., Ann. Mycol. 12: 554. 1914 = *Sphaerella ditissima* (Syd. & P. Syd.) Trotter, Syll. Fung. 24: 817. 1928.— Fig. 271.

Type — Philippines: Luzon, Benguet, Pauai. On upper and lower surface of dead squamules of *Lycopodium flexuosum* (Lycopodiaceae). Copeland no. 1522, V 1913 (S, holotype).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. filicum*, with ascospores large, asci broadly cylindrical, ascospores $16-19 \times 3-4 \mu\text{m}$.

Sphaerella ditricha (Fr.) Auersw., in Rabenh., Fungi Europaea Exsiccati, ed. nov., ser. 2 no. 943. 1866 = *Sphaeria ditricha* Fr., Systema Mycol. 2: 515. 1823 = *Vermicularia ditricha* (Fr.) Fr., Summa Veg. Scand.: 420. 1849 = *Venturia ditricha* (Fr.) P. Karst., Bidrag Kännedom Finlands Natur Folk 23: 188. 1873 = *Endostigme ditricha* (Fr.) Syd., Ann. Mycol. 21: 173. 1923.— Fig. 272.

Type — Finland: *Betula alba* (Betulaceae).

Accepted as *Venturia ditricha* (Fr.) P. Karst. by Sivanesan (1977), with which the material examined (Germany, Münster, on upper and lower surface of dead leaves of *Fraxinus excelsior*, Nitschke, 1863, distributed in Rabenhorst, Fungi Europaei Exsiccati, ed. nov., ser. 2 no. 943, L) agrees, with asci clavate, ascospores pale brown, asymmetrically septate, $12-15 \times 4.5-5.5 \mu\text{m}$.

Mycosphaerella dodartiae Nevod., Bot. Mater. Otd. Sporov. Rast. Bot. Inst. Komarova Akad. Nauk SSSR 14: 170. 1961.

Type — Russia: *Dodartia orientalis* (Plantaginaceae).

No material was studied as the type was not included in loans from LE or LEP.

Mycosphaerella dodonaeae Sivan. & R.G. Shivas, Mycol. Res. 106: 357. 2002.

Type — Australia: *Dodonaea triquetra* (Sapindaceae).

No material was studied of this recently described species.

Mycosphaerella dolichospora (Sacc. & Fautrey) Wehm., Mycologia 38: 162. 1946 = *Sphaerella dolichospora* Sacc. & Fautrey, in Fautrey, Rev. Mycol. (Toulouse) 19: 143. 1897.— Fig. 273.

Type — France: Côte d'Or. On dead stems of *Laserpitium gallicum* (Apiaceae). Fautrey (PAD, holotype).

This belongs to section *Longissima*, and is morphologically indistinguishable from *M. latebrosa*, with asci cylindrical, ascospores $25-29 \times 3-4 \mu\text{m}$.

Mycosphaerella dominicana (Gonz. Frag. & Cif.) Cif., Cuadernos 19: 233. 1961 = *Sphaerella dominicana* Gonz. Frag. & Cif., Bol. Soc. Esp. Hist. Nat. 25: 446. 1925.— Fig. 274.

Type — Dominican Republic: Haina. On white spots with brown margins on upper and lower surface of living leaves

of *Celtis iguana* (Cannabaceae). Ciferri no. 37, VII 1920 (BPI, lectotype, here designated).

This is a parasitic species, with asci cylindrical, ascospores $6-9 \times 2-3 \mu\text{m}$.

Mycosphaerella donacis Munjal, Chona & Kapoor, Indian Phytopathol. 12: 177. 1960 ["1959"].

Type — India: *Arundo donax* (Poaceae).

No material was studied and the preservation of the type is uncertain.

Sphaerella dorycnii (Mont.) Ces. & De Not., Comment. Soc. Crittog. Ital 1(4): 237. 1863 \equiv *Sphaeria dorycnii* Mont., Ann. Sci. Nat. Bot., ser. 4, 14: 176. 1860.

Type — France: *Dorycnium suffruticosum* (Fabaceae).

No material was studied as the type was not included in a loan from PC nor found in any of the herbaria consulted.

Sphaerella dracaenae Tassi, Bull. Lab. Orto Bot. Reale Univ. Siena 2: 27. 1899.

Type — Italy: Siena, Botanical Garden. On white spots on lower surface of living leaves of *Dracaena reflexa* (Asparagaceae). Tassi, I 1899 (SIENA, holotype).

The type contains no fungal tissue at all.

Mycosphaerella dracocephali Urries, Anales Inst. Bot. Cavanilles 14: 160. 1956 ["1955"].— Fig. 275.

Type — Canary Islands: La Palma, Baranco del Agua. On brown spots on upper and lower surface of living leaves of *Dracocephalum canariense* (Lamiaceae). Urries no. 12941, IV 1954 (MA, lectotype, here designated); also Baranco de los Galgos, no. 12932 (MA, paratype).

This is a parasitic species, with asci cylindrical, ascospores $11-13 \times 2.5-3 \mu\text{m}$.

Mycosphaerella dracocephalicola Ziling, in Murashk. & Ziling, Trudy Omsk. Selskokh. Inst. 9: 230. 1928.

Type — Russia: *Dracocephalum ruyschianae* (Lamiaceae).

Anamorph: Associated with *Septoria fide* Ziling (op. cit.).

Cited as synonymous with *M. leptasca* by Tomilin (1979). No material was studied as no type material is preserved in BPI or W, and it was not included in loans from LE or LEP.

Mycosphaerella drimydis (Berk.) Rehm, Ann. Mycol. 5: 469. 1907 [as "*drymidis*"] \equiv *Sphaeria drimydis* Berk., in Curr., Trans. Linn. Soc. London 22: 333. 1859 [as "*Drymidis*"] \equiv *Sphaerella drimydis* (Berk.) Cooke, J. Bot. 21: 109. 1883 [as "*drymidis*"].— Fig. 276.

Type — Chile: Juan Fernandez. On white spots on upper surface of dead leaves of *Drimys* ["*Drymis*"] (Winteraceae). V 1830 (K, holotype).

This is a parasitic species, with asci cylindrical, ascospores $11-13 \times 3-3.5 \mu\text{m}$.

Sphaerella drobnjakensis Bubák, Bot. Közlem. 1915: 56. 1915.— Fig. 277.

Type — Montenegro: Dobsi. On dead stems and calyces of *Linum laevis* (Linaceae). Bubák, VIII 1905 (BPI, holotype). Already cited as synonymous with *Davidiella allicina* (as *Mycosphaerella allicina*) by Tomilin (1979), with which the

type agrees well, with asci pyriform, ascospores thick-walled, $12.5-14 \times 4.5-6.5 \mu\text{m}$.

Sphaerella drobnjakensis var. *confinium* Bubák, Bot. Közlem. 1915: 57. 1915.— Fig. 278.

Type — Montenegro: Maglič. On dead stems of *Linum laevis* (Linaceae). Rohlena, VII 1905 (BPI, holotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. clandestina*, with asci pyriform, ascospores thick-walled, $21-24.5 \times 6.5-7.5 \mu\text{m}$.

Mycosphaerella droserae (Tassi) Tomilin, Opredelitel' gribov roda *Mycosphaerella* Johans.: 98. 1979 \equiv *Sphaerella droserae* Tassi, Bull. Lab. Orto Bot. Reale Univ. Siena 3: 14. 1900.— Fig. 279.

Type — Italy: Siena, Botanical Garden. On dead stems and peduncles of *Drosera rotundifolia* (Droseraceae). Tassi, I 1900 (SIENA, holotype).

This belongs to *Davidiella* and is morphologically indistinguishable from *D. ammophilae*, with asci pyriform, ascospores $10-12 \times 2-3 \mu\text{m}$.

Mycosphaerella dryadicola (Rostr.) Munk, in Möller, Fungi of the Faeröes 2: 58. 1958 \equiv *Sphaerella dryadicola* Rostr., Fungi from the Faeröes, in Botany of the Faeröes, part 1: 310, 1901.

Type — Faeröes: Paglö. On upper and lower surface of dead leaves of *Dryas octopetala* (Rosaceae). Hartz & Ostenfeld, VIII 1897 (C, holotype).

The type is partly immature, partly too old. It was cited as synonymous with *Mycosphaerella biberwierensis* by Tomilin (1979).

Mycosphaerella dryadis (Auersw.) Migula, in Thomé, Fl. Deutschl. Österr. Schweiz. X, I. Kryptog.-Fl. III, 3(1): 281. 1912 ["1913"] \equiv *Sphaerella dryadis* Auersw., in Gonn. & Rabenh., Mycol. Europaea 5-6: 8. 1869.

Type — Germany: *Dryas octopetala* (Rosaceae).

No material was studied as the type has probably been destroyed in B.

Mycosphaerella drymariae Syd. & P. Syd., Mém. Soc. Sci. Nat. Neuchâtel 5: 435. 1912 \equiv *Sphaerella drymariae* (Syd. & P. Syd.) Trotter, Syll. Fung. 24: 855. 1928.— Fig. 280.

Type — Colombia: Andes, Angelopolis, La Camelia. On lower surface of living and dying leaves of *Drymaria cordata* (Caryophyllaceae). Mayor, VIII 1910 (S, holotype). This is morphologically indistinguishable from *Glomerella cingulata* (Stoneman) Spauld. & H. Schrenk, with paraphyses present, ascospores non-septate, $13-16 \times 7-10 \mu\text{m}$.

Mycosphaerella drimydis (Berk.) Rehm. See *Mycosphaerella drimydis* (Berk.) Rehm.

Sphaerella dryophila Cooke & Harkn., Grevillea 9: 86. 1881 \equiv *Leptosphaeria dryophila* (Cooke & Harkn.) Sacc., Syll. Fung. 2: 51. 1883.

Type — USA: California, Marin Co., San Rafael. On ill-defined pale spots with brown margins on upper surface of

living leaves of *Quercus agrifolia* (Fagaceae). Harkness no. 1471, IV 1880 (BPI, isotype).

The isotype contains *Pseudomassaria agrifolia* (Ellis & Everh.) Arx, but it is a not certain to be a synonym of this species, as it disagrees with the protologue. The isotype contains also *Monochaeta bicornis* (Durieu & Mont.) Sacc.

Mycosphaerella dubia Miles, Trans. Illinois State Acad. Sci. 10: 250. 1917 = *Sphaerella dubia* (Miles) Trotter, Syll. Fung. 24: 887. 1928.— Fig. 281.

Type — Puerto Rico: Maricao. On brown spots on lower surface of living leaves of *Solanum* (Solanaceae). Stevens, IV 1913, Porto Rican Fungi no. 750 (K, holotype).

This belongs to *Davidiella* and is morphologically indistinguishable from *D. ammophilae*, with asci pyriform to globose, ascospores $9-11 \times 2.5-3.5 \mu\text{m}$.

Mycosphaerella dubia F.A. Wolf. See *Mycosphaerella confusa* F.A. Wolf.

Sphaerella duchartrei (Crié) Sacc., Syll. Fung. 1: 495. 1882 = *Depazea duchartrei* Crié, Ann. Sci. Nat. Bot., ser. 6, 7: 45. 1878.

Type — France: *Vinca minor* (Apocynaceae).

Anamorphs: Associated with *Phoma* and *Septoria fide* Saccardo (op. cit.).

This is no *Mycosphaerella* according to Tomilin (1979). No material was studied, as the type was not included in a loan from PC.

Mycosphaerella dummeri Hansf., Proc. Linn. Soc. London 153: 21. 1941.

Type — Uganda: *Vernonia* (Asteraceae).

No material was studied as the holotype is not in IMI or K.

Mycosphaerella dunbariae Syd., in Syd. & Petr., Ann. Mycol. 29: 197. 1931.— Fig. 282.

Type — Philippines: Luzon, Cagayan, Tuguegarao. On brown spots on upper surface of living leaves of *Dunbaria cumingiana* (Fabaceae). (K, 2 isotypes).

This is a parasitic species, with asci cylindrical, ascospores $16-19 \times (2.5-)3-4 \mu\text{m}$.

Mycosphaerella earliana (G. Winter) Tomilin, Novosti Sist. Nizsh. Rast. 1968: 165. 1968 = *Sphaerella earliana* G. Winter, J. Mycol. 1: 101. 1885.— Fig. 283.

Type — USA: Illinois, Annapolis. On upper and lower surface of dead leaves of *Fragaria vesca* (Rosaceae). Earle, IV 1884, distributed in Rabenhorst-Winter, Fungi Europaei Exsiccati no. 3349 (L, isotype), also distributed in Ellis & Everhart, North American Fungi 1677 (L, isotype).

As already cited by Tomilin (1979), this is morphologically indistinguishable from *M. punctiformis*, with ascomata in dense groups (like in the synonymous *M. maculiformis*), asci cylindrical, ascospores $8-10 \times 2-2.5 \mu\text{m}$.

Mycosphaerella ebuli (Richon) Tomilin, Opredelitel' gribov roda *Mycosphaerella* Johans.: 81. 1979 = *Sphaerella ebuli* Richon, Catalog. Champ. Marne: 273. 1889.— Fig. 284.

Type — France. On dead stems of *Sambucus ebulus* (Adoxaceae).

Material studied (Germany, Reinhard, V 1919, B) belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradicans*, with asci cylindrical, ascospores $10-12 \times 3-4 \mu\text{m}$.

Mycosphaerella ebulina Petr., Ann. Mycol. 13: 46. 1915 = *Sphaerella ebulina* (Petr.) Trotter, Syll. Fung. 24: 855. 1928.— Fig. 285.

Type — Czech Republic: Weißkirchen, Hranice, Ungersdorf. On upper and lower surface of dead leaves of *Sambucus ebulus* (Adoxaceae). Petrak, X 1914, distributed in Reliquiae Petrakianae no. 860 (H, L, B, isotypes).

The isotype in H belongs to section *Caterva*, and shows that this is morphologically indistinguishable from *M. subradicans*, with asci cylindrical, ascospores $11-13 \times 3.5-4.5 \mu\text{m}$. The isotype in L is immature, the isotype in B overmature.

Mycosphaerella ecdysantherae Sawada, Taiwan Univ. Coll. Agric. Special Publ. 8: 62. 1959.

Type — Taiwan: *Ecdysanthera utilis* (Apocynaceae).

No material was studied as the type was not present in BPI, where many Sawada isotypes are kept.

Sphaerella echinophila Auersw., in Gonn. & Rabenh., Mycol. Europaea 5-6: 3, 1869, based on *Sphaeria echinophila* Ces., Unio Itin. Crypt. no. 21. 1866, nomen nudum (not validly published, Article 32) = *Laestadia echinophila* (Auersw.) Sacc., Syll. Fung. 1: 425. 1882.

Type — Italy: *Castanea vesca* (Fagaceae).

No material was studied, as the species was already excluded by Saccardo (op. cit.).

Sphaerella echinophila (Schwein.) Cooke, J. Bot. 21: 69. 1883, later homonym (illegitimate, Article 53) = *Sphaeria echinophila* Schwein., Trans. Amer. Philos. Soc., ser. 2, 4: 223. 1832.

Type — USA: Philadelphia, Bethlehem. On spines on the fruits of *Castanea* (Fagaceae). Schweinitz (PH, holotype).

The type contains only a coelomycete.

Mycosphaerella edelbergii Petr., Sydowia 7: 89. 1953.— Fig. 286.

Type — Afghanistan: Eastern Nuristan. On dead culms of *Sedum* (Crassulaceae). Edelberg no. 1064, VI 1948 (W, holotype).

This belongs to section *Longispora*, and is morphologically indistinguishable from *M. latebrosa*, with asci cylindrical, ascospores $26-30 \times 3-4.5 \mu\text{m}$.

Mycosphaerella effigurata (Schwein.) House, New York State Mus. Bull. 233-234: 27. 1921 = *Sphaeria effigurata* Schwein., Trans. Amer. Philos. Soc., ser. 2, 4: 224. 1832 = *Sphaerella effigurata* (Schwein.) Cooke, J. Bot. 21: 107. 1883.

Type — USA: Philadelphia, Bethlehem. On lower surface of dead leaves of *Fraxinus acuminatus* (Oleaceae). Schweinitz (PH, holotype).

Spermatial state and anamorph: *Asteromella fraxini* (Berk. & M.A. Curtis) Petr. and *Marssonina fraxini* Ellis & Davis *fide* Wolf & Davidson (1941).

The holotype is immature, but shows that this is morphologically indistinguishable from *M. punctiformis*. Additional material studied (California, Klamath River at Walker, on *F. oregona*, Wheeler no. 3263, IX 1934, B; also Canada, Nonwatin Lake, Thunder Bay District, Ontario, on spots on lower surface of living leaves of *F. nigra*, Parmelee *et al.* no. 4789, IX 1973, L) contains only the spermatial state *Asteromella fraxini* (Berk. & M.A. Curtis) Petr.

Sphaerella effusa Sacc. & P. Syd. See *Mycosphaerella adusta* (Fuckel) Jacz.

Mycosphaerella elaeagnicola N.P. Golovina, Bot. Mater. Otd. Sporov. Rast. Bot. Inst. Komarova Akad. Nauk SSSR 12: 156. 1959.

Type — Kazakhstan: *Elaeagnus angustifolia* (Elaeagnaceae).

No material was studied as type material was not included in loans from LE or LEP.

Mycosphaerella elaeidis (Beeli) Hendr., Publ. Inst. Natl. Étude Agron. Congo Belge, Sér. Sci. 35: 7. 1948 = *Sphaerella elaeidis* Beeli, Rev. Zool. Bot. Africaine, Suppl. Bot. 11(2), B: 10. 1923 = *Oxydothis elaeidis* (Beeli) Sivan., Trans. Brit. Mycol. Soc. 54: 496. 1970.— Fig. 287.

Type — Zaire: Leopoldville. On pale spots with brown margins on upper surface of living leaves of *Elaeis* (Arecaceae). Ghesquière no. 1124, III 1922 (BR, holotype). Accepted by Hyde (1994) as *Oxydothis elaeidis* (Beeli) Sivan., with which the type agrees well, with eccentric ostioles and ascospores fusiform, 38-46 × 7-9 µm.

Mycosphaerella elasticae Koord., Verh. Kon. Ned. Akad. Wetensch., Afd. Natuurk., Tweede Sect. 13: 189. 1907 = *Sphaerella elasticae* (Koord.) Sacc. & Traverso, Syll. Fung. 20: 820. 1911.

Type — Indonesia: *Ficus elastica* (Moraceae).

No material was studied as type material was not found in L or in any of the other herbaria consulted; its preservation is uncertain.

Mycosphaerella elatior (Sacc. & Speg.) Tomilin, Novosti Sist. Nizsh. Rast., 1968: 165. 1968 = *Sphaerella elatior* Sacc. & Speg., Michelia 1: 379. 1878.— Fig. 288.

Type — Italy. On upper surface of dead leaves of *Liriodendron tulipifera* (Magnoliaceae). Spegazzini (PAD, holotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. clandestina*, with asci pyriform, ascospores 25-30 × 8-9 µm.

Sphaerella elatior var. *apula* Sacc. & D. Sacc. See *Mycosphaerella apula* (Sacc. & D. Sacc.) Tomilin.

Mycosphaerella elatostemae Thirum. & Govindu, Sydowia 8: 345. 1954 [as “*elatostemmae*”].

Type — India: *Elatostema* (Urticaceae).

No material was studied as type material was not found in MAH or in any of the herbaria consulted; its preservation is uncertain.

Mycosphaerella ellipsoidea Crous & M.J. Wingf., Mycologia 88: 452. 1996.

Type — South Africa: Western Cape, Pampoenvlei. On leaves of *Eucalyptus cladocalyx* (Myrtaceae). Crous, XI 1994 (PREM 51924, holotype, not seen).

Anamorph: *Uwebraunia ellipsoidea* Crous & M.J. Wingf. *vide* Crous (1998).

No material was studied of this recently described species.

Mycosphaerella elodis (A.L. Sm. & Ramsb) Tomilin, Novosti Sist. Nizsh. Rast. 6: 120. 1970 [“1969, as *helodis*”] = *Sphaerella elodis* A.L. Sm. & Ramsb., Trans. Brit. Mycol. Soc. 5: 423. 1917 [as “*elodes*”].— Fig. 289.

Type — United Kingdom: Hampshire, New Forest. On upper and lower surface of dead leaves of *Hypericum elodes* (Hypericaceae). Ramsbottom, IX 1916 (K, holotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. ammophilae*, with asci pyriform, ascospores 11-13 × 3.5-3.5 µm.

Mycosphaerella elymi (Unamuno) M. Morelet, Ann. Soc. Sci. Nat. Archéol. Toulon & Var 20: 105. 1968 = *Sphaerella elymi* Unamuno, Asoc. Esp. Progr. Ci. 1929: 12. 1929.

Type — Spain: *Elymus caput-medusae* (Poaceae).

No material was studied as type material was not found in MA or in any of the herbaria consulted; its preservation is uncertain.

Mycosphaerella elymifoliae Munk, Dansk Bot. Ark. 14(8): 2. 1952.

Type — Denmark: *Elymus arenarius* (Poaceae).

Cited as synonymous with *Davidiella allicina* (as *Mycosphaerella allicina*) by Tomilin (1979). No material was studied as type material was not found in C or in any of the herbaria consulted; its preservation is uncertain, as Munk sometimes used up all material for his descriptions.

Mycosphaerella embothrii (Speg.) Tomilin, Novosti Sist. Nizsh. Rast. 10: 101. 1973 = *Sphaerella embothrii* Speg., Bol. Acad. Nac. Ci. 11: 205. 1888 [“1887”].

Type — Argentina: Tierra del Fuego, Canal de Beagle, Isla Aicina. On lower surface of dead leaves of *Embothrium coccineum* (Proteaceae). Spegazzini no. 6184, V 1882 (LPS, holotype).

The holotype contains empty ascomata which are developing below the cuticle, which is later flaking off; it is unidentifiable, but not referable to *Mycosphaerella*.

Mycosphaerella emeri (Ces.) Cruchet, Bull. Soc. Vaud. Sci. Nat. 55: 45. 1923 = *Sphaerella emeri* Ces., in Rabenh., Fungi Europaei Exsiccati, ed. nov., ser. 2 no. 1950. 1875.— Fig. 290.

Type — Italy: Brescia, Patrocínio. On upper and lower surface of dead leaves of *Coronilla emeris* (Fabaceae). Cesati, 1875, distributed in Rabenhorst, Fungi Europaei Exsiccati no. 1950 (L, isotype).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradicans*, with asci cylindrical, ascospores 11-13 × 2-2.5 µm.

Mycosphaerella endophytica Crous & H. Sm.ter, in Crous, Mycologia Memoir 21: 54. 1998.

Type — South Africa: Western Cape, Stellenbosch, Devon Valley. On leaves of *Eucalyptus* (Myrtaceae). Crous, VI 1995 (PREM 54398, holotype, not seen).

Anamorph: *Pseudocercospora endophytica* Crous & H. Sm. *vide* Crous (1998).

No material was studied of this recently described species.

Mycosphaerella endospermi Syd. & P. Syd., Ann. Mycol. 15: 206. 1917 = *Sphaerella endospermi* (Syd.) Trotter, Syll. Fung. 24: 862. 1928.— Fig. 291.

Type — Philippines: Luzon, Kalinga. On white spots on upper surface of living leaves of *Endospermum peltatum* (Euphorbiaceae). Yates, III 1915, Bureau of Science no. 25329 (BPI, isotype).

This is a parasitic species, with asci cylindrical, ascospores with rounded ends, 16-18 × 3-4.5 µm.

Mycosphaerella engleriana Reichert, Bot. Jahrb. Syst. 56: 670. 1921.

Type — Egypt: *Noaea mucronata* (Amaranthaceae).

No material was studied as type material was not found in B or in any of the other herbaria consulted; its preservation is uncertain.

Mycosphaerella entadae Sawada, Trans. Nat. Hist. Soc. Taiwan 32: 340. 1942, lacking Latin description (not validly published, Article 36.1).

Type — Taiwan: *Entada phaceoloides* (Fabaceae).

No material was studied as the type was not present in BPI, where many Sawada isotypes are kept.

Mycosphaerella enteleae (Dingley) Sivan., Biblioth. Mycol. 59: 115. 1977 = *Venturia enteleae* Dingley, New Zealand J. Agric. Res. 8: 907. 1965.— Fig. 292.

Type — New Zealand: Auckland, Cascades Kauri Park, Waitakere Stream. On dead, mainly marginal, brown parts of living leaves of *Entelea arborescens* (Malvaceae). Dingley, XI 1961 (IMI no. 173339, isotype slides).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradians*, with asci cylindrical, ascospores 10-12 × 2.5-3.5 µm. Additional material (Auckland, White's Stream, Piha, Dingley, IX 1970, IMI no. 173338) agrees.

Mycosphaerella ephedrae (Hollós) Bechet, Contr. Bot. 1960: 86. 1960 = *Sphaerella ephedrae* Hollós, Ann. Hist.-Nat. Mus. Natl. Hung. 4: 331. 1906 = *Phaeosphaerella ephedrae* (Hollós) Petr., Ann. Mycol. 32: 327. 1934.

Type — Hungary. On dead stems of *Ephedra distachya* (Ephedraceae).

The type is not in BP and may have been destroyed during the war. Material studied (Italy, Trieste, Verucca, Parta, VI 1898, BPI) is overmature.

Sphaerella ephedrae Golovin, Trudy Sreedneaz. Gosud. Univ., Nov. Ser., Vyp. 14, Biol. Nauki, Kniga 5: 9. 1950, later homonym (illegitimate, Article 53).

Type — Uzbekistan: *Ephedra* (Ephedraceae).

No material was studied as the type was not included in loans from LE or LEP.

Mycosphaerella ephedricola Butin, Sydowia 27: 276. 1975 ["1973-1974"].— Fig. 293.

Type — Chile: La Serena. On dead branches of *Ephedra americana* (Ephedraceae). Butin, X 1968 (ZT, holotype).

This belongs to *Davidiella*, with asci pyriform, ascospores 15-18 × 3-4 µm. Therefore the following new combination is made: **Davidiella ephedricola** (Butin) Aptroot comb. nov., **MB 500352**. **Basionym:** *Mycosphaerella ephedricola* Butin, Sydowia 27: 276. 1975 ["1973-1974"].

Sphaerella epicymatia (Wallr.) Speg., Michelia 1: 456. 1879 = *Sphaeria epicymatia* Wallr., in Fr., Elenchus Fungorum 2: 103. 1828 = *Epicymatia vulgaris* Fuckel, Jahrb. Nassauischen Vereins Naturk. 23-24: 118. 1870 = *Arthopyrenia epicymatia* (Wallr.) Müll. Arg., Rev. Mycol. (Toulouse) 6: 20. 1884 = *Pharcidia epicymatia* (Wallr.) G. Winter, in Rabenh., Kryptog.-Fl. Deutschl., Österr. Schweiz 2, 1(2): 342. 1887 = *Sphaerella vulgaris* (Fuckel) Masee, Grevillea 19: 44. 1890, later homonym (illegitimate, Article 53) and superfluous (illegitimate, Article 52) combination, based on *Epicymatia vulgaris* Fuckel.

Type — Germany: *Lecanora* (Ascomycota, Lecanoraceae). Cited as synonymous with *Muellerella pygmaea* (Koerber) D. Hawksw. by Roux & Triebel (1994: 485), although the basionym *epicymatia* is much older. The epithet *vulgaris* is occasionally incorrectly seen as heterotypic, and Roux & Triebel (1994) have even chosen a lectotype for it and cite it as synonymous with *Stigmidium congestum* (Koerber) Triebel. Material studied (Leipzig, on apothecium discs of *Lecanora chlarotera* [as "*L. subfusca*"], Auersw., B) agrees with the latter.

Mycosphaerella epilobii (Crié) Tomilin, Mikol. Fitopatol. 2: 394. 1968 = *Depazea epilobii* Crié, Ann. Sci. Nat. Bot., ser. 6, 7: 44. 1878 = *Sphaerella epilobii* (Crié) Sacc., Syll. Fung. 1: 503. 1882, later homonym (illegitimate, Article 53).— Fig. 294.

Type — France: *Epilobium* (Onagraceae).

Anamorphs: Associated with *Phoma* and *Septoria* *vide* Saccardo (1882).

Cited as synonymous with *Venturia maculiformis* (Desm.) G. Winter by Sivanesan (1977), with which the material examined (Switzerland, Neuchâtel. On spots on upper surface of living leaves of *Epilobium montanum*, Morthier, VIII 1879, distributed in Thümen, Mycotheca Universalis no. 1652, L) agrees, with asci surrounded by paraphyses, ascospores pale brown, asymmetrically septate, 12-14 × 4-5 µm.

Sphaerella epilobii (Wallr.) Auersw., in Gonn. & Rabenh., Mycol. Europaea 5-6: 14. 1869 = *Sphaeria epilobii* Wallr., Fl. Cryptog. Germ. 2: 771. 1833 = *Laestadia epilobii* (Wallr.) Sacc., Syll. Fung. 1: 431. 1882 = *Guignardia epilobii* (Wallr.) Lindau, in Engl. & Prantl, Natürlichen Pflanzenf. 1(1): 423. 1897 = *Discosphaerina epilobii* (Wallr.) Petr., Ann. Mycol. 38: 251. 1940.

Type — Germany: *Chamaenerion* ["*Epilobium*"] *angustifolium* (Onagraceae).

Anamorph: *Selenophoma epilobii* Petr. *fide* Eriksson (1992).

Accepted as *Guignardia epilobii* (Wallr.) Lindau by Eriksson (1992).

Sphaerella epilobii Fuckel. See *Sphaerella fuckelii* Pass.

Sphaerella epilobii f. *toletana* Gonz. Frag., Trab. Mus. Nac. Ci. Nat., Ser. Bot. 9: 30. 1916.— Fig. 295.

Type — Spain: Toledo, S. Pable de los Montes. On brown spots on upper and lower surface of living leaves of *Epilobium hirsutum* (Onagraceae). Cogolludo, no. 1205, VII 1915 (MA, holotype).

This is morphologically indistinguishable from *Venturia maculiformis* (Desm.) G. Winter, with asci cylindrical, surrounded by hamathecium filaments, ascospores greenish brown, 12-14 × 5-7 µm.

Mycosphaerella epilobii-montani Lobik, Bolezni Rast. 17: 163. 1928 = *Phaeosphaerella epilobii-montani* (Lobik) Tomilin, Opređelitel' gribov roda *Mycosphaerella* Johans.: 285. 1979.— Fig. 296.

Type — Russia: On green spots on upper surface of living leaves of *Epilobium montanum* (Onagraceae). (LE 34981, holotype).

This is morphologically indistinguishable from *Venturia adusta* (Fuckel) E. Müll., with asci surrounded by paraphyses, ascospores olive brown, asymmetrically septate, 13-16 × 5-6 µm.

Mycosphaerella epimedii (Sacc.) Jaap, Ann. Mycol. 14: 13. 1916 = *Sphaerella epimedii* Sacc., Syll. Fung. 1: 500. 1882.

Type — Italy: Treviso, Montello. On lower surface of living leaves of *Epimedium alpinum* (Berberidaceae). Saccardo, IX 1902, Mycotheca Italica no. 1032 (B, BPI, topotypes).

The topotypes studied are immature, but belongs probably to *M. punctiformis*, with asci cylindrical. Additional material studied (Venezia, Hrubby, VI 1918, CBS, L) is also immature.

Mycosphaerella epiphylla (Kirschst.) Tomilin, Opređelitel' gribov roda *Mycosphaerella* Johans.: 131. 1979 = *Sphaerella epiphylla* Kirschst., Kryptog.-Fl. Mark Brandenburg 7(3): 338. 1938, nomen novum (Article 58) for *Stigmatea quercina* Rehm (1906), non *Sphaerella quercina* Jacz. (1896).— Fig. 297.

Type — Germany: Königstein. On upper and lower surface of dead leaves of *Quercus* (Fagaceae). Krieger, Fungi Saxonici no. 1967, IV 1901 (B, lectotype, here designated; also B, isolectotype).

This is morphologically indistinguishable from *Guignardia punctoidea* (Cooke) J. Schröt., with asci clavate, ascospores rhomboidal, simple, 12-13 × 5-5.5 µm.

Mycosphaerella epiphylla D. Gupta, Padhi & Chowdhry, Indian Phytopathol. 33: 500. 1981 [“1980”], later homonym (illegitimate, Article 64).

Type — India: *Epiphyllum ackermannii* (Cactaceae).

No material was studied and the preservation of the type is uncertain.

Mycosphaerella epistroma (Cooke) Tomilin, Novosti Sist. Nizsh. Rast. 1967: 189. 1967 = *Sphaerella epistroma* Cooke, J. Bot. 21: 137. 1883.

Type — United Kingdom: on dead leaves of cultivated Poaceae (straw). Cooke no. 372 (K, holotype).

The holotype contains only stromata without ascospores, belonging to an unidentifiable ascomycete.

Sphaerella epitea Auersw., in Gonn. & Rabenh., Mycol. Europaea 5-6: 4. 1869, nomen nudum (not validly published, Article 32).

Authentic material — Germany: *Salix amygdaloides* (Salicaceae).

Cited as synonym of *Sphaerella punctiformis* [as “*sparsa*”] by Auerswald (op. cit.).

Mycosphaerella equiseti (Fuckel) J. Schröt., in Cohn, Kryptog.-Fl. Schlesien 3(2): 341 1894 [“1893”] = *Sphaerella equiseti* Fuckel, Jahrb. Nassauischen Vereins Naturk. 23-24: 102. 1870.— Fig. 298.

Type — Germany: Budenheim. On dead stems of *Equisetum palustre* (Equisetaceae). Fuckel, Fungi Rhenani Exsiccati no. 2241 (L, isotype).

In the isotype studied only a *Phoma* could be found. Additional material studied (Brandenburg, Glindow, Sydow, VII 1937, distributed in Mycotheca Germanica no. 3106, L) belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradicans*, with large ascospores, asci cylindrical, ascospores 12-15 × 3-3.5 µm. Further additional material studied (Netherlands, Nunspeet, Beins × 1900, L) belongs to *Davidiella*, but is *D. allicina*, with asci pyriform, ascospores 18-22 × 5-6 µm.

Mycosphaerella equiseti f. *equiseti-arvensis* Thüm., Mycotheca Universalis no. 263.

Type — Germany: Sachsen, Eisleben. On dead stems of *Equisetum arvensis* (Equisetaceae). Winter, 1874, distributed in Thümen, Mycotheca Universalis no. 263 (L, 4 isotypes).

The isotypes studied are immature, but probably belong to section *Caterva*, with asci cylindrical.

Mycosphaerella equiseticola Bond.-Mont., Bot. Mater. Inst. Sporov. Rast. Glavn. Bot. Sada 2: 18. 1923.— Fig. 299.

Type — Russia: On dead branches of *Equisetum sylvaticum* (Equisetaceae). (LE 34978, holotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. ammophilae*, with asci small, pyriform, ascospores 9-11 × 3.5-4.5 µm.

Mycosphaerella equisetina Syd., Ann. Mycol. 19: 139. 1921 = *Didymella equisetina* (Syd.) Petr., Ann. Mycol. 29: 358. 1931.— Fig. 300.

Type — Germany: Brandenburg, Zossen, Spereberg. Immersed in dead stems of *Equisetum hiemale* (Equisetaceae). Sydow, VII 1917, distributed in Reliquiae Petrakianae no. 2051 and Mycotheca Germanica no. 1540 (L, isotype).

This is morphologically indistinguishable from *D. allicina*, with asci clavate, ascospores 18-21 × 6-7 µm, although the

species was still accepted as *Didymella equisetina* (Syd.) Petr. by Eriksson (1992).

Sphaerella equisetina Kirschst., Kryptog.-Fl. Mark Brandenburg 7(3): 354. 1938.

Type — Germany: *Equisetum hiemale* (Equisetaceae). No material was studied as the type was not found in B, where many other Kirschstein types were preserved.

Mycosphaerella eragrostidis Castell. & Ciccaroni, in Cif. & Baldrati, Il “Teff”, Bibl. Agr. Colon. Firenze 1939, R. Ist. Agron. Africa Ital.: 97. 1939.

Type — Africa: *Eragrostis* (Poaceae).
Anamorph: Associated with *Septoria eragrostidis* Castell. & Ciccaroni *vide* Ciferri & Baldrati (op. cit.).
No material was studied as the type was not included in a loan from PC and its preservation is uncertain.

Mycosphaerella erechthitidina Petr. & Cif., Ann. Mycol. 28: 391. 1930 [as “*erechthitidina*”].— Fig. 301.

Type — Dominican Republic: Lome La Vuide. On dead spots on upper and lower surface of living leaves of *Erechtites hieracifolia* (Asteraceae). Ciferri no. 2917, VIII 1929 (W, holotype; B, isotype).

This is a species of *Didymella*, with anastomosing pseudoparaphyses, asci cylindrical, ascospores $9-15 \times 3-4.5 \mu\text{m}$. Additional material studied (Ecuador, Tungurahua, Baños, on upper and lower surface of dead leaf tips and sectors of *Erechtites prenanthoides*, Sydow, XII 1937, Fungi Exotici Exsiccati no. 1189, L, W) belongs to *Davidiella* and is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores $15-17 \times 4.5-5.5 \mu\text{m}$.

Mycosphaerella ericae-ciliaris (Unamuno) M. Morelet, Ann. Soc. Sci. Nat. Archéol. Toulon & Var 20: 105. 1968 \equiv *Sphaerella ericae-ciliaris* Unamuno, Bol. Soc. Esp. Hist. Nat. 29: 117. 1929.

Type — Spain: *Erica ciliaris* (Ericaceae).
No material was studied as the type was not found in MA, where many other Unamuno types were preserved.

Mycosphaerella eriodendri Kuijper, Recueil Trav. Bot. Néerl. 11: 49. 1914.

Type — Surinam: *Ceiba* [“*Eriodendron*”] *anfractuosa* (Malvaceae).
No material was studied as the type was not found in L, and its preservation is uncertain.

Mycosphaerella eriophila (Niessl) Lindau, Hilfsb. Sammeln Ascomyceten: 12. 1903 \equiv *Sphaerella eriophila* Niessl, Oesterr. Bot. Z. 25: 86. 1875.— Fig. 302.

Type — Austria: Innsbrück. On dead stems and upper and lower surface of dead leaves of *Artemisia mutellina* (Asteraceae). Niessl (M, holotype).

The holotype is overmature, but the species was already synonymised with *Davidiella allicina* (as *Mycosphaerella tassiana*) by von Arx (1949), with which additional materials studied (Inntal, on *Artemisia eriantha* [“*baumgarten*”], Niessl, M; also Switzerland, Albula-Pass, Rhätische Alpen, Winter, VIII 1882, distributed in Rabenhorst-Winter, Fungi Europaei Exsiccati no. 3145, L,

M (2 \times); also Wallis, on *Artemisia glacialis*, Niessl, M; also Poland, Tatra, on *Artemisia danngarteri*, Niessl, M; also Zermatt, On *Artemisia glacialis*, Niessl, M) agree, with asci pyriform, ascospores immature, ca. $16 \times 5 \mu\text{m}$.

Sphaerella erlangeae Baccl., Ann. Bot. (Rome) 14: 133. 1917.— Fig. 303.

Type — Ethiopia: Amhara-Dernba. On lower surface of dead leaves of *Erlangea abyssinica* (Asteraceae). Baccarini (PAD, holotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores $17-21 \times 4.5-5.5 \mu\text{m}$.

Sphaerella errabunda (Roberge) Auersw., in Rabenh., Fungi Europaei Exsiccati, ed. nov., ser. 2 no. 944. 1866 \equiv *Sphaeria errabunda* Roberge, in Desm., Cryptog. Fr., ed. 2 no. 1441 \equiv *Gnomonia errabunda* (Roberge) Auersw., in Gonn. & Rabenh., Mycol. Europaea 5-6: 25. 1869 \equiv *Laestadia errabunda* (Roberge) Rehm, Hedwigia 41: 202. 1902 \equiv *Apiognomonium errabunda* (Roberge) Höhn., Ann. Mycol. 16: 51. 1918.

Type — France: *Fagus* (Fagaceae).
Anamorphs: *Gloeosporium fagi* (Desm. & Roberge) Westend. *vide* Barr (op. cit.) \equiv *Discula umbrinella* (Berk. & Broome) M. Morelet *vide* Morelet (1989).

Accepted as *Apiognomonium errabunda* (Roberge) Höhn. by Barr (1978). Material studied (Germany, Thüringen, on lower surface of dead leaves of *Corylus avellana*, Fleischhack, distributed in Rabenhorst, Fungi Europaei Exsiccati, ed. nov., ser. 2 no. 944, CBS) is immature.

Mycosphaerella eryngii (Fr.) Johanson ex Oudem., Rév. Champ. Pays-Bas 2: 213. 1897 \equiv *Sphaeria eryngii* Fr., in Duby, in DC., Botan. Gallicum, ed. 2, 2: 710. 1830 \equiv *Sphaerella eryngii* (Fr.) Fuckel, Jahrb. Nassauischen Vereins Naturk. 23-24: 105. 1870.

Type — France: Vosges. On upper and lower surface of dead leaves of *Eryngium campestre* (Apiaceae). Mougeot no. F-03568 and Calvados, Caen. Roberge no. F-03651 (UPS-FRIES, syntypes).

Spermatial state: *Asteroma eryngii* Auersw. *vide* Fuckel (op. cit.).

Both syntypes contain several ascomycetes, most of which are immature, the only with ascospores is *Pleospora herbarum* (Pers. : Fr.) Rabenh. Additional material studied (Germany, Doemitz, Fiedler, distributed in Klotzsch, Herbarium Vivum Mycologicum no. 1639 and Fuckel, Enumeratio Fungi Nassauae no. 525, L) belongs to section *Longispora*, and is morphologically identical to *M. millegrana*, with asci cylindrical, ascospores $15-18 \times 2.5-3.5 \mu\text{m}$. Additional material on *Eryngium maritimum* (Netherlands, Katwijk, L) is immature.

Sphaerella eryngii f. *eryngii-maritimi* Roum., Fungi Gallici Exsiccati no. 2742.

Type — Belgium: Oostende. On upper and lower surface of dead leaves of *Eryngium maritimum* (Apiaceae). Bommer & Rousseau, distributed in Roumeguère, Fungi Gallici Exsiccati no. 2742 (L, isotype).

The isotype studied contains a coelomycete and an immature ascomycete.

Sphaerella eryngii var. *libanotidis* Fuckel. See *Mycosphaerella libanotidis* (Fuckel) Lind.

Mycosphaerella eryngicola (Speg.) Tomilin, Novosti Sist. Nizsh. Rast. 7: 203. 1970 = *Sphaerella eryngicola* Speg., Revista Fac. Agron. Vet., ser. 2, 6: 53. 1910.

Type — Chile: *Eryngium paniculatum* (Apiaceae).

No material was studied as the type was not included in a loan from LPS.

Mycosphaerella eryngina (Gonz. Frag.) Tomilin, Novosti Sist. Nizsh. Rast. 7: 202. 1970 = *Sphaerella eryngina* Gonz. Frag., Bol. Soc. Esp. Hist. Nat. 18: 366. 1918.— Fig. 304.

Type — Spain: Madrid, Ribas del Iarama. On upper and lower surface of dead leaves of *Eryngium campestre* (Apiaceae). Vicioso no. 3040, IV 1908 (MA, holotype).

Already cited as synonymous with *Davidiella allicina* (as *Mycosphaerella allicina*) by Tomilin (1979), but the holotype shows that it is morphologically indistinguishable from *D. clandestina*, with asci clavate to pyriform, ascospores 20-23 × 6-8 µm.

Mycosphaerella erysiphina (Berk. & Broome) Kirchn., Die Krankheiten und Beschädigungen unserer landwirtschaftlichen Kulturpflanzen, ed. 3: 319. 1923 = *Sphaeria erysiphina* Berk. & Broome, in Berk., J. Hort. Soc. London 9: 65. 1855 = *Sphaerella erysiphina* (Berk. & Broome) Cooke, J. Bot. 4: 250. 1866.

Type — United Kingdom: *Humulus lupulus* (Urticaceae).

No material was studied as the type was not found in K, and its preservation is uncertain.

Mycosphaerella erysiphoides (Sacc.) Miles, Trans. Illinois State Acad. Sci. 10: 249. 1917 = *Sphaerella erysiphoides* Sacc., Michelia 2: 604. 1882.

Type — France: Saintes. On lower surface of dead leaves of *Tecoma radicans* (Bignoniaceae). Brunaud (PAD, holotype).

Anamorph: *Phyllosticta erysiphoides* Sacc. fide Saccardo (1891).

This is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores 7-9 × 2-2.5 µm.

Mycosphaerella erythrinae Koord., Verh. Kon. Ned. Akad. Wetensch., Afd. Natuurk., Tweede Sect. 13: 189. 1907 = *Sphaerella erythrinae* (Koord.) Sacc. & Traverso, Syll. Fung. 20: 820. 1911.

Type — Indonesia: *Erythrina ovalifolia* (Fabaceae).

The type was not found in L, and its preservation is uncertain. Material studied (Surinam, Paramaribo, on spots on upper surface of living leaves of *Erythrina glauca*, van Suchtelen, 1956, CBS) contains *Asteromella*, *Phyllosticta* and *Septoria*.

Mycosphaerella erythrinae F. Stevens. See *Mycosphaerella stevensii* Tomilin.

Mycosphaerella erythrincola Syd., Ann. Mycol. 28: 80. 1930.— Fig. 306.

Type — Venezuela: El Limón, Puerto La Cruz. On spots on upper surface of living leaves of *Erythrina micropteryx* (Fabaceae). Sydow, I 1928, distributed in Fungi Exotici Exsiccati no. 812, also distributed in Reliquiae Petrakianae no. 651 (H, L (3×), isotypes).

This is a parasitic species, with asci cylindrical, ascospores 15-17 × 2.5-4.5 µm. It was cited as synonymous with *Mycosphaerella erythrinae* Koord. by Tomilin (1979).

Mycosphaerella erythroxyli (Speg.) M. Morelet, Ann. Soc. Sci. Nat. Archéol. Toulon & Var 20: 105. 1968 = *Sphaerella erythroxyli* Speg., Anales Soc. Ci. Argent. 90: 27. 1921 [“1920”].— Fig. 307.

Type — Argentina: On brown spots on upper surface of living leaves of *Erythroxyllum coca* (Rhizophoraceae).

Anamorph (presumed): *Phyllosticta erythroxyli* Graziani fide Spegazzini (op. cit.).

The type was not included in a loan from LPS. Material studied (Bolivia, intercepted in New York, Wong & Musniicky, X 1971, BPI) is a parasitic species, with asci cylindrical, ascospores 13.5-15 × 4.4-5.5 µm.

Mycosphaerella escalloniae (Speg.) Sivan. & R.G. Shivas, Mycol. Res. 106: 357. 2002 = *Sphaerella escalloniae* Speg., Anales Mus. Nac. Hist. Nat. Buenos Aires 19 (ser. 3, 12): 351. 1909.

Type — Argentina: *Escallonia rosea* (Saxifragaceae).

No material was studied as the type was not included in a loan from LPS.

Sphaerella etrusca Tognini, Atti. Ist. Bot. Univ. Pavia, n. ser. 3: 52. 1893.

Type — Italy: *Castanea vesca* (Fagaceae).

No material was studied as the type was not found in any of the herbaria consulted.

Mycosphaerella eucalypti (Wakef.) Hansf., Proc. Linn. Soc. New South Wales 82: 218. 1957 = *Hypospila eucalypti* Wakef., Bull. Misc. Inform. 1912: 190. 1912.

Type — Australia. On leaves of *Eucalyptus* (Myrtaceae). Wakefield (K, holotype).

Better disposed as *Hypospila eucalypti* Wakef.

Sphaerella eucarpa P. Karst., Öfvers. Förh. Kongl. Svenska Vetensk.-Akad. 1872(2): 103. 1872 = *Wettsteinina eucarpa* (P. Karst.) E. Müll. & Arx, Ber. Schweiz. Bot. Ges. 60: 335. 1950.

Type — Sweden: *Polygonum viviparum* (Polygonaceae).

Accepted as *Wettsteinina eucarpa* (P. Karst.) E. Müll. & Arx by Eriksson (1992).

Mycosphaerella eugeniae Rehm, Hedwigia 44: 4. 1905 = *Sphaerella eugeniae* (Rehm) Sacc. & D. Sacc., Syll. Fung. 17: 637. 1905.

Type — Brazil: On white spots with black margins on upper surface of living leaves of *Eugenia michelii* (Myrtaceae).

The type could not be found in S or in any of the herbaria consulted. Material studied (Dominica, Hillsborough

station, Critchett, III 1972, IMI no. 165165) contains a *Guignardia*.

Mycosphaerella eugenicola Crous, Alfenas & R.W. Barreto, Mycotaxon 64: 425. 1997.

Type — Brazil: Rio de Janeiro, Cabo Frio. On leaves of *Eugenia uniflora* (Myrtaceae). Viégas & Krug (IACM 2997, holotype, not seen).

Anamorph: *Pseudocercospora sphaerellae-eugeniae* (Sacc.) Crous, Alfenas & R.W. Barreto *vide* Crous, Alfenas & Barreto (op. cit.).

Synonymised with *Mycosphaerella eugeniae* Rehm by the original author of the new species, Crous (1999).

Mycosphaerella eulaliae (Pass.) Tomilin, Novosti Sist. Nizsh. Rast. 1967: 189. 1967 = *Sphaerella eulaliae* Pass., Atti Reale Accad. Lincei, Mem. Cl. Sci. Fis., ser. 4, 3: 9. 1887.

Type — Italy: *Eulalia japonica* (Poaceae).

Cited as synonymous with *Davidiella allicina* (as *Mycosphaerella allicina*) by Tomilin (1979). No material was studied as the type was not found in any of the herbaria consulted.

Sphaerella eumorpha (Berk. & M.A. Curtis) Cooke, J. Bot. 21: 137. 1883 = *Sphaeria eumorpha* Berk. & M.A. Curtis, in Berk., Grevillea 4: 145. 1876 = *Leptosphaeria eumorpha* (Berk. & M.A. Curtis) Earle, Bull. Torrey Bot. Club 25: 361. 1898.

Type — USA: *Arundinaria* (Poaceae). (K, isotype).

Already accepted as *Leptosphaeria eumorpha* by Saccardo, and therefore no type material studied.

Mycosphaerella eumusae Crous & Mourichon, Sydowia 54: 36. 2002.

Type — Réunion: *Musa* (Musaceae).

Anamorph: *Pseudocercospora eumusae* Crous & Mourichon *vide* Crous & Mourichon (op. cit.).

No material was studied of this recently described species.

Mycosphaerella euodiae J.F. Lue & P.K. Chi, in P.K. Chi, Fungal Diseases of Cultivated Medicinal Plants in Guangdong Province: 141. 1994 [as “*evodiae*”].

Type — China: *Euodia rutaecarpa* (Rutaceae).

No material was studied as the location of the type is unknown and its preservation uncertain.

Mycosphaerella euonymi (G. Kunze) J. Schröt., in Cohn, Kryptog.-Fl. Schlesien 3(2): 334. 1894 [“1893”] = *Sphaeria euonymi* G. Kunze, in Fr., Systema Mycol. 2: 439. 1823 [as “*evonymi*”] = *Sphaerella euonymi* (G. Kunze) Rabenh., Fungi Europaei Exsiccati, ed. nov., ser. 2. no. 657. 1864.— Fig. 308.

Type — France: Calvados, Caen. On upper and lower surface of dead leaves of *Euonymus europaeus* (Celastraceae). Roberge, F-03649, X 1864 (UPS-FRIES, holotype).

The holotype is an immature ascomycete. Additional material studied (Czech Republic, Starnislan Czarny, Petr., IV 1918, L) is *M. punctiformis*, with asci cylindrical, ascospores 10-12 × 2.5-3 µm.

Mycosphaerella eupatorii W.Y. Yen, Cah. Pacifique 13: 272. 1969.

Type — Malaysia: *Eupatorium odoratum* (Asteraceae).

No material was studied as the location of the type is unknown and its preservation uncertain.

Mycosphaerella eupatoriicola Höhn., in Strasser, Verh. Zool.-Bot. Vereins Wien 69: 361. 1919 [as “*eupatoriicola*”] = *Sphaerella eupatoriicola* (Höhn.) Trotter, Syll. Fung. 24: 857. 1928.

Type — Austria: Niederösterreich, Sonntagsberg. On upper and lower surface of dead leaves of *Eupatorium cannabinum* (Asteraceae). Strasser, V 1916 (FH-Höhnel, holotype; W, 4 isotypes).

This is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores 8-10 × 2.5-3.5 µm. It was cited as synonymous with *Mycosphaerella serratulae* by Tomilin (1979).

Mycosphaerella eupatoriicola Petr., Ann. Mycol. 19: 277. 1921, later homonym (illegitimate, Article 53).— Fig. 309.

Type — Czech Republic: Hrabuvka. On dead stems of *Eupatorium cannabinum* (Asteraceae). Petr., VI 1917 (L, isotype).

Anamorph: Probably *Septoria eupatorii* Roberge & Desm. *vide* Cash (1972).

The type and additional material studied (Czech Republic, Weißkirchen, Hranice, Petrak, VI 1916, distributed in Reliquiae Petrakianae no. 2473, L) belong to section *Caterva*, and show that this is morphologically indistinguishable from *M. subradicans*, with asci cylindrical, ascospores 11-13 × 3.5-4 µm.

Mycosphaerella euphorbiae Niessl ex J. Schröt., in Cohn, Kryptog.-Fl. Schlesien 3(2): 338. 1894 [“1893”].— Fig. 311.

Type — Poland. On dead stems of *Euphorbia cyparissias* (Euphorbiaceae).

Material studied (Germany, Oberbarnim, Choriner Forst, Kirschstein, VI 1917, B) is *M. punctiformis*, with asci cylindrical, ascospores 8-9 × 2.5-3 µm.

Sphaerella euphorbiae Phillips & Plowr., Grevillea 6: 28. 1877 = *Physalospora euphorbiae* (Phillips & Plowr.) Sacc., Syll. Fung. 1: 436. 1882.— Fig. 310.

Type — United Kingdom: Herefordshire, Dinmore. On dead stems of *Euphorbia amygdaloides* (Euphorbiaceae). Plowright, 1876, distributed in Sphaeriacei Britannici no. 97 (K, holotype; B, isotype).

This is morphologically indistinguishable from *Glomerella cingulata* (Stoneman) Spauld. & H. Schrenk, for which it would represent an earlier epithet, with paraphysoids and aseptate ascospores of 15-18 × 5-7 µm.

Mycosphaerella euphorbiae-canariensis Tomilin, Novosti Sist. Nizsh. Rast. 6: 120. 1970 [“1969, as *euphorbiae canariensis*”], nomen novum (Article 58) for *Sphaerella canariensis* Tassi, Bull. Lab. Orto Bot. Reale Univ. Siena 3: 118. 1900.

Type — China: *Euphorbia canariensis* (Euphorbiaceae).

No material was studied as the type was not found in SIENA, where many other Tassi types were preserved.

Mycosphaerella euphorbiae-exiguae (Unamuno) Tomilin, Novosti Sist. Nizsh. Rast. 6: 120. 1970 ["1969"] = *Sphaerella euphorbiae-exiguae* Unamuno, Bol. Soc. Esp. Hist. Nat. 30: 383. 1930.

Type — Spain: *Euphorbia serrata* (Euphorbiaceae).

No material was studied as the type was not found in MA, where many other Unamuno types were preserved.

Mycosphaerella euphorbiae-spinosae (De Not.) Lindau, in Engl. & Prantl, *Natürlichen Pflanzenf.* 1(1): 424. 1897 = *Sphaerella euphorbiae-spinosae* De Not., *Comment. Soc. Crittog. Ital.* 2(3): 487. 1867.

Type — Italy: Iglesias. On dead branches of *Euphorbia spinosa* (Euphorbiaceae). 1835 (RO, holotype).

This is morphologically indistinguishable from *Didymosphaeria futilis* (Berk. & Broome) Rehm, with anastomosing paraphysoids, brown, ornamented, uniseriate ascospores of 8-10 × 4-5 µm.

Sphaeria euphorbiicola Schwein., *Trans. Amer. Philos. Soc.*, ser. 2, 4: 207. 1832 = *Sphaerella euphorbiicola* (Schwein.) Ellis & Everh., *N. Amer. Pyrenomyc.*: 745. 1892.

Type — USA: Philadelphia, Bethlehem. On dead stems of *Euphorbia* (Euphorbiaceae). Schweinitz (PH, holotype).

The holotype contains only an *Ascochyta*.

Sphaerella euphrasiae Pass., *Atti Reale Accad. Lincei*, *Mem. Cl. Sci. Fis.*, ser. 4, 3: 7. 1887.

Type — Italy: *Euphrasia lutea* (Orobanchaceae).

No material was studied as the type was not found in any of the herbaria consulted.

Mycosphaerella euryae Theiss., *Ann. Mycol.* 16: 187. 1918 = *Sphaerella euryae* (Theiss.) Trotter, *Syll. Fung.* 24: 888. 1928.

Type — Japan: *Eurya chinensis* (Theaceae).

No material was studied as the type was not found in any of the herbaria consulted.

Mycosphaerella euryptami Kohlm., *Volk.-Kohlm. & O.E. Erikss.*, *Botan. Mar.* 42: 505. 1999.

Type — USA: *Juncus roemerianus* (Juncaceae).

No material was studied of this recently described species.

Mycosphaerella evansiae (Pat.) Lindau, in Engl. & Prantl, *Natürlichen Pflanzenf.* 1(1): 426. 1897 = *Sphaerella evansiae* Pat., *Rev. Mycol. (Toulouse)* 8: 83. 1886.

Type — Tibet: Province de Moupin. On upper surface of dead leaves of "*Evansia*" *fimbriata* (Iridaceae, not Hepaticae, Cephaloziellaceae). Abbé David, 1870 (FH-Patouillard, holotype).

The holotype contains only an empty ascomycete.

Mycosphaerella evernia (Syd.) Petr., distributed in Reliquiae Petrakianae no. 652 = *Haplodothis evernia* Syd., *Ann. Mycol.* 37: 374. 1939.— Fig. 312.

Type — Ecuador: Prov. Tungurahua, Baños. On lower surface of brown areas on dead leaves of *Myrica pubescens* (Myricaceae). Sydow, distributed in Reliquiae Petrakianae no. 652, (L, NY, isotypes).

This is probably a species of Parmulariaceae, with ascomata delineated in stromata, ostioles linear, asci cylindrical, with short paraphyses, ascospores 30-35 × 3-3.5 µm.

Mycosphaerella exaci T.S. Ramakr. & K. Ramakr., *Proc. Indian Acad. Sci. Sect. B*, 32: 205. 1950.

Type — India: *Exacum wightianum* (Gentianaceae).

No material was studied as the location of the type is unknown and its preservation uncertain.

Mycosphaerella exarida (G. Winter) Starbäck, *Ark. Bot.* 5(7): 21. 1905 = *Sphaerella exarida* G. Winter, *Rev. Mycol. (Toulouse)* 7: 207. 1885.

Type — Paraguay: Sapindaceae.

No material was studied as the type was not found in any of the herbaria consulted.

Mycosphaerella exigua Syd. & P. Syd., *Ann. Mycol.* 11: 58. 1913 = *Sphaerella exigua* (Syd. & P. Syd.) Trotter, *Syll. Fung.* 24: 886. 1928.— Fig. 313.

Type — Japan: Mino, Kawanyu-mura. On upper and lower surface of dead leaves of *Schizophragma hydrangioides* (Saxifragaceae). Hara, IV 1912 (S, holotype).

This is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores 7-9 × 2-2.5 µm.

Mycosphaerella exitialis (Morini) Lind, *Danish Fungi*: 205. 1913 = *Sphaerella exitialis* Morini, *Nuovo Giorn. Bot. Ital.* 18: 37. 1886 = *Didymella exitialis* (Morini) E. Müll., *Phytopathol. Z.* 19: 407. 1952.

Type — Italy: *Triticum vulgare* (Poaceae).

Anamorph: *Ascochyta fide* Müller (op. cit.).

Material studied (Austria, Tirol, Längenfeld, VIII 1897, B) contains no fungi.

Mycosphaerella exutans (Cooke) Miles, *Trans. Illinois State Acad. Sci.* 10: 251. 1917 = *Sphaerella exutans* Cooke, *Ann. New York Acad. Sci.* 1: 187. 1879.

Type — USA: Texas, Harris Co. On brown spots on upper surface of dead leaves of *Persea carolinensis* (Lauraceae). Ravenel no. 46, III 1869 (K, holotype; NY, isotype).

The holotype contains only two other fungi, a Meliolaceae and *Eustegia affinis*, the isotype belongs to section *Caterva*, and is morphologically indistinguishable from *M. subadians*, with ascomata erumpent, lifting off flaps of the epidermis, asci cylindrical, ascospores 10-12 × 2.5-3 µm. None of these species agree with the protologue.

Mycosphaerella fagi (Auersw.) Lindau, in Engl. & Prantl, *Natürlichen Pflanzenf.* 1(1): 424. 1897 = *Sphaerella fagi* Auersw., in Gonn. & Rabenh., *Mycol. Europaea* 5-6: 6. 1869.

Type — Germany: On upper and lower surface of dead leaves of *Fagus sylvatica* (Fagaceae).

Material studied (Brandenburg, Eberswalde, Spechthausen, Sydow, VII 1917, Mycotheca Germanica no. 1541, L) is

probably *M. punctiformis*, with asci cylindrical, ascospores immature.

Sphaerella fagicola (Fr.) Auersw., in Gonn. & Rabenh., Mycol. Europaea 5-6: 7. 1869 = *Sphaeria fagicola* Fr., Systema Mycol. 2: 529. 1823.

Type — Sweden: On upper and lower surface of dead leaves of *Fagus sylvatica* (Fagaceae).

Material studied (Germany, Oestrich, Fuckel, Fungi Rhenani no. 1935, L) is probably *M. punctiformis*, with asci cylindrical, ascospores immature.

Sphaerella faginea Cooke & Plowr., J. Bot. 21: 68. 1883 = *Laestadia faginea* (Cooke & Plowr.) Sacc., Syll. Fung. 2: XXXI. 1883.

Type — United Kingdom: Lynn. On upper and lower surface of dead leaves of *Fagus sylvatica* (Fagaceae). Plowright, 1878, distributed in Sphaeriacei Britannici no. 100 (K, holotype; B, isotype).

The holotype and isotype contain various immature species, probably including *M. punctiformis* and *Glomerella cingulata* (Stoneman) Spauld. & H. Schrenk.

Mycosphaerella fagraeae W.Y. Yen, Bull. Soc. Mycol. France 96: 23. 1980.

Type — Malaysia: *Fagraea fragans* (Loganiaceae). No material was studied as the location of the type is unknown and its preservation uncertain.

Mycosphaerella falcariae Syd., Ann. Mycol. 40: 202. 1942.— Fig. 314.

Type — Germany: ["Poland"], Brandenburg, Tamsel, Küstrin. On dead stems of *Falcaria rivinii* (Apiaceae). Vogel, distributed in Sydow, Mycotheca Germanica no. 3504, also distributed in Reliquiae Petrakianae no. 2474, V 1937 (H, L, isotypes).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradians*, with asci cylindrical, ascospores 11-13 × 2.5-3.5 µm. It was cited as synonymous with *Mycosphaerella lysimachiae* by Tomilin (1979).

Sphaerella fallax Auersw., in Gonn. & Rabenh., Mycol. Europaea 5-6: 6. 1869, nomen nudum (illegitimate, Article 32).

Authentic material — Germany: *Fagus sylvatica* (Fagaceae).

This was cited as synonymous with *Mycosphaerella fagi*, which is morphologically indistinguishable from *M. punctiformis*, by Auerswald (op. cit.). No material was studied as none was found in B or in any of the other herbaria consulted.

Mycosphaerella familiaris (Auersw.) Lindau, Hilfsb. Sammeln Ascomyceten: 93. 1903 = *Sphaerella familiaris* Auersw., in Gonn. & Rabenh., Mycol. Europaea 5-6: 6. 1869.

Type — Germany: *Quercus* (Fagaceae).

This is no *Mycosphaerella* according to Tomilin (1979). No material was studied as the type was not found in B or in any of the other herbaria consulted.

Mycosphaerella feijoae Artemiev, Sovetsk. Subtrop. (Sukhumi) 7: 62. 1935, lacking Latin description (not validly published, Article 36.1).

Type — Georgia: *Feijoa sellowiana* (Myrtaceae).

Anamorphs: *Phyllosticta* and *Septoria fide* Tomilin (1979).

No material was studied as the location of the type is unknown and its preservation uncertain.

Mycosphaerella ["*Mycosphaerium*"] *fendlerae* Clem., Cryptogamae Formationum Coloradensium no. 419. 1908, nomen herbariorum (not validly published, Article 32).— Fig. 315. Authentic material — USA: Colorado, Durango. On upper and lower surface of dead leaves of *Fendlera rupicola* (Saxifragaceae). Clements, VI 1907, Cryptogamae Formationum Coloradensium no. 419 (BPI).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. clandestina*, with asci pyriform, ascospores thick-walled, 21-25 × 7-8.5.

Mycosphaerella fendleri Tracy & Earle, in Greene, Plantae Bakerianae 1(1): 33. 1901 = *Sphaerella fendleri* (Tracy & Earle) Sacc. & P. Syd., Syll. Fung. 16: 1133. 1902.— Fig. 316.

Type — USA: Colorado, La Plata Mts., Mt. Herperus, Botanical Creek. On dead stems of *Thalictrum fendleri* (Ranunculaceae). Baker no. 1091, VI 1898 (NY, 2 isotypes).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores 14-17 × 5.5-7 µm.

Mycosphaerella fennica (P. Karst.) Tomilin, Novosti Sist. Nizsh. Rast. 6: 120. 1970 ["1969"] = *Sphaerella fennica* P. Karst., Bidrag Kännedom Finlands Natur Folk 23: 173. 1873 = *Didymella fennica* (P. Karst.) Arx, in E. Müll. & Arx, Beitr. Kryptogamenfl. Schweiz 11(2): 366. 1962.— Fig. 317.

Type — Finland: Tavastia australis, Tammela, Mustiala. On dead petioles of *Tilia parvifolia* (Malvaceae). Karsten, Fungi Fennici Exsiccati no. 3134, V 1866 (H, lectotype).

This is indeed a species of *Didymella*, and could be accepted as *Didymella fennica* (P. Karst.) Arx, with ostiole papillate, sparse paraphysoids, asci cylindrical, ascospores 16-19 × 5-6.6 µm.

Mycosphaerella ferruginea (Fuckel) Cruchet, Bull. Soc. Vaud. Sci. Nat. 55: 47. 1923 = *Sphaerella ferruginea* Fuckel, Jahrb. Nassauischen Vereins Naturk. 27-28: 20. 1873.

Type — Germany: Oestrich. On lower surface of living leaves of *Artemisia vulgaris* (Asteraceae). Fuckel, Fungi Rhenani no. 496 (BPI, isotype); also no. 2435 (L, isotype).

Anamorphs: *Cercospora ferruginea* Fuckel (= *Passalora ferruginea* (Fuckel) U. Braun & Crous) and *Phoma fide* Fuckel (op. cit.).

The isotypes studied contain only a *Phoma* and other coelomycetes.

Mycosphaerella ferulae (Maffei) Tomilin, Novosti Sist. Nizsh. Rast. 7: 203. 1970 = *Sphaerella ferulae* Maffei, Atti Ist. Bot. Univ. Pavia, ser. 2, 11: 30. 1905.— Fig. 979.

Type — Italy: Liguria, Isla Gallinara. On dead stems of *Ferula communis* (Apiaceae). Ex herbario Ciferri (BPI, possible isotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. ammophilae*, with asci pyriform, ascospores $7-9 \times 1.5-2.5 \mu\text{m}$. The dimensions are not in accordance with the protologue.

Sphaerella festucae (Lib.) Auersw., in Gonn. & Rabenh., Mycol. Europaea 5-6: 16. 1869 \equiv *Sphaeria festucae* Lib., Plantae Cryptogamicae Arduennae, cent. 3 no. 246. 1834 \equiv *Physalospora festucae* (Lib.) Sacc., Syll. Fung. 1: 434. 1882.— Fig. 318.

Type — Belgium: Malmedy. On upper and lower surface of dead leaves of *Festuca sylvatica* (Poaceae). Libert, Plantae Cryptogamicae Arduennae no. 246 (BR, holotype).

This is morphologically indistinguishable from *Glomerella cingulata* (Stoneman) Spauld. & H. Schrenk, for which it would represent an earlier epithet, with paraphysoids and aseptate ascospores of $20-25 \times 10-12 \mu\text{m}$.

Mycosphaerella fibrillosa (Syd. & P. Syd.) Joa.E. Taylor & Crous, Mycol. Res. 107: 657. 2003 \equiv *Teratosphaeria fibrillosa* Syd. & P. Syd., Ann. Mycol. 10: 40. 1912.

Type — South Africa: *Protea grandiflora* (Proteaceae). No material was studied of this recently described species.

Mycosphaerella fici-ovatae Hansf., Proc. Linn. Soc. London 153: 22. 1941.

Type — Uganda: *Ficus ovata* (Moraceae).

No material was studied as the type is not in IMI or K.

Mycosphaerella fici-wightianae Sawada, Rep. Gov. Res. Inst. Formosa 85: 33. 1943, lacking Latin description (not validly published, Article 36.1).

Type — Taiwan: *Ficus wightiana* (Moraceae).

Anamorph: *Cercospora* fide Sawada (op. cit.).

No material was studied as the type is not in BPI, where many Sawada isotypes are kept.

Mycosphaerella ficophila (G. Winter) Cruchet, Bull. Soc. Vaud. Sci. Nat. 55: 43. 1923 [as “*sycophila*”] \equiv *Sphaerella ficophila* G. Winter, Hedwigia 25: 101. 1886.

Type — Namibia: *Ficus* (Moraceae).

No material was studied as the type was not found in any of the herbaria consulted.

Mycosphaerella ficus (Traverso & Spessa) Tomilin, Novosti Sist. Nizsh. Rast. 1968: 165. 1968 \equiv *Sphaerella ficus* Traverso & Spessa, Bol. Soc. Brot. 25: 170. 1910.— Fig. 319.

Type — Portugal: Coimbra. On lower surface of dead leaves of *Ficus macrophylla* (Moraceae). Moller, XII 1903 (PAD, holotype).

This is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores $8-10 \times 2-2.5 \mu\text{m}$.

Mycosphaerella fijiensis M. Morelet, Ann. Soc. Sci. Nat. Archéol. Toulon & Var 21: 105. 1969.

Type — Hawaii: Moanoa, Honolulu. On white to brown spots on upper surface of living to decaying leaves of *Musa* “Dwarf Cavendish” (Musaceae). Meredith, XII 1968 (BPI, IMI no. 136696, isotypes).

Anamorph: *Paracercospora fijiensis* (M. Morelet) Deighton fide Deighton (1979) (= *Pseudocercospora fijiensis* (M. Morelet) Deighton).

The isotypes contain only the anamorph.

Mycosphaerella fijiensis var. *difformis* Miller & Stover, Trans. Brit. Mycol. Soc. 67: 82. 1976.— Fig. 320.

Type — Honduras: La Lima. On white spots with black margins on upper surface of living leaves of *Musa* (Musaceae). Stover, III 1974 (IMI no. 183747, isotype).

Anamorph: *Paracercospora fijiensis* var. *difformis* (J.M. Mulder & Stover) Deighton fide Deighton (1979) (= *Pseudocercospora fijiensis* (M. Morelet) Deighton).

This is a parasitic species, with asci cylindrical, ascospores $12-14 \times 3-4 \mu\text{m}$. Currently accepted as synonym of *M. fijiensis* based on morphology and DNA analyses.

Mycosphaerella filicum (Desm.) Starbäck, Bih. Kongl. Svenska Vetensk.-Akad. Handl. 15(3, 2): 9. 1889 \equiv *Sphaeria filicum* Desm., Plantes Cryptog. France, fasc. 20 no. 283. 1839 \equiv *Sphaerella filicum* (Desm.) Ces. & De Not., Comment. Soc. Crittogam. Ital. 1(4): 237. 1863.— Fig. 321.

Type — France: On upper and lower surface of dead leaves of *Asplenium adiantum-nigrum* (Polypodiaceae). Desmazières, Plantes Cryptog. France, fasc. 20 [“Fungi Exsiccati”] no. 283 (L, isotype).

This belongs to section *Caterva*, with asci cylindrical, ascospores $11-16 \times 3-4 \mu\text{m}$. Additional material (Netherlands, Putten, on upper and lower surface of dead fronds of *Dryopteris filix-mas* (Polypodiaceae), Oudemans, 1884, L) is *M. punctiformis*.

Mycosphaerella filipendulae Tomilin. See *Mycosphaerella maculans* Lindau.

Mycosphaerella filipendulae-denudatae Kamilov, Mikol. Fitopatol. 7: 360. 1973.

Type — Russia: Leningrad, Petrodvorets. On upper and lower surface of dead leaves of *Filipendula ulmaria* var. *denudata* (Rosaceae). Kamilov, X 1969 (LEP, holotype).

Anamorphs: *Phyllosticta* and *Ramularia spiraeae* Peck fide Tomilin (1979).

The type is mostly moulded and only contains a *Phoma*.

Mycosphaerella filipendulae-ulmariae Tomilin. See *Mycosphaerella maculans* Lindau.

Mycosphaerella firmianae (Pass.) Tomilin, Novosti Sist. Nizsh. Rast. 6: 120. 1970 [“1969”] \equiv *Sphaerella firmianae* Pass., Atti Reale Accad. Lincei, Mem. Cl. Sci. Fis., ser. 4, 3: 5. 1887.

Type — Italy: *Firmiana platanifolia* (Malvaceae).

No material was studied as the type was not found in any of the herbaria consulted.

Mycosphaerella flagellariae Alcorn, Queensland Naturalist 22: 58. 1978.— Fig. 322.

Type — Australia: Hinchinbrook Island. On white spots with black margins on upper surface of living leaves of *Flagellaria indica* (Flagellariaceae). Simmonds, VIII 1975 (IMI no. 201920, isotype).

This is a parasitic species, with asci cylindrical, ascospores $10\text{-}12 \times 2.5\text{-}3 \mu\text{m}$.

Mycosphaerella flageoletiana (Sacc. & Traverso) Tomilin, Novosti Sist. Nizsh. Rast. 1968: 165. 1968 = *Sphaerella flageoletiana* Sacc. & Traverso, in Sacc., Rendiconti Congr. Bot. Palermo 1902: 51. 1902.— Fig. 323.

Type — France: Rigny. On upper and lower surface of dead leaves of *Fagus sylvatica* (Fagaceae). Flageolet (PAD, holotype).

This belongs to section *Longispora*, and is morphologically indistinguishable from *M. millegrana*, with asci cylindrical, ascospores $17\text{-}20 \times 2\text{-}2.5 \mu\text{m}$. Additional material studied (Czech Republic, Weißkirchen, Ungersdorf, Petrak, Flora Bohemia et Moravia exsiccata no. 1239, V 1914, L) is *M. punctiformis*, with asci cylindrical, ascospores $7\text{-}9 \times 2.5\text{-}3.5 \mu\text{m}$.

Mycosphaerella flexuosa Crous & M.J. Wingf., Mycologia Memoir 21: 58. 1998.

Type — Colombia: La Selva. On leaves of *Eucalyptus globulus* (Myrtaceae). Wingfield, V 1995 (PREM 54401, holotype, not seen).

No material was studied of this recently described species.

Sphaerella foeniculacea (Mont.) Cooke, J. Bot. 21: 70. 1883 = *Sphaeria foeniculacea* Mont., Ann. Sci. Nat. Bot., ser. 3, 11: 40. 1849 = *Physalospora foeniculacea* (Mont.) Sacc., Syll. Fung. 1: 445. 1882 = *Guignardia foeniculacea* (Mont.) Arx & E. Müll., Beitr. Kryptogamenfl. Schweiz. 11(1): 48. 1954 [as “foeniculata”].

Type — France: *Anethum foeniculum* (Apiaceae).

Accepted as *Guignardia foeniculacea* (Mont.) Arx & E. Müll. by von Arx & Müller (op. cit.).

Mycosphaerella foeniculi Komirn., Uchen. Zap. Moskovsk. Gosud. Univ. 35: 138. 1952 = *Mycosphaerella foeniculicola* Khokhr., Opredelel'boleznei rastenii: 291, 1956, superfluous (illegitimate, Article 52) nomen novum (Article 58).

Type — Russia: *Anethum foeniculum* [as “*Foeniculum officinale*”] (Apiaceae).

Anamorphs: *Cercospora depressa* f. *foeniculi* Komirnaya (= *Passalora punctum* (Delacr.) S. Petzoldt) and *Phoma anethi* (Pers. : Fr.) Sacc. fide Komirnaya (op. cit.).

No material was studied as the type was not included in loans from LE or LEP.

Mycosphaerella foeniculi (Speg.) Tomilin, Novosti Sist. Nizsh. Rast. 7: 203. 1970, later homonym (illegitimate, Article 53) = *Sphaerella foeniculi* Speg., Anales Mus. Nac. Hist. Nat. Buenos Aires 19 (ser. 3, 12): 353. 1909.

Type — Argentina: *Foeniculum piperitum* (Apiaceae).

No material was studied as the type was not included in a loan from LPS.

Mycosphaerella foeniculicola Khokhr. See *Mycosphaerella foeniculi* Kormirnaya.

Mycosphaerella foeniculina (Speg.) Tomilin, Novosti Sist. Nizsh. Rast. 7: 203. 1970 = *Sphaerella foeniculina* Speg., Revista Fac. Agron. Univ. Nac. Vet. La Plata, ser. 2, 6: 54. 1910.

Type — Chile: *Foeniculum piperitum* (Apiaceae).

No material was studied as the type was not included in a loan from LPS.

Mycosphaerella formosana T.Y. Lin & W.Y. Yen, Rev. Mycol. (Paris) 35: 323. 1971.

Type — Taiwan: *Musa* (Musaceae).

No material was studied as the location of the type is unknown and its preservation uncertain.

Mycosphaerella fragariae (Schwein.) Johanson ex Oudem., Rév. Champ. Pays-Bas 2: 216. 1897 = *Sphaeria fragariae* Schwein., Trans. Amer. Philos. Soc., ser. 2, 4: 223. 1834 (non Tul., Ann. Sci. Nat. Bot., ser. 4, 5: 112. 1856) = *Stigmatea fragariae* (Schwein.) Tul., Selecta Fungorum Carpologia 2: 286. 1863 = *Sphaerella fragariae* (Schwein.) Sacc., Syll. Fung. 1: 505. 1882.— Fig. 324.

Type — USA: Philadelphia, Bethlehem. On lower surface of dead leaves of *Fragaria vesca* (Rosaceae). Schweinitz no. 1769 (BPI, isotype).

Anamorphs: *Ramularia brunnea* Peck fide Sivanesan (1984), *Ramularia tulasnei* Sacc. fide Eriksson (1992). According to Tomilin (1979), associations have also been reported with *Ascochyta fragariae* Lib., *Cylindrosporium grevilleanum* Tul. (= *Ramularia grevilleana* (Oudem.) Jørst.), *Depazea fragariaecola* Wallr., *Graphium phyllogenum* Desm., *Phyllosticta fragariaecola* Desm. and *Septoria fragariae* (Lib.) Desm.

The holotype was not found in PH, but an isotype was still preserved in BPI. It is however overmature. Therefore the only mature material found amongst dozens of specimens preserved under this name is selected here as epitype:

Epitype — Germany: Oestrich. Fuckel, Fungi Rhenani no. 2018 (L, epitype, here designated). It is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores $16\text{-}21 \times 5\text{-}6.5 \mu\text{m}$. Most additional material attributed to this species (e.g. Netherlands, Baarn, van Steenis, VIII 1923, L) contains only coelomycetes or even no fungi at all (just wilt).

Mycosphaerella frankeniae (Unamuno) Cash, Syll. Fung. 26: 339. 1972 = *Sphaerella frankeniae* Unamuno, Bol. Soc. Esp. Hist. Nat. 30: 292. 1930.

Type — Spain: Huelva, La Rabida. On upper surface of dead leaves of *Frankenia boissieri* (Frankeniaceae). Unamuno no. 9175, IV 1929 (MA, holotype).

The type contains only a *Phyllosticta*.

Mycosphaerella frauxii M. Morelet, Bull. Soc. Sci. Nat. Archéol. Toulon & Var 175: 5. 1968, nomen novum (Article 58) for *Mycosphaerella cassiae* F. Stevens, Illinois Biol. Monogr. 11: 199. 1927, later homonym (illegitimate, Article 53).— Fig. 326.

Type — Costa Rica: Siquirris. On brown spots with black margins on upper surface of living leaves of *Cassia* (Fabaceae). Stevens no. 724, VIII 1923 (BPI, holotype; BPI, isotype).

This belongs to *Davidiella*, of which it represents a parasitic species, and is morphologically indistinguishable from *D. ariadna*.

Sphaerella fraxinea Peck, Annual Rep. New York State Mus. 35: 145. 1884.

Type — USA: New York, Helderberg Mts. On upper and lower surface of dead leaves of *Fraxinus americana* (Oleaceae). Peck, V (NY, isotype).

Cited as synonymous with *Mycosphaerella effigurata* by Tomilin (1979). The isotype is immature, but shows that this is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical.

Mycosphaerella fraxini (Niessl) Lindau, Hilfsb. Sammeln Ascomyceten: 47. 1903 = *Sphaerella fraxini* Niessl, in Linhart, Fungi Hungarici no. 162. 1883.— Fig. 325.

Type — Hungary: Altenburg (Markt-Au). On lower surface of dead leaves of *Fraxinus excelsior* (Oleaceae). Linhart, V 1883 (M, holotype), also distributed in Linhart, Fungi Hungarici no. 162 (M, isotype), also distributed in Rehm, Ascomyceten no. 738 (M, isotype).

Anamorphs: *Cylindrosporium fraxini* (Ellis & Kellerm.) Ellis & Everh. (= *Pseudocercospora fraxini* (Ell. & Kellerm.) U. Braun), *Phyllosticta* and *Septoria fraxini* Desm. *vide* Tomilin (1979).

The types and other material studied (Czech Republic, Weißkirchen, Hrabuvka, Petrak, Flora Bohemiae et Moraviae exsiccata no. 780, V 1913, L) belong to section *Longispora*, and it is morphologically indistinguishable from *M. latebrosa*, with ascomata in stromata, asci cylindrical, ascospores 23-28 × 2.5-3 µm.

Mycosphaerella fraxinicola (Schwein.) House, New York State Mus. Bull. 233-234: 27. 1921 = *Sphaeria fraxinicola* Schwein., Trans. Amer. Philos. Soc., ser. 2, 4: 224. 1832 [as “*fraxicola*”] = *Sphaerella fraxinicola* (Schwein.) Cooke, J. Bot. 21: 107. 1883.

Type — USA: Philadelphia, Bethlehem. On lower surface of dead leaves of *Fraxinus americana* (Oleaceae). Schweinitz (BPI, isotype).

Anamorph: *Phyllosticta viridis* Ellis & Kellerm. *vide* Wolf (1939).

No type material was preserved in PH, but an isotype was still present in BPI. The isotype is partly immature, partly overmature, but shows that this is morphologically indistinguishable from *M. punctiformis*, with ascomata in groups.

Mycosphaerella frenumbensis (Speg.) de Barros, Garcia de Orta, ser. Estud. Agron. 1: 33. 1973 = *Sphaerella frenumbensis* Speg., Revista Fac. Agron. Univ. Nac. La Plata 15 (or ser. 2, 2): 19. 1908.

Type — Brazil: Phoenix [“Palma”] *pinnatifida* (Arecaceae). Reported by Fröhlich & Hyde (1998) as morphologically indistinguishable from *Mycosphaerella cocoës*, which is a

later synonym at species level. No material was studied as the type was not included in a loan from LPS.

Mycosphaerella freycinetiae F. Stevens, Bernice P. Bishop Mus. Bull. 19: 103. 1925.— Fig. 327.

Type — Hawaii: Oahu, Kaliki Valley. On white spots with brown margins on upper surface of living leaves of *Freycinetia arnotti* (Pandanaeae). Stevens no. 3, XII 1908 (NY, isotype).

This is a parasitic species, with asci cylindrical, ascospores 24-38 × 4-6 µm.

Mycosphaerella friesii Tomilin. See *Mycosphaerella acilegna* M. Morelet.

Sphaerella frigida Ellis & Everh., Bull. Torrey Bot. Club 24: 461. 1897.— Fig. 328.

Type — USA: Colorado, San Juan Mts. On dead leaf (the veins still visible) overlying dead wood. Bethel no. 361, VII 1897 (NY, holotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores 15-20 × 5-5.5 µm. The substratum is not bleached and weather-beaten wood as indicated in the protologue and the membranaceous stroma [“Dermatostroma”] mentioned in accordance the protologue is the remnant of the host leaf.

Sphaerella fructinex Kirschst., Kryptog.-Fl. Mark Brandenburg 7(3): 393. 1938.— Fig. 329.

Type — Germany: Bayern, Eisenstein. On dry, but still attached fleshy cones of *Juniperus communis* (Cupressaceae). Kirschstein, VI 1935 (B, holotype).

This is morphologically indistinguishable from *M. juniperina*, with ascomata closed, breaking through the the epidermis of the fruit, wall (including stromatic tissue) 20-30 µm wide above and at the sides, asci cylindrical, ascospores 9-10 × 2.5-3.5 µm.

Mycosphaerella fruticum Starbäck, Bih. Kongl. Svenska Vetensk.-Akad. Handl. 14(3, 5): 17. 1889 = *Sphaerella fruticum* (Starbäck) Sacc., Syll. Fung. 9: 643. 1891.— Fig. 330.

Type — Scandinavia: On dead stems of *Rubus idaeus* (Rosaceae).

The type was not found in S, or in any of the herbaria consulted. Material studied (Austria, Niederösterreich, Zumpoldskirchen, Petr., VI 1940, B; also Lunz, Seehof, Petrak, Mycotheca Generalis no. 584, VI 1943, B (2×), IMI no. 30484; also Lunz, Seebachtal, Petrak., VII 1939, IMI no. 21114) contain different fungi: The first two collections contain *Schizothyrium pomi* (Mont. : Fr.) Arx, with peltate ascomata and ascospores 9-11 × 4-5 µm, the last collection belongs to *Davidiella* and is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores 12-15 × 3.5-4.5 µm.

Mycosphaerella fuchsicola (Speg.) Cash, Syll. Fung. 26: 340. 1972 = *Sphaerella fuchsicola* Speg., Bol. Acad. Nac. Ci. 27: 361. 1924.

Type — Argentina: *Fuchsia magellanica* (Onagraceae).

No material was studied as the type was not included in a loan from LPS.

Sphaerella fuckelii Pass., Erbario Crittogamico Italiano, ser. 2, 1-50. 1868, nomen novum (Article 58) for *Sphaerella epilobii* Fuckel, Jahrb. Nassauischen Vereins Naturk. 23-24: 103. 1870, later homonym (illegitimate, Article 53), nomen novum (Article 58) for *Dothidea maculiformis* Desm., Ann. Sci. Nat. Bot., ser. 6, 8: 176. 1847.

Type — France: *Epilobium* (Onagraceae).

Accepted as *Venturia maculiformis* (Desm.) G. Winter by Barr (1968).

Sphaerella fuegiana Speg., Bol. Acad. Nac. Ci. 11: 206. 1888 [“1887”].— Fig. 331.

Type — Argentina: Isla de Los Estados, Shammacus. On upper surface of dead, but still green leaf of *Acaena adscendens* (Rosaceae). Spegazzini no. 6180, V 1882 (LPS, holotype).

This belongs to the Microthyriaceae, with ascoma wall cells meandric, ascomata conical, superficial, asci pyriform, ascospores 10-13 × 3.5-4.5 µm.

Mycosphaerella fujiensis Hara, A List of Japanese fungi hitherto known: 399, 1954.

Type — Japan: Suruga kajima. On white spots on upper surface of living leaves of *Pyrus sinensis* (Rosaceae). Hara, VII 1920 (TNS 209265, holotype; TNS 209911, isotype).

The types contain only coelomycetes.

Mycosphaerella fumaginea (Cattaneo) Tomilin, Novosti Sist. Nizsh. Rast. 6: 120. 1970 [“1969”] = *Sphaerella fumaginea* Cattaneo, Rendiconti Reale Ist. Lombardo Sci., ser. 2, 10: 205. 1877 [as “*fumagina*”].

Type — Italy: *Vitis vinifera* (Vitaceae).

Anamorph: *Cladosporium fasciculatum* Corda fide Cattaneo (op. cit.) (= *Cladosporium herbarum* (Pers. : Fr.) Link).

No material was studied as the type was not found in any of the herbaria consulted.

Mycosphaerella fusca (Pass.) Tomilin, Novosti Sist. Nizsh. Rast. 1967: 189. 1967 = *Sphaerella fusca* Pass., Atti Reale Accad. Lincei, Mem. Cl. Sci. Fis., ser. 4, 3: 8. 1887.

Type — Italy: *Gladiolus segetum* (Iridaceae).

No material was studied as the type was not found in any of the herbaria consulted.

Sphaerella fuscata Ellis, Bull. Torrey Bot. Club 8: 125. 1881.— Fig. 332.

Type — USA: Utah, Carbon Co., Pleasant Valley. On dead herbaceous stems. Harkness no. 83, 1881 (NY, holotype).

This belongs to section *Longispora*, and is morphologically indistinguishable from *M. millegrana*, with asci cylindrical, ascospores 17-20 × 3.5-4.5 µm. Additional material studied (USA, Colorado, Larimer Co, on dead stems of *Pentstemon* (Plantaginaceae), Baker, distributed in Ellis & Everhart, North American Fungi no. 3321, VII 1895, L) belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores 15-18 × 5-6 µm.

Sphaerella fuscata Tassi. See *Sphaerella fuscomaculans* Sacc. & P. Syd.

Sphaerella fuscomaculans Sacc. & P. Syd., Syll. Fung. 16: 468. 1902, nomen novum (Article 58) for *Sphaerella fuscata* Tassi, Bull. Lab. Orto. Bot. Reale Univ. Siena, 2: 140. 1899, later homonym (illegitimate, Article 53).

Type — Australia: Sydney. On upper and lower surface of dead leaves of *Drimys dipetala* (Winteraceae). IX 1899 (SIENA, holotype).

The holotype is very sparse and the ascomata studied are totally empty.

Mycosphaerella fushinoki Miura, Industr. Contr. 27: 165. 1928.

Type — China: *Rhus javanicum* (Anacardiaceae).

No material was studied as the location of the type is unknown and its preservation uncertain.

Mycosphaerella fuisispora (Fuckel) Jacz., Opredelitel' gribov 2: 616. 1917 = *Sphaerella fuisispora* Fuckel, in Heuglin, Reisen nach dem Nordpolarmeer 3: 320. 1874.— Fig. 333.

Type — Russia: On dead culms of *Ranunculus pygmaeus* (Ranunculaceae). Heuglin (G, holotype).

Synonymised with *Mycosphaerella ranunculi* by von Arx (1949). The type is very sparse, but shows that it is morphologically indistinguishable from *D. clandestina*, with asci pyriform, ascospores ca. 25 × 7 µm.

Sphaerella fuisispora var. *groenlandica* Allesch., in Allesch. & Henn., Biblioth. Bot. 42: 46. 1897.

Type — Greenland: *Ranunculus nivalis* (Ranunculaceae).

Synonymised with *Mycosphaerella ranunculi* by von Arx (1949).

No material was studied as the type was not found in B and its preservation uncertain.

Mycosphaerella galanthina (Tassi) Tomilin, Novosti Sist. Nizsh. Rast. 1967: 189. 1967 = *Sphaerella galanthina* Tassi, Bull. Lab. Orto Bot. Reale Univ. Siena 2: 140. 1899.

Type — Italy: Siena, Botanical garden, on dead bulb sheaths of *Galanthus nivalis* (Alliaceae). Tassi, VII 1877 (SIENA, holotype).

The holotype contains empty, superficial ascomata, and does not belong to a *Mycosphaerella*.

Mycosphaerella galatea (Sacc.) Jacz., Opredelitel' gribov 2: 612. 1917 = *Sphaerella galatea* Sacc., Nuovo Giorn. Bot. Ital. 7: 304. 1875.— Fig. 334.

Type — Italy: Valezia. On dead stems of *Valeriana officinalis* (Caprifoliaceae). Saccardo (PAD, holotype; PAD, isotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. ammophilae*, with asci pyriform, ascospores 7-10 × 2-3 µm.

Sphaerella galatea var. *centranthi* Gonz. Frag., Mem. Real Soc. Esp. Hist. Nat. 11: 86. 1919 [as “*centranthii*”].

Type — Spain: Tarragona, Montsaut. On dead stems of *Centranthus angustifolium* var. *longicalcaratum*

(Caprifoliaceae). Font Zuer no. 3450, VI 1918 (MA, holotype).

The holotype contains only a *Phoma* anamorph.

Sphaerella galatea var. *valerianellae* Fautrey, Rev. Mycol. (Toulouse) 13: 125. 1891.

Type — France: Côte-d'Or, Jardin de Noidan. On dead stems of *Valerianella olitoria* (Caprifoliaceae). Fautrey, distributed in Roumeguère, Fungi Selecti Exsiccati no. 5720, X 1890 (PC, holotype).

The holotype contains only empty ascomata.

Mycosphaerella galatellae Lobik, Materialy po floristicheskim i faunisticheskim obsledovaniyam Terskogo okruga: 26. 1928.

Type — Russia: *Galatella punctata* (Asteraceae).

No material was studied as the type was not included in loans from LE or LEP.

Mycosphaerella galegae Lobik, Bolezni Rast. 17: 163. 1928.— Fig. 335.

Type — Russia: On dead stems of *Galega orientalis* (Fabaceae). Lobik (LE 34983, holotype).

This is a species of *Didymella*, with asci thick-walled, surrounded by parenchymatous tissue, ascospores 13-15 × 5-6 µm.

Mycosphaerella galii (Sacc.) Tomilin, Novosti Sist. Nizsh. Rast. 8: 150. 1971 ≡ *Sphaerella minor* var. *galii* Sacc., Syll. Fung. 1: 519. 1882 ≡ *Sphaerella galii* (Sacc.) Traverso, Fl. Ital. Cryptog. 1(11): 562. 1911.— Fig. 336.

Type — Italy. On dead stems of *Galium sylvaticum* (Rubiaceae). Saccardo V 1877 (PAD, holotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. ammophilae*, with asci pyriform, ascospores 8-10 × 2.5-3 µm.

Mycosphaerella galii-elliptici Petr., Bot. Jahrb. Syst., Beibl. 142: 119. 1928.— Fig. 337.

Type — Canary Islands: Tenerife. On upper and lower surface of dead leaves of *Galium ellipticum* (Rubiaceae). Ade, V 1926 (W 10126, holotype; W, isotype).

Cited as synonymous with *Mycosphaerella hermonie* by Tomilin (1979). This is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores 8-10 × 2.5-3 µm.

Mycosphaerella gallae (Ellis & Everh.) Tomilin, Novosti Sist. Nizsh. Rast. 7: 203. 1970 ≡ *Sphaerella gallae* Ellis & Everh., Bull. Torrey Bot. Club 11: 75. 1884.— Fig. 338.

Type — USA: New Jersey, Newfield. On galls on *Vaccinium corymbosum* (Ericaceae). Ellis no. 499e, VII 1884 (NY, holotype).

Cited as synonymous with *Mycosphaerella harthensis* by Tomilin (1979). However, this is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores 7-9 × 2-2.5 µm. Additional material studied on galls (USA, Massachusetts, on galls on *Rosa* sp. (Rosaceae), Clarke, III 1886, NY) agrees. Further additional material studied on *Vaccinium* (Washington, Marysville, on upper and lower surface of dead leaves of *Vaccinium* sp.,

Grant, 1928, IMI no. 26953, NY) contains only a coelomycete.

Sphaerella galtoniae Hollós, Ann. Hist.-Nat. Mus. Natl. Hung. 6: 528. 1908.

Type — Hungary: *Galtonia candicans* (Asparagaceae).

No material was studied as the type is not in BP and may have been destroyed during the war.

Sphaerella gangraena (Fr.) P. Karst., Bidrag Kännedom Finlands Natur Folk 23: 185. 1873 ≡ *Sphaeria gangraena* Fr., in Duby, in DC., Botan. Gallicum, ed. 2, 2: 695. 1830 ≡ *Dothidea gangraena* (Fr.) Fr., Summa Veg. Scand., sect. post.: 387. 1849 ≡ *Telimenella gangraena* (Fr.) Petr., Sydowia 1: 79. 1947.

Type — Sweden: *Poa* (Poaceae).

Accepted as *Telimenella gangraena* (Fr.) Petr. by M.E. Barr (1977) and therefore not studied.

Mycosphaerella garciniae Z.D. Jiang & P.K. Chi, in P.K. Chi, Fungal Diseases of Cultivated Medicinal Plants in Guangdong Province: 79. 1994.

Type — China: *Garcinia tinctoria* (Clusiaceae).

No material was studied as the location of the type is unknown and its preservation uncertain.

Mycosphaerella gardeniae (Cooke) Henn., Bot. Jahrb. Syst. 28: 273. 1900 ≡ *Sphaerella gardeniae* Cooke, J. Bot. 21: 108. 1883.— Fig. 339.

Type — USA: South Carolina. On white spots on upper surface of living leaves of *Gardenia florida* (Rubiaceae). (K, holotype; K, 2 isotypes).

Anamorph: *Phyllosticta gardeniae* Cooke *vide* Cooke (op. cit.).

The type and additional material (Hong Kong, Castle Peak Farm, on *G. jasminoides*, Leatler, IV 1967, IMI 128353) belong to section *Plaga*, with asci cylindrical, ascospores 12-15 × 3-3.5 µm. Most material distributed under the name *Sphaerella gardeniae* is from the host *Gordonia* (Theaceae), with dentate leaves, instead of from *Gardenia* (Rubiaceae). See *Mycosphaerella gordoniae* for this material.

Mycosphaerella garganica (Sacc.) Tomilin, Novosti Sist. Nizsh. Rast. 1967: 189. 1967 ≡ *Sphaerella garganica* Sacc., Ann. Mycol. 2: 14. 1904.— Fig. 340.

Type — Italy: Gargano, S. Nuando. On lower surface of dead leaves of *Smilax aspera* (Smilacaceae). Martelli, V 1893 (PAD, holotype).

The type shows that this belongs to section *Caterva*, and it is morphologically indistinguishable from *M. subradians*, with asci cylindrical, ascospores 10-12 × 3-3.5 µm. Also present on the type material is *D. ammophilae*, with asci pyriform, ascospores 9-10 × 2.5-3 µm. Additional material studied (Sardinia, Ulassai, Santa Barbara, Gams, V 1971, CBS) is *M. punctiformis*, with asci cylindrical, ascospores 9-11 × 3-3.5 µm.

Mycosphaerella gastonis (Sacc.) Lindau, in Engl. & Prantl, Natürlichen Pflanzenf. 1(1): 426. 1897 ≡ *Sphaerella gastonis* Sacc., Rev. Mycol. (Toulouse) 7: 158. 1885.— Fig. 341.

Type — Tahiti [“Haïti, Océanie”]. On veins on upper surface of leaves of *Cocos nucifera* (Arecaceae). Brunaud, 1884, distributed in Roumeguère, Fungi Gallici Exsiccati no. 3438 [as “*gastonix*”] (L, isotype).

This is probably a species of *Physolepora*, with ascospores simple, not septate, 12-15 × 4-5 µm.

Mycosphaerella gaubae Arx & Constant., in von Arx, Proc. Kon. Ned. Akad. Wetensch. C 86: 34. 1983 = *Eruptio gaubae* (Arx & Constant.) Crous, Mycol. Res. 103: 617. 1999.— Fig. 342.

Type — Australia: New South Wales, between Adaminaby and Kiandra. On upper surface of dead leaves of *Callistemon sieberi* (Myrtaceae). Gauba, I 1950 (IMI no. 288072, isotype).

Anamorph: *Lecanosticta gaubae* (Petr.) Arx & Constant. *fide* von Arx & Constantinescu (op. cit.).

Accepted as *Eruptio gaubae* by Crous (op. cit.). However, the hamathecium contains of copious cells, and this is in disagreement with the type species of *Eruptio*. The ascospores are ornamented and brownish when old, 25-27(-30) × 7-9 µm. The species rather belongs to the Parmulariaceae.

Mycosphaerella gaultheriae (Cooke & Peck) House, New York State Mus. Bull. 233-234: 27. 1921 = *Sphaerella gaultheriae* Cooke & Peck, in Cooke & Ellis, Grevillea 7: 42. 1878.— Fig. 343.

Type — USA: New Jersey, Newfield. On pale spots with brown margins on upper and lower surface of living leaves of *Gaultheria procumbens* (Ericaceae). Ellis no. 312, V 1878 (NY, holotype), also distributed in North American Fungi no. 799 (L, NY (2×), isotypes).

This is a parasitic species, with asci cylindrical, ascospores 14-16 × 3-4(-4.5) µm.

Mycosphaerella gaveensis Henn., Hedwigia 43: 84. 1904 = *Sphaerella gaveensis* (Henn.) Sacc. & D. Sacc., Syll. Fung. 17: 640. 1905.

Type — Brazil: *Plantago* (Plantaginaceae).

No material was studied as the type was not found in B and was probably destroyed.

Sphaerella geicola Kalchbr. & Cooke, Grevillea 9: 30. 1880.

Type — South Africa: Cape of Good Hope. On dark green spots on lower surface of living leaves of *Geum capensis* (Rosaceae). MacOwen no. 1148 (K, holotype; K, isotype).

The holotype contains an immature ascomycete.

Mycosphaerella geniostomatis Hansf., nomen herbariorum (not validly published, Article 32).— Fig. 344.

Authentic material — New Zealand, Auckland, Piha. On white spots with black margins on upper surface of living leaves of *Geniostoma ligustrifolium* (Loganiaceae). Dingley, Herbarium of the Plant Diseases Division no. 15902, I 1951 (IMI no. 74208).

This material belongs to *Davidiella*, of which it represents a parasitic species, with asci pyriform, ascospores 13-15 × 4.5-5.5 µm.

Mycosphaerella gentianae (Niessl) Lindau, in Engl. & Prantl, Natürlichen Pflanzenf. 1(1): 424. 1897 = *Sphaerella gentianae* Niessl, Oesterr. Bot. Z. 25: 128. 1875.— Fig. 345.

Type — Austria: On dead stems of *Gentiana asclepias* (Gentianaceae). Niessl (M, holotype).

Spermatial state and anamorph: *Asteroma gentianellae* (C. Massal.) Petr. and *Ramularia evanida* (Kühn) Sacc. *fide* Tomilin (1979).

The type and additional material studied (Ukraine, Galicia, Akna-Slatina, Hruby, distributed in Reliquiae Petrakianae no. 2475, VII 1921, L) belong to *Davidiella*, and it is morphologically indistinguishable from *D. ammophilae*, with asci small, pyriform, ascospores 9-10 × 2-2.5 µm. It was cited as synonymous with *Mycosphaerella galatea* by Tomilin (1979).

Mycosphaerella genuflexa (Auersw.) Migula, in Thomé, Fl. Deutschl., Österr. Schweiz X, I. Kryptog.-Fl. III, 3(1): 295. 1912 [“1913”] = *Sphaerella genuflexa* Auersw., in Gonn. & Rabenh., Mycol. Europaea 5-6: 8. 1869.— Fig. 346.

Type — Germany: Arnstadt. On lower surface of dead leaves of *Salix alba* (Salicaceae). Auersw., V 1969 (B, isotype).

This is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores 7-8 × 2-2.5 µm. Surprisingly, it was cited as synonymous with *Davidiella allicina* (as *Mycosphaerella allicina*) by Tomilin (1979).

Sphaerella genuflexa var. *polaris* P. Karst. See *Mycosphaerella polaris* (P. Karst.) Lindau.

Mycosphaerella gibelliana (Pass.) Jacz., Trudy Tiflissk. Bot. Sada 11: 143. 1910 = *Sphaerella gibelliana* Pass., in Thüm., Mycotheca Universalis, cent. 5 no. 462. 1876.— Fig. 347.

Type — Italy: Lago Maggiore, Cannaro. On upper and lower surface of dead leaf tips of *Citrus limon* (Rutaceae). Gibelli, VIII 1874, distributed in Thümen, Mycotheca Universalis no. 462 (L, NY, isotypes), also distributed in Herbarium Mycologicum Oeconomicum no. 425 (L, isotype).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradians*, with asci cylindrical, ascospores 11-12 × 2.5-3 µm.

Mycosphaerella gibsonii Evans, Mycol. Pap. 153: 61. 1984.— Fig. 348.

Type — Tanzania: Lindi, Rondo and Rondo Forest Reserve. On dead needles of *Pinus radiata* (Pinaceae). Gibson 825, III 1962 (IMI no. 92286a, holotype).

Anamorph and spermatial state: *Cercoseptoria pini-densiflorae* (Hori & Nambu) Deighton (= *Pseudocercospora pini-densiflorae* (Hori & Nambu) Deighton) and *Asteromella fide* Evans (op. cit.).

This is morphologically indistinguishable from *Mycosphaerella juniperina* (= *Eruptio acicola* (Dearn.) M.E. Barr), with ascomata partly gregarious on cellular stromata, asci cylindrical, ascospores 9-11 × 2-2.5 µm. Additional materials (Phillipines, Luzon, Abra Province, on

P. caribaea var. *hondurensis*, Speechly, VII 1980, IMI no. 250111, paratype; also Vietnam, Than Long, on *P. merkusii*, Gibson, VIII 1980, IMI no. 281637, paratype; also Nepal, Katmandu, Kaulitana nursery, Napier, IX 1984, IMI no. 297483) agree well.

Mycosphaerella gifuensis Hara, J. Pl. Protect. 5: 616. 1918.
Type — Japan.

No material was studied as the type was not included in a loan from TNS and its preservation uncertain.

Mycosphaerella glauca (Cooke) Woron., Vestn. Tiflissk. Bot. Sada 35: 6. 1914 = *Sphaerella glauca* Cooke, Hedwigia 17: 39. 1878.— Fig. 349.

Type — USA: South Carolina, Aiken. On white spots with black margins on upper surface of living leaves of *Magnolia glauca* (Magnoliaceae). Ravenel, distributed in Fungi Americani Exsiccati no. 94 (NY, 2 isotypes).

Anamorph (presumed): *Phyllosticta magnoliae* Sacc. fide Woronichin (op. cit.).

This is a parasitic species, with asci cylindrical, ascospores 18-22 × 2.5-3.5 µm. Additional material (Florida, Martin, 1878, distributed in Ellis, North American Fungi no. 1350, L) agrees, but has up to 4 µm wide ascospores.

Sphaerella glaucescens Cooke, Grevillea 7: 54. 1878 = *Laestadia glaucescens* (Cooke) Sacc., Syll. Fung. 1: 429. 1882.

Type — USA: South Carolina, Aiken. On lower and upper surface of dead leaves of *Acer rubrum* (Sapindaceae). Ravenel, distributed in Fungi Americani Exsiccati no. 381 (NY, 2 isotypes).

In the isotypes only a coelomycete was present.

Mycosphaerella glechomatis (Sacc. & Flageolet) Tomilin, Novosti Sist. Nizsh. Rast. 7: 203. 1970 [as “*glechomae*”] = *Sphaerella glechomatis* Sacc. & Flageolet, in Sacc., Syll. Fung. 17: 642. 1905 [as “*Glechomae*”].— Fig. 350.

Type — France: Rigny. On lower surface of dead leaves of *Glechoma hederacea* (Lamiaceae). Flageolet, 1904 (PAD, holotype).

Anamorph: Probably *Phyllosticta glechomae* Sacc. fide Saccardo & Flageolet (op. cit.).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradians*, with asci cylindrical, ascospores 11-12 × 3-3.5 µm. It was already cited as synonymous with *Mycosphaerella jurineae* (which is also *M. subradians*) by Tomilin (1979).

Mycosphaerella gleicheniae T.S. Ramakr. & K. Ramakr., Proc. Indian Acad. Sci., Sect. B, 32: 205. 1950.— Fig. 351.

Type — India: On upper surface of dead fronds of *Gleichenia linearis* (Polypodiaceae).

The location of the type is unknown and its preservation uncertain. Material studied (Malaysia, Peninsular Malaysia, Cameron Highlands, Federal Experiment Station, Johnston, X 1953, IMI no. 54878) belongs to section *Caterva*, and is morphologically indistinguishable from *M. filicum*, with asci broad-cylindrical, ascospores 13-16 × 3.5-4.5 µm.

Mycosphaerella glochidionis Sivan. & R.G. Shivas, Mycol. Res. 106: 358. 2002.

Type — Australia: *Glochidion sumatrana* (Euphorbiaceae). No material was studied of this recently described species.

Mycosphaerella glycosmae Tracy & Earle, in E.L. Green, Plantae Bakerianae 1(1): 33. 1901 = *Sphaerella glycosmae* (Tracy & Earle) Sacc. & P. Syd., Syll. Fung. 16: 1133. 1902.— Fig. 352.

Type — USA: Colorado, La Plata Mts., Mt. Herperus, Botanical Creek. On dead stems of *Glycosma occidentalis* (Apiaceae). Baker, Earle & Tracy, VI 1898, distributed in Plants of Southern Colorado no. 1047 (NY, 2 isotypes).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores 15-20 × 5-6.5 µm.

Mycosphaerella glycyrrhizae Lebedeva, Mater. Mikol. Fitopatol. Rossii 5(3): 2. 1922 [“1921”].— Fig. 353.

Type — Russia: On upper and lower surface of dead leaves of *Glycyrrhiza glabra* (Fabaceae). (LE 34984, lectotype, here designated; LE 34985, isotype).

Already cited as synonymous with *Davidiella allicina* (as *Mycosphaerella allicina*) by Tomilin (1979), but the types indicates that it is morphologically indistinguishable from *D. clandestina*, with asci pyriform, ascospores 20-24 × 6-8 µm.

Mycosphaerella gneticola Syd., Ann. Mycol. 21: 100. 1923.— Fig. 354.

Type — Philippines: Luzon, Bataan Province, Lamao. On white spots with black margins on upper surface of living leaves of *Gnetum indicum* (Gnetaceae). Reyes, V 1921, Bureau of Science no. 39325 (IMI no. 16759, isotype).

This is a parasitic species, with asci cylindrical, ascospores 12-14 × 3-4 µm.

Sphaerella gnidii Maire, Bull. Soc. Hist. Nat. Afrique N. 8: 167. 1917.

Type — Mauritania: *Daphne gnidium* (Thymelaeaceae). No material was studied as the location of the type is unknown and its preservation uncertain.

Sphaerella goodeniae Tassi, Bull. Lab. Orto Bot. Reale Univ. Siena 5: 108. 1902.— Fig. 355.

Type — Australia: Sydney, Botanical Garden. On dead branches of *Goodenia stelligera* (Goodeniaceae). 1904 (SIENA, holotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. ammophilae*, with asci pyriform, ascospores 11-13 × 2.5-3.5 µm.

Mycosphaerella goodiaefolia (Cooke) Aptroot comb. nov., **MB 500506**. **Basionym:** *Sphaerella goodiaefolia* Cooke, Grevillea 21: 38. 1893 [“1892”].— Fig. 356.

Type — Australia: Victoria, Frankston. On grey spots with brown margins on upper surface of living leaves of *Goodia latifolia* (Fabaceae). IX 1891 (K, holotype).

This is a parasitic species, with asci cylindrical, ascospores 12-14 × 3-4 µm.

Mycosphaerella gordoniae (Cooke) J.H. Miller, Mycologia 33: 80. 1941 = *Sphaerella gordoniae* Cooke, J. Bot. 21: 109. 1883.— Fig. 357.

Type — USA: Georgia, Darien. On lower surface of dead leaves of *Gordonia* [as “*Gardenia*”] *lasianthus* (Theaceae). XI 1881 (K, holotype; 4 isotypes), also distributed in Ravenel, Fungi Americani Exsiccati no. 799 (K, NY, isotypes [sub “*gardeniae*”]).

This is probably a *Hyponectria*, with asci clavate, no apparent hamothecium, ascospores simple, in bad shape, ca. 10-12 × 3-4 µm. Most material distributed under the name *Sphaerella gardeniae* belongs here, as it is from the host *Gordonia* (Theaceae), with dentate leaves, instead of from *Gardenia* (Rubiaceae), with entire leaf margins.

Mycosphaerella gossypina (G.F. Atk.) Earle, Exp. Sta. Bull. (Auburn, Alabama) 107: 309. 1900 = *Sphaerella gossypina* G.F. Atk., Bull. Torrey Bot. Club 18: 300. 1891.

Type — USA: On upper and lower surface of dead leaves of *Gossypium herbaceum* (Malvaceae).

Anamorph: *Cercospora gossypina* Cooke *vide* Atkinson (op. cit.).

The type was not found in any of the herbaria consulted. Material studied (Uganda, Masaka District, Bowden, I 1962, IMI no. 93556; also Zimbabwe, Kadoma, Hillocks, III 1985, IMI no. 296919) suggests that it is not a *Mycosphaerella*, as the specimen from Uganda is a *Didymella* and the specimen from Zimbabwe an *Ascochyta*, which is its anamorph.

Mycosphaerella gracilis Crous & Alfenas, Mycologia 87: 123. 1995.

Type — Indonesia: Sumatra. On spots on upper surface of living leaves of *Eucalyptus urophylla* (Myrtaceae). Alfenas, XI 1993 (IMI no. 361582 and 361583, isotypes).

Anamorph: *Pseudocercospora gracilis* Crous & Alfenas *vide* Crous & Alfenas (op. cit.).

The isotypes studied contain only the anamorph.

Mycosphaerella graeca Petr., Ann. Mycol. 34: 214. 1936.— Fig. 358.

Type — Greece: Achaia, Chelmos. On upper and lower surface of dead leaves of *Arenaria stygia* (Caryophyllaceae). Ade, VI 1931 (W, holotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores 13-20 × 3.5-5.5 µm.

Mycosphaerella graminicola (Fuckel) J. Schröt., in Cohn, Kryptog.-Fl. Schlesien 3(2): 340. 1894 [“1893”] = *Sphaeria graminicola* Fuckel, Fungi Rhenani Exsiccati no. 1578 = *Sphaerella graminicola* (Fuckel) Fuckel, Jahrb. Nassauischen Vereins Naturk. 23-24: 101. 1870.— Fig. 359.

Type — Germany: Oestrich. On upper and lower surface of dead leaves of *Triticum repens* (Poaceae). Fuckel, Fungi Rhenani Exsiccati no. 1578 (L, isotype; IMI no. 59187, isotype slide).

Anamorph: *Septoria tritici* Roberge *vide* Sivanesan (1984).

The isotype in L is immature. The isotype in IMI shows that this belongs to section *Caterva*, with asci cylindrical, ascospores 15-17 × 3-4 µm.

Mycosphaerella graminicola f. *alpina* (Rehm) Magnus, in Dalla Torre & Sarnth., Fl. Tirol, Vorarlberg, Leichtenstein, III, Pilze: 464. 1905. *Sphaerella graminicola* f. *alpina* Rehm, Ascomyceten no. 794.— Fig. 360.

Type — Austria: Tirol, Ortler Alpen. On upper and lower surface of dead leaves of Poaceae. Rehm, VIII 1884, distributed in Ascomyceten no. 794 (BPI), also distributed in Rabenhorst-Winter, Fungi Europaei Exsiccati no. 3446 (L, isotype).

Already synonymised with *Davidiella allicina* (as *Mycosphaerella tassiana*) by von Arx (1949), with which the isotypes studied agrees well, with asci pyriform, ascospores 16-20 × 4-6 µm.

Mycosphaerella graminis (Sacc.) Tomilin, Novosti Sist. Nizsh. Rast. 1966: 172. 1966 = *Sphaerella graminis* Sacc., Boll. Soc. Bot. Ital.: 152. 1913.— Fig. 361.

Type — Italy. On dead culms of *Stipa tenacissima* (Poaceae). Pamparini, IV 1913 (PAD, holotype; PAD, isotype).

Anamorph: *Asteromella graminis* Westend. *vide* Saccardo (op. cit.).

The species was already synonymised with *M. recutita* sensu von Arx, which is morphologically indistinguishable from *Davidiella disseminata*, by von Arx (1949), with which the type materials agree, with asci pyriform, ascospores 12-14 × 4-5 µm.

Mycosphaerella graminum (Sacc. & Scalia) Lavrov, Trudy Tomsk. Gosud. Univ. Kujbyševa 110: 174. 1951 = *Sphaerella graminum* Sacc. & Scalia, in Sacc., Peck & Trelease, The fungi of Alaska. Harriman Alaska Exped. 5: 32. 1904.

Type — USA: Alaska, Shumgrin Is. On dead leaves of *Poa stenantha* (Poaceae). Harrington, 1871-1872 (PAD, holotype).

Nothing identifiable was found in the type.

Mycosphaerella grandis Carnegie & Keane, Mycol. Res. 98: 414. 1994.

Type — Australia: Victoria, East Gippsland, Nowa Nowa, Old Orbost Road, Tostaree Pilot Farm, On leaf of *Eucalyptus grandis* (Myrtaceae). Carnegie, XI 1990 (IMI 353729a, holotype).

Reported by Crous (1998) to be synonymous with *M. parva* R.F. Park & Keane, with which the type agrees.

Mycosphaerella grandispora Bubák, in Hand.-Mazz., Ann. K.K. Naturhist. Hofmus. 23: 103. 1909 = *Sphaerella grandispora* (Bubák) Sacc. & Traverso, Syll. Fung. 20: 822. 1911.— Fig. 362.

Type — Turkey: Pons Region, Trapezunti, Elchu, Eseli. On upper and lower surface of dead leaves of *Narthecium balansa* (Asparagaceae). Handel-Mazzetti no. 921, VII 1907 (BPI, holotype).

This species was synonymised with *Davidiella allicina* (as *Mycosphaerella tassiana*) by von Arx (1949), probably on the basis of the description and without checking any material. It belongs indeed to *Davidiella*, but is instead morphologically indistinguishable from *D. disseminata*, with asci pyriform, ascospores 12-15 × 3.5-4.5 µm.

Mycosphaerella granulata (Ellis & Everh.) Lindau, in Engl. & Prantl, *Natürlichen Pflanzenf.* 1(1): 424. 1897 = *Sphaerella granulata* Ellis & Everh., *J. Mycol.* 2: 102. 1886.— Fig. 363.

Type — USA: New Jersey, Newfield. On dead stems of *Baptisia tinctoria* (Fabaceae). IV 1886 (NY, holotype), also distributed in Ellis & Everhart, *North American Fungi* no. 1798, V 1886 (L, isotype).

Cited as synonym with *Mycosphaerella oenotherae* by Barr (1972). According to the types studied, this belongs to section *Longispora*, and is morphologically indistinguishable from *M. latebrosa*, with asci cylindrical, ascospores 19-24 × 3-3.5 µm.

Mycosphaerella greenei Tomilin, *Novosti Sist. Nizsh. Rast.* 7: 201. 1970, nomen novum (Article 58) for *Mycosphaerella calamagrostidis* H.C. Greene, *Amer. Midl. Naturalist* 34: 265. 1945, later homonym (illegitimate, Article 53) [as “*calamagrostis*”].

Type — USA: Wisconsin, Dane Co., Madison. On dead culms of *Calamagrostis canadensis* (Poaceae). Greene, VIII 1945 (BPI, isotype).

The isotype contains only a coelomycete.

Sphaerella gregaria Auersw., in Gonn. & Rabenh., *Mycol. Europaea* 5-6: 2. 1869, nomen nudum (not validly published, Article 32).

Authentic material — Germany: *Quercus* (Fagaceae).

Cited as synonym of *Sphaerella cookeana* Auersw. by Auerswald (op. cit.). No material was studied as it was not found in B and was probably destroyed.

Mycosphaerella gregaria Carnegie & Keane, *Mycological Research* 101: 843. 1997 nom. nov. pro *Mycosphaerella aggregata* Carnegie & Keane, *Mycol. Res.* 98: 415. 1994, later homonym (illegitimate, Article 53) non *M. aggregata* (Schwein.) Stev. (1918).— Fig. 364.

Type — Australia: Victoria, Nowa Nowa. On dead marginal parts of living leaves of *Eucalyptus grandis* (Myrtaceae). Carnegie, XI 1990 (IMI no. 353729b, holotype).

This is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores 9-10 × 2.5-3 µm. It is particularly unfortunate that this material has been given first an illegitimate name and next an epithet that was already taken in *Sphaerella*, especially as the material turns out to be morphologically indistinguishable from the type and oldest species in the genus *Mycosphaerella*, described about 2 centuries earlier.

Mycosphaerella grevilleae Munjal, Chona & Kapoor, *Indian Phytopathol.* 12: 177. 1960 [“1959”].

Type — India: *Grevillea robusta* (Proteaceae).

No material was studied as the location of the type is unknown and its preservation uncertain.

Mycosphaerella grisea (Boyer & Jacz.) Höhn., *Sitzungsber. Kaiserl. Akad. Wiss., Math.-Naturwiss. Kl., Abt. 1*, 126: 355. 1917 = *Sphaerella grisea* Boyer & Jacz., *Bull. Soc. Bot. France* 39, suppl.: CCLXXXIII. 1893.— Fig. 365.

Type — France: On dead stems of *Scrophularia canina* (Scrophulariaceae).

The location of the type is unknown and its preservation uncertain. Material studied (Var, Dardennes, Crozals, VI 1927, BPI) belongs to *Davidiella*, and is morphologically indistinguishable from *D. ammophilae*, with asci pyriform, ascospores 10-11 × 3-3.5 µm, surrounded by a 1 µm thick gelatinous sheath.

Mycosphaerella grossulariae (Fr. : Fr.) Lindau, in Engl. & Prantl, *Natürlichen Pflanzenf.* 1(1): 424. 1897 = *Sphaeria grossulariae* Fr., *Scleromyceti Sueciae* no. 57. 1819, sanctioned by Fr., *Systema Mycol.* 2: 521. 1823 = *Sphaerella grossulariae* (Fr. : Fr.) Auersw., in Gonn. & Rabenh., *Mycol. Europaea* 5-6: 11. 1869 = *Pleospora grossulariae* (Fr. : Fr.) Fuckel, *Jahrb. Nassauischen Vereins Naturk.* 23-24: 133. 1870.— Fig. 366.

Type — Sweden: On upper and lower surface of dead leaves of *Ribes grossularia* (Grossulariaceae).

Anamorphs: *Phyllosticta grossulariae* Sacc. and *Septoria ribis* (Lib.) Desm. *fide* Tomilin (1979).

This is morphologically indistinguishable from *Pleospora herbarum* (Pers. : Fr.) Rabenh., according to Eriksson (1992). Additional materials studied (Germany, Brandenburg, Prignitz, Triglitz, Jaap, IV 1905, distributed in *Fungi Selecti Exsiccati* no. 154B, L; also Brandenburg, Tamsel, Vogel, IV 1906, distributed in *Sydow, Mycotheca Germanica* no. 581, L) belong to section *Longispora*, and are *M. latebrosa*, with asci cylindrical, ascospores 23-27 × 2-2.5 µm. Dutch material studied (Netherlands, Culemborg, Hulsebosch, 1878, L) is immature.

Sphaerella grossulariae var. *salicella* Sacc. & Scalia, in Sacc., Peck & Trelease, *The fungi of Alaska, Harriman Alaska Exped.* 5: 32. 1904.

Type — USA: *Salix* (Salicaceae).

No material was studied, as the type was not found in any of the herbaria consulted.

Mycosphaerella groveana (Sacc.) Arx, in E. Müll. & Arx, *Beitr. Kryptogamenfl. Schweiz* 11(2): 358. 1962 = *Scirrhia groveana* Sacc., *Atti Reale Ist. Veneto Sci., ser. 6*, 3: 733. 1885 = *Scirrhia groveana* (Sacc.) Theiss. & Syd., *Ann. Mycol.* 13: 626. 1915.

Type — United Kingdom: Langler Poal. On upper and lower surface of dead leaves of *Typha latifolia* (Typhaceae). Grove, VIII 1884 (K, holotype).

The stromatic ascomata exclude this species from *Mycosphaerella*, but suggest that it could better be accepted as *Scirrhia groveana* or, alternatively, classified in *Cymadothea*.

Mycosphaerella grumiformis (P. Karst.) Starbäck, *Bih. Kongl. Svenska Vetensk.-Akad. Handl.* 16(3, 5): 3. 1890 = *Sphaerella grumiformis* P. Karst., *Bidrag Kännedom Finlands Natur Folk* 23: 173. 1873 = *Gibbera grumiformis* (P. Karst.) M.E. Barr, *Canad. J. Bot.* 39: 314. 1961 = *Phaeosphaerella grumiformis* (P. Karst.) Tomilin, *Opredelitel' gribov roda Mycosphaerella* Johans.: 285. 1979.

Type — Finland: *Arctostaphylos alpina* (Ericaceae).

Accepted as *Gibbera grumiformis* (P. Karst.) M.E. Barr by Eriksson (1992), and therefore not studied.

Mycosphaerella guadarramica (Gonz. Frag.) M. Morelet, Ann. Soc. Sci. Nat. Archéol. Toulon & Var 20: 105. 1968 = *Sphaerella guadarramica* Gonz. Frag., Bol. Soc. Esp. Hist. Nat. 24: 444. 1924.— Fig. 367.

Type — Spain: Madrid, Cercedilla, Estacion Alpina de Biologia. On living needles of *Juniperus communis* (Cupressaceae). Fragoso no. 5252, VIII 1922 (MA, holotype).

This is morphologically indistinguishable from *Seynesiella juniperi* (Desm.) Arnaud, with ascomata conical, asci bitunicate, ascospores $20\text{--}25 \times 6\text{--}8 \mu\text{m}$.

Mycosphaerella guettardina Petr. & Cif., Ann. Mycol. 30: 211. 1932.— Fig. 368.

Type — Dominican Republic: Llano Costero, El Manielito. On upper surface of dead leaves of *Guettarda scabra* (Rubiaceae). Ciferri, I ["IP"] 1930, distributed in Reliquiae Petrakianae no. 2052 (H, L, NY (2 \times), isotypes).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. ammophilae*, with asci small, pyriform, ascospores $7.5\text{--}9.5 \times 2.5\text{--}3.5 \mu\text{m}$.

Mycosphaerella guineensis Kranz, Nova Hedwigia 18: 235. 1969.— Fig. 369.

Type — Africa: Guinea: IRF, Kindia. On brown spots on upper surface of living leaves of *Cassia alata* (Fabaceae). Kranz, III 1964 (IMI no. 106556, holotype).

This is a parasitic species, with asci cylindrical, ascospores $10\text{--}12 \times 2.5\text{--}3 \mu\text{m}$.

Sphaerella gunnerae Speg., Bol. Acad. Nac. Ci. 11: 206. 1888 ["1887"].— Fig. 370.

Type — Tierra del Fuego: Isla Basket. On upper and lower surface of dead leaves of *Gunnera lobata* (Gunneraceae). Spegazzini no. 6151, V 1882 (LPS, holotype, sub "gumerae").

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradians*, with asci cylindrical, ascospores $11\text{--}13 \times 3.5\text{--}4 \mu\text{m}$.

Mycosphaerella guttiferiae Miles, Trans. Illinois State Acad. Sci. 10: 250. 1917.— Fig. 371.

Type — Puerto Rico: Maricao. On pinkish spots with brown margins on upper surface of living leaves of *Clusia gaudlachia* (Clusiaceae). Stevens no. 286, I 1912 (NY, isotype, sub "gutterae").

This is a parasitic species, with asci cylindrical, ascospores $19\text{--}25 \times 4\text{--}6 \mu\text{m}$.

Mycosphaerella gypsophilae (Fuckel) Feltgen, Recueil Mém. Trav. Soc. Bot. Grand-Duché Luxembourg 14: 287. 1899 = *Sphaerella gypsophilae* Fuckel, Jahrb. Nassauischen Vereins Naturk. 23-24: 101. 1870 = *Sphaeria gypsophilae* Lasch, in Rabenh., Klotzsch" Herb. Vivum Mycol. cent. 11 no. 1050. 1846, nomen nudum (not validly published, Article 32).

Type — Germany: Driessen. On dead stems and calyces, and on upper and lower surface of dead leaves of *Gypsophila muralis* (Caryophyllaceae). Lasch, distributed in Rabenhorst, Klotzschii Herbarium Vivum Mycologicum no. 1050 (L, isotype).

This is no *Mycosphaerella* according to Tomilin (1979). The isotype studied contains only a *Septoria* anamorph, with conidia 1-septate, $25\text{--}35 \times 3\text{--}3.5$, which agrees somewhat with the description, and with additional material (Oestrich, Fuckel, Fungi Rhenani Exsiccati no. 849, L).

Sphaerella gypsophilae f. *scleranthi* Fuckel, Jahrb. Nassauischen Vereins Naturk. 23-24: 101. 1870.

Type — Germany: Hattenheim. On dead stems and upper and lower surface of dead leaves of *Scleranthus annuus* (Caryophyllaceae). Fuckel, Fungi Rhenani no. 1573 (L, isotype).

The isotype studied contains only a *Septoria* anamorph.

Mycosphaerella gypsophilicola (Hollós) Petr., Flora Bohemiae et Moraviae Exsiccata, vo. 43 no. 2119. 1925 = *Sphaerella gypsophilicola* Hollós, Ann. Hist.-Nat. Mus. Natl. Hung. 5: 44. 1907 [as "gypsophilaecola"].— Fig. 372.

Type — Hungary: *Gypsophila paniculata* (Caryophyllaceae).

The type is not in BP and may have been destroyed during the war. Material studied (Czech Republic, Datschitz, on dead stems, fruits and upper and lower surface of dead leaves of *Spergularia rubra* (Caryophyllaceae), Hruby, VII 1925, distributed in Petrak, Flora Bohemiae et Moraviae Exsiccata no. 2119, BPI) belongs to *Davidiella*, and is morphologically indistinguishable from *D. clandestina*, with asci pyriform, ascospores thick-walled, $20\text{--}25 \times 6.5\text{--}8 \mu\text{m}$.

Mycosphaerella gypsophilina Naumov, nomen herbariorum (not validly published, Article 32).

Authentic material — Russia: On dead calyces and upper and lower surface of dead leaves of *Gypsophila muralis* (Caryophyllaceae). Naumov (LE 34986).

The material studied contains only a *Septoria* anamorph.

Sphaerella haematites (Roberge) Cooke, J. Bot. 21: 109. 1883 = *Sphaeria haematites* Roberge, in Desm., Ann. Sci. Nat. Bot., ser. 3, 16: 311. 1851 = *Leptosphaeria haematites* (Roberge) Niessl, in G. Winter, in Rabenhorst, Fungi Europaei Exsiccati, ed. nov., ser. 2, cent. 28 no. 2761. 1882.

Type — France: *Clematis vitalba* (Ranunculaceae).

Anamorph: *Phoma fide* Lucas & Webster (1967).

Accepted as *Leptosphaeria haematites* (Roberge) Niessl by Lucas & Webster (1967) and therefore not studied.

Sphaerella haematodes Berk. & Cooke, in Cooke, J. Bot. 21: 70. 1883.

Type — USA: *Kalmia glauca* (Ericaceae).

No material was studied, as the type was not found in any of the herbaria consulted.

Sphaerella hageniae Rehm, Flora 55: 523. 1872 = *Pharcidia hageniae* (Rehm) Rehm, Ber. Naturhist. Vereins Augsburg 26: 12. 1881 = *Epicymatia hageniae* (Rehm) Sacc., Syll. Fung. 1: 573. 1882 = *Stigmidium hageniae* (Rehm) Hafellner, in R. Sant., Thunbergia 6: 8. 1988.— Fig. 373.

Type — Germany: Franken, Obernesselbach. On upper surface of thallus of *Anaptychia* ["*Hagenia*"] *ciliaris*

(Ascomycota, Physciaceae). Rehm, *Ascomyceten* no. 32, 1870 (L, isotype).

This is a species of *Stigmidium*, with asci pyriform, surrounded by sparse paraphysoids, ascospores 11-13 × 4-5 µm, surrounded by a gelatinous sheath. It is accepted as *Stigmidium hageniae* (Rehm) Hafellner by Clauzade, Diederich & Roux (1989).

Mycosphaerella halimodendri Jacz., *Bull. Soc. Mycol. France* 38: 207. 1922 ≡ *Mycosphaerella halimodendri* Murashk., *Trudy Omsk. Selskokh. Inst.* 3: 123. 1924, later homonym (illegitimate, Article 53), superfluous (illegitimate, Article 52) nomen novum (Article 58) ≡ *Mycosphaerella murashkii* M. Morelet, *Ann. Soc. Nat. Archéol. Toulon & Var* 20: 106. 1968, superfluous (illegitimate, Article 52) nomen novum (Article 58).— Fig. 374.

Type — Russia: Omsk. On upper and lower surface of dead leaves of *Halimodendron argenteum* (Fabaceae). Murashkinsky, V 1921 (LEP, holotype).

Anamorph: *Phoma halimodendri* Murashk. *vide* Murashkinsky (op. cit.).

This is morphologically indistinguishable from *M. punctiformis*, with ascomata in groups, asci cylindrical, ascospores 9-10 × 2.5-3 µm.

Mycosphaerella halophila (E. Bommer, M. Rousseau & Sacc.) O.E. Erikss., *Non-lichenized Pyrenomycetes of Sweden*: 7. 1992 ≡ *Lizonia halophila* E. Bommer, M. Rousseau & Sacc., in Sacc., *Syll. Fung.* 9: 680. 1891 ≡ *Sphaerulina halophila* (E. Bommer, M. Rousseau & Sacc.) Starbäck, *Bih. Kongl. Svenska Vetensk.-Akad. Handl.* 21(3, 2): 21. 1896 ≡ *Lizoniella halophila* (E. Bommer, M. Rousseau & Sacc.) Sacc. & D. Sacc., *Syll. Fung.* 17: 661. 1905.

Type — Belgium: *Honckenya peploides* (Caryophyllaceae). Synonymised with *Davidiella allicina* (as *Mycosphaerella tassiana*) by von Arx (1949). No material was studied as the type was not found in BR or any of the herbaria consulted.

Mycosphaerella hambergii (Romell & Sacc.) Petr., *Acta Horti Gothob.* 17: 137. 1947 ≡ *Sphaerella confinis* subsp. *hambergii* Romell & Sacc., in Sacc., *Syll. Fung.* 11: 296. 1895.

Type — Sweden: *Arabis planisiliqua* ["gerardii"] (Brassicaceae). No material was studied as the type was not found in S or any of the herbaria consulted.

Mycosphaerella handelii Crous & U. Braun, *Mycosphaerella* and its anamorphs: 1. Names published in *Cercospora* and *Passalora*: 211. 2003.

Type — The Netherlands: Bilthoven. On upper surface of pale spots on living leaves of *Rhododendron* (Ericaceae). M. Crous & Crous, III 2003 (CBS, holotype).

Anamorph: *Cercospora handelii* Bubák *vide* Crous & Braun (op. cit.) (= *Pseudocercospora handelii* (Bubák) Deighton). The type of this recently described species was not further investigated.

Mycosphaerella haraeana Syd. & P. Syd., *Ann. Mycol.* 11: 59. 1913 ≡ *Sphaerella haraeana* (Syd. & P. Syd.) Trotter, *Syll. Fung.* 24: 870. 1928.— Fig. 375.

Type — Japan: Mino, Kawaye-mura. On upper and lower surface of dead leaves of *Pterocarya* (Juglandaceae). Hara, V 1912 (S, holotype).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradicans*, with asci cylindrical, ascospores 11-13 × 3.5-4 µm.

Mycosphaerella hariotiana (Speg.) Tomilin, *Novosti Sist. Nizsh. Rast.* 1967: 189. 1967 ≡ *Sphaerella hariotiana* Speg., *Anales Mus. Nac. Hist. Nat. Buenos Aires* 6: 265. 1898.

Type — Argentina: *Juncus scheuchzerioides* (Juncaceae). No material was studied as the type was not included in a loan from LPS.

Mycosphaerella harknessii (Sacc.) Tomilin, *Novosti Sist. Nizsh. Rast.* 7: 203. 1970 ≡ *Sphaerella harknessii* Sacc., *Syll. Fung.* 1: 511. 1882, nomen novum (Article 58) for *Sphaerella brachytheca* Cooke & Harkn., *Grevillea* 9: 8. 1880, later homonym (illegitimate, Article 53).— Fig. 376.

Type — USA: California, Mt. Jamalpais. On dead stems of *Calystegia* ["*Convolvulus*"] *sepium* (Convolvulaceae). Harkness no. 1382, IV 1880 (K, holotype; BPI, isotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. ammophilae*, with asci pyriform, ascospores 11-13 × 2.5-3.5 µm.

Mycosphaerella harthensis (Auersw.) Migula, in Thomé, *Fl. Deutschl., Österr. Schweiz. X, 1. Kryptog.-Fl. III, 3(1)*: 289. 1913 ["1912"] ≡ *Sphaerella harthensis* Auersw., in Gonn. & Rabenh., *Mycol. Europaea* 5-6: 9. 1869.

Type — Germany: On lower surface of dead leaves of *Betula alba* (Betulaceae).

The type was not found in B and is probably destroyed. Material studied (France, Pyrenees centrales, Luchon, Roumeguère, distributed in *Fungi Gallici Exsiccati* no. 3229, 1884, L) is probably *M. punctiformis*, with asci cylindrical, ascospores immature.

Mycosphaerella hawaiiensis F. Stevens & K. Young, in F. Stevens, *Bernice P. Bishop Mus. Bull.* 19: 103. 1925.— Fig. 377.

Type — Hawaii: On upper and lower surface of dead leaves of *Gunnera petaloides* (Gunneraceae).

The type was not found in any of the herbaria consulted. Topotype material studied (Molokai, Papaala Pali, Degener & Wiebke, distributed in *Reliquiae Petrakianae* no. 68, VI 1928, L, B) shows that this belongs to section *Caterva*, and is morphologically indistinguishable from *M. superflua*, with asci cylindrical, ascospores 16-18 × 3.5-4.5 µm. The topotype in L contains only empty ascomata.

Sphaerella hederæ (Sowerby) Cooke, *Handb. Brit. Fungi* 2: 921. 1871 ≡ *Sphaeria hederæ* Sowerby, *Eng. Fungi*, vol. 3, tab. 371, fig. 5. 1803 ["1802"] ≡ *Metasphaeria hederæ* (Sowerby) Sacc., *Syll. Fung.* 2: 169. 1883.— Fig. 378.

Type — United Kingdom: *Hedera helix* (Araliaceae). Sowerby (K, holotype).

This is morphologically indistinguishable from *Glomerella cingulata* (Stoneman) Spauld. & H. Schrenk, for which it would represent an earlier epithet, with paraphysoids 2 µm wide and aseptate ascospores of 10-12 × 5 µm.

Mycosphaerella hederiae-helicis Siemaszko, Acta Soc. Bot. Pol. 1: 20. 1923.

Type — Georgia: *Hedera helix* (Araliaceae).

No material was studied as the type was not found in BPI, where some Siemaszko material is preserved, nor in any of the other herbaria consulted.

Mycosphaerella hedericola (Desm.) Lindau, in Engl. & Prantl, Natürlichen Pflanzenf. 1(1): 424. 1897 = *Sphaeria hedericola* Desm., Ann. Sci. Nat. Bot., ser. 3, 14: 115. 1850 [as “*hederaecola*”] = *Sphaerella hedericola* (Desm.) Cooke, Grevillea 3: 69. 1874 [as “*hederaecola*”] = *Carlina hedericola* (Desm.) Höhn., Mitt. Bot. Lab. TH Wien, ser. 2, 3: 91. 1925 [as “*hederaecola*”].— Fig. 379.

Type — France: On spots on upper and lower surface of living leaves of *Hedera helix* (Araliaceae). Desmazières, Plantes Cryptogames de France, ser. 3 no. 362 (PC, holotype: also B-Desmazières, isotype).

Spermatial state and anamorph: *Asteromella hederiae* (Sacc. & Roum.) Petr. fide Tomilin (1979) and *Septoria hederiae* Desm. fide Desmazières (op. cit.).

The type and additional material studied (Germany, Brandenburg, Werder, Glindow, Sydow, VI 1940, distributed in Mycotheca Germanica no. 3505, L) belong to section *Plaga*, with asci cylindrical, ascospores 10.5-12.5 × 1.5-2 µm.

Mycosphaerella hedychii F. Stevens & K. Young, in F. Stevens, Bernice P. Bishop Mus. Bull. 19: 103. 1925.

Type — Hawaii: *Hedychium coronarium* (Zingiberaceae).

No material was studied as the type was not found in BPI or NY, where much of Stevens material is preserved, nor in any of the other herbaria consulted.

Mycosphaerella heimii Bouriquet ex Crous, in Crous & W.J. Swart, S. African Forest. J. 172: 2. 1995; *Mycosphaerella heimii* Bouriquet, Encyclopédie Mycologique 12: 418. 1946 (nomen nudum).

Type — Madagascar: Moramanga. On leaves of *Eucalyptus* (Myrtaceae). Crous, IV 1994 (PREM 51749, holotype, not seen).

Anamorph: *Pseudocercospora heimii* Crous fide Crous (op. cit.).

No material was studied of this recently described species.

Mycosphaerella heimioides Crous & M.J. Wingf., Canad. J. Bot. 75: 787. 1997.

Type — Indonesia: Sumatra, Lake Toba. On the leaves of *Eucalyptus* (Myrtaceae). M.J. Wingf., III 1996 (PREM 54966, holotype, not seen).

Anamorph: *Pseudocercospora heimioides* Crous & M.J. Wingf. fide Crous (1998).

No material was studied of this recently described species.

Mycosphaerella helenae Chevaugeon, Encyclopédie Mycologique 28: 36. 1956.

Type — Africa: *Manihot utilissima* (Euphorbiaceae).

No material was studied as the location of the type is unknown and its preservation uncertain.

Sphaerella hellebori (Chaillat ex Fr.) Ces. & De Not., Comment. Soc. Crittog. Ital. 1(4): 238. 1863 = *Sphaeria hellebori* Chaillat ex Fr., Systema Mycol. 2: 512. 1823 = *Didymella hellebori* (Chaillat ex Fr.) Sacc., Syll. Fung. 1: 553. 1882.

Type — Sweden: *Helleborus* (Ranunculaceae).

No material was studied as it was already accepted as *Didymella hellebori* (Chaillat ex Fr.) Sacc. by Saccardo (op. cit.).

Sphaerella hellebori Roum., Fungi Gallici Exsiccati no. 1710, 1880, nomen herbariorum (not validly published, Article 32).— Fig. 380. Authentic material — France: Toulouse, on upper and lower surface of dead leaves of *Helleborus viridis* (Ranunculaceae). Roumeguère, 1880, Fungi Gallici Exsiccati no. 1710 (L).

This is *M. punctiformis*, with asci cylindrical, ascospores 11-13 × 2.5-3.5 µm.

Sphaerella heloniaefolia (Cooke & Ellis) Cooke, J. Bot. 21: 139 = *Sphaeria heloniaefolia* Cooke & Ellis, Grevillea 8: 16. 1879.

Type — USA: On leaves of *Helonias bullata* (Alliaceae). Ellis no. 3199, V 1878 (NY, isotype), also distributed in North American Fungi no. 595 (NY, 2 isotypes).

All isotypes contain only coelomycetes.

Mycosphaerella hemerocallidicola Petr., Hedwigia 74: 31. 1934.— Fig. 381.

Type — Russia: Primorsky Krai, Vladivostok. On dead stems and on upper and lower surface of dead leaves of *Hemerocallis* (Xanthorrhoeaceae). Ziling, VI 1928 (W, holotype).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradicans*, with asci cylindrical, ascospores 13-16 × 3-4 µm.

Mycosphaerella hemerocallidis (Pass.) Lindau ex Ranojevič, Ann. Mycol. 8: 360. 1910 = *Sphaerella hemerocallidis* Pass., Atti Reale Accad. Lincei, Mem. Cl. Sci. Fis., ser. 4, 3: 9. 1887.

Type — Italy: *Hemerocallis fulva* (Xanthorrhoeaceae).

Cited as synonymous with *Davidiella allicina* (as *Mycosphaerella allicina*) by Tomilin (1979). No material was studied as the type was not found in any of the herbaria consulted.

Mycosphaerella henningsii Sivan., Trans. Brit. Mycol. Soc. 84: 552. 1985, nomen novum (Article 58) for *Mycosphaerella manihotis* Ghesq. & Henrard, Rev. Zool. Bot. Africaine 12: 1. 1924, later homonym (illegitimate, Article 53) = *Mycosphaerella manihotis* f. sp. *macrospora* Chevaug., Encycl. Mycol. 28: 34. 1956, superfluous (illegitimate, Article 52) nomen novum (Article 58).— Fig. 382.

Type — Zaire: Kisangani. On spots on upper and lower surface of living leaves of *Manihot utilissima*

(Euphorbiaceae). Ghesquière no. 11924, I 1924 (BR, lectotype), also distributed in Reliquiae Petrakianae no. 656 (H, L, isotypes).

Anamorph: *Cercosporidium henningsii* (Allesch.) Deighton *vide* Sivanesan (1984).

This is a parasitic species, with asci cylindrical, ascospores 15-18 × 3-4 µm.

Sphaerella henriquesiana G. Winter, Bol. Soc. Brot. 4: 13. 1886.

Type — South Africa: *Musa* (Musaceae).

No material was studied as the type was not found in B or LISU and is probably destroyed.

Sphaerella henriquesiana Sacc., Bol. Soc. Brot. 11: 10. 1893, later homonym (illegitimate, Article 53).

Type — Portugal: *Ailanthus glandulosa* (Simaroubaceae).

No material was studied as the type was not found in LISU or any of the herbaria consulted.

Mycosphaerella hepaticae Petr., Ann. Mycol. 29: 110. 1931.— Fig. 383.

Type — Spain: Cataluña, Monserrat. On upper surface of dead leaves of *Hepatica hispanica* (Ranunculaceae). Ade, V 1929 (W, holotype).

Cited as synonymous with *Mycosphaerella galatea* by Tomilin (1979). This is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores 8-10 × 2-3 µm.

Mycosphaerella hepaticarum (Pat.) Petr., Ann. Mycol. 32: 378. 1934 ≡ *Stigmatea hepaticarum* Pat., Bull. Soc. Mycol. France 21: 121. 1905.

Type — Algeria: On upper surface of dead thallus of *Lunularia cruciata* (Hepaticae, Marchantiaceae). Trabut, IV 1904 (FH-Patouillard, holotype).

This is morphologically indistinguishable from *Anisomeridium polypori* (Ellis & Everh.) M.E. Barr. It represents the oldest palearctic record of this species, which is thought to be a neophyte in Europe. See Aptroot (2000) for a discussion of hosts, distribution and further synonyms. The specimen mainly consists of its anamorph, *Sarcinulella banksiae* B. Sutton & Alcorn.

Mycosphaerella heraclei (Fr.) Petr., Ann. Mycol. 19: 203. 1921 ≡ *Dothidea heraclei* Fr., Systema Mycol. 2: 556. 1823 ≡ *Phyllachora heraclei* (Fr.) Fuckel, Jahrb. Nassauischen Vereins Naturk. 23-24: 219. 1870 ≡ *Sphaerella heraclei* (Fr.) Traverso, Fl. Ital. Cryptog. 1(11): 633. 1913 ≡ *Oligostroma heraclei* (Fr.) Höhn. ex Petr., Ann. Mycol. 19: 203. 1921.— Fig. 384.

Type — Russia: Kamchatka. On lower surface of dead leaves of *Heracleum sphondylium* ["*trifoliatum*"] (Apiaceae). Wormskjold (UPS-FRIES, holotype).

Anamorph: presumably *Septoria heraclei* (Libert) Desm. *vide* Eriksson (1992).

The holotype shows that this is a *Didymella*, with asci clavate, ascospores 18-22 × 4-5 µm. An *Ascochyta* anamorph is also present in the material.

Mycosphaerella heracleina Nevod., Bot. Mater. Otd. Sporov. Rast. Bot. Inst. Komarova Akad. Nauk SSSR 14: 172. 1961.

Type — Russia: *Heracleum sphondylium* (Apiaceae).

No material was studied as the type was not included in loans from LE or LEP.

Mycosphaerella hermione (Sacc.) Lindau ex Ranojevič, Ann. Mycol. 8: 361. 1910 ≡ *Sphaerella hermione* Sacc., Nuovo Giorn. Bot. Ital. 7: 301. 1875.— Fig. 385.

Type — Italy: Bologna, Casalecchio. On upper and lower surface of dead leaves of *Helleboris viridis* (Ranunculaceae). Saccardo, II 1899, distributed in Mycotheca Italica no. 484 (L, isotype).

Anamorph: *Phyllosticta helleborella* Sacc. *vide* Saccardo (1882).

This is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores 8-11 × 2.5-3 µm.

Sphaerella hertiae Pat., Bull. Soc. Mycol. France 18: 52. 1902.— Fig. 386.

Type — Algeria: Sgag, Aurés. On upper and lower surface of dead leaves of *Hertia cheirifolia* (Asteraceae). Patouillard no. 66, IV 1904 (FH-Patouillard, holotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with asci clavate, ascospores 17-22 × 6-7.5 µm.

Mycosphaerella hesperidum (Penz. & Sacc.) Jacz., Opredelitel' gribov 2: 613. 1917 ≡ *Sphaerella hesperidum* Penz. & Sacc., in Penzig, Atti Reale Ist. Veneto Sci., ser. 6, 2: 12. 1884.— Fig. 387.

Type — Italy: Ventimiglia. On white spots with brown margins on upper and lower surface of living leaves of *Citrus limonum* (Rutaceae). Penzig, IX 1882 (PAD, holotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores 19-22 × 6-7 µm. Also the anamorph, *Cladosporium herbarum* (Pers. : Fr.) Link, is present on the holotype.

Mycosphaerella heucherae (Ellis & Everh.) Petr., Sydowia 11: 340. 1958 ["1957"] ≡ *Dothidella heucherae* Ellis & Everh., Bull. Torrey Bot. Club 27: 571. 1900.— Fig. 388.

Type — USA: Washington, Waitesburg. On upper and lower surface of dead leaves of *Heuchera cylindrica* var. *glabella* (Saxifragaceae). Horner no. 1418, III 1900 (NY, holotype; IMI no. 227776, isotype).

Anamorph: *Phyllosticta heucherae* Ellis & Everh. *vide* Ellis & Everhart (op. cit.).

This belongs to section *Longispora*, and is morphologically indistinguishable from *M. millegrana*, with asci cylindrical, ascospores (15-)17-21 × 3-4 µm.

Sphaerella heufleri Niessl, Verh. Naturf. Vereins Brünn 10: 169. 1872 ["1871"] ≡ *Leptosphaeria heufleri* (Niessl) Sacc., Syll. Fung. 2: 83. 1883 ≡ *Mycotodea heufleri* (Niessl) Kirschst., Ann. Mycol. 34: 201. 1936.

Type — Austria: *Polytrichum formosum* (Musci, Polytrichaceae).

No material was studied as it was already accepted as *Leptosphaeria heufleri* (Niessl) Sacc. by Saccardo (1883).

Sphaerella heveae Petch, Ann. Roy. Bot. Gard. (Peradeniya) 6: 223. 1917.— Fig. 389.

Type — Sri Lanka: Sudayangar. On dead part of living leaves of *Hevea brasiliensis* (Euphorbiaceae). Petch, X 1916 (K, isotype; IMI no. 255141 and IMI no. 31223, isotype slides).

This is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores $7-9 \times 2-2.5 \mu\text{m}$. Additional material (Brazil, Pernambuco, Beberibe, Recife, on pale spots on upper surface of living leaves of *Bowdichia virgilioides* (Euphorbiaceae), Soares, VIII 1960, IMI no. 88194) contains nothing identifiable.

Mycosphaerella heveana (Sacc.) M. Morelet, Ann. Soc. Sci. Nat. Archéol. Toulon & Var 20: 105. 1968 = *Sphaerella heveana* Sacc., Bull. Orto Bot. Regia Univ. Napoli 6: 44. 1921.— Fig. 390.

Type — Singapore: Straits Settlement. On dead parts of living leaves of *Hevea brasiliensis* (Euphorbiaceae). Baker no. 488, VIII 1917 (IMI no. 31222, isotype).

This is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores $8-10 \times 2-2.5 \mu\text{m}$. Additional material (Mauritius, Reduit, Orioux, I 1962, IMI no. 92566b) belongs to *M. subradians*, with asci cylindrical, ascospores $10-12 \times 2.5-3.5 \mu\text{m}$.

Mycosphaerella hevicola Saccas, Agron. Trop. (Nogent-sur-Marne) 8: 247. 1953.

Type — Central African Republic: *Hevea brasiliensis* (Euphorbiaceae).

No material was studied as the type was not included in a loan from PC, and its preservation is uncertain.

Mycosphaerella hibisci Gutner, Trudy Mosk. Inst. Novogo Lubyano Syr'ya 3: 71. 1933.

Type — Russia: *Hibiscus esculenta* (Malvaceae).

Anamorph: Associated with *Phyllosticta hibiscina* Ellis & Everh. *vide* Gutner (op. cit.).

No material was studied as the type was not included in loans from LE or LEP, and its preservation is uncertain.

Mycosphaerella hieracii (Sacc. & Briard) Jaap, Verh. Bot. Vereins Prov. Brandenburg 50: 36. 1908 = *Sphaerella nebulosa* var. *hieracii* Sacc. & Briard, Rev. Mycol. (Toulouse) 7: 208. 1885 = *Sphaerella prignitzensis* Kirschst., Kryptog.-Fl. Mark Brandenburg 7(3): 344. 1938, nomen novum (Article 58).— Fig. 391.

Type — France: *Hieracium* (Asteraceae).

Anamorph: *Ramularia hieracii* (Bäumler) Jaap *vide* Jaap (op. cit.).

The type was not found in any of the herbaria consulted. Materials studied (Germany, Brandenburg, Prignitz, Triglitz, on stems of *Hieracium tridentatum*, Jaap, V 1910, distributed in Fungi Selecti Exsiccati no. 263D, L; also on upper and lower surface of dead leaves of *Hieracium boreale*, Jaap, V 1907, distributed in Fungi Selecti Exsiccati

no. 263B, L) belong to section *Caterva*, and are *M. superflua*, with asci cylindrical, ascospores $15-17 \times 3-3.5 \mu\text{m}$.

Mycosphaerella hieracii (Cooke & Masee) P. Larsen, in Rosenvinge & Warming, Botany of Iceland 2(3), 9. Fungi of Iceland: 489. 1932, later homonym (illegitimate, Article 53) = *Sphaerella hieracii* Cooke & Masee, Grevillea 15: 111. 1887.

Type — United Kingdom: Inbridge Wells. On upper and lower surface of dead leaves of *Hieracium pilosella* (Asteraceae). Masee (K, holotype; NY, 2 isotypes).

The types contain only an *Ascochyta* coelomycete.

Mycosphaerella hieraciophila Petr., Hedwigia 74: 32. 1934.— Fig. 392.

Type — Russia: Siberia, Distr. Tsherepanovo. On dead stems and on upper and lower surface of dead leaves of *Hieracium umbellatum* (Asteraceae). Murashkin no. 425, VI 1925 (W, holotype).

The holotype contains two species of *Mycosphaerella*, and it shows that this is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores $8-10 \times 2-3 \mu\text{m}$. Also present (both on stems and leaves) is *M. superflua*, with asci cylindrical, ascospores $16-19 \times 3-4 \mu\text{m}$.

Mycosphaerella himantia (Pers. : Fr.) Diedicke, Ann. Mycol. 9: 538. 1911 = *Sphaeria himantia* Pers., Observ. Mycol. 2: 69. 1800 [“1799”] = *Dothidea himantia* Pers., sanctioned by Fr., Systema Mycol. 2: 559. 1823 = *Sphaerella himantia* (Pers.) P. Karst., Acta Soc. Fauna Fl. Fenn. 2(6): 66. 1885 = *Omphalospora himantia* (Pers. : Fr.) Höhn., Sitzungsber. Kaiserl. Akad. Wiss., Math.-Naturwiss. Kl., Abt. 1, 128: 601. 1919 = *Plectosphaerella himantia* (Pers. : Fr.) Kirschst., Kryptog.-Fl. Mark Brandenburg 7(3): 310. 1938.— Fig. 393.

Type — Netherlands: Apiaceae.

The type, or any other authentic material, could not be found in L. Material studied (Germany, Bayern, Gerolzhofen, Unterfranken, on stems of *Daucus carota*, Vill, II 1909, distributed in Sydow, Mycotheca Germanica no. 1091, L) belongs to *Davidiella*, and is morphologically indistinguishable from *D. ammophilae*, with asci small, pyriform, ascospores immature, ca. $8 \times 3 \mu\text{m}$. It was accepted as *Omphalospora himantia* (Pers. : Fr.) Höhn. by Müller & von Arx (1962).

Mycosphaerella hippocastani (Jaap) Kleb., Haupt- und Nebenfruchtformen der Askomyzeten: 48. 1918 = *Sphaerella maculiformis* var. *hippocastani* Jaap, Verh. Bot. Vereins Prov. Brandenburg 52: 141. 1910.— Fig. 394.

Type — Germany: Brandenburg, Prignitz, Triglitz. On lower surface of dead leaves of *Aesculus hippocastanum* (Sapindaceae). Jaap, III 1910, distributed in Fungi Selecti Exsiccati no. 423 (B, L (2×), isotypes).

Anamorph: *Septoria aesculi* (Lib.) Westend., *vide* Klebahn (l.c.).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. superflua*, with ascospores $13-16 \times 3-3.5 \mu\text{m}$. It

was cited as synonymous with *Mycosphaerella aesculi* by Tomilin (1979).

Mycosphaerella holci Tehon, Mycologia 29: 434. 1937 = *Didymella holci* (Tehon) Arx, Beih. Nova Hedwigia 87: 288. 1987.— Fig. 395.

Type — USA: Illinois, Oakland, Coles County. On *Sorghum vulgare* var. *technicum* [“*Holcus sorghum* var. *technicus*”] (Poaceae). Stout, X 1927 (IMI no. 142817, isotype slides).

Anamorph: *Phoma sorghina* (Sacc.) Boerema, Dorenb. & Kesteren *fide* Boerema, Dorenbosch & van Kesteren (1973). Accepted as *Didymella holci* (Tehon) Arx by Farr *et al.* (1989), with which the isotype studied agrees well, with copious paraphysoids, asci cylindrical, ascospores 14-16 × 4-5 µm.

Mycosphaerella holmii O.E. Erikss., Non-lichenized Pyrenomycetes of Sweden: 7. 1992, nomen novum (Article 58) for *Mycosphaerella chamaemori* L. Holm & K. Holm, Kew Bull. 31: 571. 1976, later homonym (illegitimate, Article 53).— Fig. 396.

Type — Sweden: Dalarna, Sundborn. On upper and lower surface of dead leaves of *Rubus chamaemorus* (Rosaceae). Holm no. 538b, V 1975 (UPS, holotype).

This was reported not to be *Mycosphaerella* by Tomilin (1979). However, the holotype shows that it belongs to *Davidiella* and is morphologically indistinguishable from *D. ammophilae*, with asci pyriform, ascospores 11-13 × 3.5-5 µm. An additional *Mycosphaerella* species with longer ascospores is also present on the type material, but only on the lower surface of the leaves.

Mycosphaerella holualoana Crous, J.E. Taylor & M.E. Palm, in J.E. Taylor, Crous & M.E. Palm, Mycotaxon 78: 458. 2001.

Type — Hawaii: *Leucospermum* (Proteaceae).
No material was studied of this recently described species.

Mycosphaerella holopteleae Naphade, Sydowia 24: 241. 1971 [“1970”].

Type — India: Pune. On white spots on upper and lower surface of living leaves of *Holoptelea integrifolia* (Ulmaceae). Naphade no. 1232, XI 1968 (LWG, holotype). The holotype material studied contains only a coelomycete.

Mycosphaerella homalanthi Syd. & P. Syd. See *Mycosphaerella omalanthi* Syd. & P. Syd.

Mycosphaerella honckenya Dominik, Acta Soc. Bot. Pol. 11: 240. 1934.

Type — Poland: *Honckenya peploides* (Caryophyllaceae).
Anamorph: Associated with *Alternaria* [“*Macrosporium*”] *fide* Dominik (op. cit.).

Synonymised with *Davidiella allicina* (as *Mycosphaerella tassiana*) by von Arx (1949), cited as a probable synonym of *Mycosphaerella halophila* (E. Bommer, M. Rousseau & Sacc.) O.E. Erikss. by Eriksson (1992). No material was studied as the location of the type is unknown, and its preservation is uncertain.

Mycosphaerella hondai I. Miyake, J. Coll. Agric. Imp. Univ. Tokyo 2: 245. 1910 = *Sphaerella hondai* (I. Miyake) Sacc. & Trotter, Syll. Fung. 22: 144. 1913.

Type — Japan: intercepted at San Francisco. On dead leaves of *Oryza sativa* (Poaceae). Chatterley, XI 1920 (BPI, possible topotype).

This contains only a coelomycete.

Mycosphaerella hordei (P. Karst.) Kirchn., Die Krankheiten und Beschädigungen unserer Landwirtschaftlichen Kulturpflanzen, ed. 3: 76. 1923 = *Sphaerella hordei* P. Karst., Hedwigia 29: 271. 1890.

Type — Finland: *Hordeum vulgare* (Poaceae).

Cited as synonymous with *Davidiella allicina* (as *Mycosphaerella allicina*) by Tomilin (1979). No material was studied as the type was not included in a loan from H, and its preservation is uncertain.

Sphaerella hordeicola Hara, in Tsuruda, J. Pl. Protect. 3: 532. 1916 [as “*hordicola*”].

Type — Japan: *Hordeum* (Poaceae).

No material was studied as the type was not included in a loan from TNS, and its preservation is uncertain.

Sphaerella horii Hara, J. Jap. Hort. Soc. 29: 278. 1917.

Type — Japan: *Citrus* (Rutaceae).

Cited as synonymous with *Mycosphaerella gibelliana* by Tomilin (1979).

No material was studied as the type was not included in a loan from TNS, and its preservation is uncertain.

Mycosphaerella hosackiae (Cooke & Harkn.) Bonar, Mycologia 57: 384. 1965 = *Sphaerella hosackiae* Cooke & Harkn., Grevillea 9: 86. 1881 = *Ditopella hosackiae* (Cooke & Harkn.) Sacc., Syll. Fung. 1: 451. 1882 = *Physalospora hosackiae* (Cooke & Harkn.) Cooke, Grevillea 17: 88. 1889.— Fig. 397.

Type — USA: California, Mt. Jamalpais. On dead twigs of *Hosackia* (Fabaceae). Harkness no. 1393, IV 1880 (K, holotype; BPI, isotype).

Anamorph: *Microdiplodia hosackiae* (Cooke & Harkn.) Bonar *fide* Bonar (op. cit.).

Accepted as *Ditopella hosackiae* (Cooke & Harkn.) Sacc. by Saccardo (l.c.). The types contain only the anamorph. Additional material seen (Contra Costa County, Mt. Diablo State park, Rock City Campground, on dead twigs of *Hosackia glabra*, Bonar, IV 1960, IMI no. 142273) contains *Dothidotthia ramulicola* (Peck) M.E. Barr, with a hamathecium consisting of ca. 5 µm wide isodiametric cells, asci clavate, thick-walled, ascospores pale brown, 15-20 × 7-9 µm.

Mycosphaerella hostae Syd. & P. Syd., Ann. Mycol. 11: 59. 1913 = *Sphaerella hostae* (Syd. & P. Syd.) Trotter, Syll. Fung. 24: 876. 1928.— Fig. 398.

Type — Japan: Mino, Kawanye-mura. On upper and lower surface of dead leaves of *Hosta japonica* (Asparagaceae). Hara, IV 1912 (S, holotype).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradians*, with asci cylindrical, ascospores 12-15 × 3-3.5 µm.

Mycosphaerella hranicensis Petr., Ann. Mycol. 19: 75. 1921.

Type — Czech Republic: Weißkirchen, Ribar. On dead parts of living leaves of *Carex silvatica* (Cyperaceae). Petrak, V 1919 (W, holotype), also distributed in Flora Bohemia et Moravia Exsiccata no. 1240 (W, isotype).

Cited as synonymous with *Mycosphaerella caricicola* by Tomilin (1979). The type shows that this belongs to *Davidiella*, and is morphologically indistinguishable from *D. disseminata*, with asci pyriform, ascospores 11-14 × 3-4 µm.

Mycosphaerella humuli (Hazsl.) Cruchet, Bull. Soc. Vaud. Sci. Nat. 55: 43. 1923 = *Sphaerella humuli* Hazsl., Math. Természettud. Közlem. 25: 107. 1893 [“1892”].

Type — Hungary: *Humulus lupulus* (Urticaceae).

The type is not in BP and may have been destroyed during the war.

Mycosphaerella huteriana Petr., Sydowia 16: 172. 1963.— Fig. 399.

Type — Austria: Kärnten, Gamsgrube. On upper and lower surface of dead leaves of *Saxifraga macropetala* (Saxifragaceae). Huter, VII 1887 (W, holotype).

This belongs to section *Longispora*, and is morphologically indistinguishable from *M. topographica*, with asci cylindrical, ascospores 26-30 × 4-5 µm. Additional material seen (Brennergebiet, Vennetal, on *S. biflora*, Hepp, VIII 1891, W) agrees well.

Sphaerella hyalospora (Ces.) Ces. & De Not., Comment. Soc. Crittog. Ital. 1(4): 237. 1863 = *Sphaeria hyalospora* Ces., Hedwigia 2: 24. 1859 = *Physalospora hyalospora* (Ces.) Sacc., Syll. Fung. 1: 439. 1882.

Type — Italy: *Salix* (Salicaceae).

Accepted as *Physalospora hyalospora* (Ces.) Sacc. by von Arx & Müller (1954) and therefore not studied.

Mycosphaerella hydrangeae Hara, J. Pl. Protect. 5: 615. 1918 [as “*hydrangiae*”].— Fig. 400.

Type — Japan: On dead stems of *Hydrangea* (Saxifragaceae).

The type was not included in a loan from TNS, and its preservation is uncertain. Material studied (USA, Oregon, Zeller, III 1936, NY, 2×) belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores 17-20 × 5-7 µm.

Mycosphaerella hydrocotyles-asiaticae (Pat.) Petr., Ann. Mycol. 27: 375. 1929 = *Sphaerella hydrocotyles-asiaticae* Pat., Bull. Soc. Mycol. France 34: 90. 1918 = *Mycosphaerella centellae-asiaticae* (Pat.) Petr., in Syd., Ann. Mycol. 35: 353. 1937 [in error].— Fig. 401.

Type — Madagascar. On white spots on lower surface of probably already dead leaves of *Hydrocotyle asiatica* (Araliaceae). Viguier no. 670 (FH-Patouillard, holotype).

This is a parasitic species, with asci clavate, ascospores 14-17 × 4.5-5.5 µm.

Mycosphaerella hyperici (Auersw.) Starbäck, Bih. Kongl. Svenska Vetensk.-Akad. Handl. 15(3, 2): 9. 1889 =

Sphaerella hyperici Auersw., in Gonn. & Rabenh., Mycol. Europaea 5-6: 14. 1869.

Type — Germany: On dead stems of *Hypericum perforatum* (Hypericaceae).

Anamorph: *Septoria hyperici* Roberge *vide* Eriksson (1992). The type is not in B and may have been destroyed during the war. Material studied (Germany, Brandenburg, Tamsel, Vogel, III 1929, distributed in Sydow, Mycotheca Germanica no. 2527, L) belongs to *Davidiella*, and is morphologically indistinguishable from *D. disseminata*, with asci pyriform, ascospores immature.

Mycosphaerella hypericina (Ellis) Tomilin, Opredelitel' gribov roda *Mycosphaerella* Johans.: 139. 1979 = *Sphaerella hypericina* Ellis, Bull. Torrey Bot. Club 9: 74. 1882.— Fig. 402.

Type — USA: New Jersey, Newfield. On upper and lower surface of dead leaves of *Hypericum prolificum* (Hypericaceae). Ellis, V 1880, distributed in North American Fungi no. 797 (L, NY (5×), isotypes).

This is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores 9-11 × 2-2.5 µm.

Sphaerella hyperopta Rehm, Ascomyceten no. 348. 1876, nomen herbariorum (not validly published, Article 32).— Fig. 403.

Authentic material — Germany: Sachsen, Eisleben. On *Carex acuta*. Kunze, VII 1875, distributed in Rehm, Ascomyceten no. 348 (B).

This is *Didymella proximella* (P. Karst.) Sacc., see Aptroot (1995a).

Mycosphaerella hyphiseda (Fautrey & Lambotte) Tomilin, Novosti Sist. Nizsh. Rast. 1967: 189. 1967 = *Sphaerella hyphiseda* Fautrey & Lambotte, in Fautrey, Rev. Mycol. (Toulouse) 19: 143. 1897.

Type — France: *Ornithogalum pyrenaicum* (Alliaceae).

No material was studied as the type was not included in a loan from PC nor found in any of the herbaria consulted.

Mycosphaerella hypochaeridis Morozkovsky, Bot. Zhurn. (Kiev) 2: 183. 1946.

Type — Ukraine: *Hypochaeris maculata* (Asteraceae).

No material was studied as the location of the type is unknown and its preservation uncertain.

Mycosphaerella hypodermellae Wehm., Mycologia 38: 164. 1946.

Type — USA: *Pinus murrayana* (Pinaceae).

No material was studied as the type was not found in any of the herbaria consulted.

Mycosphaerella hypostomatica Höhn., Sitzungsber. Kaiserl. Akad. Wiss., Math.-Naturwiss. Kl., Abt. 1, 111: 7. 1903 [“1902”] = *Sphaerella hypostomatica* (Höhn.) Sacc. & D. Sacc., Syll. Fung. 17: 644. 1905.— Fig. 404.

Type — Austria: Laudonpark. On upper surface of dead leaves of *Luzula campestris* (Juncaceae). Höhnel (FH-Höhnel, 2 microscopic slides serving as holotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. disseminata*, with asci pyriform, ascospores $10-13 \times 2.5-3.5 \mu\text{m}$.

Mycosphaerella hypsicola (Ellis & Everh.) Lindau, in Engl. & Prantl, *Natürlichen Pflanzenf.* 1(1): 424. 1897 = *Sphaerella hypsicola* Ellis & Everh., *Proc. Acad. Nat. Sci. Philadelphia* 46: 334. 1894.— Fig. 405.

Type — USA: Colorado, Larimer Co., Cameron Pass. On dead stems of *Trollius laxus* (Ranunculaceae). Crandall no. 4, VII 1894 (NY, holotype).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradicans*, with asci cylindrical, ascospores $11-13 \times 3-4 \mu\text{m}$.

Sphaerella hystrix Fautrey, in Fautrey & Lambotte, *Rev. Mycol. (Toulouse)* 17: 170. 1895.

Type — France: Noidan. On upper and lower surface of dead leaves of *Asperella hystrix* (Poaceae). Fautrey, III 1895, distributed in Roumeguère, *Fungi Selecti Exsiccati* no. 6885 (PC, holotype).

The holotype contains only an *Ascochyta*, which may have led to the description of the *Sphaerella*.

Mycosphaerella idaeina (Hazsl.) Lindau, *Hilfsb. Sammeln Ascomyceten*: 102. 1903 = *Sphaerella idaeina* Hazsl., *Math. Természettud. Közlem.* 25: 104. 1893 [“1892”].— Fig. 406.

Type — Hungary. On upper surface of dead leaves of *Rubus idaeus* (Rosaceae).
The type is not in BP and may have been destroyed during the war. Material studied (Germany, Hessen, Dillkreis, Donsbach, Ludwig, V 1939, B) is a *Guignardia* sp. with ascospores rhomboid, simple, $10-12 \times 4-5 \mu\text{m}$.

Mycosphaerella idesiae Hara, *J. Pl. Protect.* 5: 617. 1918.

Type — Japan.

No material was studied as the type was not included in a loan from TNS.

Mycosphaerella ignobilis (Auersw.) Lindau, *Hilfsb. Sammeln Ascomyceten*: 50. 1903 = *Sphaerella ignobilis* Auersw., in Gonn. & Rabenh., *Mycol. Europaea* 5-6: 17. 1869.— Fig. 407.

Type — Germany: Leipzig. On dead leaves of *Glyceria spectabilis* (Poaceae). Auerswald, IV 1866 (B, holotype; B, isotype).

Both type specimens contain only a *Phaeosphaeria* sp. indet., with ascospores 3-septate, old, shriveled, brown, $23-25 \times 5-7 \mu\text{m}$. Additional material studied (on *Dactylis glomerata*, Winter no. 873, L) belongs to *Davidiella*, and is morphologically indistinguishable from *D. disseminata*, with asci pyriform, ascospores immature, ca. $13 \times 3.5 \mu\text{m}$.

Sphaerella ignobilis f. *bromi* Gonz. *Frag., Trab. Mus. Nat. Ci. Nat., Ser. Bot.* 10: 80. 1916.

Type — Spain: *Bromus mollis* (Poaceae).

No material was studied as the type was not included in a loan from MA.

Mycosphaerella ikedai Hara, *Tea J.* 14: 10. 1919 = *Sphaerella ikedai* (Hara) Trotter, *Syll. Fung.* 24: 888. 1928.— Fig. 408.

Type — Japan. On upper and lower surface of dead leaves of *Camellia* [“*Thea*”] *sinensis* (Theaceae). Hara, IX 1918 (TNS 209267, holotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores $14-17 \times 5-6 \mu\text{m}$. Additional material (Malaya, Bidur, Parak, Johnston, XII 1952, IMI no. 62203) contains nothing identifiable.

Mycosphaerella ilicella (Cooke) Feltgen, *Recueil Mém. Trav. Soc. Bot. Grand-Duché Luxembourg* 15: 256. 1903 = *Sphaerella ilicella* Cooke, *Grevillea* 8: 119. 1880.

Type — USA: New York. *Ilex opaca* (Aquifoliaceae). 267 (K, holotype; K, 2 isotypes).

This is morphologically indistinguishable from *Aulographum hederæ* Lib. It was not studied in detail.

Mycosphaerella ilicicola (Maubl.) M. Morelet, *Bull. Soc. Sci. Nat. Archéol. Toulon & Var* 176: 7. 1968 = *Sphaerella ilicicola* Maubl., in Rangel, *Bol. Agric. (São Paulo)* 16: 313. 1915 [as “*illicicola*”].

Type — Brazil: *Ilex paraguariensis* (Aquifoliaceae).

No material was studied as the type was not included in a loan from PC.

Mycosphaerella ilicis (Ellis) Lindau, in Engl. & Prantl, *Natürlichen Pflanzenf.* 1(1): 425. 1897 = *Sphaerella ilicis* Ellis, *Amer. Naturalist* 17: 317. 1883.— Fig. 409.

Type — USA: New Jersey, Newfield. On white spots on upper and lower surface of living leaves of *Ilex glabra* (Aquifoliaceae). Ellis, VI 1882 (NY, holotype), also distributed in Ellis & Everhart, *North American Fungi* no. 1351 (L, NY (3×), isotypes).

This is a parasitic species, with asci cylindrical, ascospores $12-15 \times 3-3.5 \mu\text{m}$.

Mycosphaerella ilicis-canariensis Petr., *Bot. Jahrb. Syst.* 62, *Beibl.* 142: 120. 1928.— Fig. 410.

Type — Canary Islands: Tenerife, Anaga-mountains. On white spots with black margins on upper surface of living leaves of *Ilex canariensis* (Aquifoliaceae). Ade, V 1926 (W no. 10058, holotype; W no. 26890, isotype).

This is a parasitic species, with asci cylindrical, ascospores $12-17 \times 3-4 \mu\text{m}$.

Mycosphaerella immersa Dearn., *Rep. Canad. Arctic Exped. 1913-1918* 4, C: 6. 1923.

Type — Canada: *Cassiope tetragona* (Ericaceae).

Cited as synonym of *Discostroma hyperborea* (P. Karst.) O.E. Erikss. by Barr (1959) and therefore not studied.

Mycosphaerella impatientina Syd. & Hara, in Syd. & P. Syd., *Ann. Mycol.* 11: 58. 1913 = *Sphaerella impatientina* (Syd. & Hara) Trotter, *Syll. Fung.* 24: 852. 1928.— Fig. 411.

Type — Japan: Mino, Kawanye-mura. On dead stems of *Impatiens* (Balsaminaceae). Hara, III 1912 (S, holotype).

Cited as synonymous with *M. subradians* by Tomilin (1979). However, the holotype shows that this is a species of *Discostroma*, with paraphyses 2 µm wide, asci cylindrical, ascospores 11-13 × 3.5-4 µm.

Mycosphaerella impatientis (Peck & Clinton) House, New York State Mus. Bull. 233-234: 28. 1921 ≡ *Sphaerella impatientis* Peck & Clinton, Annual Rep. New York State Mus. 30: 67. 1878 [“1877”].— Fig. 412.

Type — USA: New York, Adirondack Mountains. On upper and lower surface of probably still living leaves of *Impatiens fulva* (Balsaminaceae). Peck, VIII 1876, distributed in Thümen, Mycotheca Universalis no. 963 (L, NY, isotypes).

Anamorph: *Septoria fide* Sivanesan (1984).

This is a parasitic species, with asci cylindrical, ascospores 15-18 × 4-5 µm.

Mycosphaerella imperatae Sawada, Rep. Gov. Res. Inst. Formosa 87: 24. 1944, lacking Latin description (not validly published, Article 36).

Type — Taiwan: *Imperata cylindrica* var. *koenigii* (Poaceae).

No material was studied as the type was not found in BPI.

Mycosphaerella implexae (Pass.) M. Morelet, Ann. Soc. Sci. Nat. Archéol. Toulon & Var 17: 103. 1965 ≡ *Sphaerella implexae* Pass., Atti Reale Accad. Lincei, Mem. Cl. Sci. Fis., ser. 4, 3: 7. 1887 [as “*implexa*”].

Type — Italy: On brown spots with black margins on upper surface of living leaves of *Lonicera implexa* (Adoxaceae).

Material studied (Malta, Caruana, 1913-1914, PAD) is sterile.

Sphaerella implexae var. *implexicola* (Maire) Sacc. See *Mycosphaerella implexicola* (Maire) Jaap.

Mycosphaerella implexicola (Maire) Jaap, Ann. Mycol. 14: 14. 1916 ≡ *Sphaerella implexicola* Maire, in Maire, Dumée & Lutz, Bull. Soc. Bot. France 48: CXCVIII. 1901 ≡ *Sphaerella implexae* var. *implexicola* (Maire) Sacc., Nuovo Giorn. Bot. Ital., ser. 2, 22: 35. 1915.— Fig. 413.

Type — France: On white spots with black margins on upper and lower surface of living leaves of *Lonicera implexa* (Adoxaceae).

The type was not included in a loan from PC. Already cited as synonymous with *Mycosphaerella clymenia* by Tomilin (1979). Materials studied (Malta, Caruana, 1913-1914, PAD; also Corsica, Ajaccio, Höhnel, IV 1905, distributed in Rehm, Ascomyceten no. 1619, BPI, 2×) agree with this and belong to section *Plaga*, with asci cylindrical, ascospores 9.5-10.5 × 2-2.5 µm.

Mycosphaerella inaequalis Hino & Katum., Bull. Fac. Agric. Yamagata Univ. 8: 657. 1957 ≡ *Pseudomassaria inaequalis* (Hino & Katum.) Hino & Katum., J. Jap. Bot. 40: 84. 1965.

Type — Japan: Honshu, Nagato. On leaves of *Sasa tyungokensis* (Poaceae). Katumoto, VIII 1956 (YAM, holotype, not seen).

No material was studied as the species was already excluded from the genus by the original authors.

Sphaerella inaequalis Cooke, J. Bot. 4: 248. 1866 ≡ *Didymosphaeria inaequalis* (Cooke) Niessl, in Rabenh., Fungi Europaei Exsiccati no. 2663. 1881 ≡ *Venturia inaequalis* (Cooke) G. Winter, Hedwigia 36: 81. 1897 ≡ *Endostigme inaequalis* (Cooke) Syd., Ann. Mycol. 21: 171. 1923 ≡ *Spilosticta inaequalis* (Cooke) Petr., Ann. Mycol. 38: 193. 1940.

Type — United Kingdom: Shere. On lower surface of dead leaves of *Sorbus* [“*Pyrus*”] *torminalis* (Rosaceae). Cooke, Fungi Britannici Exsiccati no. 173, IV 1866 (K, 2 isotypes; B, 2 isotypes).

Accepted as *Venturia inaequalis* (Cooke) G. Winter by Sivanesan (1977), with which additional material studied (Czech Republic, Brno [“Brünn”], Niessl, distributed in Rabenhorst, Fungi Europaei Exsiccati no. 2663, III 1913, L) agrees well.

Sphaerella inaequalis var. *salicis* Cooke, Fungi Britannici Exsiccati no. 690. 1874.

Type — United Kingdom. On upper surface of dead leaves of *Salix* (Salicaceae). Cooke, Fungi Britannici Exsiccati no. 690 (B, isotype).

The type is overmature, but shows that this is morphologically indistinguishable from *M. punctiformis*.

Mycosphaerella incanescens (Schwein.) Tomilin, Novosti Sist. Nizsh. Rast. 6: 121. 1970 [“1969”] ≡ *Sphaeria incanescens* Schwein., Trans. Amer. Philos. Soc., ser. 2, 4: 225. 1832 ≡ *Sphaerella incanescens* (Schwein.) Cooke, J. Bot. 21: 107. 1883.— Fig. 414.

Type — USA: Philadelphia, Bethlehem. On lower surface of dead leaves of *Tilia* (Malvaceae). Schweinitz (PH, holotype; K, PH, isotypes).

Already cited as synonymous with *M. punctiformis* by Tomilin (1979), with which the type specimens agree well, with asci cylindrical, ascospores 10-12 × 2-2.5 µm.

Sphaerella incisa Ellis & G. Martin, J. Mycol. 1: 99. 1885.

Type — USA: *Sabal serrulata* (Arecaceae).

No material was studied, as no type material was found in NY or any of the other herbaria consulted. This is reported by Fröhlich and Hyde (1998), who did not examine any material, to be possibly *Oxydothis sabalensis* (Cooke) Petr.

Mycosphaerella incomperta Podl.-Ruz. & Svrcek, Česká Mykol. 24: 130. 1970.

Type — Czech Republic: *Alnus viridis* (Betulaceae).

No material was studied as the type was not found in any of the herbaria consulted.

Sphaerella inconspicua (Desm.) Ces. & De Not., Comment. Soc. Crittog. Ital. 1(4): 237. 1863 ≡ *Sphaeria inconspicua* Desm., Ann. Sci. Nat. Bot., ser. 2, 19: 354. 1843.

Type — France: On bark of *Platanus orientalis* (Platanaceae) [in publication as “*Acer*” (Sapindaceae)]. Desmazières, Plantes Cryptogames de France, ser. 1, no. 1270 (PC, holotype).

The holotype contains only a coelomycete, as already annotated on the material by Desmazières in 1849.

Mycosphaerella inconspicua Vestergr., Svensk Bot. Tidskr. 3: 53. 1909, nomen novum (Article 58) for *Sphaerella inconspicua* J. Schröt., Jahresber. Schles. Ges. Vaterl. Cult. 58: 173. 1880, later homonym (illegitimate, Article 53) = *Mycosphaerella cassiopes* M.E. Barr, Contr. Inst. Bot. Univ. Montréal 73: 14. 1959, superfluous (illegitimate, Article 52) nomen novum (Article 58).— Fig. 415.

Type — Sweden: On dead stems and fruits of *Cassiope* ["*Andromeda*"] *tetragona* (Ericaceae).

The type was not included in a loan from S. Material studied (Lapland, Lula, Sarjek, Vestergren, VIII 1901, distributed in Micromycetes Rariores Selecti, BPI, 2×) belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores 12-18 × 4-5.5 µm.

Mycosphaerella indica T.S. Viswan., Mycopathol. Mycol. Appl. 12: 157. 1960.

Type — India: *Morus alba* (Moraceae).

No material was studied as the type was not found in IMI or LWG.

Mycosphaerella indistincta (Peck) Lindau, in Engl. & Prantl, Natürlichem Pflanzenf. 1(1): 426. 1897 = *Sphaerella indistincta* Peck, Annual Rep. New York State Mus. 28: 81. 1877.— Fig. 416.

Type — USA: New York, Albany, Kamer. On upper and lower surface of dead fronds of *Pteridium aquilinum* (Polypodiaceae). Peck, VI 1874 (NY, isotype); also VI 1976, distributed in Thümen, Mycotheca Universalis no. 759 (L, topotype).

This belongs to section *Longispora*, and is morphologically indistinguishable from *M. millegrana*, with asci cylindrical, ascospores 16-21 × 2-2.5 µm.

Mycosphaerella inflata (Penz.) Tomilin, Opredelitel' gribov roda *Mycosphaerella* Johans.: 239. 1979 = *Sphaerella inflata* Penz., Michelia 2: 412. 1882.

Type — Italy: *Citrus aurantium* (Rutaceae).

No material was studied as the type was not found in any of the herbaria consulted.

Mycosphaerella infuscans (Ellis & Everh.) M.E. Barr, Contr. Univ. Michigan Herb. 9: 592. 1972 = *Sphaerella infuscans* Ellis & Everh., Bull. Torrey Bot. Club 25: 504. 1898.— Fig. 417.

Type — USA: West Virginia, Fayette Co., Nuttallburg. On dead petioles of *Liriodendron tulipifera* (Magnoliaceae). Nuttall no. 937, V 1898 (NY, holotype; NY, 2 isotypes), also distributed in Ellis & Everhart, Fungi Columbiani no. 1330 (H, NY (4×), isotypes).

This belongs to section *Longispora*, and is morphologically indistinguishable from *M. millegrana*, with ascospores 15-22 × 2.5-3 µm. Cited as synonymous with *Mycosphaerella tulipifera* by Tomilin (1979).

Sphaerella innata (Nyl.) Arnold, Flora 53: 488. 1870 = *Verrucaria innata* Nyl., Flora 48: 358. 1865 = *Pharcidia*

allogena f. *innata* (Nyl.) Keissl., in Rabenh., Kryptog.-Fl. Deutschl., Österr., Schweiz 2, 8: 363. 1930.

Type — Germany: *Psora decipiens* (Ascomycota, Lecideaceae).

Cited as synonymous with *Stigmatidium allogenum* (Nyl.) D. Hawksw. by Clauzade, Diederich & Roux (1989).

Mycosphaerella innumerella (P. Karst.) Starbäck, Bih. Kongl. Svenska Vetensk.-Akad. Handl. 15(3, 2): 9. 1889 = *Sphaerella innumerella* P. Karst., Fungi Fenniae Exsiccati no. 965. 1870.— Fig. 418.

Type — Finland: Tavastia Australis, Tammela, Mustiala. On upper and lower surface of dead leaves of *Comarum palustre* (Rosaceae). Karsten, Fungi Fenniae Exsiccati no. 965 (H, holotype), also distributed as no. 3646 (H, isotype). Cited as synonymous with *Mycosphaerella ranunculi* (P. Karst.) Lind by Barr (1972). The type specimens are immature, but show that it is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical and ascospores immature but small. Additional material studied (Czech Republic, Eger in Böhmen, Konradsgrün, Jahn, distributed in Petrak, Flora Bohemiae et Moraviae Exsiccata no. 1242, 1916, L) belongs to section *Longispora*, and is morphologically indistinguishable from *M. millegrana*, with asci cylindrical, ascospores 14-17 × 2.5-3 µm.

Sphaerella innumerella var. *rubi* Hazsl., Math. Természettud. Közlem. 25: 104. 1892.

Type — Hungary: *Rubus fruticosus* (Rosaceae).

The type is not in BP and may have been destroyed during the war.

Mycosphaerella ["*Mycosphaerium*"] *insidens* Clem., Cryptogamae Formationum Coloradensium no. 221. 1906, nomen herbariorum (not validly published, Article 32).

Authentic material — USA: Colorado, Minnehaha. On dead stems of *Thalictrum sparsiflorum* (Ranunculaceae). Clements, VII 1906, Cryptogamae Formationum Coloradensium no. 221 (BPI).

The material is immature.

Mycosphaerella insignita Syd., Ann. Mycol. 37: 371. 1939.— Fig. 419.

Type — Ecuador: Prov. Tungurahua, Baños, Hacienda San Antonio. On upper surface of dead leaf parts of living fronds of *Pteridium arachnoideum* (Polypodiaceae). Sydow, XII 1937 (NY, isotype), also distributed in Fungi Exotici Exsiccati no. 1190 (L, isotype), also distributed in Reliquiae Petrakianae no. 1250 (H, L (2×), isotypes).

This is a species of the Parmulariaceae, with ascomata multi-layered, with schizolytic, slit-like ostioles, asci thick-walled, clavate, surrounded by copious paraphysoids, ascospores 32-40 × 11-13 µm.

Mycosphaerella insulana Bubák & Syd., Ann. Mycol. 13: 7. 1915 = *Sphaerella insulana* (Bubák & Syd.) Trotter, Syll. Fung. 24: 882. 1928.— Fig. 420.

Type — Germany: Wadden Sea Island Sylt. On dead stems of *Rumex* (Polygonaceae). Sydow, VIII 1911 (BPI, holotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. ammophilae*, with asci pyriform, ascospores $11-13.5 \times 3-4 \mu\text{m}$.

Sphaerella insularis (Wallr.) Fuckel, Jahrb. Nassauischen Vereins Naturk. 23-24: 107. 1870 = *Sphaeria insularis* Wallr., Fl. Cryptog. Germ. 2: 814. 1833.

Type — Germany: Oestrich. On lower surface of dead leaf parts of living leaves of *Alnus glutinosa* (Betulaceae).

Cited as synonymous with *M. punctiformis* by Tomilin (1979). Materials studied (Fuckel, Fungi Rhenani Exsiccati no. 827, L; also Netherlands, Rotterdam, Botanical garden, on *Pyrus communis*, Oudemans, VIII 1850, L) contain only an immature ascomycete with an *Asteromella* anamorph.

Sphaerella intercellularis (Berk. & M.A. Curtis) Cooke, J. Bot. 21: 137. 1883 = *Sphaeria intercellularis* Berk. & M.A. Curtis, Grevillea 4: 153. 1876.

Type — USA: *Typha* (Typhaceae).

No material was studied as the type was not found in K or NY.

Mycosphaerella intermedia M. Dick & K. Dobbie, New Zeal. J. Bot. 39: 272. 2001.

Type — New Zealand: *Eucalyptus saligna* (Myrtaceae).

No material was studied of this recently described species.

Mycosphaerella intermixta Lindau, Hilfsb. Sammeln Ascomyceten: 23. 1903, nomen novum (Article 58) for *Sphaerella intermixta* Niessl, Oesterr. Bot. Z. 31: 346. 1881, later homonym (illegitimate, Article 53) = *Mycosphaerella niesslii* Tomilin, Novosti Sist. Nizsh. Rast. 8: 151. 1971, superfluous (illegitimate, Article 52) nomen novum (Article 58).

Type — Austria: *Campanula* (Campanulaceae).

No material was studied as the type was not included in a loan from M.

Sphaerella intermixta (Berk. & Broome) Ces. & De Not., Comment. Soc. Crittog. Ital. 1(4): 237. 1863 = *Sphaeria intermixta* Berk. & Broome, Ann. Mag. Nat. Hist., ser. 2, 9: 327. 1852 = *Sphaerulina intermixta* (Berk. & Broome) Sacc., Fungi Italici Autographice Delineati, tab. 347. 1878.

Type — United Kingdom: *Rosa* (Rosaceae). Massee (K, holotype).

Reported to be morphologically indistinguishable from *Sacothecium sepincola* (Fr.) Fr. by Cannon *et al.* (1985).

Mycosphaerella ipiranguensis (Speg.) Tomilin, Novosti Sist. Nizsh. Rast. 8: 150. 1971 = *Sphaerella ipiranguensis* Speg., Bol. Acad. Nac. Ci. 23: 469. 1919.

Type — Brazil: *Tabebuia* (Bignoniaceae).

No material was studied as the type was not included in a loan from LPS.

Mycosphaerella ipomoeae (Ferraris) Dunin ex Tomilin, Opredelitel' gribov roda *Mycosphaerella* Johans.: 89. 1979 = *Sphaerella ipomoeae* Ferraris, in Ferraris & Massa, Ann. Mycol 10: 286. 1912.

Type — Italy: *Ipomoea purpurea* (Convolvulaceae).

Anamorph: Associated with *Sphaeropsis ipomoeae* Ellis & Everh. *vide* Ferraris & Massa (op. cit.).

No material was studied as the type was not found in any of the herbaria consulted.

Mycosphaerella iridis (Auersw.) J. Schröt. in Cohn, Kryptog.-Fl. Schlesien 3(2): 339. 1894 ["1893"] = *Sphaerella iridis* Auersw., in Gonn. & Rabenh., Mycol. Europaea 5-6: 18. 1869 (not *Didymellina iridis* (Desm.) Höhn., Ann. Mycol. 16: 66. 1918 = *Dothidea iridis* Desm., Ann. Sci. Nat. Bot., ser. 3, 8: 176. 1847 = *Metasphaeria iridis* (Desm.) Sacc., Syll. Fung. 2: 178. 1883 = *Laestadia iridis* (Desm.) Berl. & Voglino, Syll. Fung., Addit. ad vol. 1-4: 61. 1886).— Fig. 421.

Type — Germany: On upper and lower surface of dead leaf parts of living leaves of *Iris pumila* (Iridaceae).

Anamorph: *Cladosporium iridis* (Fautrey & Roum.) G.A. de Vries *vide* David (1997).

The type was not found in B and may be lost. Materials studied (France, Hermanville, Roberge, B-Desmazières; also Czech Republic, Mähren, Unterwald, on *Iris pseudacorus*, Zimmerman, distributed in Petrak, Flora Bohemiae et Moraviae Exsiccata no. 151, VI 1912, L) belong to *Davidiella*, and are morphologically indistinguishable from *D. allicina*, with asci globose to pyriform, ascospores $13-16 \times 5-7 \mu\text{m}$.

Sphaerella iridis Cooke, Grevillea 13: 99. 1885, later homonym (illegitimate, Article 53).

Type — United Kingdom: *Iris germanica* (Iridaceae).

No material was studied as the type was not found in K.

Sphaerella iridis var. *anceps* Sacc., Ann. Mycol. 7: 435. 1909.— Fig. 422.

Type — Germany: Brandenburg, Berlin. On upper and lower surface of dead leaves of *Iris pseudacorus* (Iridaceae). Sydow, V 1900 (B, isotype).

The type and additional material studied (Germany, Brandenburg, Berlin, Syd., Mycotheca Germanica no. 786, V 1909, L) belong to section *Caterva*, and show that this is morphologically indistinguishable from *M. superflua*, with asci cylindrical, ascospores $16-20 \times 3.5-4.5 \mu\text{m}$.

Sphaerella iridis var. *ancipitella* Sacc., Ann. Mycol. 11: 15. 1913.

Type — France: *Iris pseudacorus* (Iridaceae).

No material was studied as the type was not found in any of the herbaria consulted.

Mycosphaerella irregulariramosa Crous & M.J. Wingf., Canad. J. Bot. 75: 785. 1997.

Type — South Africa: Northern Province, Tzaneen. On the leaves of *Eucalyptus* (Myrtaceae). Wingfield, III 1996 (PREM 54964, holotype, not seen).

Anamorph: *Pseudocercospora irregulariramosa* Crous & M.J. Wingf. *vide* Crous (1998).

No material was studied of this recently described species.

Mycosphaerella isariphora (Desm.) Johanson, Öfvers. Förh. Kongl. Svenska Vetensk.-Akad. 41(9): 165. 1884 = *Sphaeria isariphora* Desm., Ann. Sci. Nat. Bot., ser. 2, 19:

358. 1843 = *Sphaerella isariphora* (Desm.) Ces. & De Not., Comment. Soc. Crittog. Ital. 1(4): 237. 1863 = *Leptosphaeria isariphora* (Desm.) Auersw., in Gonn. & Rabenh., Mycol Europaea 5-6: fig. 170. 1869.— Fig. 423. Type — France: On upper and lower surface of dead leaves of *Stellaria holostea* (Caryophyllaceae). Desmazières, Fungi Exsiccati no. 791 (L, isotype), also Plantes Cryptogames de France no. 1291 (G, isotype).

Anamorphs: *Cephalotrichum pallescens* (Fuckel) Sacc., *Cephalotrichum pusillum* (Fuckel) Sacc. and *Septoria stellariae* (Roberge & Desm.) Westend. *vide* Saccardo (1882).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradicans*, with asci cylindrical, ascospores 11-15 × 3.5-4.5 µm. Additional material seen (Netherlands, Nunspeet, Beins, 1898, L) contains only a coelomycete.

Sphaerella isariphora f. *stellariae-mediae* Thüm., Fungi Austriaci no. 1257. 1873.

Type — Germany: Bohemia, Königswald. On upper and lower surface of dead leaves of *Stellaria media* (Caryophyllaceae). Thümen, Fungi Austriaci no. 1257, 1873 (BPI, isotype).

The isotype is immature, but morphologically indistinguishable from the nominal forma, and thus morphologically indistinguishable from *M. subradicans*.

Mycosphaerella isatidis Kalymb., Trudy Inst. Bot. Akad. Nauk Kazakh. SSR 7: 326. 1959.

Type — Kazakhstan: *Isatis tinctoria* (Brassicaceae). No material was studied as the type was not included in loans from LE or LEP.

Sphaerella ischnosperma Kirschst., Hedwigia 80: 123. 1941.— Fig. 424.

Type — Germany: Oberbayern, Heilbrunn. On dead stems of *Galium silvaticum* (Rubiaceae). Kirschstein, VI 1938 (B, holotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. ammophilae*, with asci pyriform, ascospores 8-10 × 2-3 µm.

Mycosphaerella isoplexidis Petr., Bot. Jahrb. Syst. 62, Beibl. 142: 121. 1928.— Fig. 425.

Type — Canary Islands: Tenerife, Guimar. On upper and lower surface of dead leaves of *Isoplexis canariensis* (Plantaginaceae). Ade, V 1926 (W no. 10050, holotype; W no. 26924, isotype).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradicans*, with asci cylindrical, ascospores 13-16(-18) × 3-3.5 µm.

Mycosphaerella ixanthi Petr., Bot. Jahrb. Syst. 62, Beibl. 142: 122. 1928.— Fig. 426.

Type — Canary Islands: Tenerife, Anaga-mountains. On bark of *Ixanthus viscosus* (Gentianaceae). Ade, V 1926 (W no. 10251, holotype; W no. 26865, isotype).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradicans*, with asci cylindrical, ascospores 11-14 × 2.5-3.5 µm.

Mycosphaerella ixodiae Hansf., Proc. Linn. Soc. New South Wales 81: 35. 1956.

Type — Australia. On dead stems and upper surface of dead leaves of *Ixodia achilleoides* (Asteraceae).

Anamorph: *Septoria ixodiae* Hansf. *vide* Hansford (op. cit.). The holotype is not in IMI or K; additional material (Australia, Victoria, Longford, VIII 1990, IMI no. 342640) contains only the anamorph.

Mycosphaerella ixorae Bat. & Peres, in Bat., Peres & Poroca, Atas Inst. Micol. 5: 93. 1967.

Type — Jamaica: On yellowish spots with brown margins on upper and lower surface of living leaves of *Ixora* (Rubiaceae). Batista exs. no. 20147 (URM no. 28211, holotype).

The holotype contains several fungi: an *Asteromella*, a *Pestalotiopsis*, a *Phoma* and an *Uredo*.

Mycosphaerella jaapiana (Kirschst.) Tomilin, Opredelitel' gribov roda *Mycosphaerella* Johans.: 228. 1979 = *Sphaerella jaapiana* Kirschst., Kryptog.-Fl. Mark Brandenburg 7(3): 392. 1938.— Fig. 427.

Type — Germany: Triglitz, Ostprignitz. On upper and lower surface of dead leaves of *Rosa alba* (Rosaceae). Jaap, IV 1906 (B, holotype).

This is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores 7-7.5 × 2-2.5 µm.

Mycosphaerella jaczewskii Potebnia, Ann. Mycol. 8: 50. 1910 = *Sphaerella jaczewskii* (Potebnia) Sacc. & Traverso, Syll. Fung. 20: 822. 1911.— Fig. 428.

Type — Russia: Charkow. On upper and lower surface of dead leaves of *Caragana arborescens* (Fabaceae). Potebnia, Tranzchel & Serebrianikow, Mycotheca Rossica no. 271, V 1911 (L, LEP, topotypes).

Anamorphs: *Phyllosticta borszczowii* Thüm. (= *Asteromella borszczowii* (Thüm.) Aa) and *Septoria caraganae* Henn. *vide* Tomilin (1979).

The topotypes show that this belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores 19-23 × 5-6 µm.

Mycosphaerella jaffueli (Speg.) Cash, Syll. Fung. 26: 343. 1972 = *Sphaerella jaffueli* Speg., Bol. Acad. Nac. Ci. 25: 59. 1921.

Type — Chile: *Senecio alnicornis* (Asteraceae). No material was studied as the type was not included in a loan from LPS.

Mycosphaerella janus (Berk. & M.A. Curtis) Petr., Sydowia 11: 340. 1958 = *Sphaeria janus* Berk. & M.A. Curtis, Grevillea 4: 154. 1876 = *Leptosphaeria janus* (Berk. & M.A. Curtis) Sacc., Syll. Fung. 2: 85. 1883 = *Metasphaeria janus* (Berk. & M.A. Curtis) Berl., Icones Fung. 1: 88. 1894 = *Dothidella janus* (Berk. & M.A. Curtis) Höhn., Sitzungsber. Kaiserl. Akad. Wiss., Math.-Naturwiss. Kl., Abt. 1, 124: 67. 1915.— Fig. 429.

Type — USA. On pale spots with brown margins on upper surface of living leaves of *Quercus* (Fagaceae). Curtis (K, holotype).

Spermatial state: *Asteromella castanicola* (Ellis & Everh.) Petr. *vide* Barr (1972).

Additional material studied: USA, California, Siskiyou Co., Mt. Shasta, on *Castanopsis sempervirens* (Fagaceae), Costa, II 1937 (B). Also Mendocino County, east of Van Damme State Park, Bonar, VI 1937 (IMI no. 158270). The type and additional material studied shows that this is indeed a *Dothidella* species, morphologically indistinguishable from *Dothidella castanopsidis* Dearn. and to be accepted as *Dothidella janus* (Berk. & M.A. Curtis) Höhn., with ascoma wall massive, hamathecium consisting of up to 4 µm wide pseudoparaphyses, asci cylindrical, ascospores 20-25 × 3.5-4.5 µm.

Mycosphaerella japonica (Pass.) Tomilin, Novosti Sist. Nizsh. Rast. 6: 121. 1970 ["1969"] = *Sphaerella japonica* Pass., Atti Reale Accad. Lincei, Mem. Cl. Sci. Fis., ser. 4, 3: 6. 1887.— Fig. 434.

Type — Portugal: Coimbra. On upper and lower surface of dead leaves of *Euonymus japonica* (Celastraceae). Moller (PAD, holotype).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradicans*, with asci cylindrical, ascospores 11-13 × 2.5-3 µm. It was previously cited as synonymous with *Davidiella allicina* (as *Mycosphaerella allicina*) by Tomilin (1979).

Mycosphaerella jasminicola T.S. Viswan., in T.S. Viswan. & Tilak, Sydowia 14: 309. 1960 ["1959"].

Type — India: *Jasminum primulinum* (Oleaceae).

No material was studied as the type was not found in LWG or IMI.

Mycosphaerella jasmini-officinalis Siemaszko, Acta Soc. Bot. Pol. 1: 20. 1923.

Type — Georgia: *Jasminum officinale* (Oleaceae).

Anamorph: *Septoria jasmini* Roum. *vide* Siemaszko (op. cit.).

No material was studied as the type was not found in BPI.

Mycosphaerella jenensis (J. Kunze) Lindau, Hilfsb. Sammeln Ascomyceten: 61. 1903 = *Stigmatea jenensis* J. Kunze ex Sacc., Syll. Fung. 1: 543. 1882 = *Stigmatea jenensis* f. *laserpitii-latifolii* J. Kunze, Fungi Selecti Exsiccati no. 239. 1879 = *Sphaerella jenensis* (J. Kunze) G. Winter, in Rabenh., Kryptog.-Fl. Deutschl., Österr. Schweiz 1(2): 367. 1887.— Fig. 430.

Type — Germany: Thüringen, Jena. On upper and lower surface of dead leaves of *Laserpitium latifolium* (Apiaceae). Kunze, Fungi Selecti Exsiccati no. 239, VI 1879 (B, UPS, isotypes) and no. 284 (UPS, isotype).

All isotypes show that this is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores 8-10 × 2-2.5 µm.

Stigmatea jenensis f. *laserpitii-latifolii* J. Kunze. See *Mycosphaerella jenensis* (J. Kunze) Lindau.

Mycosphaerella jenkinsii Tomilin. See *Mycosphaerella arachidis* Deighton.

Mycosphaerella joerstadii Arx, Acta Bot. Neerl. 6: 337. 1957.

Type — Norway: *Rubus chamaemorus* (Rosaceae).

Anamorph: *Septoria rubi* (Duby) Westend. *vide* von Arx (op. cit.).

No material was studied as the type was not found in CBS or in any other herbarium consulted.

Mycosphaerella jonkershoekensis Van Wyk, Marasas & Knox-Davies, J. S. African Bot. 41: 234. 1975.— Fig. 431.

Type — South Africa: *Protea repens* (Proteaceae).

The type was not found in any of the herbaria consulted. Material seen (South Africa, intercepted at Atlanta, on upper surface (not in spots) of living leaves of *Protea* sp., Eldridge, V 1985, BPI) is no *Mycosphaerella*, but probably an *Anthracostroma*, with ascomata superficial, ascospores brown, 11-12 × 5-5.5 µm.

Mycosphaerella juglandis Kessler, Mycologia 76: 363. 1984.— Fig. 432.

Type — USA: Illinois, Alexander. On upper and lower surface of dead leaves of *Juglans nigra* (Juglandaceae). Kessler, V 1979 (NY, isotype).

Anamorph: *Cylindrosporium juglandis* F.A. Wolf *vide* Kessler (op. cit.).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradicans*, with asci cylindrical, ascospores 10-12 × 2.5-3.5 µm. The material is in very bad shape, as it has been totally decayed.

Mycosphaerella juncaginearum (Lasch) J. Schröt., in Cohn, Kryptog.-Fl. Schlesien 3(2): 342. 1894 ["1893"] = *Dothidea juncaginearum* Lasch, in Rabenh., Herb. Vivum Mycol. no. 672. 1844 = *Diaporthe juncaginearum* (Lasch) Rostr., Bot. Tidsskr. 19: 216. 1893 = *Phaeosphaerella juncaginearum* (Lasch) Sacc., Syll. Fung. 11: 312. 1895 = *Leptosphaeria juncaginearum* (Lasch) Munk, Dansk Bot. Ark. 17(1): 377. 1957 = *Venturia juncaginearum* (Lasch) M.E. Barr, Canad. J. Bot. 46: 82. 1968.

Type — Germany: *Triglochin palustre* (Juncaginaceae).

Accepted as *Venturia juncaginearum* (Lasch) M.E. Barr by Sivanesan (1977) and therefore not studied.

Mycosphaerella juncellina Munk, Dansk Bot. Ark. 17(1): 315. 1957.— Fig. 433.

Type — Denmark: Laesø. On dead culms of *Juncus squarrosus* (Juncaceae). Lind, VII 1902 (C, holotype).

Cited as synonymous with *Mycosphaerella najas* by Eriksson (1992). The holotype shows that this is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores 17-21 × 4-6 µm.

Mycosphaerella junciginea (Cooke) Lindau, in Engl. & Prantl, Natürlichen Pflanzenf. 1(1): 426. 1897 [as "*juncigenea*"] = *Sphaerella junciginea* Cooke, Grevillea 19: 3. 1890.

Type — New Zealand: Stewart Island. On dead culms of *Juncus vaginatus* (Juncaceae). Kirk no. 207 (K, holotype; K, isotype).

The holotype contains only coelomycetes, e.g. an *Ascochyta* with hyaline, 1-septate conidia, which could have led to the original description.

Sphaerella juncina (Auersw.) Auersw., in Gonn. & Rabenh., Mycol. Europaea 5-6: 18. 1869 ≡ *Sphaeria juncina* Auersw., in Rabenh., Fungi Europaei Exsiccati, ed. nov., ser. 2 no. 748. 1865 ≡ *Leptosphaeria juncina* (Auersw.) Sacc., Syll. Fung. 2: 66. 1883.— Fig. 435.

Type — Germany: Leipzig, Doemitz. On dead culms of *Juncus effusus* (Juncaceae). Auerswald (B, holotype); also distributed in Rabenhorst, Fungi Europaei Exsiccati, ed. nov., ser. 2 no. 748, VII 1862 (B, isotype); also Winter, VI 1871 (B, topotype).

The types show that this belongs to *Davidiella*, and is morphologically indistinguishable from *D. ammophilae*, with asci pyriform, ascospores 10-11.5 × 2.5-3.5 µm. Additional material studied (Stralsund, Fischer, 1869, distributed in Rabenhorst, Fungi Europaei Exsiccati, ed. nov., ser. 2 no. 1344, CBS) is a species of *Sphaerulina*, with asci pyriform, ascospores 3-septate, 14-28 × 4-5 µm.

Mycosphaerella juniperi (Fautrey & Roum.) Woron., Trudy Tiflissk. Bot. Sada, ser. 2, 3: 137. 1923 ≡ *Sphaerella juniperi* Fautrey & Roum., in Roum., Rev. Mycol. (Toulouse) 13: 166. 1891.— Fig. 436.

Type — France: Côte-d'Or, Noidan. On upper surface of dead needles of *Juniperus communis* (Cupressaceae). Fautrey, distributed in Roumeuguère, Fungi Selecti Exsiccati no. 5833, III 1891 (PC, holotype).

Cited as synonymous with *Mycosphaerella juniperina* by Tomilin (1979). The type shows that this is indeed morphologically indistinguishable from *Mycosphaerella juniperina*, with ascomata often aggregated below the cortex, asci cylindrical, ascospores 8-12 × 2-2.5 µm. Additional material studied (Italy, Treviso, Saccardo, Mycotheca Italica no. 289, VI 1898, L) is a species of *Delphinella*, with ascomata with schizolytic, slit-like ostioles, asci cylindrical, surrounded by copious paraphysoids, ascospores 9-10 × 2.5-3 µm.

Mycosphaerella juniperina (Ellis) Tomilin, Novosti Sist. Nizsh. Rast. 6: 117. 1970 [“1969”] ≡ *Sphaerella juniperina* Ellis, Amer. Naturalist 17: 317. 1883.— Fig. 437.

Type — USA: Iowa, Decorah. On upper and lower surface of dead leaves of *Juniperus communis* (Cupressaceae). Holway, V 1882 (NY, holotype).

Anamorph: presumably *Phoma juniperi* (Desm.) Sacc. *vide* Eriksson (1992).

This is an accepted species in the genus *Mycosphaerella*, with ascomata schizolytic, asci cylindrical, ascospores 10-13 × 3.5-4.5 µm. It represents the earliest epithet for the species now called *Eruptio acicola*, the phylogenetic position of which seems to be inside the *Mycosphaerella* clade.

Mycosphaerella jurineae (Fuckel) Starbäck, Bih. Kongl. Svenska Vetensk.-Akad. Handl. 15(3, 2): 9. 1889 ≡ *Sphaeria jurineae* Fuckel, Fungi Rhenani Exsiccati no. 839. 1863 ≡ *Sphaerella jurineae* (Fuckel) Auersw., in Gonn. & Rabenh., Mycol. Europaea 5-6: 13. 1869.— Fig. 438.

Type — Germany: Budenheim. On upper surface of dead leaves of *Jurinea cynaroides* [“cyanoides”] (Asteraceae). Fuckel, Fungi Rhenani Exsiccati no. 839 (L, isotype).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradians*, with asci cylindrical, ascospores 12-16 × 3.5-4.5 µm. Also present in the type material is a *D. ammophilae*, with pyriform asci, ascospores 2-septate, 11-13 × 3.5-4.5 µm.

Mycosphaerella jutlandica Munk, Dansk Bot. Ark. 17(1): 331. 1957.— Fig. 439.

Type — Denmark: Jutland, Vissing. *Chamaenerion* [“*Epilobium*”] *angustifolium* (Onagraceae). Lind, V 1925 (C, holotype).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradians*, with asci cylindrical, ascospores 15-19 × 3-4 µm.

Mycosphaerella juvenis Crous & M.J. Wingf., Mycologia 88: 453. 1996.

Type — South Africa: Kwazulu-Natal Province, Pietermaritzburg. On leaves of *Eucalyptus nitens* (Myrtaceae). Wingfield, I 1995 (PREM 51910, holotype, not seen).

Anamorph: *Uwebraunia juvenis* Crous & M.J. Wingf. *vide* Crous (1998).

No material was studied of this recently described species. Cited as synonymous with *M. nubilosa* (Cooke) Hansf. by Crous *et al.* (2004).

Mycosphaerella kabocha Hara, A List of Japanese fungi hitherto known: 399. 1954.

Type — Japan: *Cucurbita moschata* (Cucurbitaceae).

No material was studied as the type was not included in a loan from TNS.

Mycosphaerella kaduae F. Stevens & K. Young, in F. Stevens, Bernice P. Bishop Mus. Bull. 19: 103. 1925.— Fig. 440.

Type — Hawaii: On white spots with black margins on upper surface of living leaves of *Gouldia* [as “*Kadua*”] (Rubiaceae).

The type was not found in NY or BPI. Topotype material studied (Oahu, Shear, II 1928, B; BPI) shows that this is a parasitic species, with asci cylindrical, ascospores 17-20 × 4-5 µm.

Mycosphaerella kakomensis Esfandiari, Sydowia 5: 366. 1951.— Fig. 441.

Type — Iran: Fars, Kakom Mountains. On dead stems of Apiaceae. Behkoudi no. 340, VII 1949 (W, holotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores 15-18 × 5-6 µm.

Mycosphaerella kandawanica Petr., Ann. Naturhist. Mus. Wien 50: 435. 1940 [“1939”].— Fig. 442.

Type — Iran: Kandawan, Elburs. On upper and lower surface of dead leaves of *Carex* (Cyperaceae). Reehinger, no. 2407, 1937 (W no. 15987, holotype; W no. 21453, isotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores $16-19 \times 5.5-7.5 \mu\text{m}$.

Mycosphaerella kankeshwarensis C.H. Ramesh, Current Sci. 55: 368. 1986.

Type — India: *Murraya paniculata* (Rutaceae).

No material was studied as the type was not found in IMI or LWG.

Mycosphaerella karajacensis (Allesch.) Tomilin, Opredelitel' gribov roda *Mycosphaerella* Johans.: 178. 1979 = *Sphaerella karajacensis* Allesch., in Allesch. & Henn., Biblioth. Bot. 42: 46. 1897.

Type — Greenland: *Papaver nudicaule* (Papaveraceae).

No material was studied as the type was not found in B. Synonymised with *Davidiella allicina* (as *Mycosphaerella tassiana*) by von Arx (1949).

Mycosphaerella karakulinii Tomilin, Novosti Sist. Nizsh. Rast. 1968: 166. 1968, nomen novum (Article 58) for *Mycosphaerella orobi* Karakulin, in Karakulin & Lobik, Mater. Mikol. Obsl. Rossii 2: 71. 1915, later homonym (illegitimate, Article 53).

Type — Russia: *Lathyrus* ["*Orobus*"] *vernus* (Fabaceae).

Anamorph: *Cylindrosporium orobicolum* (Sacc.) Bubák fide Tomilin (1979).

No material was studied as the type was not included in loans from LE or LEP.

Sphaerella karsteniana Speg., Michelia 1: 229. 1878.— Fig. 443, 966, 967.

Type — Italy: Conegliano. On waste paper. Spegazzini, IX 1877 (PAD, holotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores $12-15 \times 4.5-5.5 \mu\text{m}$. The type collection contains of a piece of paper with still readable letter print, a beautiful illustration of the saprobic life style of the species. Additional species present on the type are *Epicoccum nigrum* Link and *Didymosphaeria oblitescens* (Berk. & Broome) Fuckel, with ascospores brown, ornamented, ca. $15 \times 5 \mu\text{m}$.

Mycosphaerella kawanensis Hara, J. Pl. Protect. 5: 462. 1918.

Type — Japan. On upper and lower surface of dead leaves of *Malus* (Rosaceae). Hara, V 1914 (TNS 209269, holotype).

This is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores $7-9 \times 2-2.5 \mu\text{m}$.

Mycosphaerella keissleri Tomilin, Opredelitel' gribov roda *Mycosphaerella* Johans.: 12. 1979, nomen novum (Article 58) for *Mycosphaerella puerariae* (Keissl.) Petr., Sydowia 1: 362. 1947, later homonym (illegitimate, Article 53) = *Sphaerella puerariae* Keissl., Akad. Wiss. Wien Sitzungsber., Math.-Naturwiss. Kl., Abt. 1, 61(2): 13. 1924.— Fig. 736.

Type — China: Yunnan, Tonkin, Manhao. On white spots with brown margins on lower surface of living leaves of *Pueraria* (Fabaceae). Handel-Mazzetti no. 5755, II 1915 (W, holotype).

This is a parasitic species, with asci cylindrical, ascospores $13-16 \times 3-4 \mu\text{m}$.

Mycosphaerella kenienis Crous & T. Coutinho, in Crous, Mycologia Memoir 21: 74. 1998.

Type — Kenya: On leaves of *Eucalyptus grandis* (Myrtaceae). Coutinho, V 1995 (PREM 54402, holotype, not seen).

No material was studied of this recently described species.

Mycosphaerella kerguelensis (Henn.) Tomilin, Novosti Sist. Nizsh. Rast. 1966: 172. 1966 = *Sphaerella kerguelensis* Henn., Deutsche Südpolar-Exped. 1901-1903, Bot., Pilze 8: 9. 1907 ["1906"].— Fig. 444.

Type — Kerguelen Islands: On dead leaves of *Festuca kerguelensis* (Poaceae).

Cited as synonymous with *M. recutita*, which is morphologically indistinguishable from *Davidiella disseminata*, by Tomilin (1979).

The type has been destroyed in B. Material studied (Kerguelen Islands, Mosley, IMI no. 24198) shows that this belongs to *Davidiella*, and is morphologically indistinguishable from *D. clandestina*, with asci pyriform, ascospores $(17-20-25 \times 4.5-5.5 \mu\text{m})$.

Mycosphaerella khayae Sivan. & R.G. Shivas, Mycol. Res. 106: 358. 2002.

Type — Australia: *Khaya senegalense* (Meliaceae).

No material was studied of this recently described species.

Mycosphaerella killianii Petr., Ann. Mycol. 39: 324. 1941 [often as "*killiani*"], nomen novum (Article 58) for *Sphaeria trifolii* Pers., Synopsis Meth. Fung.: 30. 1801, sanctioned by Fr., Systema Mycol. 2: 435. 1823 = *Dothidea trifolii* (Pers. : Fr.) Fr., Summa Veg. Scand.: 387. 1849 = *Phyllachora trifolii* (Pers. : Fr.) Fuckel, Jahrb. Nassauischen Vereins Naturk. 23-24: 218. 1870 = *Plowrightia trifolii* (Pers. : Fr.) Kill., Rev. Pathol. Vég. Entomol. Agric. France 10: 219. 1923 = *Cymadothea trifolii* (Pers. : Fr.) F.A. Wolf, Mycologia 27: 71. 1935 non *Mycosphaerella trifolii* (P. Karst.) Jacz. (1916) non *Didymella trifolii* (Fuckel) Sacc.

Type — Netherlands: On lower surface of living leaves of *Trifolium pratense* (Fabaceae). (L-Persoon, holotype).

Anamorphs: *Polythrincium trifolii* G. Kunze and *Placosphaeria trifolii* (Pers.) Traverso fide Sivanesan (1984).

Accepted as *Cymadothea trifolii* (Pers. : Fr.) F.A. Wolf. The holotype is immature, but agrees well with the description by Barr (1972), with the ascomata aggregated in thick, black stromata.

Mycosphaerella kirschsteinii Tomilin, Opredelitel' gribov roda *Mycosphaerella* Johans.: 154. 1979, nomen novum (Article 58) for *Sphaerella pascuorum* Kirschst., Kryptog.-Fl. Mark Brandenburg 7(3): 376. 1938, later homonym (illegitimate, Article 53).

Type — Germany: *Armeria vulgaris* (Plumbaginaceae).

No material was studied as the type was not found in B.

Mycosphaerella koae Petr., Sydowia 7: 390. 1953.— Fig. 445.

Type — Hawaii: N. Kona. On dead fruits (pods) of *Acacia koa* (Fabaceae). Shear, I 1928 (W no. 11839, holotype; W, isotype).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradicans*, with asci cylindrical, ascospores 13-15 × 3-4 µm.

Mycosphaerella kochiae Tomilin, Novosti Sist. Nizsh. Rast. 1966: 146. 1966.

Type — Russia: *Kochia prostrata* (Amaranthaceae).

No material was studied as the type was not included in loans from LE or LEP. Cited as synonymous with *Davidiella allicina* (as *Mycosphaerella allicina*) by Tomilin (1979).

Mycosphaerella koldingensis Munk, Dansk Bot. Ark. 14(8): 1. 1952.— Fig. 446.

Type — Denmark: Kolding Skov. On dead stems of *Geum rivale* (Rosaceae). Larsen, IV 1926 (C, holotype).

Cited as synonymous with *Mycosphaerella ushuvaiensis* by Tomilin (1979). The type shows that this is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores 8-10 × 2-2.5 µm.

Mycosphaerella konae Crous, J.E. Taylor & M.E. Palm, in J.E. Taylor, Crous & M.E. Palm, Mycotaxon 78: 459. 2001.

Type — Hawaii: *Leucadendron* (Proteaceae).

Anamorph: *Pseudocercospora fide* Taylor, Crous & Palm (op. cit.).

No material was studied of this recently described species.

Sphaerella kriegeri Kirschst., Kryptog.-Fl. Mark Brandenburg 7(3): 342. 1938.

Type — Germany: *Mercurialis perennis* (Euphorbiaceae).

No material was studied as the type was not found in B and might be lost. Cited as synonymous with *Mycosphaerella leptasca* by Tomilin (1979).

Mycosphaerella krigiae (Ellis & Everh.) H.C. Greene, Trans. Wisconsin Acad. Sci. 35: 128. 1944 ≡ *Sphaerella krigiae* Ellis & Everh., N. Amer. Pyrenomyc.: 280. 1892.— Fig. 447.

Type — USA: Wisconsin, Racine. On brown spots on upper and lower surface of living leaves of *Krigia amplexicaulis* (Asteraceae). Davis no. 9019, VI 1890 (NY, holotype).

Anamorph: *Septoria krigiae* Ellis & Everh. occurs on the same spots *fide* Ellis & Everhart (op. cit.).

This is a parasitic species, with asci cylindrical, ascospores 8-12 × 3-3.5 µm. The anamorph is also present in the holotype.

Mycosphaerella laburni (Pass.) Lindau, in Engl. & Prantl, Natürlichen Pflanzenf. 1(1): 424. 1897 ≡ *Sphaerella laburni* Pass., Erbario Crittogamico Italiano, ser. 2 no. 1069. 1881.— Fig. 448, 964.

Type — Italy: Parma, Botanical Garden. On dead marginal zones on lower surface of living leaves of *Laburnum*

anagyroides (Fabaceae). Passerini, Erbario Crittogamico Italiano, ser. 2, no. 1069 (B, isotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores 16-18 × 4.5-5.5 µm.

Sphaerella laburni var. *eribotryae* Scalia, Atti Accad. Gioenia Sci. Nat. Catania, ser. 4, 14(a): 18. 1901.

Type — Italy: *Eriobotrya japonica* (Rosaceae).

No material was studied as the type was not found in any of the herbaria consulted.

Mycosphaerella lachesis (Sacc.) Tomilin, Novosti Sist. Nizsh. Rast. 1968: 166. 1968 ≡ *Sphaerella lachesis* Sacc., Nuovo Giorn. Bot. Ital. 7: 302. 1875.— Fig. 449, 987.

Type — Italy: Conegliano. On upper and lower surface of dead leaves of *Helleborus viridis* (Ranunculaceae). Spegazzini, III 1878, distributed in Decades Mycologiae Italicae no. 11 (BPI, topotype).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradicans*, with ascospores large, asci cylindrical, ascospores 9-11 × 2-3 µm. Additional material studied (France, Roumeguère, Fungi Gallici Exsiccati no. 3439, 1884, L) is *M. punctiformis*, with asci cylindrical, ascospores 8-10 × 2-2.5 µm.

Mycosphaerella lachmannii M. Morelet, Ann. Soc. Sci. Nat. Archéol. Toulon & Var 17: 105. 1965.

Type — France: *Nigella damascena* (Ranunculaceae).

No material was studied as the type was not included in a loan from PC.

Mycosphaerella lactucae (Ellis & Kellerm.) Tomilin, Opredelitel' gribov roda *Mycosphaerella* Johans.: 63. 1979 ≡ *Sphaerella lactucae* Ellis & Kellerm., Bull. Torrey Bot. Club 11: 123. 1884.— Fig. 450.

Type — USA: Kansas, Manhattan. On grey spots with reddish brown margins on upper and lower surface of living leaves of *Lactuca canadensis* (Asteraceae). Kellerman no. 619, VIII 1884 (NY, holotype).

This belongs to *Davidiella*, of which it represents a parasitic species, with asci pyriform, ascospores 13-15 × 4.5-5.5 µm. Therefore the following new combination is made: **Davidiella lactucae** (Ellis & Kellerm.) Aptroot comb. nov., **MB 500353**. **Basionym:** *Sphaerella lactucae* Ellis & Kellerm., Bull. Torrey Bot. Club 11: 123. 1884. A coelomycete is also present in the holotype.

Mycosphaerella lageniformis Rehm, Pomona Coll. J. Econ. Bot. 1: 196. 1911 ≡ *Sphaerella lageniformis* (Rehm) Trotter, Syll. Fung. 24: 884. 1928.— Fig. 451.

Type — USA: Carona. On upper and lower surface of dead leaves and dead parts of living leaves of *Citrus aurantium* (Rutaceae). Metz, I 1911 (S, holotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores 18-23 × 6-8 µm.

Sphaerella lagunensis Syd. & P. Syd., Ann. Mycol. 15: 206. 1917.— Fig. 452.

Type — Philippines: Luzon, Prov. Laguna, Los Baños. On upper and lower surface of dead leaves of *Dendrochilum* (Orchidaceae). Baker no. 4321, VI 1916 (S, holotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores 16-19 × 4-5 µm.

Sphaerella lamprocarpi Pass., Erbario Crittogamico Italiano, ser. 2 no. 392. 1871 ≡ *Leptosphaeria lamprocarpi* (Pass.) Sacc., Syll. Fung. 3: 66. 1883.— Fig. 453.

Type — Italy: Parma. On dead inflorescences of *Juncus articulatus* ["*lamprocarpus*"] (Juncaceae). Passerini, 1871, distributed in Rabenhorst, Fungi Europaei Exsiccati, ed. nov., ser. 2 no. 2442 (CBS, isotype).

Anamorph: *Coniothyrium scirpi* Trail *fide* Sivanesan (1984).

Already synonymised by Eriksson (1967) with *Paraphaeosphaeria michotii* (Westend.) O.E. Erikss., with which the type agrees well, with ascospores 1-2-septate, ornamented, 24-30 × 5-6 µm.

Mycosphaerella lantanae (Nitschke) Migula, in Thomé, Fl. Deutschl. Österr. Schweiz. X, 1. Kryptog.-Fl. III, 3(1): 290. 1912 ["1913"] ≡ *Sphaerella lantanae* Nitschke ex Auersw., in Gonn. & Rabenh., Mycol. Europaea 5-6: 6. 1869 ≡ *Sphaeria lantanae* Nitschke, in Fuckel, Fungi Rhenani Exsiccati no. 1786. 1866, nomen nudum (not validly published, Article 32) ≡ *Sphaerella viburni* var. *lantanae* (Nitschke) Cif., Ann. Mycol. 20: 39. 1922.— Fig. 454.

Type — Germany: Östlich. On lower surface of dead leaves of *Viburnum lantana* (Adoxaceae) Fuckel, Fungi Rhenani Exsiccati no. 1786 (BPI, L, isotypes).

Isotype materials studied are immature but show that this is morphologically indistinguishable from *M. punctiformis*, with tiny cylindrical asci. Additional material studied (Thüringen, Arnstadt, Sydow, Mycotheca Germanica no. 483, V 1906, L) is mature *M. punctiformis*, with asci cylindrical, ascospores 8-10 × 2-2.5 µm.

Sphaerella lapageriae Speg., Revista Fac. Agron. Univ. Nac. La Plata, ser. 2, 6: 54. 1910.

Type — Chile: *Lapageria rosea* (Asparagaceae).

No material was studied as the type was not included in a loan from LPS.

Mycosphaerella lapathi (Laib.) Petr., Sydowia 1: 196. 1947 ≡ *Ovosphaerella lapathi* Laib., Centralbl. Bakteriell., 1. Abt., Originale 55: 293. 1922.

Type — Germany: *Rumex* (Polygonaceae).

Anamorph: *Ovularia obovata* (Fuckel) Sacc. ≡ *Ovularia obliqua* Cooke *fide* von Arx (1983) or *Ramularia rubella* (Bonord.) Nannfeldt *fide* Jørstad (1963).

No material was studied as the type was probably not preserved.

Sphaerella lapponum De Not., Comment. Soc. Crittog. Ital. 2(3): 456. 1867 ≡ *Didymella lapponum* (De Not.) Sacc., Syll. Fung. 1: 548. 1882.— Fig. 455.

Type — Italy: Riva. On corticate branches of *Salix lapponum* (Salicaceae). Carestia no. 13, VII 1859 (RO, holotype).

This is morphologically indistinguishable from *Arthopyrenia fraxini* A. Massal., with ascomata KOH-negative, pseudoparaphyses 1.5 µm wide, asci clavate, ascospores 15-20 × 4.5-6 µm, surrounded by a gelatinous sheath.

Sphaerella lardizabala Speg., Revista Fac. Agron. Univ. Nac. La Plata, ser. 2, 6: 55. 1910.— Fig. 456.

Type — Chile: Concepción, Cerro Caracol. On upper and lower surface of dead leaves of *Lardizabala biternata* (Lardizabalaceae). Spegazzini no. 6171, I 1909 (LPS, holotype).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradians*, with asci cylindrical, ascospores 10-12 × 2.5-3 µm.

Mycosphaerella laricina (Hartig) Migula, in Thomé, Fl. Deutschl. Österr. Schweiz. X, 1. Kryptog.-Fl. III, 3(1): 301. 1912 ["1913"] ≡ *Sphaerella laricina* Hartig, Forstl.-Naturwiss. Z. 4: 445. 1895.

Type — Germany: *Larix europaea* (Pinaceae).

Anamorphs: *Cercoseptoria* (= *Pseudocercospora*) *fide* Farr *et al.* (1989) and *Leptostroma* *fide* Tomilin (1979).

No material was studied as the type was not found in any of the herbaria consulted.

Mycosphaerella laricis-leptolepidis K. Itô, Satô & Ota, Bull. Gov. Forest Exp. Sta. 96: 84. 1957 [as "*laricis-leptolepis*"].

Type — Japan: *Larix leptolepis* (Pinaceae).

Anamorphs: *Leptostroma* *fide* Saccardo & P. Sydow (1899) and *Phyllosticta laricis* Sawada *fide* Itô, Satô & Ota (op. cit.).

No material was studied as the type was not included in a loan from TNS.

Mycosphaerella larsenii Munk, Dansk Bot. Ark. 14(8): 3. 1952.— Fig. 457.

Type — Denmark: Kolding Skov. On dead stems of *Geum urbanum* (Rosaceae). Larsen, V 1935 (C, holotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. ammophilae*, with asci pyriform, ascospores 8-10 × 2-2.5 µm.

Mycosphaerella lasiana (Sacc.) Aptroot comb. nov., **MB 500505**. **Basionym:** *Sphaerella lasiana* Sacc., Bull. Orto Bot. Regia Univ. Napoli 6: 44. 1921.— Fig. 458.

Type — Singapore. On pale spots with brown margins on upper and lower surface of living leaves of *Lasia heterophylla* (Araceae). Baker, X 1917, distributed in Fungi Malayana no. 489 (BPI, 2 isotypes).

This is a parasitic species, with asci cylindrical, ascospores 15.5-19.5 × 3.5-4 µm.

Mycosphaerella latebrosa (Cooke) J. Schröt., in Cohn, Kryptog.-Fl. Schlesien 3(2): 334. 1894 ["1893"] ≡ *Sphaerella latebrosa* Cooke, J. Bot. 4: 248. 1866 ≡ *Carlia latebrosa* (Cooke) Höhn., Hedwigia 62: 73. 1920.— Fig. 459.

Type — United Kingdom: Surrey, Shere. On upper and lower surface of dead leaves of *Acer pseudoplatanus* (Sapindaceae). Cooke (K, holotype).

Anamorphs: *Cylindrosporium platanoides* (Allesch.) Diedicke, *Phleospora aceris* (Lib.) Sacc. and *Phyllosticta platanoides* Sacc. *vide* Tomilin (1979).

The type and additional material studied (Germany, Westfalen, Siegen, Müsen, Ludwig, distributed in Sydow, Mycotheca Germanica no. 2331, V 1924, L, also Netherlands, Apeldoorn, Oudemans, 1897, L) belong to section *Longispora*, with asci cylindrical, ascospores $17-23 \times 2-3 \mu\text{m}$.

Mycosphaerella lateralis Crous & M.J. Wingf., Mycologia 88: 454. 1996.

Type — South Africa: Northern Province, Tzaneen. On leaves of *Eucalyptus grandis* \times *saligna* (Myrtaceae). Kemp, X 1994 (PREM 51926, holotype, not seen).

Anamorph: *Uwebraunia lateralis* Crous & M.J. Wingf. *vide* Crous (1998) (= *Dissoconium dekkeri* de Hoog & Hijwegen).

No material was studied of this recently described species.

Mycosphaerella lathyri Potebnia, K istorii razvitiya nekotorykh askomitsetov. 1. *Mycosphaerella*. 2. *Gnomonia*, *Glomerella* i *Pseudopeziza*: 80. 1908 = *Sphaerella lathyri* (Potebnia) Sacc. & Traverso. Syll. Fung. 20: 823. 1911.— Fig. 460.

Type — Russia. On upper and lower surface of dead, but still green, leaves of *Lathyrus pisiformis* (Fabaceae). Potebnia (PAD, isotype).

Anamorphs: Probably *Phleospora caraganae* var. *lathyri* Potebnia and *Phyllosticta minussinensis* Thüm. *vide* Saccardo & Traverso (op. cit.).

The type and additional material studied (Czech Republic, Weißkirchen, Hrabuvka, Petrak, IV 1916, distributed in Flora Bohemiae et Moraviae exsiccata no. 1626, L) belong to *Davidiella*, and show that this is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores $14-17 \times 4-5.5 \mu\text{m}$.

Sphaerella lathyrina (Berk. & M.A. Curtis) Cooke, J. Bot. 21: 136. 1883 = *Sphaeria lathyrina* Berk. & M.A. Curtis, in Berk., Grevillea 4: 152. 1876 = *Didymella lathyrina* (Berk. & M.A. Curtis) Sacc., Syll. Fung. 1: 555. 1882.— Fig. 461.

Type — USA: Pennsylvania. On dead stems of *Lathyrus latifolia* (Fabaceae). Ellis no. 173 (K, holotype).

The holotype contains a *Didymella* with hamathelial filaments, ascospores $18-22 \times 4-6$, and an *Ascochyta* anamorph. It could be accepted as *Didymella lathyrina* (Berk. & M.A. Curtis) Sacc.

Mycosphaerella laureolae (Chevallier) Feltgen, Recueil Mém. Trav. Soc. Bot. Grand-Duché Luxembourg 1901: 181. 1901 = *Asteroma laureolae* Chevallier, Flore de Paris 1: 448 = *Sphaerella laureolae* (Chevallier) Auersw., in Gonn. & Rabenh., Mycol. Europaea 5-6: 9. 1869.— Fig. 462.

Type — Germany: On spots on upper and lower surface of living and dead leaves of *Daphne laureola* (Thymelaeaceae).

Anamorph: *Phyllosticta laureolae* Desm. *vide* Auerswald (op. cit.).

The type was not included in a loan from PC. Material studied (Switzerland, Aarburg, von Arx, distributed in Reliquiae Petrakianae no. 1251, V 1948, L) is a parasitic species, with asci cylindrical, ascospores $12-15 \times 3-3.5 \mu\text{m}$.

Mycosphaerella lavrovii Naumov, nomen herbariorum (not validly published, Article 32).— Fig. 463.

Authentic material — Russia: On dead stems of *Saussurea frolovii* (Asteraceae). Lavrov (LE 34991).

This is a species of *Phaeosphaeria*, probably *Phaeosphaeria donacina* (Sacc.) Shoemaker & C.E. Babc., with ascospores brown, 3-septate, $23-25 \times 6-7 \mu\text{m}$.

Mycosphaerella lebedevae Tomilin. See *Mycosphaerella phlomidis* Lebedeva.

Sphaerella leersiae Pass., in Rabenh., Fungi Europaei Exsiccati, ed. nov., ser. 2 no. 2342. 1876 = *Metasphaeria leersiae* (Pass.) Sacc., Syll. Fung. 2: 173. 1883.— Fig. 464.

Type — Italy: Parma. On upper and lower surface of dead leaves of *Leersia oryzoides* (Poaceae). Passerini, 1876, distributed in Thümen, Mycotheca Universalis no. 965 (L, NY, isotypes), also distributed in Rabenhorst, Fungi Europaei Exsiccati no. 2342 (L, isotype).

This is morphologically indistinguishable from *Glomerella cingulata* (Stoneman) Spauld. & H. Schrenk, with asci clavate, surrounded by paraphyses, ascospores not septate, $19-21 \times 3.5-4 \mu\text{m}$, for which it would be an older name. The name *Glomerella cingulata* has been widely used for one of the most common ascomycetes, which, for instance, is present with the highest number of specimens in the IMI herbarium.

Mycosphaerella leguminis-cytisi (Desm.) Lindau, in Engl. & Prantl, Natürlichem Pflanzenf. 1(1): 425. 1897 = *Sphaeria leguminis-cytisi* Desm., Ann. Sci. Nat. Bot., ser. 2, 19: 358. 1843 = *Sphaerella leguminis-cytisi* (Desm.) Ces. & De Not., Comment. Soc. Crittog. Ital. 1(4): 237. 1863.— Fig. 465.

Type — France. On dead fruits (pods) of *Laburnum* (Fabaceae). Desmazières, Fungi Exsiccati no. 792 (L, isotype). The isotype and additional material studied (Netherlands, Groningen, van Hall, IV 1844, L) contain only the *Ascochyta* anamorph, with conidia 1-septate, $11-13 \times 3.5-4 \mu\text{m}$, of which the dimensions agree with the reported ascospores. Therefore, the name may have been based on the *Ascochyta*.

Mycosphaerella leguminosarum Upadhyay, Publ. Inst. Micol. Univ. Fed. Pernambuco 411: 8. 1964.— Fig. 466.

Type — Brazil: On white spots with black margins on lower surface of living leaves of *Drepanocarpus paludicola* (Fabaceae). Batista exs. no. 11946 (URM 38683, holotype).

Spermatial state and anamorph: *Asteroma* and *Ramularia* *vide* Tomilin (1979).

This belongs to *Davidiella*, of which it represents a parasitic species, and is morphologically indistinguishable from *D. nemorosa* (see below), with asci pyriform, ascospores $14-18 \times 3-3.5 \mu\text{m}$.

Sphaerella leightonii (Berk. & Broome) Cooke, J. Bot. 4: 250. 1866 [as “*leightoni*”] = *Sphaeria leightonii* Berk. & Broome, Ann. Mag. Nat. Hist., ser. 2, 9: 287. 1852 [as “*leightoni*”] = *Sphaerulina leightonii* (Berk. & Broome) Sacc., Syll. Fung. 2: 188. 1883 [as “*leightoni*”].— Fig. 467.

Type — United Kingdom: Glen Dale. On upper surface of dead leaves of *Linnaea borealis* (Caprifoliaceae). Clover, VIII 1837 (K, holotype; K, 3 isotypes).

This is an older synonym of *Gnomonia linnaeae* Auersw. (1889), with large immersed ascomata with long ostioles, ascospores $17-20 \times 4.5-5.5 \mu\text{m}$. Therefore the following combination is proposed: ***Gnomonia leightonii*** (Berk. & Broome) Aptroot comb. nov., **MB 500368**. **Basionym:** *Sphaeria leightonii* Berk. & Broome, Ann. Mag. Nat. Hist., ser. 2, 9: 287. 1852 (British Fungi no. 659) [as “*leightoni*”].

Mycosphaerella lenticula (Cooke) Tomilin, Opređelitel' gribov roda *Mycosphaerella* Johans.: 226. 1979 \equiv *Sphaerella lenticula* Cooke, J. Bot. 21: 108. 1883.— Fig. 468.

Type — USA: South Carolina, Aiken. On dark spots on lower surface of leaves of *Prunus* [“*Cerasus*”] *caroliniana* (Rosaceae). Ravenel, VIII 1881 (K, holotype; K, 9 isotypes).

This is a species of *Cymadothea*, with ascomata in dense groups in stromata, ascospores $11-13 \times 3.5-4.5 \mu\text{m}$.

Sphaerella lepidiotae Anzi, Atti Soc. Ital. Sci. Nat. 11: 27. 1868 \equiv *Metasphaeria lepidiotae* (Anzi) Sacc., Syll. Fung. 2: 184. 1883 \equiv *Sphaerulina lepidiotae* (Anzi) Vainio, Természettud. Füzet. 22: 342, 1899 \equiv *Sphaeria lepidiotae* (Anzi) H. Olivier, Bull. Acad. Int. Géogr. Bot. 17: 171. 1907.— Fig. 469.

Type — Italy: Bormio. On thallus of *Parmeliella praetermissa* [“*Pannaria lepidiota*”] on rocks (Ascomycota, Pannariaceae). Anzi, 1865, distributed in Lichenes Rariores Longobardi Exsiccati no. 440 (B, BPI, isotypes).

Accepted as *Sphaerulina lepidiotae* (Anzi) Vainio by Clauzade, Diederich & Roux (1989). However, this species is not closely related to the type of the genus *Sphaerulina*, viz. *S. myriadea*, as it has large ascomata with desintegrated hamathecium filaments, asci unitunicate, old, ascospores 8 per ascus, hyaline, dextrinoid, only 1-septate, $27-30 \times 5-6.5 \mu\text{m}$, without gelatinous sheath, upper cell shorter and broader than lower cell. This all suggests that it belongs to the Verrucariales, possibly to the currently unnaturally delimited genus *Thelidium*.

Mycosphaerella lepidospermatis Hansf., Proc. Linn. Soc. New South Wales 82: 218. 1957.— Fig. 470.

Type — Australia: Meningie. On *Lepidosperma gladiatum* (Cyperaceae). Williams, XI 1957 (IMI no. 74255, isotype slide).

Anamorph: possibly *Septoria lepidospermatis* Cooke & Massee fide Hansford (op. cit.).

The identity of the fungus is difficult to judge from the slide only but it is not a *Mycosphaerella*. The asci are pyriform, the ascospores contain 4 oil globules and possibly sometimes a septum, and measure $9-11 \times 2-2.5 \mu\text{m}$.

Mycosphaerella leptasca (Auersw.) J. Schröt., in Cohn, Kryptog.-Fl. Schlesien 3(2): 337. 1894 [“1893”] \equiv *Sphaerella leptasca* Auersw., in Gonn. & Rabenh., Mycol. Europaea 5-6: 15. 1869 [as “*leptoascus*”].— Fig. 471.

Type — Germany: Apiaceae.

Anamorph: *Septoria leptidea* Sacc. fide Saccardo (1882).

The type was not found in B and is possibly lost. Material studied (Netherlands, Lochem, on stems of *Angelica*, van Luyk, III 1914, L) belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradicans*, with asci cylindrical, ascospores $11-13 \times 3-3.5 \mu\text{m}$.

Sphaerella leptidea (Fr. : Fr.) Fuckel, Jahrb. Nassauischen Vereins Naturk. 23-24: 100. 1870 \equiv *Sphaeria leptidea* Fr., Observ. Mycol. Suecicam 2: 333. 1818, sanctioned by Fr., Systema Mycol. 2: 522. 1823 \equiv *Myxothyrium leptideum* (Fr. : Fr.) Bubák & Kabát, in Bubák, Svensk Bot. Tidskr. 9: 379. 1915.

Type — Sweden: On lower surface of dead leaves of *Vaccinium vitis-idaea* (Ericaceae). (L, isotype).

Anamorph: *Myxothyrium leptideum* (Fr. : Fr.) Bubák & Kabát fide Bubák & Kabát (op. cit.).

Accepted as the coelomycete *Myxothyrium leptideum* (Fr. : Fr.) Bubák & Kabát by Eriksson (1992), with which the isotype studied agrees, with conidia $3.5-4.5 \times 1-2 \mu\text{m}$.

Mycosphaerella leptopleura (De Not.) Earle, in Mohr, Contr. U.S. Natl. Herb. 6: 174. 1901 \equiv *Sphaerella leptopleura* De Not., Comment. Soc. Crittog. Ital. 2(3): 488. 1867.— Fig. 472.

Type — Italy: Poregio, Val Intrasin. On dead culms of *Secale cereale* (Poaceae). De Notaris, X 1862 (RO, holotype; B, isotype).

The holotype contains a immature specimen of a *Phaeosphaeria*, with badly preserved hamathecium, ascospores 1-3-septate, $15-19 \times 4-5 \mu\text{m}$. The isotype contains a badly preserved *Didymella*.

Sphaerella leptosperma Speg., Revista Fac. Agron. Univ. Nac. La Plata, ser. 2, 6: 55. 1910.— Fig. 473.

Type — Chile: Concepción, Cerro Caracol. On upper surface of dead leaves of *Proustia pirifolia* (Asteraceae). Spegazzini no. 6170, I 1909 (LPS, holotype).

This belongs to section *Longispora*, and is morphologically indistinguishable from *M. millegrana*, with asci cylindrical, ascospores $16-19 \times 3-4 \mu\text{m}$.

Sphaerella leptosphaerioides Auersw., in Gonn. & Rabenh., Mycol. Europaea 5-6: 13. 1869 \equiv *Leptosphaeria auerswaldii* Sacc. & Traverso, Syll. Fung. 19: 1096. 1910, superfluous (illegitimate, Article 58) nomen novum (Article 58).

Type — Germany: *Arenaria ciliata* (Caryophyllaceae).

No material was studied as the type was not found in B and is possibly lost.

Mycosphaerella leptospora (Sacc. & Scalia) Tomilin, Novosti Sist. Nizsh. Rast. 1967: 189. 1967 \equiv *Sphaerella leptospora* Sacc. & Scalia, in Sacc., Peck & Trelease, The Fungi of Alaska. Harriman Alaska Expedition 5: 31. 1904.

Type — USA, Alaska, Yes Bay. On dead leaves of *Carex mertensiana* (Cyperaceae). Howell, VII 1895 (PAD, holotype). The type contains no *Mycosphaerella*, only an immature ascomycete with large ascomata.

Mycosphaerella lethalis R. Stone, Ann. Mycol. 10: 587. 1912 \equiv *Sphaerella lethalis* (R. Stone) Trotter, Syll. Fung. 24: 873. 1928 \equiv *Didymella lethalis* (R. Stone) Sivan., Bitunicate Ascomycetes and their anamorphs: 424. 1984.

Type — USA: New York, Ithaca. On dead stems of *Melilotus alba* (Fabaceae). Chupp no. 7878, VI 1915 (BPI, topotype).

Anamorph: *Ascochyta lethalis* Ellis & Barthol. fide Sivanesan (op. cit.).

Accepted as *Didymella lethalis* (R. Stone) Sivan. by Sivanesan (op. cit.) but cited as synonymous with *Didymella pinodes*

(Berk. & A. Bloxam) Petr. by Corbaz (1956). In the topotype, only the *Ascochyta* anamorph was found.

Mycosphaerella leucophaea (Ellis & Kellerm.) Tomilin, Novosti Sist. Niz Rast. 6: 121. 1970 ["1969"] ≡ *Sphaerella leucophaea* Ellis & Kellerm., J. Mycol. 2: 3. 1886.— Fig. 474. Type — USA: Kansas, Manhattan. On upper and lower surface of dead leaves of *Baptisia leucophaea* (Fabaceae). Kellerman no. 752, VI 1885 (NY, holotype), also distributed in Ellis & Everhart, North American Fungi no. 1675 (L, NY (2×), isotypes).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. superflua*, with asci cylindrical, ascospores 17-20 × 3.5-4.5 μm.

Mycosphaerella leucospila Syd., Leaf. Philipp. Bot. 9: 3121. 1925.— Fig. 475.

Type — Philippines: Luzon, Prov. Sorsogon, Irosin. On spots on upper surface of living leaves of *Ficus celebica* (Moraceae). Elmer, Philippine Islands Plants no. 15025, VIII 1916 (L, isotype).

This is a parasitic species, with asci cylindrical, ascospores 13-17 × 3-3.5 μm.

Mycosphaerella leucothoës Miles, Mycologia 18: 164. 1926.

Type — USA: Missouri, Poplarville. On pale spots with brown margins on the upper surface of living leaves of *Leucothoë axillaris* (Ericaceae). Miles no. 447, IV 1921 (NY, isotype).

This is morphologically indistinguishable from the heterotypic *Pseudomassaria leucothoës* (Cooke) Petr. & Syd., with asci clavate, ascospores asymmetrically septate.

Sphaerella leucothoës Cooke, J. Bot. 21: 70. 1883 ≡ *Laestadia leucothoës* (Cooke) Sacc., Syll. Fung. 9: 581. 1891 ≡ *Pseudomassaria leucothoës* (Cooke) Petr. & Syd., Ann. Mycol. 22: 385. 1924.— Fig. 476.

Type — USA: South Carolina, Pinopolis. On white spots with brown margins mainly occupying the dead tips of the upper surface of living leaves of *Leucothoë* (Ericaceae). Ravenel, Fungi Americani Exsiccati no. 687 (NY, isotype).

Accepted as *Pseudomassaria leucothoës* (Cooke) Petr. & Syd. by Müller & von Arx (1962), with which the additional material studied (South Carolina, Seaboard, Ravenel, distributed in Ellis, North American Fungi no. 1348, VI 1884, L) agrees well, with asci clavate, ascospores asymmetrically septate, 11-13 × 5-6 μm. In the isotype only the coelomycetes *Pestalotiopsis* and *Septoria* were found.

Sphaerella leucothoës var. *terminalis* Ellis, nomen herbariorum (not validly published, Article 32).

Authentic material — USA: South Carolina, Lew Co. On white spots mainly occupying the dead tips of the upper surface of living leaves of *Leucothoë* (Ericaceae). Ravenel no. 4072, 1884 (NY).

As the nominal variety, this is *Pseudomassaria leucothoës* (Cooke) Petr. & Syd., with asci clavate, ascospores asymmetrically septate, 17-20 × 5-6.5 μm.

Mycosphaerella liabi Petr., Sydowia 2: 343. 1948.— Fig. 477.

Type — Ecuador: Prov. Tungurahua, Baños. On upper and lower surface of dead leaf margins of *Liabum pellarangense*

(Asteraceae). Sydow, XII 1937, distributed in Reliquiae Petrakianae no. 655 (H, L, isotypes).

This belongs to *Davidiella*, with asci pyriform, ascospores 30-35 × 8-11 μm. Therefore the following new combination is made: **Davidiella liabi** (Petr.) Aptroot comb. nov., **MB 500354**. **Basionym:** *Mycosphaerella liabi* Petr., Sydowia 2: 343. 1948.

Mycosphaerella libanotidis (Fuckel) Lind, Danish Fungi: 203. 1913 ≡ *Sphaerella eryngii* var. *libanotidis* Fuckel, Jahrb. Nassauischen Vereins Naturk. 23-24: 105. 1870 [as "*libanotis*"].

Type — Switzerland: *Libanotis montana* (Apiaceae).

No material was studied as the type was not included in a loan from G.

Mycosphaerella ligea (Sacc.) Zeller, Phytopathology 27: 1004. 1937 ≡ *Sphaerella ligea* Sacc., Nuovo Giorn. Bot. Ital. 7: 300. 1875.— Fig. 478.

Type — Italy. On brown spots on upper and lower surface of living leaves of *Rubus fruticosus* (Rosaceae). Saccardo (PAD, holotype; PAD, isotype).

Anamorph: *Septoria rubi* (Duby) Westend. *vide* Saccardo (1882).

This is a parasitic species, with asci cylindrical, ascospores 15-17 × 2.5-3 μm.

Mycosphaerella ligea f. *rubi-caesii* Sacc., Mycotheca Veneta no. 692. 1876.— Fig. 479.

Type — Italy: Treviso, Montello. On brown spots on upper and lower surface of living leaves of *Rubus caesius* (Rosaceae). Saccardo, IX 1875, distributed in Mycotheca Veneta no. 692 (BPI, isotype).

This is a parasitic species, with asci cylindrical, ascospores 12.5-15 × 3.5-4.5 μm.

Mycosphaerella lignicola (Munk) Munk, Dansk Bot. Ark. 17(1): 319. 1957 ≡ *Sphaerella lignicola* Munk, Dansk Bot. Ark. 4(12): 15. 1948 ≡ *Chaetosphaeria lignicola* (Munk) Tomilin, Novosti Sist. Nizsh. Rast. 9: 120. 1972.— Fig. 480.

Type — Denmark: Jylland, Mols. On soft, dead wood of *Quercus* (Fagaceae). Munk, IV 1943 (C, holotype).

The holotype contains two fungi: The one that belongs to *Davidiella* and is morphologically indistinguishable from *D. ammophilae*, with asci pyriform, ascospores 8-10 × 2-2.5 μm should be regarded as the holotype, as it is in accordance with the protologue. The other species is a *Chaetosphaeria*, with a firm ostiolar beak, ascospores 15-19 × 4-5 μm, which has inspired Tomilin to make his new combination.

Mycosphaerella ligulicola K.F. Baker, Dimock & Davis, Phytopathology 39: 799. 1949 ≡ *Didymella ligulicola* (K.F. Baker, Dimock & Davis) Arx, in E. Müll. & Arx, Beitr. Kryptogamenfl. Schweiz 11(2): 364. 1962.— Fig. 481.

Type — USA: North Carolina, Pittsboro. On dead stems of *Chrysanthemum morifolium* (Asteraceae). Baker, VI 1948 (BPI, isotype).

Anamorph: *Phoma ligulicola* Boerema *vide* Boerema et al. (2004); *Ascochyta chrysanthemi* F. Stevens *vide* Sivanesan (1984).

Accepted as *Didymella ligulicola* (K.F. Baker, Dimock & Davis) Arx by Eriksson (1992), with which the type agrees well, with asci clavate, surrounded by copious pseudoparaphyses, ascospores immature, ca. $12 \times 5 \mu\text{m}$.

Mycosphaerella ligustri (Roberge) Lindau, in Engl. & Prantl, *Natürlichen Pflanzenf.* 1(1): 424. 1897 \equiv *Sphaeria ligustri* Roberge, in Desm., *Ann. Sci. Nat. Bot.*, ser. 2, 19: 360. 1843 \equiv *Sphaerella ligustri* (Roberge) Cooke, *J. Bot.* 4: 249. 1866.

Type — France: On upper surface of dead leaves of *Ligustrum vulgare* (Oleaceae). Desmazières, *Fungi Exsiccati* no. 796 (L, isotype, IMI no. 243624, isotype slides); also *Plantes Cryptogames de France* no. 12961 (BPI, isotype).

Anamorph: probably *Cercoseptoria ligustri* (Boerema) Arx *fide* von Arx (1983) (= *Theadgonia ligustrina* (Boerema) B. Sutton).

All isotypes studied contain only a *Septoria* anamorph. The protologue mentions opaque spores 3-4/ascus, which is not in accordance with *Mycosphaerella*. ITS DNA sequence data place this species outside *Mycosphaerella* (Crous & Braun, 2003).

Mycosphaerella limbalis (Pers.) Arx, *Sydowia* 3: 87. 1949, based on an anamorph (illegitimate, Article 59) \equiv *Phyllosticta limbalis* Pers., in Wallr., *Cryptogamae Germaniae* no. 3706. 1818.— Fig. 482.

Type — Germany: On spots on lower surface of living leaves of *Buxus sempervirens* (Buxaceae).

Anamorph: *Phyllosticta limbalis* Pers. *fide* von Arx (1949).

The type was not found in L. Cited as synonym of *Mycosphaerella buxicola* by Eriksson (1992). Materials seen (Switzerland, Slothal, Oberbuchsiten, Arx, VII 1962, CBS and Egerkingen, Jokobsleiter, von Arx, VI 1974, CBS) belong to section *Longispora*, and is morphologically indistinguishable from *M. latebrosa*, with the ascomata mostly pale, not carbonized, asci cylindrical, ascospores $21-25 \times 3-3.5 \mu\text{m}$.

Mycosphaerella limonis Tomilin, *Novosti Sist. Nizsh. Rast.* 1968: 166. 1968, nomen novum (Article 58) for *Sphaerella citricola* McAlpine, *Fungus Diseases of Citrus Trees in Australia*: 85. 1899.

Type — Australia: *Citrus* (Rutaceae).

No material was studied as the type was not included in loans from LE or LEP.

Mycosphaerella linariae Vestergr., *Bih. Kongl. Svenska Vetensk.-Akad. Handl.* 22 (3, 6): 15. 1896 \equiv *Sphaerella linariae* (Vestergr.) Sacc. & P. Syd., *Syll. Fung.* 14: 530. 1899.— Fig. 483.

Type — Sweden: On dead stems and upper and lower surface of dead leaves of *Linaria vulgaris* (Plantaginaceae).

The type was not included in a loan from S. Material studied (Germany, Brandenburg, Nieder-Barnim, Prenden, Sydow, IV 1918, distributed in Mycotheca Germanica no. 1544, L) belongs to section *Caterva*, and is morphologically indistinguishable from *M. superflua*, with asci cylindrical, ascospores $14-17 \times 3-3.5 \mu\text{m}$.

Mycosphaerella lindaviana Staritz, *Verh. Bot. Vereins Prov. Brandenburg* 55: 74. 1913 \equiv *Sphaerella lindaviana* (Staritz) Trotter, *Syll. Fung.* 24: 891. 1928.

Type — Germany: *Valerianella olitoria* (Caprifoliaceae).

No material was studied as the type was not found in B and might be lost. Cited as synonymous with *Mycosphaerella isariphora* by Tomilin (1979).

Mycosphaerella lindiana Jaap, *Verh. Bot. Vereins Prov. Brandenburg* 59: 30. 1917 \equiv *Sphaerella lindiana* (Jaap) Trotter, *Syll. Fung.* 24: 858. 1928.— Fig. 484.

Type — Germany: Brandenburg, Triglitz in der Prignitz. On upper and lower surface of dead leaves of *Tanacetum vulgare* (Asteraceae). Jaap, *Fungi Selecti Exsiccati* no. 771, V 1915 (L, 2 isotypes).

Cited as synonymous with *Mycosphaerella lysimachiae* by Tomilin (1979). This belongs to section *Caterva*, and is morphologically indistinguishable from *M. superflua*, with asci cylindrical, ascospores $14-17 \times 3-3.5 \mu\text{m}$.

Mycosphaerella lindiana Munk. See *Mycosphaerella danica* Tomilin.

Mycosphaerella lindingeri Werdermann, in Syd. & Werdermann, *Ann. Mycol.* 22: 184. 1924.— Fig. 485.

Type — Canary Islands: Laguna. On white spots on upper surface of dead leaves of *Viburnum rugosum* (Adoxaceae). Ade, V 1926 (S, holotype).

This is a parasitic species, with asci cylindrical, ascospores $20-24 \times 4.5-5.5 \mu\text{m}$.

Sphaerella linearis (Rehm) Höhn., *Sitzungsber. Kaiserl. Akad. Wiss., Math.-Naturwiss. Kl., Abt. 1*, 119: 920. 1910 \equiv *Hypomyces linearis* Rehm, *Hedwigia* 39: 223. 1900.

Type — Brazil: *Manettia* (Rubiaceae).

No material was studied as the type was not included in a loan from S.

Sphaerella lineata (Roberge) Ces. & De Not. See *Sphaerella lineolata* (Roberge) Ces. & De Not.

Mycosphaerella lineata Clem., *Bull. Torrey Bot. Club* 30: 84. 1903, [as “*Mycosphaerium lineatum*”] \equiv *Sphaerella lineata* (Clem.) Sacc. & D. Sacc., *Syll. Fung.* 17: 641. 1905.

Type — USA: *Pedicularis procera* (Orobanchaceae).

No material was studied as the type was not found in NY or BPI. Cited as synonymous with *Davidiella allicina* (as *Mycosphaerella allicina*) by Tomilin (1979).

Mycosphaerella lineolata (Roberge) J. Schröt., in Cohn, *Kryptog.-Fl. Schlesien* 3(2): 339. 1894 [“1893”] \equiv *Sphaeria lineolata* Roberge, in Desm., *Ann. Sci. Nat. Bot.*, ser. 2, 19: 351. 1843 \equiv *Sphaerella lineolata* (Roberge) Ces. & De Not., *Comment. Soc. Crittog. Ital.* 1(4): 237. 1863 \equiv *Carlina lineolata* (Roberge) Höhn., *Mitt. Bot. Lab. TH Wien*, ser. 2, 3: 90. 1925 [often also erroneously as “*lineata*”].— Fig. 486.

Type — France: On upper and lower surface of dead leaves of *Carex* (Cyperaceae). Desmazières, *Fungi Exsiccati* no. 763 (L, isotype).

The type and additional material (Netherlands, Vogelenzang, on dead leaves of *Ammophila arenaria* (Poaceae), Oudemans, IV 1878, L) belong to section *Caterva*, and show that this is morphologically indistinguishable from *M. subradians*, with ascomata aggregated in dense groups, asci cylindrical, ascospores $11-15 \times 3.5-4 \mu\text{m}$.

Mycosphaerella linhartiana (Niessl) Starbäck, Bih. Kongl. Svenska Vetensk.-Akad. Handl. 15(3, 2): 9. 1889 = *Sphaerella linhartiana* Niessl, in Linhart, Fungi Hungarici no. 461. 1883.— Fig. 487.

Type — Hungary: Altenburg (“Markttau”). On dead stems of *Melilotus alba* (Fabaceae). Niessl, Fungi Hungarici no. 461, VI 1884 (M, holotype; M, isotype).

The types show that this belongs to section *Caterva* and is morphologically indistinguishable from *M. subradians*, with asci cylindrical, ascospores $11-13 \times 2.5-3.5 \mu\text{m}$.

Mycosphaerella [*“Mycosphaerium”*] *lini* Clem., Cryptogamae Formationum Coloradensium no. 420. 1908, nomen herbariorum (not validly published, Article 32).— Fig. 488. Authentic material — USA: Colorado, Sulphur Springs. On dead stems of *Linum lewisii* (Linaceae). Clements, VII 1907, Cryptogamae Formationum Coloradensium no. 420 (BPI).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores thick-walled, $15-20 \times 4.5-5.5 \mu\text{m}$.

Mycosphaerella lini Hulea & Neagu-Tirc., Stud. Cercet. Biol. (Bucharest) 13: 548. 1961, later homonym (illegitimate, Article 53) = *Sphaerella lini* (Hulea & Neagu-Tirc.) Sandu, Ciuperici Pyrenomycetes-Sphaeriales din România: 133. 1971.

Type — Romania: *Linum* (Linaceae).

No material was studied as the type was not found in any of the herbaria consulted. Cited as synonymous with *Davidiella allicina* (as *Mycosphaerella allicina*) by Tomilin (1979).

Mycosphaerella linicola Naumov, Mater. Mikol. Fitopatol. Rossii 5: 2. 1926.

Type — Russia: Leningrad Prov., Detskoe, Selo. On dead stems of *Linum usitatissimum* (Linaceae). Naumov, IX 1925 (BPI, isotype).

Anamorphs: *Phoma linicola* Naumov *vide* Naumov (op. cit.) and *Septoria linicola* (Speg.) Garass. *vide* Sivanesan (1984).

This is an immature ascomycete with copious pseudoparaphyses, probably a species of *Didymella*. Additional materials seen (United Kingdom, Kent, Ashford, on brown spots on living stems, Bryan, IX 1991, IMI no. 350442; also Yorkshire, Darlington, Croft, Ottway, VIII 1985, IMI no. 297208) contain only the anamorph.

Sphaerella linicola Wollenw. See *Mycosphaerella linorum* (Wollenw.) Garcia Rada & Stev.

Mycosphaerella linicola var. *latispora* Rothers, La Defense des Plantes 4: 3. 1927.

Type — Russia: *Linum usitatissimum* (Linaceae).

No material was studied as the type was not included in a loan from PC.

Mycosphaerella lini-perennis Lobik, Materialy po floristicheskim i faunisticheskim obsledovaniyam Terskogo okruga: 26. 1928.— Fig. 489.

Type — Russia: On stems of *Linum perenne* (Linaceae). (LE 34992, holotype).

This is a species of *Guignardia*, with asci thick-walled, without hamathecial filaments, ascospores not septate, $14-19 \times 5-6 \mu\text{m}$.

Mycosphaerella linnaeae M.E. Barr, in Bigelow & Barr, Rhodora 68: 187. 1966.— Fig. 491.

Type — USA: New Hampshire, White Mts. National Forest, Jefferson. On upper surface of dead leaves of *Linnaea borealis* var. *americana* (Caprifoliaceae). Barr no. 3831 & Bigelow, VI 1963 (NY, holotype; NY, isotype).

This belongs to section *Longispora*, and is morphologically indistinguishable from *M. latebrosa*, with the ascomata mostly immature, asci cylindrical, ascospores $19-22 \times 2-2.5 \mu\text{m}$.

Mycosphaerella linorum (Wollenw.) Garcia Rada & Stev., Minist. Formento, Dirección Agric. Ganad. Peru 54: 17. 1942 = *Sphaerella linorum* Wollenw., Lilloa 2: 493. 1938 [as *“linicola”*].— Fig. 490.

Type — Argentina: Buenos Aires. On dead culms of *Linum usitatissimum* (Linaceae). Garassini, 1935 (B, holotype).

The name was given as *linicola* throughout the original publication and in the title of Wollenweber (op. cit.), but corrected in a postscriptum. This belongs to section *Caterva*, and is morphologically indistinguishable from *M. superflua*, with asci cylindrical, ascospores $15-17 \times 3-3.5 \mu\text{m}$.

Mycosphaerella lippiae (Cif. & Gonz. Frag.) Cif., Quaderno 19: 234. 1961 = *Sphaerella lippiae* Cif. & Gonz. Frag., Bol. Soc. Esp. Hist. Nat. 25: 360. 1925.

Type — Dominican Republic: Haina. On dead, but still tomentose, branches of *Lippia* [*“Lautonia”*] *reticulata* (Verbenaceae). Ciferri no. 84, VI 1925 (BPI, isotype).

In the type only a coelomycete could be found.

Mycosphaerella lirioidendri (Cooke) Woron., Vestn. Tiflissk. Bot. Sada 35: 6. 1914 = *Sphaerella lirioidendri* Cooke, J. Bot. 21: 108. 1883.

Type — USA: Georgia, Darien. On brown spots with black margins on upper surface of leaves of *Liriodendron tulipifera* (Magnoliaceae). Ravenel, XI 1881 (K, holotype; K, 2 isotypes).

Anamorph: *Phyllosticta lirioidendri* Cooke *vide* Cooke (op. cit.).

The types contain nothing identifiable.

Mycosphaerella lithospermi Ellis & Everh., Bull. Torrey Bot. Club 27: 52. 1900 = *Sphaerella lithospermi* (Ellis & Everh.) Sacc. & P. Syd., Syll. Fung. 16: 472. 1902 = *Didymella lithospermi* (Ellis & Everh.) Tomilin, Opredelitel' gribov roda *Mycosphaerella* Johans.: 285. 1979.— Fig. 492.

Type — Canada: Ontario, Gillivray. On dead stems of *Lithospermum officinale* (Boraginaceae). Dearness no. 2862, VI 1898 (NY, holotype; NY, isotype), also distributed in Ellis & Everh., Fungi Columbiani no. 1327 (NY, isotype).

This is a species of *Didymella*, with ascomata large, wall layered, with large cells outside and smaller cells inside, asci cylindrical, with sparse hamathecium, ascospores constricted, $13-16 \times 7-9 \mu\text{m}$.

Mycosphaerella lithreae (Speg.) M. Morelet, Ann. Soc. Sci. Nat. Archéol. Toulon & Var 20: 105. 1968 [as *“litreae”*] =

Sphaerella lithraeae Speg., Revista Chilena Hist. Nat. 28: 27. 1924 [as “*litreae*”].

Type — Chile: *Lithrea* [“*Litrea*”] *caustica* (Anacardiaceae). No material was studied as the type was not included in a loan from LPS.

Mycosphaerella liukiensis Sawada, Taiwan Univ. Coll. Agric. Special Publ. 8: 63. 1959.

Type — Taiwan: Kaohshung, Kuraru. On ill-defined white spots on upper surface of living leaves of *Musa liukiensis* (Musaceae). Sawada, VII 1910 (BPI, isotype).

This is a parasitic species, but the isotype is immature.

Mycosphaerella lobeliae Petr., in Syd. & Petr., Ann. Mycol. 29: 198. 1931.— Fig. 493.

Type — Philippines: Luzon, Benguet, Baguio. On upper and lower surface of dead leaves of *Lobelia nicotianaefolia* (Lobeliaceae). Clemens, Philippine Fungi no. 1538, VI 1923 (W, holotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores 15-20 × 5-8 µm.

Sphaerella lobeliae Petch. See *Mycosphaerella petchii* M. Morelet.

Mycosphaerella loefgreni F. Noack, Z. Pflanzenkrankh. 10: 333. 1901 ≡ *Sphaerella loefgreni* (F. Noack) Sacc. & P. Syd., Syll. Fung. 16: 470. 1902.— Fig. 494.

Type — Brazil: *Citrus aurantium* (Rutaceae).

Anamorph: *Septoria loefgreni* F. Noack *vide* Noack (op. cit.).

The type could not be found in SP and is probably lost. Material seen (Mexico, intercepted at Chicago, on white spots with brown margins on upper and lower surface of living leaves of *Citrus* sp., McNulty, VII 1981, BPI) is a parasitic species, with asci cylindrical, ascospores 10.5-12 × 2-2.5 µm.

Mycosphaerella loliacea (Pass.) Tomilin, Novosti Sist. Nizsh. Rast. 1966: 172. 1966 ≡ *Sphaerella loliacea* Pass., Atti Reale Accad. Lincei, Mem. Cl. Sci. Fis., ser. 4, 3: 10. 1887.

Type — Italy: *Lolium perenne* (Poaceae).

No material was studied as the type was not found in any of the herbaria consulted.

Mycosphaerella longibasalis Crous & M.J. Wingf., Mycologia Memoir 21: 79. 1998.

Type — Colombia: La Selva. On leaves of *Eucalyptus grandis* (Myrtaceae). Wingfield, VI 1994 (PREM 54403, holotype, not seen).

No material was studied of this recently described species.

Mycosphaerella longispora (Penz. & Sacc.) Miles, Trans. Illinois State Acad. Sci. 10: 250. 1917 ≡ *Sphaerella longispora* Penz. & Sacc., Malpighia 11: 397. 1897.— Fig. 495.

Type — Indonesia: Java, Tjibodas. On pale brown spots with brown margins on upper surface of living leaves of

Araliaceae (? host identity questionable). Penzig no. 346 (PAD, holotype).

This is a parasitic species, with asci clavate, ascospores 19-23 × 4.5-5.5 µm, not 25-28 × 4 µm as given in the protologue.

Mycosphaerella longissima (Fuckel) Lindau, Hilfsb. Sammeln Ascomyceten 1903: 20. 1903 ≡ *Sphaerella longissima* Fuckel, Jahrb. Nassauischen Vereins Naturk. 23-24: 107. 1870, nomen novum (Article 58) for *Sphaeria longissima* Fuckel, Fungi Rhenani Exsiccati no. 821. 1863, later homonym (illegitimate, Article 53), non Pers. (1801) ≡ *Sphaerella bromi* Auersw., in Gonn. & Rabenh., Mycol. Europaea 5-6: 17. 1869, superfluous (illegitimate, Article 58) nomen novum (Article 58).— Fig. 496.

Type — Germany: Oestrich. On upper and lower surface of dead leaves and dead leaf sectors of living leaves of *Bromus asper* (Poaceae). Fuckel, Fungi Rhenani Exsiccati no. 821 (L, isotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. ammophilae*, with asci small, pyriform, ascospores immature, 10-12 × 2.5-3.5 µm.

Mycosphaerella lorantheri Syd. & P. Syd., Ann. Mycol. 12: 264. 1914 ≡ *Sphaerella lorantheri* (Syd. & P. Syd.) Trotter, Syll. Fung. 24: 264. 1928.

Type — South Africa: *Loranthus dregei* (Loranthaceae).

No material was studied as the type was not included in a loan from S.

Mycosphaerella louisianae Plakidas, Phytopathology 31: 236. 1941.— Fig. 497.

Type — USA: Louisiana, Hammond. On upper and lower surface of dead leaves of *Fragaria* (Rosaceae). Plakidas no. 3264, V 1938 (NY, holotype).

Already cited as synonymous with *M. punctiformis* by Tomilin (1979), with which the holotype agrees well, with asci cylindrical, ascospores 8-10 × 2-3 µm.

Sphaerella lucillae Sacc., Syll. Fung. 3: 487. 1884, nomen nudum (not validly published, Article 32).

Authentic material — France: *Pyrus* (Rosaceae).

No material was studied as the type was not found in any of the herbaria consulted.

Mycosphaerella ludwigiana (Sacc. & Har.) Tomilin, Novosti Sist. Nizsh. Rast. 8: 150. 1971 ≡ *Sphaerella ludwigiana* Sacc. & Har., in Sacc., Ann. Mycol. 4: 490. 1907 [“1906”].

Type — France: Lardy. On dead stem of *Globularia vulgaris* (Plantaginaceae). Ludwig, 1906 (PAD, holotype).

The type contains only a coelomycete, however, its 1-septate conidia of ca. 9 × 2.5 µm fit the description of the ascospores in the protologue.

Mycosphaerella ludwigii Syd., Ann. Mycol. 22: 260. 1924.— Fig. 498.

Type — Germany: Brandenburg, Nieder-Barnim. On upper and lower surface of dead leaves of *Chamaenerion* [as “*Epilobium*”] *angustifolium* (Onagraceae). Sydow,

Mycotheca Germanica no. 2113, IV 1923 (B, L (2×), isotypes).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradicans*, with asci cylindrical, ascospores 13-16 × 3.5-4.5 µm. Cited as synonymous with *Mycosphaerella epilobii* (Crié) Tomilin, the type of which is a *Venturia*, by Tomilin (1979).

Mycosphaerella lumae Syd., Ann. Mycol. 26: 108. 1928.

Type — Chile: *Myrtus luma* (Myrtaceae).

No material was studied as the type was not included in a loan from S.

Mycosphaerella lupini W.J. Kaiser & Crous, Mycologia 90: 727. 1998.— Fig. 499.

Type — USA: Idaho, Sawtooth National Forest, near Ketchum. On upper and lower surface of dead leaves of *Lupinus argenteus* (Fabaceae). Kaiser, IV 1990 (BPI 806257 p.p., lectotype, here designated).

Anamorph: *Theadgonia lupini* (Davis) U. Braun (op. cit.).

The original publication cites collection BPI 806257 as holotype, but this packet contains three different collections with different collecting dates. Besides the lectotype, which was placed in the soil on 10 IX 1989 and collected on 5 IV 1990, it contains two paratypes, one placed in the soil on 20 IX 1988 and collected on 9 V 1989, and one placed in the soil on 7 VIII 1988 and collected on 3 VII 1989. All specimens belong to *Davidiella*, and are morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores 13.5-15 × 4.5-5.5 µm.

Mycosphaerella lupulina (Kirschst.) M. Morelet, Ann. Soc. Sci. Nat. Archéol. Toulon & Var 20: 105. 1968 ≡ *Sphaerella lupulina* Kirschst., Ann. Mycol. 37: 103. 1939.— Fig. 500.

Type — Germany: Osthavelland, Bredower Forst. On dead stems of *Humulus lupulus* (Urticaceae). Kirschstein, 1916 (B, holotype).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradicans*, with asci cylindrical, ascospores 13-15 × 3-3.5 µm.

Sphaerella lusitanica Auersw., in Gonn. & Rabenh., Mycol. Europaea 5-6: 15. 1869 ≡ *Laestadia lusitanica* (Auersw.) Sacc., Syll. Fung. 1: 425. 1882.

Type — Germany: *Dianthus lusitanicus* (Caryophyllaceae).

No material was studied as the type was not found in B and might be lost.

Mycosphaerella luzonensis T. Kobay., Trans. Mycol. Soc. Japan 21: 311. 1980.

Type — Philippines: *Gardenia phylastrei* (Rubiaceae).

Anamorph: *Cercospora gardeniae* Boedijn fide Kobayashi (op. cit.) (= *Pseudocercospora gardeniae* (Boedijn) Deighton).

No material was studied as the type was not included in a loan from TNS.

Mycosphaerella luzonica Syd., in Syd. & Petr., Ann. Mycol. 29: 199. 1931.

Type — Philippines: *Quisqualis indica* (Combretaceae).

No material was studied as the type was not included in a loan from S.

Mycosphaerella luzulae (Cooke) Höhn., Sitzungsber. Kaiserl. Akad. Wiss., Math.-Naturwiss. Kl., Abt. 1, 111: 8. 1902 ≡ *Sphaerella luzulae* Cooke, Grevillea 6: 31. 1877.— Fig. 501.

Type — Czech Republic: Bohemia, Bodenbach. On dead stems and upper and lower surface of dead leaves of *Luzula albida* (Juncaceae). Thümen, Fungi Austriaci no. 785, 1872 [sub "*Hendersonia typhoidearum* f. *luzulae albidae*"] (K, holotype).

This is morphologically indistinguishable from *Oletheriostrigula papulosa* (Durieu & Mont.) Huhndorf & R.C. Harris, with cellular pseudoparaphyses, ascospores (2-) 4-septate, 22-26 × 4.5-5.5 µm. Additional material studied (Czech Republic, High Tatra, Mlinica-valley. Hruby, distributed in Reliquiae Petrakianae no. 2478, VII 1925, L) belongs to section *Fusispora*, and is morphologically indistinguishable from *M. caricicola*, with asci pyriform, ascospores 12-15 × 2.5-3.5 µm.

Mycosphaerella lychnidicola Syd. & P. Syd., Ann. Mycol. 11: 58. 1913 ≡ *Sphaerella lychnidicola* (Syd. & P. Syd.) Trotter, Syll. Fung. 24: 856. 1928.— Fig. 502.

Type — Japan: Mino, Kawanye-mura. On upper and lower surface of dead leaves of *Lychnis miqueliana* (Caryophyllaceae). Hara, V 1912 (S, holotype).

The type shows that this belongs to section *Caterva* and is morphologically indistinguishable from *M. subradicans*, with asci cylindrical, ascospores 11-13 × 3-3.5 µm.

Mycosphaerella lycii (Ellis & Everh.) Miles, Trans. Illinois State Acad. Sci. 10: 250. 1917 ≡ *Sphaerella lycii* Ellis & Everh., Proc. Acad. Nat. Sci. Philadelphia 45: 135. 1893.— Fig. 503.

Type — Canada: London. On white spots with thickened margins on upper and lower surface of living leaves of *Lycium vulgare* (Solanaceae). Dearness no. 1984, VIII 1892 (NY, holotype).

This is a species of *Leptosphaeria*, with asci cylindrical, ascospores consistently 5-septate, yellowish brown, 20-25 × 4-5 µm.

Mycosphaerella lycopodii (Peck) House, New York State Mus. Bull. 233-234: 28. 1921 ≡ *Sphaerella lycopodii* Peck, Annual Rep. New York State Mus. 39: 51. 1886.

Type — USA: *Lycopodium clavatum* (Lycopodiaceae).

No material was studied as the type was not found in NY or BPI.

Mycosphaerella lycopodii-annotini Petr., Kryptog. Forsch. 2: 167. 1931.— Fig. 504.

Type — Germany: Schwaben, Sesbachmoor. On upper and lower surface of dead scales of *Lycopodium annotinum* (Lycopodiaceae). Ade, VI 1928 (IMI no. 21129, isotype).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradicans*, with asci cylindrical, ascospores 10-12 × 2.5-3.5 µm.

Mycosphaerella lycopodiicola Moesz & Smarods, in Moesz, Bot. Közlem. 34: 60. 1937.

Type — Latvia: *Lycopodium annotinum* (Lycopodiaceae). No material was studied as the type was not found in BP and might be lost.

Mycosphaerella lycopodina (P. Karst.) Starbäck, Bih. Kongl. Svenska Vetensk.-Akad. Handl. 14(3, 5): 9. 1889 = *Sphaerella lycopodina* P. Karst., Fungi Fenniae Exsiccati, cent. 6 no. 569. 1866 = *Pseudapiozpora lycopodina* (P. Karst.) L. Holm, Fungi Exsiccati Suecici no. 2292. 1954 = *Pseudomassaria lycopodina* (P. Karst.) Arx, in E. Müll. & Arx, Beitr. Kryptogamenfl. Schweiz 11(2): 686. 1962.— Fig. 505.

Type — Finland: Tavastia australis, Tammela, Mustiala. On upper surface of decaying leaves of *Lycopodium complanatum* (Lycopodiaceae). Karsten, Fungi Fenniae Exsiccati no. 569, (H, lectotype).

Accepted as *Pseudomassaria lycopodina* (P. Karst.) Arx by Müller & von Arx (op. cit.), with which the lectotype and additional material studied (Germany, Ostpreussen, Samland, Rauschen, on upper and lower surface of dead leaves of *Lycopodium annotinum*, Sydow, VII 1914, distributed in Mycotheca Germanica no. 1233, L) agrees well, with asci cylindrical, ascospores asymmetrically septate, 18-23 × 3.5-6.5 µm.

Mycosphaerella lygei Petr., Ann. Mycol. 29: 111. 1931.— Fig. 506.

Type — Spain: Almeria, Nijar. On dead culms of *Lygeum spartum* (Poaceae). Ade, V 1929 (W, holotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. disseminata*, with asci pyriform, ascospores 13-15 × 3-4 µm.

Mycosphaerella lysimachiae (Höhn.) Höhn., Ann. Mycol. 3: 556. 1905 = *Sphaerella lysimachiae* Höhn., in Strasser, Verh. K.K. Zool.-Bot. Ges. Wien 55: 605. 1905.— Fig. 507.

Type — Austria: Rosenau, Sonntagberg. On upper and lower surface of dead leaves of *Lysimachia vulgaris* (Myrsinaceae). Straßner, Kryptogamae Exsiccatae no. 1151 (L, isotype).

Anamorphs: *Phyllosticta lysimachiae* Allesch. and *Ramularia lysimachiae* Thüm. fide Tomilin (1979). The isotype studied contains only empty ascomata. Additional material studied (Germany, Brandenburg, Triglitz in der Prignitz, Fungi Selecti Exsiccati, Jaap no. 375B, V 1906, L) belongs to section *Caterva*, and is morphologically indistinguishable from *M. superflua*, with asci cylindrical, ascospores 14-16 × 2.5-3.5 µm.

Mycosphaerella lysimachiicola Hino & Katum., J. Jap. Bot. 39: 363. 1964.

Type — Japan: *Lysimachia japonica* (Myrsinaceae). No material was studied as the type was not included in a loan from TNS.

Mycosphaerella lythracearum F.A. Wolf, J. Agr. Res. 35: 468. 1927.— Fig. 508.

Type — USA: *Punica granatum* (Lythraceae).

Anamorphs: *Cercospora lythracearum* Heald & F.A. Wolf fide Wolf (op. cit.) (= *Pseudocercospora lythracearum* (Heald & F.A. Wolf) X.J. Liu & Y.L. Guo) and *Pseudocercospora punicae* (Henn.) Deighton fide Deighton (1976).

No material was studied as the type was not found in BPI and might be lost.

Mycosphaerella lythri Syd., Ann. Mycol. 33: 368. 1935.

Type — Latvia: *Lythrum salicaria* (Lythraceae).

No material was studied as the type was not included in a loan from S. Cited as synonymous with *Mycosphaerella montellica* by Tomilin (1979).

Mycosphaerella macedonica Petr., Ann. Mycol. 34: 215. 1936.— Fig. 509.

Type — Greece: Makedonia, Rudoka planina. On upper and lower surface of dead leaves of *Narthecium scardicum* (Asparagaceae). Ade, II 1931 (W no. 5629, holotype; W no. 9471, isotype).

Already synonymised with *M. recutita* sensu von Arx, which is morphologically indistinguishable from *Davidiella disseminata*, by von Arx (1949), with which the material agrees well, with asci pyriform, ascospores 12-14 × 3.5-4.5 µm.

Mycosphaerella machaerii Bat. & Peres, in Batista *et al.*, Atas Inst. Micol. 3: 224. 1966.— Fig. 510.

Type — Brazil: On white spots on upper surface of living leaves of *Machaerium* (Fabaceae). Batista exs. no. 22493 (URM 45686, holotype).

This is a parasitic species, with asci cylindrical, ascospores 20-24 × 3.5-5.5 µm.

Mycosphaerella macleayae Shira & Hara, Bot. Mag. (Tokyo) 25: 71. 1911 = *Sphaerella macleayae* (Shira & Hara) Trotter, Syll. Fung. 24: 882. 1928 [as “*macleayae*”].

Type — Japan: *Macleaya cordata* (Papaveraceae).

No material was studied as the type was not included in a loan from TNS.

Mycosphaerella macluriae (Ellis & Everh.) Tomilin, Novosti Sist. Nizsh. Rast. 1968: 166. 1968 = *Sphaerella macluriae* Ellis & Everh., Proc. Acad. Nat. Sci. Philadelphia 1890: 231. 1890.— Fig. 511.

Type — USA: Missouri, Saline Co., Emma. On brown spots with black margins on upper surface of living leaves of *Maclura aurantiaca* (Moraceae). Demetrio no. 251, VIII 1889 (NY, holotype).

This is a parasitic species, with asci cylindrical, ascospores 12-15 × 3-4 µm.

Sphaerella macowaniana G. Winter, Flora 67: 267. 1884.

Type — South Africa: *Melianthus major* (Melianthaceae).

No material was studied as the type was not found in B.

Mycosphaerella macrospora (Kleb.) Jørst., Meld. Stat. Plantepatol. Inst. 1: 20. 1945 = *Didymellina macrospora* Kleb., Ber. Deutsch. Bot. Ges. 42: 60. 1925 [“1924”] = *Davidiella macrospora* (Kleb.) Crous & U. Braun, in U. Braun *et al.*, Mycol. Progress 2: 10. 2003.

Type — Germany: *Iris* (Iridaceae).

Anamorph: *Cladsporium iridis* (Fautrey & Roum.) G.A. de Vries (= *Heterosporium gracile* Sacc.) *fide* Sivanesan (1984).

The Klebahn types have not been kept. Material studied (Scotland, West Isles, St. Kilda, on living leaves of *Iris pseudacorus*, Minter, VIII 1988, IMI no. 328172) contains only the anamorph.

Mycosphaerella maculans Lindau, Hilfsb. Sammeln Ascomyceten: 46. 1903, nomen novum (Article 58) for *Sphaerella maculans* Sacc. & Roum., in Roum. & Sacc., Rev. Mycol. (Toulouse) 3: 46. 1881, later homonym (illegitimate, Article 53) = *Mycosphaerella filipendulae* Tomilin, Novosti Sist. Nizsh. Rast. 1968: 165. 1968, superfluous (illegitimate, Article 58) nomen novum (Article 58) = *Mycosphaerella filipendulae-ulmariae* Tomilin, Opredelitel' gribov roda *Mycosphaerella* Johans.: 72. 1979, superfluous (illegitimate, Article 52) nomen novum (Article 58).

Type — Belgium: Malmedy. On lower surface of dead leaves of *Filipendula* ["*Spiraea*"] *ulmaria* (Rosaceae). Libert, distributed in Roumeguère, Fungi Gallici Exsiccati no. 1605 (L, isotype).

Cited as synonymous with *Mycosphaerella innumerella* by Tomilin (1979). The isotype studied is immature.

Mycosphaerella maculans Rick, Brotéria, Ciênc. Nat. 2 (4): 200. 1933, later homonym (illegitimate, Article 53).

Type — Brazil: Myrtaceae.

No material was studied as the type was not found in any of the herbaria consulted.

Sphaerella maculans Auersw., in Gonn. & Rabenh., Mycol. Europaea 5-6: 18. 1869 = *Sphaeria maculans* Sowerby, in Berk. & Broome, Ann. Mag. Nat. Hist., ser. 2, 9: 378. 1852, later homonym (illegitimate, Article 53), non Desm. (1846) = *Sphaeria disseminata* var. *paludosa* Nees von Esenbeck ex Fr., Systema Mycol. 2: 513. 1823 = *Pleospora sowerbyi* Fuckel, Jahrb. Nassauischen Vereins Naturk. 25-26: 301. 1871, non *Pleospora maculans* (Desm.) Fuckel (1870) = *Leptosphaeria sowerbyi* (Fuckel) Sacc., Syll. Fung. 2: 78. 1883, non *Leptosphaeria maculans* (Desm.) Ces. & De Not. (1863) = *Phaeosphaeria sowerbyi* (Fuckel) L. Holm, Symb. Bot. Upsal. 14(3): 130. 1957, superfluous (illegitimate, Article 52) nomen novum (Article 58).

Type — United Kingdom: *Scirpus lacustris* (Cyperaceae). Accepted by Shoemaker & Babcock (1989) as *Phaeosphaeria sowerbyi* (Fuckel) L. Holm and therefore not studied.

Sphaerella maculans Pass. See *Sphaerella subcrassa* Sacc. & P. Syd.

Mycosphaerella macularis (Fr. : Fr.) J. Schröt., in Cohn, Kryptog.-Fl. Schlesien 3(2): 335. 1894 ["1893"] = *Sphaeria macularis* Fr., Observ. Mycol. 1: 186. 1815, sanctioned by Fr., Systema Mycol. 2: 502. 1823 = *Perisporium maculare* (Fr. : Fr.) Fr., Systema Mycol. 3: 251. 1829 = *Sphaerella macularis* (Fr. : Fr.) Auersw., in Gonn. & Rabenh., Mycol. Europaea 5-6: 9. 1869 = *Phaeosphaerella macularis* (Fr. :

Fr.) Traverso, Atti Soc. Veneto-Trentina Sci. Nat. 5: 8. 1912 = *Venturia macularis* (Fr. : Fr.) E. Müll. & Arx, Ber. Schweiz. Bot. Ges. 60: 366. 1950 = *Sphaerella maculosa* Sacc., Syll. Fung. 1: 487. 1882, superfluous (illegitimate, Article 52) nomen novum (Article 58) for *Sphaerella macularis* (Fr. : Fr.) Auersw. ["P. Karst., Bidrag Kännedom Finlands Natur Folk 23: 174. 1873"] = *Phaeosphaerella maculosa* (Sacc.) P. Karst., Meddel. Soc. Fauna Fl. Fenn. 16: 28. 1888.

Type — Sweden: *Populus* (Salicaceae).

Anamorph: *Pollaccia radiosa* (Lib.) E. Bald. & Cif. *fide* Sivanesan (1984) (= *Fusicladium radiosum* (Lib.) Lind).

Accepted as *Venturia macularis* (Fr. : Fr.) E. Müll. & Arx by Sivanesan (1977), with which material studied (Germany, Oestrich, on lower surface of dead leaves of *Populus tremula*, Fuckel, Fungi Rhenani no. 830, L) agrees well, although it is not fully mature. Additional material (Netherlands, Groningen, on *Acer pseudoplatanus* (Sapindaceae), L) contains only coelomycetes.

Sphaerella macularis f. *populi-tremulae* J. Kunze, Fungi Selecti Exsiccati no. 245.— Fig. 512.

Type — Germany: Sachsen, Eisleben, Wolfersode. On lower surface of dead leaves of *Populus tremula* (Salicaceae). Kunze, V 1879, distributed in Fungi Selecti Exsiccati no. 245 (L, holotype).

This is morphologically indistinguishable from *Venturia macularis* (Fr. : Fr.) E. Müll. & Arx, like the nominal forma, with ascospores brown, asymmetrically septate, 11-13 × 5-6 µm.

Sphaerella maculata Fautrey, Rev. Mycol. (Toulouse) 13: 166. 1891.— Fig. 513.

Type — France: On upper and lower surface of dead leaves of *Prunus mahaleb* (Rosaceae). Fautrey, III 1891, distributed in Herbar Cryptogamique de la Cote d'Or no. 53 (L, isotype).

This is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores 8-10 × 2-2.5 µm. It was cited as synonymous with *Mycosphaerella cerasina* by Tomilin (1979).

Mycosphaerella maculicola (G. Winter) Tomilin, Opredelitel' gribov roda *Mycosphaerella* Johans.: 62. 1979, nom. inval. (Article 33) = *Sphaerella maculicola* G. Winter, Hedwigia 24: 28. 1885.

Type — South Africa: Cape Province. *Helichrysum* (Asteraceae).

No material was studied as the type was not found in B.

Mycosphaerella maculiformis (Pers. : Fr.) J. Schröt., in Cohn, Kryptog.-Fl. Schlesien 3(2): 333. 1894 ["1893", as "*maculaeformis*"] = *Sphaeria maculiformis* Pers., Tent. Disposit. Method. Fung.: 52. 1797, superfluous nomen novum (illegitimate, Article 52) for *Sphaeria corylea* Pers., Ann. Bot. (Usteri) 11: 26. 1794, but sanctioned by Fr., Systema Mycol. 2: 524. 1823 [as "*maculaeformis*"] = *Sphaeria maculiformis* var. *corylea* (Pers.) Pers., Tent. Disposit. Method. Fung.: 52. 1797 = *Sphaerella maculiformis* (Pers. : Fr.) Auersw., in Gonn. & Rabenh., Mycol. Europaea 5-6: 5. 1869 = *Carlia maculiformis* (Pers. :

Fr.) Höhn., Hedwigia 62: 70. 1920, later homonym (illegitimate, Article 53), non Bonord. (1864) = *Sphaerella vulgaris* P. Karst., Bidrag Kännedom Finlands Natur Folk 23: 168. 1873, superfluous nomen novum (illegitimate, Article 52) = *Mycosphaerella vulgaris* (P. Karst.) Johanson, Öfvers. Förh. Kongl. Svenska Vetensk.-Akad. 41(9): 164. 1884, illegitimate combination, because basionym illegitimate.— Fig. 514.

Type — Netherlands: On lower surface of dead leaves of *Corylus avellana* (Betulaceae). (L-Persoon, holotype).

Anamorphs: *Phyllosticta ludoviciana* Ellis & G. Martin *vide* Tomilin (1979), *Septoria quercina* Desm. and probably *Septoria castanicola* Desm. *vide* Saccardo (1882).

Cited as synonymous with *M. punctiformis* by Barr (1972), from which the species traditionally has been separated by the aggregated ascospores. The holotype is immature, but morphologically indistinguishable from *M. punctiformis*, as is most additional material studied. However, this character is very inconstant, even on one leaf. *Sphaeria corylea* is the oldest name available for this well-known species, but the name *Sphaeria maculiformis* has been sanctioned. This name and the name *Sphaeria punctiformis* have been published simultaneously and the choice is therefore open to use either one for the taxon. Here, the choice for the name *Mycosphaerella punctiformis*, as earlier made by e.g. Barr (1972) is followed here and by Verkley *et al.* (2004).

Mycosphaerella maculiformis f. *aceris* (Höhn.) Petr., Sydowia 1: 227. 1947 [as “*maculaeformis* f.”] = *Carlia maculiformis* f. *aceris* Höhn., Hedwigia 62: 70. 1920 [as “*maculaeformis* f.”].

Type — Germany: *Acer* (Sapindaceae).

No type material was preserved in FH.

Sphaerella maculiformis f. *aceris-pseudoplatani* Rabenh., Fungi Europaei Exsiccati no. 1159. 1868 [as “*maculaeformis* f.”].

Type — Germany: Württemberg, Donnstetten. On lower surface of dead leaves of *Acer pseudoplatanus* (Sapindaceae). Rabenh., Fungi Europaei Exsiccati no. 1159, X 1867 (L, isotype).

The isotype studied contains only an *Asteromella* anamorph, but this is morphologically indistinguishable from *M. punctiformis*.

Sphaerella maculiformis f. *alni* Roum., Fungi Gallici Exsiccati 2292 [as “*maculaeformis* f.”].

Type — France: Toulouse. On lower surface of dead leaves of *Alnus glutinosa* (Betulaceae). Roumeguère, Fungi Gallici Exsiccati no. 2292, 1882 (L, isotype).

The isotype studied is immature, but this is morphologically indistinguishable from *M. punctiformis*.

Sphaerella maculiformis f. *carpini* Rabenh., Fungi Europaei Exsiccati, ed. nov., ser. 2 no. 542. 1863 [as “*maculaeformis* f.”].

Type — Germany: Driesen. On lower surface of dead leaves of *Carpinus betulus* (Betulaceae). Lasch, distributed in Rabenhorst, Fungi Europaei Exsiccati, ed. nov., ser. 2 no. 542 (L, 2 isotypes).

The isotype studied contains only empty ascospores, but this is morphologically indistinguishable from *M. punctiformis*.

Sphaerella maculiformis f. *castaneae* Thüm., Fungi Austriaci no. 760. 1873 [as “*maculaeformis* f.”].

Type — Czech Republic: Bohemia, Teplitz. On upper and lower surface of dead leaves of *Castanea sativa* [“*vesca*”] (Fagaceae). Thümen, 1872, distributed in Fungi Austriaci no. 760 (BPI, isotype).

This is immature, but morphologically indistinguishable from *M. punctiformis*.

Sphaerella maculiformis f. *castaneae* Westend., nomen herbariorum (not validly published, Article 32) also later homonym (illegitimate, Article 53) [as “*maculaeformis* f.”].

Authentic material — Belgium: Bruxelles. On lower surface of dead leaves of *Castanea sativa* (Fagaceae). Westendorp, distributed in Roumeguère, Fungi Gallici Exsiccati no. 3034 (L).

This is immature, but morphologically indistinguishable from *M. punctiformis*.

Sphaerella maculiformis f. *castaneae-vescae* Sacc., Mycotheca Veneta no. 693. 1876. [as “*maculaeformis* f.”].

Type — Italy: Treviso, Vittorio. On lower surface of dead leaves of *Castanea sativa* [“*vesca*”] (Fagaceae). Sacc., X 1875, distributed in Mycotheca Veneta no. 693 (BPI, isotype).

This is immature, but morphologically indistinguishable from *M. punctiformis*.

Sphaerella maculiformis f. *comari-palustris* Rabenh., Fungi Europaei Exsiccati no. 1042. 1868 [as “*maculaeformis* f.”].— Fig. 515.

Type — Germany: Sachsen, Chemnitz. On upper and lower surface of dead leaves of *Comarum palustre* (Rosaceae). Delitsch, distributed in Rabenhorst, Fungi Europaei Exsiccati no. 1042, VII (L, isotype).

This belongs to section *Longispora*, and is morphologically indistinguishable from *M. millegrana*, with asci cylindrical, ascospores 14-16 × 2-2.5 µm. It was cited as synonymous with *Mycosphaerella innumerella* by Tomilin (1979).

Mycosphaerella maculiformis f. *congesta* Rehm, Ascomyceten no. 1343a. 1900.— Fig. 516, 962.

Type — Germany: München, Neufriedenheim. On upper and lower surface of dead leaves of *Quercus*. Rehm, Ascomyceten no. 1343a, VI 1900 (B, isotype).

This is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores 8-10 × 2-3 µm.

Mycosphaerella maculiformis f. *dispersa* Rehm, Ascomyceten no. 1343b. 1900.— Fig. 517.

Type — Germany, München, Neufriedenheim. On upper and lower surface of dead leaves of *Quercus*. Rehm, Ascomyceten no. 1343b, VI 1900 (B, isotype).

This is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores 9-11 × 2-3 µm.

Sphaerella maculiformis f. *fraxini-excelsioris* Thüm., Mycotheca Universalis no. 853. 1877 [as “*maculaeformis* f.”].— Fig. 518.

Type — Germany: Bayreuth. On lower surface of dead leaves of *Fraxinus excelsior* (Oleaceae). Thümen, Mycotheca Universalis no. 853, 1876 (L, isotype).

This is morphologically indistinguishable from *Davidiella allicina*, with asci pyriform, ascospores $19-22 \times 7-8 \mu\text{m}$.

Sphaeria maculiformis f. *myrtilli* Klotzsch, Herb. Vivum Mycol., cent. 11 no. 1040. 1846 [as “*maculaeformis* f.”].

Type — Germany: Leipzig. On upper and lower surface of dead leaves of *Vaccinium myrtillus* (Ericaceae). Auerswald, distributed in Klotzsch, Herbarium Vivum Mycologicum no. 1040 (L, isotype).

This belongs to section *Longispora*, and is morphologically indistinguishable from *Mycosphaerella vaccinii*, which is morphologically indistinguishable from *M. millegrana*, with asci cylindrical, ascospores $12-15 \times 1.5-2 \mu\text{m}$.

Sphaerella maculiformis f. *pseudoplatani* Pass. & Brunaud, Rev. Mycol. (Toulouse) 8: 205. 1886 [as “*maculaeformis* f.”].

Type — France: *Acer pseudoplatanus* (Sapindaceae).

No material was studied as the type was not found in any of the herbaria consulted.

Sphaerella maculiformis f. *quercus* Thüm., Fungi Austriaci no. 160. 1871 [as “*maculaeformis* f.”].

Type — Austria, Krems. On living leaves of *Quercus pedunculata*. Thümen, Fungi Austriaci no. 160, IX 1871 (B, isotype).

No fungus is present in this material.

Sphaerella maculiformis f. *quercus-roboris* J. Kunze, Fungi Selecti Exsiccati no. 244. 1878.— Fig. 519.

Type — Germany: Sachsen, Eisleben. On lower surface of dead leaves of *Quercus robur* (Fagaceae). Kunze, V 1879, distributed in Fungi Selecti Exsiccati no. 244 (L, holotype).

This is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores $8-10 \times 2-2.5 \mu\text{m}$.

Sphaerella maculiformis f. *senecionis* Roum., Fungi Gallici Exsiccati no. 1604. 1881 [as “*maculaeformis* f.”].

Type — France: On lower surface of dead leaves of *Senecio bannelieri* (Asteraceae). Libert, 1881, distributed in Roumeguère, Fungi Gallici Exsiccati no. 1604 (L, isotype).

The isotype studied is immature, but this is morphologically indistinguishable from *M. punctiformis*.

Sphaerella maculiformis f. *sorbi* Roum., Fungi Gallici Exsiccati no. 2096. 1882 [as “*maculaeformis* f.”].

Type — Belgium: Malmedy. On lower surface of dead leaves of *Acer campestre* [sic] (Sapindaceae). Libert, distributed in Roumeguère, Fungi Gallici Exsiccati no. 2096 (L, isotype).

The isotype studied is immature, but this is morphologically indistinguishable from *M. punctiformis*.

Sphaerella maculiformis f. *spermogonifera* Rabenh., Fungi Europaei Exsiccati, ed. nov., ser. 2 no. 761. 1863 [as “*maculaeformis* f.”].

Type — Germany: Salem. On lower surface of dead leaves of *Acer platanoides* (Sapindaceae). Rabenhorst, Fungi Europaei Exsiccati no. 761, 1863 (L, 2 isotypes).

The isotype studied is immature, but this is morphologically indistinguishable from *M. punctiformis*, of which it represents the *Asteromella* spermatial state.

Sphaerella maculiformis var. *aequalis* Cooke. See *Sphaerella aequalis* (Cooke) Auersw.

Sphaerella maculiformis var. *alni* Cooke, Fungi Britannici Exsiccati, ser. 2, no. 274. 1875.— Fig. 520.

Type — British Isles: Gomshall. On lower surface of dead leaves of *Alnus* (Betulaceae). Cooke, Fungi Britannici Exsiccati, ser. 2, no. 274 (B, isotype).

This is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores $9-11 \times 2-2.5 \mu\text{m}$.

Sphaerella maculiformis var. *arcana* (Cooke) Auersw. See *Sphaerella arcana* Cooke.

Sphaerella maculiformis var. *carpini* Fuckel, Jahrb. Nassauischen Vereins Naturk. 23-24: 107. 1870 [as “*maculaeformis* var.”].

Type — Germany: Oestrich. On upper and lower surface of dead leaves of *Carpinus betulus* (Betulaceae). Fuckel, Fungi Rhenani no. 1783 (L, isotype).

This is indeed morphologically indistinguishable from *M. punctiformis*, with ascospores immature.

Sphaerella maculiformis var. *castaneae* Cooke, Fungi Britannici Exsiccati, ser. 2, no. 275. 1875 [as “*maculaeformis* var.”].— Fig. 521.

Type — British Isles: Kent, Darenth. On lower surface of dead leaves of *Castanea vesca* (Fagaceae). Cooke, Fungi Britannici Exsiccati, ser. 2, no. 275 (B, isotype).

This is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores $10-12 \times 2-3 \mu\text{m}$.

Sphaerella maculiformis var. *castaneaecola* (Fr.) Poetsch & Schied., Syst. Aufzählung, Herausgegeben von der K.K. Zool.-Bot. Ges. Wien. 143 no. 1475. 1872 [as “*maculaeformis* var.”] \equiv *Sphaeria castaneaecola* Fr., Systema Mycol. 2: 530. 1823 \equiv *Sphaeria maculiformis* f. *castaneaecola* (Fr.) Rabenh., Fungi Europaei Exsiccati no. 1948, 1875.

Type — Sweden: On lower surface of dead leaves of *Castanea sativa* (Fagaceae).

The material studied (Austria, Kremsmünster, Poetsch, distributed in Rabenhorst, Fungi Europaei Exsiccati no. 1948, L) is immature, but this is morphologically indistinguishable from *M. punctiformis*.

Sphaerella maculiformis var. *centigrana* Cooke, J. Bot. 4: 246. 1899 [as “*maculaeformis* var.”].— Fig. 522.

Type — United Kingdom: Kent, Darent. On lower surface of dead leaves of *Castanea vesca* (Fagaceae). Cooke, Fungi Britannici Exsiccati no. 169, IV 1866 (K, holotype; B, 2 isotypes).

This is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores $9-11 \times 2-3 \mu\text{m}$.

Sphaeria maculiformis var. *corylea* (Pers.) Pers. See *Mycosphaerella maculiformis* (Pers. : Fr.) J. Schröt.

Sphaerella maculiformis var. *coryli* Cooke, Fungi Britannici Exsiccati, ser. 2, no. 273. 1875 [as “*maculaeformis* var.”].— Fig. 523.

Type — United Kingdom: Kent, Darent. On lower surface of dead leaves of *Corylus avellana* (Betulaceae). Cooke, Fungi Britannici Exsiccati, ser. 2, no. 273 (B, isotype).

This is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores $7-8 \times 2-2.5 \mu\text{m}$.

Sphaerella maculiformis var. *fici* Roum., Fungi Selecti Gallici Exsiccati no. 1086. 1881 [as “*maculaeformis* var.”].

Type — France: Anduze. On lower surface of dead leaves of *Ficus carica* (Moraceae). Therry, X 1878, distributed in Roumeguère, Fungi Selecti Gallici Exsiccati no. 1086 (L, isotype).

The isotype studied is immature, but this is morphologically indistinguishable from *M. punctiformis*.

Mycosphaerella maculiformis var. *herbaceae* L. Holm & K. Holm, Symb. Bot. Upsal. 31(3): 287. 1996.

Type — Norway: *Salix herbacea* (Salicaceae). Holm & Holm 652b (UPS, holotype, not seen).

No material was studied of this recently described variety, which undoubtedly belongs to *M. punctiformis*.

Sphaerella maculiformis var. *hippocastani* Jaap. See *Mycosphaerella hippocastani* (Jaap) Kleb.

Sphaerella maculiformis var. *oblivia* (Cooke) Auersw. See *Sphaerella oblivia* Cooke.

Sphaerella maculiformis var. *quercinae* Cooke, Fungi Britannici Exsiccati, ser. 2, no. 276. 1875 [as “*maculaeformis* var.”].

Type — United Kingdom: Kent, Darent. On lower surface of dead leaves of *Quercus* (Fagaceae). Cooke, Fungi Britannici Exsiccati, ser. 2, no. 276 (B, isotype).

The isotype studied is immature, but this is morphologically indistinguishable from *M. punctiformis*.

Sphaerella maculiformis var. *rubi* Fuckel, Jahrb. Nassauischen Vereins Naturk. 23-24: 107. 1870 [as “*maculaeformis* var.”].

Type — Germany. On lower surface of dead leaves of *Rubus caesius* (Rosaceae). Fuckel, Fungi Rhenani no. 1782 (B, L, isotypes).

This is immature, but morphologically indistinguishable from *M. punctiformis*.

Sphaerella maculiformis var. *tiliae* Fuckel, Jahrb. Nassauischen Vereins Naturk. 23-24: 107. 1870 [as “*maculaeformis* var.”].

Type — Germany: Vollrath. On lower surface of dead leaves of *Tilia parvifolia* (Malvaceae). Fuckel, Fungi Rhenani no. 1781 (L, isotype).

This is immature, but morphologically indistinguishable from *M. punctiformis*.

Sphaeria maculiformis var. *tiliae* Pers., nomen herbariorum (not validly published, Article 32).

Authentic material — Netherlands: On lower surface of dead leaves of *Tilia* (Malvaceae). Pers. (L-Pers.).

This is immature, but morphologically indistinguishable from *M. punctiformis*.

Mycosphaerella maculiseda Nevod., nomen herbariorum (not validly published, Article 32).— Fig. 524.

Authentic material — Ukraine: Kiev. On upper surface of dead leaves of *Phragmites australis* (Poaceae). Nevodovski, V 1915 (LE 34998, LE 34999, NY).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, but the ascomata are immature, with asci pyriform, ascospores $18-22 \times 5-8 \mu\text{m}$.

Sphaerella maculosa Sacc. See *Mycosphaerella macularis* (Fr. : Fr.) J. Schröt.

Mycosphaerella maderensis Petr., Bot. Jahrb. Syst. 62, Beibl. 142: 122. 1928.— Fig. 525.

Type — Madeira: Funchal. On upper and lower surface of dead leaves of *Sibthorpia perennis* (Plantaginaceae). Ade, VI 1926 (W no. 10046, holotype; W no. 26923, isotype).

This belongs to section *Longispora*, and is morphologically indistinguishable from *M. millegrana*, with asci cylindrical, ascospores $12-14 \times 2-2.5 \mu\text{m}$.

Mycosphaerella madeirae Crous & Denman in Crous *et al.*, Stud. Mycol. 50: 204. 2004.

Type — Madeira: Party Farm, on leaves of *Eucalyptus globulus* (Myrtaceae). Denman, IV 2000 (CBS 9898, holotype); cultures ex-type CPC 3745 = CBS 112895, CPC 3747 = CBS 112301.

Anamorph: *Pseudocercospora fide* Crous *et al.* (op. cit.).

No material was studied of this recently described species.

Mycosphaerella maesae Crous & U. Braun, Sydowia 46: 210. 1994.

Type — South Africa: *Maesa lanceolata* (Myrsinaceae).

No material was studied of this recently described species.

Mycosphaerella magellanica (Speg.) Tomilin, Novosti Sist. Nizsh. Rast. 7: 204. 1970 = *Sphaerella magellanica* Speg., Bol. Acad. Nac. Ci. 11: 40. 1888 [“1887”].

Type — Argentina: *Armeria andina* (Plumbaginaceae).

No material was studied as the type was not included in a loan from LPS.

Mycosphaerella magellanicola (Speg.) Cash, Syll. Fung. 26: 345. 1972 = *Sphaerella magellanicola* Speg., Bol. Acad. Nac. Ci. 27: 362. 1924.

Type — Argentina: *Tetroncium magellanicum* (Juncaginaceae); also mentioned in the protologue from *Libocedrus tetragona* (Cupressaceae).

No material was studied as the syntypes were not included in a loan from LPS.

Mycosphaerella magnoliae (Ellis) Petr., Sydowia 5: 247. 1951 = *Sphaerella magnoliae* Ellis, Bull. Torrey Bot. Club 9: 74. 1882.— Fig. 526.

Type — USA: New Jersey, Newfield. On upper and lower surface of dead leaves of *Magnolia glauca* (Magnoliaceae). VI 1880 (NY, holotype), also distributed in North American Fungi no. 800 (L, NY (2×), isotypes), also distributed in Rabenhorst, Fungi Europaei Exsiccati no. 2755 (L, isotype), also distributed in Roumeguère, Fungi Gallici Exsiccati no. 2453 (L, isotype).

Additional authentic material studied: New Jersey, Newfield. On living leaves of *M. glauca*. Ellis & Harkness, XI 1880 (IMI no. 201400, topotype). As already indicated by Barr (1977), this is a later synonym of *Hyponectria magnoliae* (Schwein.) M.E. Barr, with which types and additional material studied (Maryland, Beltsville, on upper surface of dead leaves, but in leaf spot-like groups, of *Magnolia virginiana*, Petrak, distributed in Reliquiae Petrakianae no. 1253, V 1950, L) agree well, with ascomata mostly immersed, flattened above, asci cylindrical, ascospores not septate, 9-12 × 2-3 µm.

Mycosphaerella magnusiana Jaap, Ann. Mycol. 6: 209. 1908 = *Sphaerella magnusiana* (Jaap) Sacc. & Trotter, Syll. Fung. 22: 128. 1913.

Type — Austria: *Astragalus alpinus* (Fabaceae).

No material was studied as the type was not found in B and might be lost.

Sphaerella maidenii Tassi, Bull. Lab. Orto Bot. Reale Univ. Siena 3: 94. 1900.— Fig. 527.

Type — Australia: On dead branches of *Myoporum acuminatum* (Scrophulariaceae). Tassi, 1900 (SIENA, holotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. ammophilae*, with asci pyriform, ascospores 11-13 × 2.5-3.5 µm.

Mycosphaerella major (Auersw.) Lindau, Hilfsb. Sammeln Ascomyceten: 86. 1903 = *Sphaerella major* Auersw., in Gonn. & Rabenh., Mycol. Europaea 5-6: 8. 1869.

Type — Germany: *Populus tremula* (Salicaceae).

Cited as synonymous with *Venturia macularis* (Fr. : Fr.) E. Müll. & Arx by Sivanesan (1977).

Sphaerella majuscula Cooke, J. Bot. 21: 138. 1883.— Fig. 528.

Type — New Zealand: Stewart Island. On upper surface of dead leaves of *Senecio rotundifolius* (Asteraceae). Kirk (K, holotype; K, 2 isotypes).

This is a species of *Planistromella*, with ascomata aggregated with 2-5 in stromata, breaking through the cortex, asci cylindrical, surrounded by sparse parenchymatous cells, ascospores 11-13 × 2.5-3.5 µm. Therefore the following new combination is proposed:

Planistromella majuscula (Cooke) Aptroot comb. nov.; **MB 500373**. **Basionym:** *Sphaerella majuscula* Cooke, J. Bot. 21: 138. 1883.

Mycosphaerella malinverniana (Catt.) Jacz., Opredelitel' gribov 2: 615. 1917 = *Sphaerella malinverniana* Catt., Arch. Triennale Lab. Bot. Crittog. 2: 127. 1877.— Fig. 529.

Type — Italy: Pavia. On dead leaves of *Oryza sativa* (Poaceae). Ex laboratorio crittogamico (BPI, possible isotype). This belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores 14-16 × 5-6 µm. Additional material studied (Pavia, Marcignago, Trovamala, 1891, IMI no. 16771, topotype) contains only a hyphomycete.

Mycosphaerella malvina Nevod., Bot. Mater. Otd. Sporov. Rast. Bot. Inst. Komarova Akad. Nauk SSSR 14: 173. 1961.

Type — Kazakhstan: *Malva* (Malvaceae).

No material was studied as the type was not included in loans from LE or LEP.

Mycosphaerella mandshurica Miura, Industr. Contr. 27: 161. 1928.

Type — Manchuria: *Populus laurifolia* (Salicaceae).

Anamorphs: *Phyllosticta* and unidentified coelomycete *vide* Miura (op. cit.).

No material was studied as the preservation of the type is uncertain.

Mycosphaerella manganottiana (C. Massal.) Tomilin, Novosti Sist. Nizsh. Rast. 6: 121. 1970 [“1969”] = *Sphaerella manganottiana* C. Massal., Mem. Accad. Agric. Verona, ser. 3, 65: 140. 1889.

Type — Italy: *Vitis vinifera* (Vitaceae).

No material was studied as the type was not found in any of the herbaria consulted.

Mycosphaerella mangiferae C.H. Ramesh, Current Sci. 55: 369. 1986.

Type — India: *Mangifera indica* (Anacardiaceae).

No material was studied as the type was not found in IMI or LWG.

Mycosphaerella manginii Savul. & Sandu, Recueil Trav. Cryptog. dédiés à Louis Mangin: 1. 1931 [as “mangini”] = *Sphaerella manginii* (Savul. & Sandu) Sandu, Ciuperici Pyrenomycetes-Sphaeriales din România: 134. 1971.— Fig. 530.

Type — Romania: Distr. Prahova, Busteni. On spots on upper and lower surface of living leaves of *Spiraea chamaedryfolia* [“ulmifolia”] (Rosaceae). Savulescu & Sandu-Ville, Herbarium Mycologicum Romanicum no. 252, VIII 1930, (L, isotype).

This is a parasitic species, with asci nearly cylindrical, ascospores 10-15 × 3-4 µm.

Mycosphaerella manihotis Syd. & P. Syd., Bull. Herb. Boissier, ser. 2, 1: 78. 1901 = *Sphaerella manihotis* (Syd. & P. Syd.) Sacc. & P. Syd., Syll. Fung. 16: 472. 1902 = *Mycosphaerella manihotis* f. sp. *microspora* Chevaug.,

Encycl. Mycol. 28: 34. 1956, superfluous (illegitimate, Article 52) nomen novum (Article 58).— Fig. 531.

Type — Brazil: Santa Catharina, Itaiaty. On white spots on upper surface of living leaves of *Manihot utilissima* (Euphorbiaceae). Ule no. 610, V 1886 (S, holotype).

This is a parasitic species, with asci cylindrical, ascospores $11-13 \times 3-3.5 \mu\text{m}$.

Mycosphaerella manihotis Ghesq. & Henrard. See *Mycosphaerella henningsii* Sivan.

Mycosphaerella manihotis f. sp. *macrospora* Chevaug. See *Mycosphaerella henningsii* Sivan.

Mycosphaerella manihotis f. sp. *microspora* Chevaug. See *Mycosphaerella manihotis* Syd. & P. Syd.

Mycosphaerella maniuana Butin, Sydowia 27: 277. 1975 [“1973”].— Fig. 532.

Type — Chile: Antillanca, Osorno. On upper and lower surface of dead needles of *Saxegothea conspicua* (Podocarpaceae). Butin, III 1970 (ZT, holotype).

This is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores $8-9 \times 2-2.5 \mu\text{m}$.

Mycosphaerella manshurica Ziling, nomen herbariorum (not validly published, Article 32).

Authentic material — Manchuria: Posyet. On spots on upper surface of living leaves of *Fraxinus manshurica* (Oleaceae). Ziling, VII 1928 (LEP).

Anamorph: Associated with *Septoria manshurica* Ziling.

This contains only the anamorph *Septoria manshurica* Ziling.

Mycosphaerella mappiae (Petch) Cash, Syll. Fung. 26: 346. 1972 \equiv *Sphaerella mappiae* Petch, Ann. Roy. Bot. Gard. (Peradeniya) 7: 303. 1922.— Fig. 533.

Type — Sri Lanka: Hakgala. On white spots on lower surface of leaves of *Mappia ovata* (Icacinaceae). Petch no. 5236, IV 1917 (K, holotype).

This belongs to *Davidiella*, of which it represents a parasitic species, with asci pyriform, ascospores $18-21 \times 4.5-5.5 \mu\text{m}$. Therefore the following new combination is made:

Davidiella mappiae (Petch) Aptroot comb. nov., **MB 500355**. **Basionym:** *Sphaerella mappiae* Petch, Ann. Roy. Bot. Gard. (Peradeniya) 7: 303. 1922.

Mycosphaerella marasasii Crous & M.J. Wingf., Mycol. Res. 95: 1111. 1991.

Type — South Africa: *Syzygium cordatum* (Myrtaceae).

No material was studied of this recently described species.

Sphaerella marginata (Wallr.) G. Winter, Hedwigia 11: 145. 1872 \equiv *Sphaeria marginata* Wallr., Fl. Cryptog. Germ. 2: 770. 1833 \equiv *Leptosphaeria marginata* (Wallr.) Niessl, Verh. Naturf. Vereins Brünn 10: 173. 1872 [“1871”].

Type — Germany: *Chimaphila* [“*Pyrola*”] *umbellata* (Ericaceae).

No material was studied as the type was not found in any of the herbaria consulted.

Mycosphaerella mariae (Sacc. & E. Bommer) Lindau, Hilfsb. Sammeln Ascomyceten: 37. 1903 \equiv *Sphaerella mariae* Sacc. & E. Bommer in E. Bommer & M. Rousseau, Bull. Soc. Roy. Bot. Belgique 25: 173. 1886.— Fig. 534.

Type — Belgium: Hastière. On upper and lower surface of dead leaves of *Digitalis lutea* (Plantaginaceae). Bommer & Rousseau, V 1888, distributed in Rehm, Ascomyceten no. 997 (NY, isotype).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradians*, with asci cylindrical, ascospores $10-12 \times 3-3.5 \mu\text{m}$.

Sphaerella marii De Not., Comment. Soc. Crittog. Ital. 2(3): 488. 1867 \equiv *Laestadia marii* (De Not.) Sacc., Syll. Fung. 1: 425. 1882.

Type — Switzerland: Chiasso. On dead stems of *Nicotiana tabacum* (Solanaceae). Piratti no. 64, III 1867 (RO, holotype; B, RO, isotypes).

The types contains only various coelomycetes.

Mycosphaerella marksii Carnegie & Keane, Mycol. Res. 98: 414. 1994.— Fig. 535.

Type — Australia: Victoria, Gippsland, Nowa Nowa. On white spots with brown margins on upper surface of living leaves of *Eucalyptus botryoides* (Myrtaceae). Carnegie, II 1991 (IMI no. 353731, holotype).

Type and additional material (Same data, but on *E. grandis*, IMI no. 353730, topotype; Same data, but on *E. quadrangulata*, IMI no. 353732, topotype) belong to section *Plaga*, with asci cylindrical, ascospores $13-16 \times 3-4 \mu\text{m}$.

Mycosphaerella martagonis Arx, Sydowia 3: 81. 1949.— Fig. 536.

Type — Switzerland: Zürich, Leimbach, Entlisberg. On spots on dead leaves of *Lilium martagon* (Liliaceae). Von Arx, IV 1948 (IMI no. 226760, isotype slide).

Anamorph: *Pseudocercospora hungarica* (Bäumler) Sivan. *vide* Sivanesan (1984).

This is a parasitic species, with asci cylindrical, ascospores $15-17 \times 2.5-3 \mu\text{m}$, pointed at both ends.

Mycosphaerella martinae Hansf., Proc. Linn. Soc. New South Wales 81: 36. 1956.

Type — Australia: *Eucalyptus* (Myrtaceae).

No material studied as the holotype is not in IMI or K.

Sphaerella matthiolae Gonz. Frag., Trab. Mus. Nac. Ci. Nat., Ser. Bot. 9: 100. 1916.— Fig. 537.

Type — Spain: Madrid. On dead stems and fruits of *Matthiola annua* (Brassicaceae). Bolivar no. 1741, II 1916 (MA, holotype).

Already cited as synonymous with *Davidiella allicina* (as *Mycosphaerella allicina*) by Tomilin (1979), with which the holotype agrees well, with asci pyriform, ascospores $19-21 \times 5-6 \mu\text{m}$.

Mycosphaerella matura (Sacc.) Tomilin, Novosti Sist. Nizsh. Rast. 1967: 189. 1967 \equiv *Sphaerella matura* Sacc., Nuovo Giorn. Bot. Ital. 7: 303. 1875.— Fig. 538.

Type — Italy. On upper and lower surface of dead leaves of *Lilium candidum* (Liliaceae). Sacc. (PAD, holotype).

Anamorph: *Phyllosticta liliicola* Sacc. *vide* Saccardo (1882). This belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores 16-20 × 5-6 µm.

Mycosphaerella mauica Petr., Sydowia 7: 391. 1953.— Fig. 539.

Type — Hawaii: Maui, Haua. On brown spots with black margins on upper and lower surface of living leaves of *Cocculus* (Menispermaceae) [as “an unidentified vine”]. Shear & Stevens, I 1928 (W no. 11951, holotype; W no. 5851, isotype).

This is a parasitic species, with asci cylindrical, ascospores 11-13 × 2.5-3.5 µm.

Mycosphaerella maxima Miles, Trans. Illinois State Acad. Sci. 10: 251. 1917 = *Sphaerella maxima* (Miles) Trotter, Syll. Fung. 24: 884. 1928.

Type — Puerto Rico: Maricao. On white spots with brown margins on upper and lower surface of living leaves of a Rubiaceae. Stevens no. 754, IV 1913 (BPI, isotype).

The isotype contains only various coelomycetes, e.g. a *Septoria*.

Mycosphaerella maydina (Pass.) Tomilin, Novosti Sist. Nizsh. Rast. 1966: 172. 1966 = *Sphaerella maydina* Pass., Atti Reale Accad. Lincei, Mem. Cl. Sci. Fis., ser. 4, 3: 9. 1887.

Type — Italy: *Zea mays* (Poaceae).

No material was studied as the type was not found in any of the herbaria consulted.

Mycosphaerella maydis (Pass.) Lindau, in Engl. & Prantl, Natürlichem Pflanzenf. 1(1): 426. 1897 = *Sphaerella maydis* Pass., in Rabenh., Fungi Europaei Exsiccati, ed. nov., ser. 2 no. 1851. 1874 [as “*Sphaerella*”].— Fig. 540.

Type — Italy: Parma. On upper and lower surface of dead leaves of *Zea mays* (Poaceae). Passerini, distributed in Rabenhorst, Fungi Europaei Exsiccati, ed. nov., ser. 2 no. 1851, 1874 (L, isotype).

Already synonymised with *Davidiella allicina* (as *Mycosphaerella tassiana*) by von Arx (1949), with which the isotype studied agrees well, with asci pyriform, ascospores 15-18 × 4-6 µm.

Sphaerella mayteni Speg., Bol. Acad. Nac. Ci. 11: 206. 1888 [“1887”].

Type — Argentina: *Maytenus* (Celastraceae).

No material was studied as the type was not included in a loan from LPS.

Mycosphaerella mazzantioides (Sacc.) Tomilin, Novosti Sist. Nizsh. Rast. 6: 121. 1970 [“1969”] = *Sphaerella mazzantioides* Sacc., Bull. Soc. Mycol. France 12: 65. 1896.

Type — France: *Vitis vinifera* (Vitaceae).

No material was studied as the type was not found in any of the herbaria consulted.

Mycosphaerella medicaginicola Karimov, Bot. Mater. Otd. Sporov. Rast. Bot. Inst. Komarova Akad. Nauk SSSR 11: 120. 1956.

Type — Uzbekistan: *Medicago sativa* (Fabaceae).

Anamorph: *Cercospora medicaginicola* Karimov *vide* Karimov (op. cit.).

No material was studied as the type was not included in loans from LE or LEP.

Mycosphaerella medicaginis Karimov, Bot. Mater. Otd. Sporov. Rast. Bot. Inst. Komarova Akad. Nauk SSSR 11: 119. 1956.

Type — Uzbekistan: *Medicago sativa* (Fabaceae).

Anamorph: *Cercospora medicaginis* Ellis & Everh. *vide* Karimov (op. cit.) (= *Cercospora apii* s.lat.).

No material was studied as the type was not included in loans from LE or LEP.

Mycosphaerella mediterranea (Sacc.) Maire & Werner, Mém. Soc. Sci. Nat. Maroc 45: 27. 1938 = *Sphaerella mediterranea* Sacc., Michelia 1: 35. 1877.

Type — Malta. On white spots on upper surface of living leaves of *Nerium oleandrum* (Apocynaceae).

Anamorph: *Septoria oleandrina* Sacc. *vide* Tomilin (1979).

Authentic material (Malta, Caruana-Gatto, IMI no. 70337) contains only empty ascomata. Additional materials (Algeria, Lachiffa, Bouznad, VI 1979, IMI no. 244675, also Venezuela, Urriaga, II 1969, IMI 138193) contain only the anamorph.

Sphaerella megastoma Peck, Bot. Gaz. (Crawfordsville) 4: 231. 1879 = *Physalosporina megastoma* (Peck) Woron., Ann. Mycol. 9: 220. 1911.— Fig. 541.

Type — USA: Colorado, Colorado Springs. On upper and lower surface of dead, but still attached, leaves of *Astragalus bisulcatus* (Fabaceae). Jones, VI 1879 (NY, isotype), also distributed in Flora of Colorado no. 1573 (NY, isotype).

This is morphologically indistinguishable from *Glomerella cingulata*, with asci cylindrical, surrounded by many 3-5 µm wide paraphyses, ascospores simple, 15-20 × 8-10 µm. It represents an earlier epithet for this species. It has been synonymised with *Stigmatula astragali* (Lasch) P.F. Cannon by Cannon (1996).

Sphaerella melaena (Fr.) Auersw., in Gonn. & Rabenh., Mycol. Europaea 5-6: 16. 1869 = *Sphaeria melaena* Fr., Systema Mycol. 2: 431. 1823 = *Asterina melaena* (Fr.) Sacc., Syll. Fung. 1: 48. 1882 = *Omphalospora melaena* (Fr.) Höhn., Sitzungsber. Kaiserl. Akad. Wiss., Math.-Naturwiss. Kl., Abt. 1, 128: 603. 1919 = *Plectosphaerella melaena* (Fr.) Kirschst., Kryptog.-Fl. Mark Brandenburg 7(3): 310. 1938.

Type — Sweden. On dead stems of *Astragalus glycyphyllus* and *Lathyrus* (Fabaceae). Fries, IX 1823 (B, isosyntype), also distributed in Scleromyceti Suecicae no. 200 (B, isosyntype).

Anamorph: *Podoplaconema melaenum* (Fr.) Petr. *vide* Eriksson (1992).

Accepted as *Omphalospora melaena* (Fr.) Höhn. by Müller & von Arx (1962). The types studied contain only coelomycetes.

Mycosphaerella ["*Mycosphaerium*"] *melaenodes* Clem., Cryptogamae Formationum Coloradensium no. 421. 1908, nomen herbariorum (not validly published, Article 32).

Authentic material — USA: Colorado, Sulphur Springs. On dead stems of *Astragalus junciformis* (Fabaceae). Clements, VII 1907, Cryptogamae Formationum Coloradensium no. 421 (BPI).

This material is postmature.

Sphaerella melaleuca Berk. ex Cooke, J. Bot. 21: 70. 1883. Type — Australia: *Melaleuca* (Myrtaceae).

No material was studied as the type was not found in K and might be lost.

Mycosphaerella melaleucoides Sivan. & R.G. Shivas, Mycol. Res. 106: 359. 2002.

Type — Australia: *Melaleuca quinquenervia* (Myrtaceae).

No material was studied of this recently described species.

Sphaerella melanococca (Lév.) Berl. & Sacc., in Berl. & Voglino, Syll. Fung., Addit. ad vol. 1-4: 406. 1886 = *Sphaeria melanococca* Lév. ex Triana & Planchon, Ann. Sci. Nat. Bot., ser. 4, 20: 296. 1863.— Fig. 542, 963.

Type — Colombia. On upper surface of dead leaves of *Stylosanthes guianensis* (Fabaceae). Lindig (K, holotype; K, isotype).

This is a species of *Dothidotthia*, with ascoma wall cellular (cells 5-10 µm), hamathecium filaments ca. 2 µm wide, asci thick-walled, ascospores dark brown (not red brown), 23-27 × 6-8 µm. It differs from all known species in the genus by the foliicolous habit and the dark ascospores, and therefore the following new combination is proposed here:

Dothidotthia melanococca (Lév. ex Triana & Planchon) Aptroot comb. nov., **MB 500366**. **Basionym:** *Sphaeria melanococca* Lév. ex Triana & Planchon, Ann. Sci. Nat. Bot., ser. 4, 20: 296. 1863.

Mycosphaerella melanophora (Speg.) Tomilin, Novosti Sist. Nizsh. Rast. 11: 257. 1974 = *Sphaerella melanophora* Speg., Atti Soc. Crittog. Ital. 3: 52. 1881.

Type — Italy: *Quercus pseudosuber* ["*pseudoruber*"] (Fagaceae).

No material was studied as the type was not included in a loan from LPS.

Mycosphaerella melanoplaca (Desm.) Johanson ex Oudem., Rév. Champ. Pays-Bas 2: 214. 1897 = *Sphaeria melanoplaca* Desm., Ann. Sci. Nat. Bot., ser. 3, 18: 364. 1852 = *Sphaerella melanoplaca* (Desm.) Auersw., in Gonn. & Rabenh., Mycol. Europaea 5-6: 13. 1869.— Fig. 543.

Type — France: On upper surface of dead leaves of *Geum urbanum* (Rosaceae). Desmazières, Plantes Cryptogames de France, ser. 2, no. 1797 (NY, PC, isotypes).

The isotypes are immature, but suggest that this is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical. Additional material studied (Netherlands, L) confirms that this is *M. punctiformis*, with asci cylindrical, ascospores 7-9 × 2-2.5 µm.

Mycosphaerella melanorhabdos Petr., Sydowia 7: 22. 1953.— Fig. 544.

Type — Greece: Crete, Chania, Platania. On upper and lower surface of dead leaves of *Agave americana* (Asparagaceae). Reehinger, distributed in Reliquiae Petrakianae no. 477, IV 1942 (W, holotype; H, B, isotypes). This is didymosporous *Montagnula* species, with sparse hamathecium, ascospores brownish when old, 10-12 × 3-3.5 µm, surrounded by a 1 µm wide gelatinous sheath. Therefore the following new combination is proposed here: ***Montagnula melanorhabdos*** (Petr.) Aptroot comb. nov., **MB 500369**. **Basionym:** *Mycosphaerella melanorhabdos* Petr., Sydowia 7: 22. 1953. The species is accompanied by *Pleospora herbarum* (Pers. : Fr.) Rabenh.

Mycosphaerella melastomatacearum Bat., Cavalc. & Heringer, in Batista *et al.*, Atas Inst. Micol. 3: 226. 1966 [as "*melastomacearum*"].

Type — Brazil: *Lavoisiera* (Melastomataceae).

No material was studied as the type was not included in a loan from URM.

Mycosphaerella melconiana (Unamuno) Cash, Syll. Fung. 26: 346. 1972 = *Sphaerella melconiana* Unamuno, Assoc. Esp. Progr. Ci. 1929: 12. 1929.

Type — Spain: León, Vegarrienza. On dead leaves of *Holcus mollis* (Poaceae). Unamuno no. 8914-15, IX 1928 (MA, holotype).

Synonymised with *Mycosphaerella badensis* by Tomilin (1979).

Various fungi are present in the holotype, but no ascomycete.

Mycosphaerella meliosmae T.S. Ramakr., Srinivasan & Sundaram, Proc. Indian Acad. Sci., Sect. B, 37: 86. 1953.— Fig. 545.

Type — India: Oatacamund. On white spots with black margins on upper surface of living leaves of *Meliosma wightii* (Sabiaceae). Ramakrishnan & Srinivasan, II 1952 (BPI, isotype).

This is a parasitic species, with asci cylindrical, ascospores 13-15 × 3.5-4.5 µm. Additional material studied (Sri Lanka, Hagala, Botanical Garden, Gams, I 1973, CBS) belongs to *Davidiella*, and is morphologically identical to *D. allicina*, with asci pyriform, ascospores 16-18 × 3.5-4 µm.

Mycosphaerella meliosmae-myrianthae Hara, J. Pl. Protect. 5: 616. 1918.

Type — Japan: *Meliosma myriantha* (Sabiaceae).

No material was studied as the type was not included in a loan from TNS.

Mycosphaerella melonis (Pass.) W.F. Chiu & J.C. Walker, J. Agric. Res. 78: 589. 1949 = *Didymella melonis* Pass., Erbario Crittogamico Italiano, ser. 2, no. 1465. 1885.

Type — Italy: *Cucumis melo* (Cucurbitaceae).

Anamorph: *Ascochyta cucumis* Fautrey & Roum. *vide* Chiu & Walker (op. cit.) (= *Phoma cucurbitacearum* (Fr. : Fr.) Sacc. *vide* Boerema *et al.* 2004).

No material was studied as the type was not found in any of the herbaria consulted. Cited as synonymous with *Didymella bryoniae* (Auersw.) Rehm, which is the current

name for the teleomorph of *Ascochyta cucumis* Fautrey & Roum., by Corlett (1991).

Sphaerella melonis Ferraris, in Ferraris & Massa, Ann. Mycol. 10: 286. 1912.

Type — Italy: *Cucumis melo* (Cucurbitaceae).

No material was studied as the type was not found in any of the herbaria consulted.

Mycosphaerella melothriae T.S. Ramakr., Proc. Indian Acad. Sci., Sect. B, 34: 159. 1951.

Type — India: *Melothria mucronata* (Cucurbitaceae).

No material was studied as the type was not found in IMI or LWG.

Mycosphaerella menthae (Lambotte & Fautrey) Rehm, Ann. Mycol. 4: 409. 1906 = *Sphaerella menthae* Lambotte & Fautrey, in Fautrey & Lambotte, Rev. Mycol. (Toulouse) 17: 170. 1895.— Fig. 546.

Type — France: Pr cy. On dead stems of *Mentha silvestris* (Lamiaceae). Fautrey, V 1895, distributed in Roumeugu re, Fungi Selecti Exsiccati no. 6888 (PC, holotype).

This belongs to *Davidiella* and is morphologically indistinguishable from *D. ammophilae*, with asci pyriform, ascospores 9-11 × 2-2.5 µm. Additional material studied (Austria, Rosenau, Sonntagberg, Strasser, Kryptogamae Exsiccata no. 1150, L) is *M. punctiformis*, with asci cylindrical, ascospores 8-11 × 2-3 µm.

Mycosphaerella mercurialis (Lasch) Magnus, in Dalla Torre & Sarnth., Fl. Tirol, Vorarlberg, Liechtenstein, III, Pilze: 468. 1905, not validly published (Article 32), because basionym invalid = *Sphaerella mercurialis* Lasch, in Rabenh., Herb. Vivum Mycol., cent. 13 no. 1251. 1849, nomen nudum, not validly published (Article 32).— Fig. 547.

Authentic material — Germany: On dead stems and upper surface of dead leaves of *Mercurialis perennis* (Euphorbiaceae).

Material studied (Germany, Kirmiztal, Krieger, Fungi Saxonici no. 2111, V 1910, L) belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradians*, with asci cylindrical, ascospores 11-16 × 2.5-3.5 µm.

Mycosphaerella merrillii H.S. Yates, Philipp. J. Sci. 12: 376. 1917 = *Sphaerella merrillii* (H.S. Yates) Trotter, Syll. Fung. 24: 880. 1928.

Type — Philippines: Alabat Island. On ill-defined white spots on upper and lower surface of living leaves of *Erythralum* (Erythralaceae). Merrill, Philippines Bureau of Science no. 10542, XII 1916 (BPI (2×), K, isotypes).

All isotypes studied contain only an *Asteromella* anamorph with conidia 2-3 × 1 µm.

Mycosphaerella metrosideri F. Stevens & K. Young, in F. Stevens, Bernice P. Bishop Mus. Bull. 19: 104. 1925.— Fig. 548.

Type — Hawaii: Oahu. On white spots with broad brown margins on upper and lower surface of living leaves of

Metrosideros polymorpha (Myrtaceae). Hodges, IX 1970 (BPI, topotype).

This is a parasitic species, with asci cylindrical, ascospores 16-19 × 3-4 µm.

Mycosphaerella mexicana Crous, Mycologia Memoir 21: 81. 1998.— Fig. 549.

Type — Mexico: Puerto Negras, intercepted at Texas, Eagle Pass. On pale spots with black margins on upper and lower surface of living leaves of *Eucalyptus* (Myrtaceae). Sailors, I 1986 (BPI 602163, holotype).

This is a parasitic species, with asci cylindrical, ascospores thick-walled, yellowish, 17-19 × 5-6 µm.

Sphaerella michotii (Westend.) Auersw., in Gonn. & Rabenh., Mycol. Europaea 5-6: 18. 1869 = *Sphaeria michotii* Westend., Bull. Acad. Roy. Sci. Belgique, ser. 2, 7: 87. 1859 = *Leptosphaeria michotii* (Westend.) Sacc., Fungi Italici Autographice Delineati, tab. 279 = *Scleroplella michotii* (Westend.) H hn., Ann. Mycol. 18: 76. 1920 = *Paraphaeosphaeria michotii* (Westend.) O.E. Erikss., Ark. Bot., ser. 2, 6: 406. 1967.

Type — Belgium: *Juncus squarrosus* (Juncaceae).

Anamorph: *Coniothyrium scirpi* Trail *vide* Sivanesan (1984).

Accepted as *Paraphaeosphaeria michotii* (Westend.) O.E. Erikss. by Eriksson (1992) and therefore not studied.

Mycosphaerella miconiae Syd., Ann. Mycol. 23: 340. 1925.— Fig. 550.

Type — Costa Rica. On white spots on upper surface of living leaves of *Miconia argentea* (Melastomataceae).

The type was not included in a loan from S. Materials studied (Dominican Republic, Bonao, La Vega, on *Miconia*, Ciferri, III 1924, B, also Trinidad, Tabaquite forest, Baker, III 1947, IMI no. 17873, also Trinidad, Piarce, Clac no. 94, X 1960, IMI no. 86377) show that this is a parasitic species, with asci cylindrical to slender pyriform, ascospores 16-17.5 × 3-4 µm.

Sphaerella micromeriae Pass., Hedwigia 15: 94. 1876.

Type — Italy: *Micromeria tenuifolia* (Lamiaceae).

No material was studied as the type was not found in any of the herbaria consulted.

Mycosphaerella microscopica (Pass.) Tomilin, Novosti Sist. Nizsh. Rast. 1966: 173. 1966 = *Sphaerella microscopica* Pass., in Brunaud, J. Hist. Nat. Bordeaux 1885: 135. 1885.— Fig. 551.

Type — France: On dead leaves of *Typha angustifolia* (Typhaceae).

The type was not found in any of the herbaria consulted. Material studied (Germany, L ffelstuh!, on *T. latifolia*, XI 1935, Kirschstein, B) belongs to *Davidiella*, and suggests that this is morphologically indistinguishable from *D. disseminata*, with ascomata somewhat aggregated, asci pyriform, ascospores 10-12 × 3.5-4.5 µm.

Mycosphaerella microsora Syd., Ann. Mycol. 38: 465. 1940 = *Sphaerella microsora* (Syd.) Sandu, Ciuperci Pyrenomycetes-Sphaeriales din Rom nia: 135. 1971 [as

“*microspora*”], lacking basionym (not validly published, Article 33.2) possibly also later homonym (illegitimate, Article 53) in the spelling *microspora*.— Fig. 552.

Type — Germany: On upper and lower surface of dead leaves of *Tilia parvifolia* (Malvaceae).

Topotype material studied (Germany, Brandenburg, Lebus, Dahmsdorf, on *Tilia platyphyllos*, Sydow, IV 1941, distributed in Mycotheca Germanica no. 3506, L) belongs to section *Longispora*, and shows that this is morphologically indistinguishable from *M. millegrana*, with asci slender, cylindrical, ascospores 16-19 × 2.5-3.5 µm.

Mycosphaerella microspila (Berk. & Broome) Lind, Danish Fungi: 208. 1913 = *Sphaeria microspila* Berk. & Broome, Ann. Mag. Nat. Hist., ser. 3, 7: 556. 1861 = *Sphaerella microspila* (Berk. & Broome) Cooke, Handb. Brit. Fungi 2: 919. 1871 = *Spilosticta microspila* (Berk. & Broome) Munk, Dansk Bot. Ark. 15(2): 105. 1953, full reference to basionym not given (not validly published, Article 33).— Fig. 553.

Type — United Kingdom: Shere. On upper and lower surface of living leaves of *Epilobium montanum* (Onagraceae). Capron, X 1865 (K, holotype).

Cited as synonymous with *Venturia maculiformis* (Desm.) G. Winter by Sivanesan (1984), with which the type agrees. Additional material studied (Italy, Belluno, Agordo, on upper and lower surface of dead leaves of *Epilobium dodonaei*, Sacc., Mycotheca Italica no. 1657, IX 1905, L) belongs to *Davidiella*, and is morphologically indistinguishable from *D. ammophilae*, with asci pyriform, ascospores 10-12 × 3.5-4.5 µm.

Sphaerella microspora Auersw., in Gonn. & Rabenh., Mycol. Europaea 5-6: 16. 1869 = *Laestadia microspora* (Auersw.) Sacc., Syll. Fung. 1: 424. 1882.

Type — Germany. On upper and lower surface of dead leaves of *Milium effusum* (Poaceae).

Material studied on *Milium* (Frankfurt am Main, Bagge, B) contains a coelomycete; material on *Calamagrostis* (Winter, distributed in Rabenhorst, Fungi Europaei Exsiccati, ed. nov., ser. 2 no. 1646, CBS) contains *Pleospora herbarum* (Pers. : Fr.) Rabenh. and a *Microsphaeropsis*.

Sphaerella microspora (Syd.) Sandu. See *Mycosphaerella microsora* Syd.

Mycosphaerella microspora (Joa.E. Taylor & Crous) Joa.E. Taylor & Crous, Mycol. Res. 107: 657. 2003 = *Teratosphaeria microspora* Joa.E. Taylor & Crous, Mycol. Res. 104: 631. 2000 (as ‘*microsporum*’).

Type — South Africa: *Protea cynaroides* (Proteaceae).

Anamorph: *Trimmatostroma microspora* Joa.E. Taylor & Crous, *vide* Taylor & Crous (2000).

No material was studied of this recently described species.

Mycosphaerella midzurenensis Bubák & Ranoj., in Ranojevič, Ann. Mycol. 8: 360. 1910 = *Sphaerella midzurenensis* (Bubák & Ranoj.) Sacc. & Trotter, Syll. Fung. 22: 132. 1913.

Type — Montenegro: Muglič. On upper and lower surface of dead leaves of *Androsace caerulea* (Primulaceae). Bubák, VIII 1905 (BPI, holotype).

Already synonymised with *Davidiella allicina* (as *Mycosphaerella tassiana*) by von Arx (1949), with which the holotype seems to agree, although no mature asci were found.

Mycosphaerella mikaniae Rehm, Hedwigia 40: 111. 1901 [as “*micaniae*”] = *Sphaerella mikaniae* (Rehm) Sacc. & P. Syd., Syll. Fung. 16: 471. 1902 [as “*micaniae*”].

Type — Brazil: *Mikania* (Asteraceae).

No material was studied as the type was not included in a loan from S.

Mycosphaerella mikaniae-micranthae R.W. Barreto, in R.W. Barreto & H.C. Evans, Mycol. Res. 99: 346. 1995 = *Mycosphaerella mikanifolia* R.W. Barreto, nomen herbariorum (not validly published, Article 32).

Type — Brazil: Rio de Janeiro, Conselheiro Paulino. On white spots on upper surface of living leaves of *Mikania micrantha* (Asteraceae). Barreto, VI 1989 (IMI no. 345381, holotype, sub “*Mycosphaerella mikanifolia*”).

Anamorph: *Septoria mikaniae-micranthae* R.W. Barreto *vide* Barreto (op. cit.).

In the holotype only the coelomycete *Pestalotiopsis* could be found.

Mycosphaerella miliacei Nevod. ex Kalymb., Trudy Inst. Bot. 7: 327. 1959.

Type — Kazakhstan: *Panicum miliaceum* (Poaceae).

No material was studied as the type was not included in loans from LE or LEP.

Mycosphaerella millegrana (Cooke) J. Schröt., in Cohn, Kryptog.-Fl. Schlesien 3(2): 334. 1894 [“1893”] = *Sphaerella millegrana* Cooke, J. Bot. 4: 247. 1866.— Fig. 554.

Type — Austria: On upper and lower surface of dead leaves of *Carpinus betulus* (Betulaceae). Thümen, Fungi Austriaci no. 157 (K, holotype).

Anamorphs: *Cercospora microsora* Sacc. *vide* Cruchet (1923) (= *Passalora microsora* (Sacc.) U. Braun) and *Phyllosticta bacterioides* Vuill. *vide* Tomilin (1979) (= *Asteromella bacterioides* (Vuill.) Moesz).

This belongs to section *Longispora*, with asci cylindrical, ascospores 15-17 × 2-2.5 µm. Additional material studied (Germany, Brandenburg, Prignitz, Triglitz, on upper and lower surface of dead leaves of *Tilia cordata*, Jaap, Fungi Selecti Exsiccati no. 317B, V 1908, L) agrees, but also contains *M. punctiformis*, with asci cylindrical, ascospores 8-9 × 2-2.5 µm.

Sphaerella millegrana f. *carpini* Thüm. See *Sphaerella millegrana* f. *ulmi-effusae* Thüm.

Sphaerella millegrana f. *ulmi-effusae* Thüm., Fungi Austriaci no. 157. 1871 [“*carpini*” in handwriting on labels].— Fig. 555.

Type — Austria: Krems. On lower surface of dead leaves of *Carpinus betulus* (Betulaceae) [“*Ulmus effusus*”]. Thümen, Fungi Austriaci no. 157, IV 1871 (B, K, isotypes).

This is identical to the nominal variety, and belongs to section *Longispora*, with asci cylindrical, ascospores 14-17 × 2-2.5 µm.

Mycosphaerella millepunctata (Desm.) M.E. Barr, Mycologia 62: 382. 1970 = *Dothidea millepunctata* Desm., Ann. Sci. Nat. Bot., ser. 3, 8: 177. 1847.— Fig. 556.

Type — France: On upper and lower surface of dead leaves of *Rhododendron* (Ericaceae). Desmazières, Plantes Cryptogames de France, ser. 3, no. 91 (PC, holotype; B-Desmazières, isotype).

The types studied are immature. Additional material studied (Netherlands, Leusden, Den Treek, van der Aa no. 7022, VIII 1979, CBS) is *M. punctiformis*, with ascomata partly aggregated, asci cylindrical, ascospores 8-11 × 2-2.5 µm.

Sphaerella millepunctata Tassi, Bull. Lab. Orto Bot. Reale Univ. Siena 1: 6. 1897.— Fig. 557.

Type — Italy: Siena, Botanical Garden. On upper and lower surface of dead leaves of *Anigozanthos flavida* (Alliaceae). Tassi, II 1862 (SIENA, holotype).

This is morphologically indistinguishable from *Davidiella allicina*, with asci pyriform, ascospores 18-22 × 4.5-5.5 µm.

Mycosphaerella milleri Hodges & Haasis, Mycologia 56: 53. 1964.— Fig. 558.

Type — USA: South Carolina, Patrick. On lower surface of dead leaves of *Magnolia virginiana* (Magnoliaceae). Miller, V 1962 (IMI no. 201402; K, NY, isotypes).

Anamorph: *Cercosporidium magnoliae* (Ellis & Harkn.) Sivan. *vide* Sivanesan (1984) (= *Passalora magnoliae* (Ellis & Harkn.) U. Braun & Crous).

The isotypes studied show that this belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradians*, with asci cylindrical, ascospores 11-14 × 3.5-4.5 µm.

Mycosphaerella mimosae-pigrae H.C. Evans, G. Carrión & F. Ruiz-Belin, Mycol. Res. 99: 422. 1995.

Type — Mexico: *Mimosa pigra* (Fabaceae).

No material was studied as the type was not found in IMI.

Mycosphaerella mimosicola Henn., Hedwigia 44: 62. 1905 = *Sphaerella mimosicola* (Henn.) Sacc. & D. Sacc., Syll. Fung. 17: 636. 1905.— Fig. 559.

Type — Brazil: Amazonas, Jurua. On lower surface of dead leaves of *Mimosa asperata* (Fabaceae). Ule, Appendix Mycotheca Brasiliensis no. 23, 1901, (L, isotype).

This belongs to section *Fusispora*, with asci pyriform, ascospores 16-19 × 3.5-4.5 µm.

Mycosphaerella minabensis Petr., Sydowia 10: 5. 1957 [“1956”].— Fig. 560.

Type — Iran: Minab. On upper and lower surface of dead leaves of *Pennisetum* (Poaceae). Sharif no. 806, II 1955 (W, holotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores 17-21 × 4-6 µm.

Mycosphaerella minima (Pass.) Petr. & Syd., Ann. Mycol. 22: 358. 1924 = *Laestadia minima* Pass., Atti Reale Accad. Lincei, Mem. Cl. Sci. Fis., ser. 4, 3: 1. 1887.

Type — Italy: *Lythrum salicaria* (Lythraceae).

Anamorph: *Septoria brissaceana* Sacc. & Letell. *vide* Petrak & Sydow (op. cit.).

No material was studied as the type was not found in any of the herbaria consulted.

Mycosphaerella minimipuncta (Cooke) Kapoor & Gill, Indian Phytopathol. 14: 150. 1961, pagination of basionym not provided (not validly published, Article 33) [as “*minimaepuncta*”] = *Sphaerella minimipuncta* Cooke, J. Bot. 21: 136. 1883 [as “*minimaepuncta*”] = *Sphaeria minimipuncta* (Cooke) Ravenel, Fungi Americani Exsiccati no. 681. 1882 [as “*minimaepuncta*”].

Type — USA: South Carolina, Aiken. On dead stems of *Gladiolus* (Iridaceae). Ravenel, VIII 1880 (K, holotype), also distributed in Fungi Americani Exsiccati no. 681 (BPI, K (5×), NY, isotypes).

Cited as synonymous with *M. asteroma*, which is morphologically indistinguishable from *M. subradians* by Tomilin (1979). However, the types contain only coelomycetes (as was already indicated by Tomilin on an annotation slip in NY), e.g. a *Phomopsis*, with 18-22 × 5-7.5 large conidia that may have been mistaken for asci, as sometimes some ascospore-like differentiations can be observed within these conidia.

Mycosphaerella minoënsis Syd. & P. Syd., Ann. Mycol. 11: 58. 1913 = *Sphaerella minoënsis* (Syd. & P. Syd.) Trotter, Syll. Fung. 24: 883. 1928.— Fig. 561.

Type — Japan: Mino, Kawanye-mura. On lower surface of dead leaves of *Rubus* (Rosaceae). Hara, V 1912 (S, holotype).

This belongs to section *Longispora*, and is morphologically indistinguishable from *M. latebrosa*, with asci cylindrical, ascospores 20-24 × 2.5-3 µm.

Mycosphaerella minor (P. Karst.) Johanson, Öfvers. Förh. Kongl. Svenska Vetensk.-Akad. 41(9): 165. 1884 = *Sphaerella minor* P. Karst., Bidrag Kännedom Finlands Natur Folk 23: 171. 1873.

Type — Finland: *Chamaenerion* [as “*Epilobium*”] *angustifolium* (Onagraceae).

No material was studied as the type was not included in a loan from H.

Sphaerella minor var. *galii* Sacc. See *Mycosphaerella galii* (Sacc.) Tomilin.

Sphaerella minor var. *poterii* Parisi, Bull. Orto Bot. Regia Univ. Napoli 9: 58. 1928.

Type — Libya: *Poterium spinosum* (Rosaceae).

No material was studied as the preservation of the type is uncertain.

Mycosphaerella minor var. *reticulata* Dearn., Rep. Canad. Arctic Exped. 1913-1918 4, C: 7. 1923.— Fig. 562.

Type — USA: Alaska, Camden Bay, Kongengevik. On lower surface of dead leaves of *Salix reticulata* (Salicaceae). Johansen, VI 1914 (BPI, isotype).

Cited as synonymous with *Venturia subcutanea* Dearn. by Sivanesan (1977). However, the isotype shows that this belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradians*, with asci cylindrical, ascospores (9-)11-13 × 2.5-3 µm.

Mycosphaerella minuariae Tomilin, Novosti Sist. Nizsh. Rast. 20: 128. 1983.

Type — Russia: *Minuartia arctica* (Caryophyllaceae). No material was studied as the type was not included in loans from LE or LEP.

Sphaerella minuta Auersw., in Gonn. & Rabenh., Mycol. Europaea 5-6: 16.

Type — Germany: *Draba aizoon* (Brassicaceae). No material was studied as the type was not found in B and might be lost.

Sphaerella minutissima Auersw., in Gonn. & Rabenh., Mycol. Europaea 5-6: 3. 1869 ≡ *Laestadia minutissima* (Auersw.) Sacc., Syll. Fung. 1: 426. 1882.

Type — Germany: Grossenhain, Ponickau. On upper and lower surface of dead leaves of *Alnus glutinosa* (Betulaceae). Auerswald, IV 1869 (B, holotype). This is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores immature.

Sphaerella minutissima Peck, Annual Rep. New York State Mus. 40: 68. 1887, later homonym (illegitimate, Article 53).

Type — USA: *Alnus incana* (Betulaceae). This name was possibly meant as a reference to the previous name, but the author was omitted. Included in the synonymy of *Venturia alnea* (Fr.) E. Müll. by Sivanesan (1977) and therefore not studied. Cited as synonymous with *Mycosphaerella harthensis* by Tomilin (1979).

Mycosphaerella minutissima Tomilin. See *Mycosphaerella perparva* House.

Sphaerella mobilis (Tode : Fr.) Ces. & De Not., Comment. Soc. Crittog. Ital. 1(4): 238. 1863 ≡ *Sphaeria mobilis* Tode, Fungi Mecklenburgenses selecti, fasc. 2: 11. 1791, sanctioned by Fr., Systema Mycol. 2: 461. 1823.

Type — Germany: *Quercus* (Fagaceae). No material was studied as the type was not found in B and might be lost.

Mycosphaerella molleriana (Thüm.) Lindau, in Engl. & Prantl, Natürlichen Pflanzenf. 1(1): 424. 1897 ≡ *Sphaerella molleriana* Thüm., Revista Inst. Sci. Lit. Coimbra 28: 31. 1881.— Fig. 363.

Type — Portugal: Coimbra. On upper and lower surface of dead leaves of *Eucalyptus globulus* (Myrtaceae). Moller, distributed in Thümen, Mycotheca Universalis no. 1842, VII 1879 (L, isotype).

Anamorph: *Colletogloeopsis molleriana* Crous & M.J. Wingf. fide Crous (1998).

This is a parasitic species, with asci cylindrical, ascospores 14-16 × 3-3.5 µm, which is about double the size of the

dimensions mentioned in the original publication, and half the size of its var. *megalospora*.

Sphaerella molleriana var. *megalospora* Sousa da Câmara, Bol. Soc. Brot. 25: 9. 1910.

Type — Portugal: *Eucalyptus globulus* (Myrtaceae). No material was studied as the type was not included in a loan from LISE.

Mycosphaerella molluginis (Rehm) Tomilin, Novosti Sist. Nizsh. Rast. 8: 151. 1971 ≡ *Sphaerella molluginis* Rehm, Ann. Mycol. 7: 527. 1909.— Fig. 564.

Type — Germany: Allgäuer Alpen, Hinterstein. On dead stems of *Galium mollugo* (Rubiaceae). Rehm, Ascomyceten no. 1861, 1909 (S, holotype).

Cited as synonymous with *Mycosphaerella cruciatae* by Tomilin (1979), but that turns out to be a *Didymella*.

The type shows that it belongs to section *Caterva*, with ascomata large, asci cylindrical, ascospores 21-24 × 3.5-5 µm.

Mycosphaerella mombin Petr. & Cif., Ann. Mycol. 30: 213. 1932.

Type — Dominican Republic: Santiago, Hato de Yaqui. On white spots with brown margins on upper surface of living leaves of *Spondias mombin* (Anacardiaceae). Ekman, I 1931 (NY, lectotype, here designated).

Only an immature ascomycete is present, but the ascomata are decorated with stout hairs, and it is therefore excluded from *Mycosphaerella*.

Sphaerella monardae Ellis & Everh., North American Fungi, ser. 2 no. 2538. 1891, nomen nudum (not validly published, Article 32).— Fig. 565.

Authentic material — Canada: London. On dead stems of *Monarda* (Lamiaceae). Dearness, distributed in Ellis & Everhart, North American Fungi, ser. 2 no. 2538, V 1890 (L, NY (4×)).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradians*, with asci cylindrical, ascospores 12-15 × 2.5-3.5 µm.

Mycosphaerella monserratica Petr., Ann. Mycol. 29: 112. 1931.— Fig. 566.

Type — Spain: Cataluña, Monserrat. On dead stems of *Aphyllanthes monspeliensis* (Asparagaceae). Ade, V 1929 (W, holotype; NY, isotype).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradians*, with asci cylindrical, ascospores 12-14 × 3-4 µm. Additional material studied (France, Millau, Guyot, VIII 1947, B) agrees.

Mycosphaerella montana Naumov, Zap. Ural'sk. Obshch. Lyubit. Estestv. 35: 21. 1915.

Type — Russia: Ural. On dead stems, calyces and fruits of *Veronica chamaedrys* (Plantaginaceae). Naumov, IX 1913 (LEP, holotype).

The holotype contains only immature ascomata. Cited as synonymous with *Mycosphaerella jurineae* by Tomilin (1979).

Mycosphaerella montellica (Sacc.) Guyot, Bull. Soc. Mycol. France 62: 82. 1946 = *Sphaerella montellica* Sacc., Syll. Fung. 17: 645. 1905.— Fig. 567.

Type — Italy: Treviso, Montello. On upper and lower surface of dead leaves of *Molinia caerulea* (Poaceae). Saccardo, Mycotheca Italica no. 1478, VII 1904 (L, isotype).

Anamorph: Associated with *Cladosporium graminum* var. *molinae* Sacc. fide Saccardo (op. cit.) (= *Cladosporium graminum* var. *molinae-caeruleae* Sacc.).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. disseminata*, with asci small, pyriform, ascospores 12-16 × 3-4.5 μm.

Sphaerella montenegrina Bubák, Bull. Herb. Boissier, ser. 2, 6: 400. 1906.

Type — Montenegro: Košćiele, Rijcky. On upper and lower surface of dead leaves of *Asphodeline lutea* (Xanthorrhoeaceae). Bubák, IV 1903 (BPI, holotype).

Already synonymised with *Davidiella allicina* (as *Mycosphaerella tassiana*) by von Arx (1949), with which the type agrees well, with asci pyriform, ascospores 17-21 × 4.5-6 μm.

Mycosphaerella moquileae Bat., Anais 5 Reun. Anu. Soc. Bot. Brasil 1954: 114. 1956.

Type — Brazil: *Moquilea tomentosa* (Rosaceae).

Anamorph: *Phyllosticta moquileae* Bat. fide Batista (op. cit.).

No material was studied as the type was not included in a loan from URM.

Sphaerella moraeae Pass., Atti Reale Accad. Lincei, Mem. Cl. Sci. Fis., ser. 4, 4: 57. 1888.

Type — Italy: *Moraea chinensis* (Iridaceae).

No material was studied as the type was not found in any of the herbaria consulted.

Mycosphaerella moravica (Petr.) Tomilin, Novosti Sist. Nizsh. Rast. 8: 151. 1971 [as “*moravia*”] = *Mycosphaerellopsis moravica* Petr., Ann. Mycol. 19: 112. 1921.— Fig. 568.

Type — Czech Republic: Weißkirchen. On dead branches of *Lonicera tatarica* (Adoxaceae). Petrak (W no. 10914, holotype), also distributed in Flora Bohemiae et Moraviae Exsiccata no. 1475, II 1921 (B, W (2×), isotypes).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores 15-17 × 5-6.5 μm.

Mycosphaerella mori (Fuckel) F.A. Wolf, J. Elisha Mitchell Sci. Soc. 51: 165. 1935 = *Sphaerella mori* Fuckel, Jahrb. Nassauischen Vereins Naturk. 23-24: 106. 1870, nomen novum (Article 58) for *Sphaeria mori* Nitschke, in Fuckel, Fungi Rhenani no. 1784, nomen nudum (not validly published, Article 32).— Fig. 569.

Type — Germany: Östrich. On lower surface of dead leaves of *Morus alba* (Moraceae). Fuckel, Fungi Rhenani Exsiccata no. 1784 (L, isotype).

Anamorphs: *Cercospora mori* Peck fide Tomilin (1979), *Phloeospora maculans* (Bérenger) Allesch. fide Sivanesan

(1984) and *Phyllosticta osteospora* Sacc. fide Saccardo (1882).

This is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores 8-11 × 2-2.5 μm.

Mycosphaerella mori-albae Sawada, Taiwan Univ. Coll. Agric. Special Publ. 8: 63. 1959.

Type — Taiwan: *Morus alba* (Moraceae).

No material was studied as the type was not found in BPI.

Mycosphaerella moricola Sawada, Special Bull. Agric. Exp. Sta. Gov. Formosa 19: 294. 1919.

Type — Taiwan: *Morus alba* (Moraceae).

No material was studied as the type was not found in BPI.

Mycosphaerella morierei (Crié) Kirchn., Die Krankheiten und Beschädigungen unserer landwirtschaftlichen Kulturpflanzen, ed. 3: 109. 1923 = *Depazea morierei* Crié, Ann. Sci. Nat. Bot., ser. 6, 7: 43. 1878 = *Sphaerella morierei* (Crié) Sacc., Syll. Fung. 1: 504. 1882.

Type — France: Poaceae.

No material was studied as the type was not included in a loan from PC. Cited as synonymous with *Mycosphaerella phaseolicola* by Tomilin (1979).

Mycosphaerella morifolia (Pass.) Cruchet, Bull. Soc. Vaud. Sci. Nat. 55: 43. 1923 = *Sphaerella morifolia* Pass., Erbario Crittogamico Italiano, ser. 2, no. 1464. 1885.— Fig. 570.

Type — Italy: Parma, Villeta. On lower surface of dead leaves of *Morus alba* (Moraceae). Passerini, Erbario Crittogamico Italiano, ser. 2, no. 1464, III 1884 (NY, isotype), also distributed in Roumeguère, Fungi Gallici Exsiccata no. 4054 (NY, isotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores 18-21 × 5-6.5 μm.

Sphaerella morifolia f. *ramicola* D. Sacc., Malpighia 12: 207. 1898.

Type — Italy: *Morus alba* (Moraceae).

No material was studied as the type was not found in any of the herbaria consulted.

Mycosphaerella morindae Sivan. & R.G. Shivas, Mycol. Res. 106: 359. 2002.

Type — Australia: *Morinda citrifolia* (Rubiaceae).

No material was studied of this recently described species.

Mycosphaerella morphaea (Sacc.) Tomilin, Opredelitel' gribov roda *Mycosphaerella* Johans.: 178. 1979 = *Sphaerella morphaea* Sacc., Michelia 1: 34. 1879.

Type — Italy: *Papaver somniferum* (Papaveraceae).

No material was studied as the type was not found in any of the herbaria consulted.

Mycosphaerella morthieri (Fuckel) Petr., Ann. Mycol. 19: 203. 1921 = *Dothidea morthieri* Fuckel, Fungi Rhenani Exsiccata no. 1026. 1863 = *Phyllachora morthieri* (Fuckel) Fuckel, Jahrb. Nassauischen Vereins Naturk. 23-24: 218. 1870 = *Oligostroma morthieri* (Fuckel) Höhn. ex Petr.,

Ann. Mycol. 19: 203. 1921 = *Carlia morthieri* (Fuckel) Höhn., Mitt. Bot. Lab. TH Wien 7(3): 90. 1930.

Type — Switzerland: Jura. On lower surface of dead leaves of *Chaerophyllum aureum* (Apiaceae). Morthier, 1894, distributed in Fuckel, Fungi Rhenani Exsiccati no. 1026 (G, holotype and isotype).

The types contains only a coelomycete, probably a *Libertella*, with conidia ca. $30 \times 1 \mu\text{m}$, which however agrees with the protologue.

Mycosphaerella mougeotiana (Sacc.) Lindau, Hilfsb. Sammeln Ascomyceten: 100. 1903 = *Sphaerella mougeotiana* Sacc., Michelia 2: 57. 1880.— Fig. 571.

Type — France: Vosges. On lower surface of dead leaves of *Rubia peregrina* (Rubiaceae). Mougeot, distributed in Roumeguère, Fungi Selecti Gallici Exsiccati, no. 388 (PAD, holotype, distributed as “*Sphaeropsis atomus*”).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. superflua*, with asci cylindrical, ascospores $16-19 \times 3.5-4.5 \mu\text{m}$.

Mycosphaerella moutan Siemaszko, Acta Soc. Bot. Pol. 1: 20. 1923.— Fig. 572.

Type — Georgia: Adeharskaya, Batumi. On upper surface of dead leaves of *Paeonia moutan* (Paeoniaceae). Siemaszko, IX 1917 (BPI, isotype).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradians*, with asci cylindrical, ascospores $(7-9)-11.5 \times (1.5-2)-2.5-3.5 \mu\text{m}$.

Mycosphaerella mucosa Starbäck, Bih. Kongl. Svenska Vetensk.-Akad. Handl. 25(3, 1): 57. 1899 = *Sphaerella mucosa* (Starbäck) Sacc. & P. Syd., Syll. Fung. 16: 475. 1902 = *Phaeosphaerella mucosa* (Starbäck) Tomilin, Opređelitel' gribov roda *Mycosphaerella* Johans.: 285. 1979.

Type — Brazil: Cyperaceae.

No material was studied as the type was not included in a loan from S.

Mycosphaerella mucunae F. Stevens, Trans. Illinois State Acad. Sci. 10: 182. 1917 = *Sphaerella mucunae* (F. Stevens) Trotter, Syll. Fung. 24: 873. 1928.

Type — Puerto Rico: *Mucuna pruriens* (Fabaceae).

No material was studied as the type was not found in NY or BPI.

Mycosphaerella muelleriana M. Morelet, Bull. Soc. Sci. Nat. Archéol. Toulon & Var 201: 4. 1972, nomen novum (Article 58) for *Mycosphaerella potentillae* (E. Müll.) E. Müll., in E. Müll. & Arx, Beitr. Kryptogamenfl. Schweiz 11(2): 361. 1962, later homonym (illegitimate, Article 53) = *Phaeosphaerella potentillae* E. Müll., Rev. Mycol. (Paris) 19: 57. 1954 = *Leptosphaerulina potentillae* (E. Müll.) Crivelli, Diss. Eidgen. Techn. Hochschule 7318: 133. 1983.

Type — France: *Potentilla valderia* (Rosaceae).

Accepted as *Leptosphaerulina potentillae* (E. Müll.) Crivelli by Crivelli (op. cit.). No material was studied as the species was already redisposed.

Mycosphaerella muhlenbergiae (Ellis) Wehm., Sydowia 6: 423. 1952 = *Sphaerella muhlenbergiae* Ellis, Amer. Naturalist 17: 317. 1883.— Fig. 573.

Type — USA: New Jersey, Newfield. On upper and lower surface of dead leaves of *Muhlenbergia* (Poaceae). VII 1882 (NY, holotype); also Ellis, North American Fungi no. 1352, VI 1882 (L, topotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. ammophilae*, with asci pyriform, ascospores $12-15 \times 4-6 \mu\text{m}$; the holotype contains mostly a coelomycete.

Mycosphaerella mulgedii-alpini Petr., Ann. Mycol. 14: 159. 1916 = *Sphaerella mulgedii-alpini* (Petr.) Trotter, Syll. Fung. 24: 857. 1928.— Fig. 574.

Type — Germany: Glostzer, Schneeberg, Klessengrund. On dead stems of *Mulgedium alpinum* (Asteraceae). Hruba, VI 1913 (W, holotype).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradians*, with asci clavate, ascospores $11-13 \times 2.5-3.5 \mu\text{m}$.

Mycosphaerella multiloculata Anahosur, Sydowia 24: 287. 1971 [“1970”].— Fig. 575.

Type — India: Karnataka, Coorg. On white spots on upper and lower surface of dead leaves of *Memecylon* [“*Memycelon*”] *umbellatum* (Melastomataceae). Anahosur no. 534, X 1966 (LWG, holotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. clandestina*, with ascocarps aggregated, asci pyriform, ascospores $26-30 \times 5-6 \mu\text{m}$.

Mycosphaerella munkii Tomilin, Opređelitel' gribov roda *Mycosphaerella* Johans.: 202. 1979 = *Mycosphaerella psammae* var. *elymifloris* Munk, Dansk Bot. Ark. 17(1): 319. 1957.— Fig. 576.

Type — Denmark: Bøtø. On dead flowers and culm of *Elymus arenarius* (Poaceae). Rostrup, VIII 1880 (C, holotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. disseminata*, with asci pyriform, ascospores $13-15 \times 3.5-4.5 \mu\text{m}$.

Mycosphaerella munyangica Petr., Sydowia 8: 200. 1954.— Fig. 577.

Type — Australia: Munyang Ranges. On upper surface of dead leaves of *Richea continens* (Ericaceae). Gauba, distributed in Reliquiae Petrakianae no. 861, XII 1952 (H, L, isotypes).

This belongs to section *Fusispora*, with asci pyriform, ascospores $21-25 \times 4.5-5.5 \mu\text{m}$. Also present is an immature ascomycete, possibly *Pleospora herbarum* (Pers. : Fr.) Rabenh., with the ascomata pyriform, ascomal wall cellular, asci cylindrical, surrounded by cellular tissue with cells ca. $8 \mu\text{m}$ diam, ascospores hyaline, 1-septate, $15-19 \times 3.5-4.5$.

Mycosphaerella murashkii M. Morelet. See *Mycosphaerella halimodendri* Jacz.

Mycosphaerella murrayae Chona, Munjal & Kapoor, Indian Phytopathol. 10: 148. 1957.

Type — India: On pale spots with brown margins on upper surface of living leaves of *Murraya exotica* (Rutaceae).

Topotype material studied (India, Assam, on *M. koenigii*, Mehrotra, VII 1981, IMI no. 260397) contains only a coelomycete.

Mycosphaerella musae (Speg.) Syd. & P. Syd., Philipp. J. Sci. 8: 482. 1913 = *Sphaerella musae* Speg., Anales Mus. Nac. Hist. Nat. Buenos Aires 19 (ser. 3, 12): 354. 1909.— Fig. 578.

Type — Argentina, Jujuy, Orán. On upper surface of living leaves (not in spots) of *Musa sapientum* (Musaceae). Spegazzini no. 6163, III 1905 (IMI no. 91165, isotype, slide only).

This is a parasitic species, but with asci clavate, ascospores 10-13 × 2-2.5 µm. Additional material studied (Philippines, Mindanao, Prov. Zamboanga, on white spots on upper and lower surface of living leaves, Reyes, distributed in Sydow, Fungi Exotici Exsiccati no. 984, VI 1921, L) agrees.

Sphaerella musae Sacc., Atti Accad. Sci. Veneto, ser. 3, 10: 67. 1917, later homonym (illegitimate, Article 53).

Type — Philippines: Luzon, Prov. Laguna, Los Baños. On dead petioles of *Musa coccinea* (Musaceae). Baker, IX 1915, distributed in Fungi Malayana no. 391 (BPI, isotype). Cited as synonymous with *Mycosphaerella musae* (Speg.) Syd. & P. Sydow by Tomilin (1979). The isotype contains various immature fungi.

Mycosphaerella muscari (Hollós) Tomilin, Novosti Sist. Nizsh. Rast. 1967: 189. 1967 = *Sphaerella muscari* Hollós, Ann. Hist.-Nat. Mus. Natl. Hung. 8: 9. 1910.

Type — Hungary: *Muscari comosum* (Asparagaceae). Cited as synonymous with *Davidiella allicina* (as *Mycosphaerella allicina*) by Tomilin (1979). The type is not in BP and may have been destroyed during the war.

Mycosphaerella musicola R. Leach ex Mulder, in Mulder & Stover, Trans. Brit. Mycol. Soc. 67: 77. 1976 = *Mycosphaerella musicola* R. Leach, Trop. Agriculture 18: 92. 1941, lacking Latin description (not validly published, Article 36.1).— Fig. 579.

Type — Jamaica. On white spots on upper and lower surface of living leaves of *Musa sapientum* (Musaceae). Leach, I 1959 (IMI no. 75804a, holotype).

Anamorph: *Pseudocercospora musae* (Zimm.) Deighton *vide* Sivanesan (1984).

This is a parasitic species, but with asci clavate, ascospores 14-17 × 3-4 µm. Additional materials studied (Fiji, Viti Levu, Knowles, IMI no. 107272; also Surinam, Oort, III 1962, L) is identical.

Sphaerella mutisticola Speg., Revista Fac. Agron. Univ. Nac. La Plata 15 (or ser. 2, 2): 20. 1908.— Fig. 580.

Type — Brazil: São Paulo. On white spots on upper surface of living leaves of *Mutisia* (Asteraceae). Usteri no. 6 (LPS, holotype).

This belongs to *Davidiella*, of which it represents a parasitic species, and is morphologically indistinguishable from *D. lactucae*, with asci globose, ascospores immature, ca. 11 × 3 µm.

Mycosphaerella mycopappi A. Funk & Dorworth, Canad. J. Bot. 66: 295. 1988 = *Didymella mycopappi* (A. Funk & Dorworth) Crous, Mycologia Memoir 21: 152. 1998.

Type — Canada: *Acer macrophyllum* (Sapindaceae). Anamorphs: *Stigmina zilleri* A. Funk (= *Xenostigmina zilleri* (A. Funk) Crous) & *Mycopappus aceris* (Dearn. & Barthol.) Redhead & G.P. White *vide* Funk & Dorworth (op. cit.).

No material was studied as the type was not found in NY or BPI.

Mycosphaerella mycoparasitica Swart, Trans. Brit. Mycol. Soc. 65: 88. 1975.— Fig. 581.

Type — Australia: Victoria, Bogong High Plains. On upper surface of living leaves of *Orites lancifolia* (Proteaceae), reported to be mycoparasitic on *Thallomyces oritidis* (Ascomycota, Microthyriaceae). Ducker, I 1950 (IMI no. 163454, holotype).

This is not a mycoparasite, but the “host” mycelium belongs to the same fungus as the ascomata, as can be deduced from the developmental stages. All ascomata are positioned centrally/concentrically on the mycelium, whereas they would be at least partly acentral if they were parasites, as they would propagate independent from the host. This is a species of Parmulariaceae, with superficial, radial mycelium, asci thick-walled, pyriform, surrounded by densely anastomosing hyphae, ascospores constricted at the septum, 20-23 × 6-7.5 µm.

Mycosphaerella mycopron (Pat.) Tomilin, Novosti Sist. Nizsh. Rast. 1968: 166. 1968 = *Sphaerella mycopron* Pat., Bull. Soc. Mycol. France 36: 177. 1920.

Type — Vietnam: *Phaseolus* (Fabaceae). No type material was preserved in FH, where the Patouillard herbarium is kept.

Sphaerella mygindae G. Winter, Bol. Soc. Brot. 2: 40. 1884.

Type — Portugal: *Myginda pallens* (Celastraceae). No material was studied as the type was not found in B and might be lost.

Sphaerella myriadea (DC. : Fr.) Rabenh., Fungi Europaei Exsiccati, ed. nov., ser. 2 no. 149. 1860 = *Sphaeria myriadea* DC., Fl. Française 6: 145. 1815, sanctioned by Fr., Systema Mycol. 2: 524. 1823 = *Sphaerulina myriadea* (DC. : Fr.) Sacc., Syll. Fung. 2: 186. 1883.— Fig. 582.

Type — France: On upper and lower surface of dead leaves of *Quercus* (Fagaceae).

Accepted as *Sphaerulina myriadea* (DC. : Fr.) Sacc. by Barr (1972), with which material examined (Germany, Driesen, Lasch, distributed in Rabenhorst, Fungi Europaei Exsiccati no. 149, L) agrees well, with asci cylindrical, ascospores 3-septate, 28-33 × 2-3 µm.

Sphaerella myriadea f. *quercus-pedunculatae* Thüm., Mycotheca Universalis no. 2157. 1883.— Fig. 583.

Type — Germany: Driesen. On upper and lower surface of dead leaves of *Quercus pedunculata* (Fagaceae). Lasch, distributed in Thümen, Mycotheca Universalis no. 2157 (L, isotype).

This is morphologically indistinguishable from *Sphaerulina myriadea* (DC. : Fr.) Sacc., with asci cylindrical, ascospores 3-septate, $30\text{--}35 \times 2\text{--}3 \mu\text{m}$.

Sphaeria myriadea var. *carpini* Desm., Ann. Sci. Nat. Bot, ser. 2, 19: 359. 1838.

Type — France: On lower surface of dead leaves of *Carpinus betulus* (Betulaceae). Desmazières, Plantes Cryptogames de France no. 1294A (BPI, isotype).

The isotype studied is immature, but probably belongs to *Sphaerulina myriadea*.

Sphaeria myriadea var. *fagi* Desm., Plantes Cryptogames de France no. 1294B. 1843.

Type — France: On lower surface of dead leaves of *Carpinus betulus* (Betulaceae). Desmazières, Plantes Cryptogames de France no. 1294B (BPI, isotype).

The isotype studied is immature, but probably belongs to *Sphaerulina myriadea*.

Sphaerella myriadea var. *fagi* Roum., Fungi Selecti Gallici Exsiccati no. 2293. 1882, later homonym (illegitimate Article 53).

Type — France: Seine-et-Marne, Fontainebleau. On upper and lower surface of dead leaves of *Fagus sylvatica* (Fagaceae). Roumeguère, Fungi Selecti Gallici Exsiccati no. 2293, I 1882 (L, isotype).

The isotype studied is immature, but probably belongs to *Sphaerulina myriadea*.

Sphaerella myricae Ellis & Everh., J. Mycol. 4: 103. 1888, nomen nudum (not validly published, Article 32).

Authentic material — USA: *Myrica cerifera* (Myricaceae). No material was studied as it was not found in NY.

Mycosphaerella myricae Miles, Mycologia 18: 165. 1926.

Type — USA: Missouri, Vaneleave. On ill-defined brown spots with white margins on upper surface of dead leaves of *Myrica carolinensis* (Myricaceae). Miles no. 502, III 1921 (NY, isotype).

This is a species of Micropeltidaceae with its anamorph.

Mycosphaerella myricae Sawada, Rep. Gov. Res. Inst. Formosa 85: 33. 1943, lacking Latin description (not validly published, Article 36.1) and later homonym (illegitimate, Article 53).

Authentic material — Taiwan: Taipeh. On rusty brown spots with black margins on upper surface of living leaves of *Myrica rubra* (Myricaceae). Sawada, XII 1914 (BPI).

The authentic material contains only a coelomycete.

Mycosphaerella myricariae (Fuckel) Lindau, Hilfsb. Sammeln Ascomyceten: 70. 1903 = *Sphaeria myricariae* Fuckel, Jahrb. Nassauischen Vereins Naturk. 27-28: 22. 1873 = *Mycosphaerellopsis myricariae* (Fuckel) Höhn. ex Petr., Ann. Mycol. 19: 114. 1921.— Fig. 584.

Type — Switzerland: Ragaz. On dead branches and upper and lower surface of dead leaves of *Myricaria germanica* (Tamaricaceae). Fuckel, Fungi Rhenani Exsiccati no. 2437 (L, isotype).

This is the type of *Mycosphaerellopsis* Höhn. Although still accepted as *Mycosphaerellopsis myricariae* (Fuckel) Höhn.

ex Petr. by Barr (1996), it is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores $17\text{--}20 \times 4.5\text{--}6.5 \mu\text{m}$.

Sphaerella myrsines Kalchbr. & Cooke, Grevillea 9: 30. 1880.— Fig. 585.

Type — South Africa: Cape of Good Hope. On lower surface of living leaves of *Myrsine africana* (Myrsinaceae). Kalchbrenner no. 1348 (K, holotype).

This is a species of Microthyriaceae, probably an *Asterinella*, with ascoma wall cells meandering, asci slender pyriform, ascospores $8\text{--}10 \times 2\text{--}2.5 \mu\text{m}$.

Mycosphaerella myrticola (Speg.) M. Morelet, Ann. Soc. Sci. Nat. Archéol. Toulon & Var 20: 106. 1968 = *Sphaerella myrticola* Speg., Bol. Acad. Nac. Ci. 25: 58. 1921.— Fig. 586.

Type — Chili: Los Perales. On white spots on upper surface of living leaves of *Myrtus chequen* (Myrtaceae). Spegazzini no. 6176, 1918 (LPS, holotype).

This belongs to *Davidiella*, of which it represents a parasitic species, with asci pyriform, ascospores $15\text{--}17 \times 4.5\text{--}5.5 \mu\text{m}$. Therefore the following new combination is made: **Davidiella myrticola** (Speg.) Aptroot comb. nov., **MB 500356**. **Basionym:** *Sphaerella myrticola* Speg., Bol. Acad. Nac. Ci. 25: 58. 1921.

Sphaerella myrtilli Auersw. See *Mycosphaerella vaccinii* (Cooke) J. Schröt.

Mycosphaerella myrtillina (Pass.) Tomilin, Opredelitel' gribov roda *Mycosphaerella* Johans.: 294. 1979 = *Sphaerella myrtillina* Pass., Rev. Mycol. (Toulouse) 2: 33. 1880.

Type — Italy: *Vaccinium myrtillus* (Ericaceae).

No material was studied as the type was not found in any of the herbaria consulted.

Mycosphaerella mysorensis V.S. Seshadri, Ph.D Thesis, University of Poona, 1967, cited in Subhedar & V.G. Rao, Indian J. Mycol. Pl. Pathol. 15: 183. 1985, apparently unpublished (Article 29).— Fig. 587.

Type — India: Maharashtra. On dead, still corticate, branches of *Colebrookia oppositifolia* (Lamiaceae). Subhedar no. 3491, III 1974 (LWG, holotype).

This is morphologically indistinguishable from *Diaporthe eres* Nitschke, with ascomata immersed, asci cylindrical and gelatinizing, ascospores $9\text{--}11 \times 3\text{--}3.5 \mu\text{m}$.

Mycosphaerella najas (Sacc.) Tomilin, Novosti Sist. Nizsh. Rast. 1967: 189. 1967 = *Sphaerella najas* Sacc., Nuovo Giorn. Bot. Ital. 7: 304. 1875.— Fig. 588.

Type — Italy. On dead efflorescence of *Juncus articulatus* ["*lamprocarpus*"] (Juncaceae). Treviso, IX 1875 (PAD, holotype).

This belongs to *Davidiella* and is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores $15\text{--}19 \times 5\text{--}7.5 \mu\text{m}$. Additional material studied (Netherlands, Baarn, on upper and lower surface of dead leaves, van der Aa no. 8675, V 1983, CBS) belongs to section *Fusispora*, and is morphologically indistinguishable

from *M. alpina*, with asci pyriform, ascospores 24-28 × 4.5-5.5 µm.

Sphaerella napicola Fautrey, Rev. Mycol. (Toulouse) 12: 64. 1890.— Fig. 589.

Type — France: On dead stems of *Brassica napa* var. *oleifera* (Brassicaceae). Fautrey no. 13, 1890 (L, isotype). Although it was synonymised with *Mycosphaerella cruciferarum* by von Arx (1949), it is morphologically indistinguishable from *Davidiella allicina*, with asci pyriform, ascospores 19-22 × 5-7 µm.

Mycosphaerella nawae M. Hiura & Ikata, in M. Hiura, Res. Bull. Gifu Univ. Fac. Agric. 5(34): 2. 1929.— Fig. 590.

Type — Japan: Fukushima, Asaka-Gun, Tomida-Mura. On brown spots with black margins on upper surface of living leaves of *Diospyros kaki* var. *hachiyu* (Ebenaceae). Kawada, IX 1929 (B, IMI no. 127137, isotypes).

The isotypes contain nothing identifiable, only young leaf spots. Additional material (India, Gorakhpur, University Campus, on *D. tomentosa*, Singhaniano. 218, V 1978, IMI no. 228162a) is a parasitic species, with asci cylindrical, ascospores 14-17 × 3-4 µm.

Mycosphaerella nebulosa (Pers. : Fr.) Johanson ex Oudem., Rév. Champ. Pays-Bas 2: 213. 1897 = *Sphaeria nebulosa* Pers., Observ. Mycol. 2: 69. 1800 [“1799”], sanctioned by Fr., Systema Mycol. 2: 430. 1823 = *Exormatostoma nebulosa* (Pers. : Fr.) S.F. Gray, A natural Arrangement of British Plants 1: 522. 1821 = *Sphaeropsis nebulosa* (Pers. : Fr.) Fr., Summa Veget. Scand., sect. post. 1849 = *Phoma nebulosa* (Pers. : Fr.) Berk., Outlines of British Fungology: 314. 1860 = *Sphaerella nebulosa* (Pers. : Fr.) Sacc., Michelia 2: 56. 1880.

Type — Netherlands: On dead stems of *Urtica dioica* (Urticaceae). Persoon (L, lectotype, Boerema 1976). Accepted as the coelomycete *Phoma nebulosa* (Pers. : Fr.) Berk. by Boerema (1976), with which the lectotype agrees well. The species was originally described from several hosts. Additional material studied from other hosts (Czech Republic, Weißkirchen, Hrabuvka, on dead stems of *Solidago virgaurea*, Petrak, distributed in Reliquiae Petrakianae no. 2687, VI 1924, L) belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradians*, with asci cylindrical, ascospores 9-11 × 2-3 µm.

Sphaeria nebulosa var. *epilobii* Pers., nomen herbariorum (not validly published, Article 32).

Authentic material — France: On dead stems of *Epilobium* (Onagraceae). Chaillet (L-Persoon). This is immature.

Sphaerella nebulosa var. *euphorbiae* Sacc. & Briard, in Briard, Rev. Mycol. (Toulouse) 7: 208. 1885.

Type — France: *Euphorbia palustris* (Euphorbiaceae). No material was studied as the type was not found in any of the herbaria consulted.

Sphaerella nebulosa var. *hieracii* Sacc. & Briard. See *Mycosphaerella hieracii* (Sacc. & Briard) Jaap.

Sphaeria nebulosa var. *minor* Desm., Plantes Cryptogames de France no. 1265. 1843.

Type — France: On dead stems of *Reseda luteola* (Resedaceae). Desmazières, Plantes Cryptogames de France no. 1265 (BPI, isotype).

The isotype contains only a coelomycete (not *Phoma nebulosa*, but rather an *Asteromella*).

Sphaeria nebulosa var. *nigrescens* Pers., nomen herbariorum (not validly published, Article 32).— Fig. 591.

Authentic material — France: Paris, on herbaceous stems. Chaillet, VIII 1822. Persoon (L-Persoon).

This is a specimen of *Glomerella cingulata* (Stoneman) Spauld. & H. Schrenk, with ascospores not septate, 9-11 × 3-3.5 µm.

Sphaerella nebulosa var. *scrophulariae* Sacc. & Briard. See *Mycosphaerella scrophulariae* (Sacc. & Briard) Tomilin.

Sphaerella nebulosa-veneta De Not., in Anzi, Erbario Crittogamico Italiano, fasc. 25, no. 284 (1283). 1865 = *Laestadia nebulosa-veneta* (De Not.) Sacc., Syll. Fung. 1: 428. 1882 [as “*nebulosa*”].— Fig. 592.

Type — Italy: Bolzano. On dead stems of *Peucedanum venetum* (Apiaceae). Hausmann, distributed Erbario Crittogamico Italiano, no. 284 (1283), 1863 (RO, holotype [sub “*veneta*”]; B, 2 isotypes).

Cited as synonymous with *Guignardia foeniculacea* (Mont.) von Arx & E. Müll. by von Arx & Müller (1954). However, the holotype shows that this belongs to *Davidiella* and is morphologically indistinguishable from *D. ammophilae*, with asci pyriform, ascospores immature, 8-10 × 2-3 µm.

Mycosphaerella nectandrae (F. Stevens) Cif., Sydowia 10: 148. 1957 [“1956”] = *Guignardia nectandrae* F. Stevens, Bot. Gaz. (Crawfordsville) 69: 255. 1920.

Type — Puerto Rico: On pale brown spots with black margins on upper surface of living leaves of *Nectandra coriacea* (Lauraceae).

Material seen (Dominican Republic, Prov. Santiago, Cordillera Cental, San José de las Matas, on *N. sintenisii*, Ciferri, Mycoflora Domingensis Exsiccata no. 404, VII 1931, IMI no. 59527) contains only a coelomycete, not the anamorph of a *Mycosphaerella*.

Mycosphaerella nemesiae Dippenaar, S. African J. Sci. 28: 287. 1931.

Type — South Africa: *Nemesia* (Plantaginaceae). No material was studied as the type was not found in any of the herbaria consulted.

Mycosphaerella nemorosa (Sacc. & Speg.) Woron., Trudy Tiflissk. Bot. Sada, ser. 2, 3: 138. 1923 = *Sphaerella nemorosa* Sacc. & Speg., in Sacc., Michelia 1: 378. 1878.— Fig. 593.

Type — Italy. On pale brown spots with black margins on upper surface of dead leaves of *Lathyrus* [“*Orobus*”] *vernus* (Fabaceae). Spegazzini, 1876 (PAD, holotype).

This belongs to *Davidiella*, of which it represents a parasitic species, with asci pyriform, ascospores 16-20 × 5-7 µm. Therefore the following new combination is made:

Davidiella nemorosa (Sacc. & Speg.) Aptroot comb. nov., **MB 500357**. **Basionym:** *Sphaerella nemorosa* Sacc. & Speg., in Sacc., *Michelia* 1: 378. 1878.

Mycosphaerella nemoseridis Fairm., in Millsp. & L. Nutt., *Publ. Field Columbian Mus., Bot. Ser.* 5: 348. 1923.

Type — USA: *Rafinesquia* ["*Nemoseris*"] *californica* (Asteraceae).

No material was studied as the type was not found in any of the herbaria consulted.

Mycosphaerella nerii-odori A.K. Pande, *Maharashtra V.M. Patrika* 10: 9. 1975 [as "*nerium-odorum*"].— Fig. 594.

Type — India: Pune. On upper and lower surface of dead leaves of *Nerium odorum* (Apocynaceae). Pande no. 508, IX 1966 (LWG, holotype).

Anamorph: *Cercospora nerii-indici* W. Yamam. *fide* Sivanesan (1984) (= *Pseudocercospora kurimensis* (Fukui) U. Braun).

This is a parasitic species, with asci broad-cylindrical, ascospores 13-15 × 3-4 µm.

Sphaerella nervicola Auersw., in Gonn. & Rabenh., *Mycol. Europaea* 5-6: 11. 1869.

Type — Germany: *Rubus corylifolius* (Rosaceae).

No material was studied as the type was not found in B and might be lost.

Mycosphaerella nerviseda (Speg.) Lindau, *Hilfsb. Sammeln Ascomyceten*: 62. 1903 = *Sphaerella nerviseda* Speg., *Michelia* 1: 456. 1879.

Type — Italy: *Lathyrus* ["*Orobus*"] *vernus* (Fabaceae).

No material was studied as the type was not included in a loan from LPS.

Sphaerella nerviseda var. *microspora* Sacc., *Atti Reale Ist. Veneto Sci.*, ser. 6, 2: 456. 1884.

Type — Switzerland. On lower surface of dead leaves of *Lathyrus* ["*Orobus*"] *vernus* (Fabaceae). Morthier, III 1875 (PAD, holotype).

Anamorphs: Associated with *Septoria orobicola* Sacc. and *Phyllosticta orobella* Sacc. *fide* Saccardo (op. cit.).

On the type only the anamorphs were found.

Mycosphaerella nevodovskii Tomilin, *Novosti Sist. Nizsh. Rast.* 1966: 148. 1966, nomen novum (Article 58) for *Mycosphaerella bromi-albidis* Nevod., *Fungi Caucasici* no. 756. 1912, nomen herbariorum (not validly published, Article 32) [as "*bromi-albidi*"].

Type — Georgia: On upper and lower surface of dead leaves of *Bromus albidus* (Poaceae). Nevodovski, *Fungi Caucasici* no. 756 (LE 34960, lectotype, here designated; LE 34961, isotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores 15-19 × (4-)5-6 µm.

Mycosphaerella nicotianae (Ellis & Everh.) Miles, *Trans. Illinois State Acad. Sci.* 10: 250. 1917 = *Sphaerella nicotianae* Ellis & Everh., *Proc. Acad. Nat. Sci. Philadelphia* 1895: 420. 1895.

Type — USA: California, Hollister. On bark of woody stems of *Nicotiana* (Solanaceae). Blasdale no. 297, XII 1894 (NY, holotype; NY, 2 isotypes).

The types are mostly immature, but show that this belongs to *Davidiella*, and is morphologically indistinguishable from *D. ammophilae*, with asci pyriform, ascospores tiny (immature). In the type materials *Pleospora herbarum* is also frequent.

Sphaerella niesslii Auersw., in Niessl, *Verh. Naturf. Vereins Brünn* 10: 171. 1872 ["1871"].

Type — Germany: *Lycopodium* (Lycopodiaceae).

No material was studied as the type was not found in B and might be lost. Cited as synonymous with *Mycosphaerella lycopodina* by Tomilin (1979).

Mycosphaerella niesslii Tomilin. See *Mycosphaerella intermixta* Lindau.

Mycosphaerella nigerristigma B.B. Higgins, *Mycol. Centralbl.* 4: 192. 1913 = *Sphaerella nigerristigma* (B.B. Higgins) Trotter, *Syll. Fung.* 24: 883. 1928.

Type — USA: On pale spots with brown margins on upper surface of living leaves of *Prunus pensylvanica* (Rosaceae). Anamorph: *Septoria higginsii* Traverso *fide* Trotter (op. cit.).

No type material has been preserved in BPI and it is probably lost. Material studied (Wisconsin, Dane Co., Madison, Greene no. 2606, IX 1960, BPI) contains only the *Septoria* anamorph.

Mycosphaerella nigredo (Schwein.) Lindau, in Engl. & Prantl, *Natürlichen Pflanzenf.* 1(1): 425. 1897 = *Sphaeria nigredo* Schwein., *Trans. Amer. Philos. Soc.*, ser. 2, 4: 225. 1832 = *Sphaerella nigredo* (Schwein.) Cooke, *J. Bot.* 21: 109. 1883.

Type — USA: Philadelphia, Bethlehem. On lower surface of dead leaves of *Rhus glabra* (Anacardiaceae). Schweinitz (PH, holotype).

This is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores tiny but immature.

Mycosphaerella nigrificata (Fautrey & Roum.) Lavrov, *Trudy Tomsk. Gosud. Univ.* 110: 47. 1951 = *Sphaerella nigrificata* Fautrey & Roum., in Roum., *Rev. Mycol. (Toulouse)* 13: 166. 1891.

Type — France: Côte-d'Or, Noidan. On dead leaves of *Agrostis stolonifera* (Poaceae). Fautrey, distributed in Roumeguère, *Fungi Selecti Exsiccati* no. 5831, IV 1891 (PC, holotype).

Synonymised with *Davidiella allicina* (as *Mycosphaerella tassiana*) by von Arx (1949), with which the description agrees. The holotype now contains only empty ascomata.

Mycosphaerella nigrita (Cooke) J.H. Miller, *Mycologia* 33: 333. 1941 = *Sphaerella nigrita* Cooke, *Grevillea* 7: 13. 1878.— Fig. 595.

Type — USA: New York, Poughkeepsie. On lower surface of dead leaves of *Quercus* (Fagaceae). Gerard, distributed in

Ellis, North American Fungi no. 160 (K, holotype; K, 2 isotypes).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradians*, with ascocarps aggregated in dense groups, asci cylindrical, ascospores 12-15 × 3-4 µm.

Sphaerella nigrita Roum., Fungi Gallici Exsiccati no. 1606. 1881, nomen herbariorum (not validly published, Article 32) and later homonym (illegitimate, Article 53).

Authentic material — France: Morvan. On dead stems of *Valeriana sambucifolia* (Caprifoliaceae). Grognot, 1881, distributed in Roumeguère, Fungi Gallici Exsiccata no. 1606 (L).

This contains only coelomycetes.

Mycosphaerella nigromaculans Shear, in Shear, N.E. Stevens & Bain, Techn. Bull. U.S.D.A. 258: 11. 1931.— Fig. 596.

Type — USA: Oregon, Clatsop. On dead stems of *Vaccinium macrocarpum* (Ericaceae). Bain, I 1925 (BPI, holotype; BPI, IMI no. 226278 (slide only), isotypes).

Anamorph: *Ramularia nigromaculans* Shear *vide* Sivanesan (1984) (= possibly a species of *Ramularia* *vide* Braun, 1998).

This belongs to section *Mycosphaerella*, with asci cylindrical, ascospores 6-7 × 1.5-2 µm.

Mycosphaerella nipponica Hara, J. Pl. Protect. 5: 461.

Type — Japan.

No material was studied as the type was not included in a loan from TNS.

Sphaerella nitidula (Lév.) Berk., Fungi Braz. no. 420 = *Sphaeria nitidula* Lév., Ann. Sci. Nat. Bot., ser. 3, 3: 54. 1845 = *Dothidea nitidula* (Lév.) Cooke, J. Bot. 21: 107. 1883.

Type — French Guiana: Cayenne. On lower surface of living leaves of *Melastoma* (Melastomataceae). Léveillé (K, holotype).

This is a species of *Microthyrium*, with ascomata superficial, conical, up to 1 mm diam. Ascospores were not observed.

Sphaerella nivalis (F. Bauer) Sommerf., Mag. Naturvidensk. 4: 252. 1824.

This is a member of the Chlamydomonadaceae (Chlorophyceae): an algae growing on the surface of snowbeds.

Mycosphaerella nivalis Jacz., Opredelitel' gribov 2: 616. 1917, nomen novum (Article 58) for *Sphaerella nivalis* Oudem., Verslagen Meded. Afd. Natuurk. Kon. Akad. Wetensch., ser. 3, 2: 156. 1885, later homonym (illegitimate, Article 53).

Type — Russia: *Ranunculus nivalis* (Ranunculaceae).

No material was preserved in L, and the type is probably lost.

Sphaeria nivalis Fuckel, in Heuglin, Reisen nach dem Nordpolarmeer 3: 93. 1874 = *Didymella nivalis* (Fuckel) Berl. & Voglino, Syll. Fung., Addit. ad vol. 1-4: 88. 1886.

Type — Greenland: *Chamaenerion* ["*Epilobium*"] *latifolium* (Onagraceae).

Synonymised with *Davidiella allicina* (as *Mycosphaerella tassiana*) by von Arx (1949). No material was found in any of the herbaria consulted.

Mycosphaerella nogalesii Urries, Anales Inst. Bot. Cavanilles 14: 160. 1955 [as "*nogalesi*"].— Fig. 597.

Type — Canary Islands: La Palma, Baranco de la Galga. On white spots on upper surface of living leaves of *Cytisus stenopetalus* (Fabaceae). Urries no. 12936, IV 1954 (MA, holotype).

This is a parasitic species, with asci cylindrical, ascospores 12-14 × 2.5-3.5 µm.

Mycosphaerella nothofagi (Speg.) Spauld., Foreign Diseases of Forest Trees of the World, U.S. Dept. Agric., Agric. Handbook 197: 171. 1961 = *Sphaerella nothofagi* Speg., Anales Mus. Nac. Hist. Nat. Buenos Aires 19 (ser. 3, 12): 352. 1909 [as "*notofagi*"].

Type — Argentina: *Nothofagus* (Nothofagaceae). No material was studied as the type was not included in a loan from LPS.

Cited as synonymous with *Mycosphaerella taxodii* by Tomilin (1979).

Sphaerella novae-hollandiae Tassi, Bull. Lab. Orto Bot. Reale Univ. Siena 3: 94. 1900.

Type — Australia: Sydney. On upper surface of dead fronds of *Pteris* (Polypodiaceae). 1900 (SIENA, holotype).

The holotype is extremely scanty, and now contains only a *Pestalotiopsis*.

Sphaerella nubigena Speg., Rev. Mycol. (Toulouse) 4: 79. 1882.

Type — France: *Draba aizoides* (Brassicaceae).

No material was studied as the type was not included in a loan from LPS. Synonymised with *Mycosphaerella pyrenaica* by von Arx (1949).

Mycosphaerella nubilosa (Cooke) Hansf., Proc. Linn. Soc. New South Wales 81: 36. 1956 = *Sphaerella nubilosa* Cooke, Grevillea 19: 61. 1891.— Fig. 598.

Type — Australia: Victoria, Melbourne. On brown spots on upper surface of living leaves of *Eucalyptus* (Myrtaceae). Martin no. 584, 1891 (IMI no. 59210, isotype; K, 4 isotypes).

Anamorph and spermatial state: *Colletogloeum nubilosum* Ganapathi & Corbin and *Asteromella* *vide* Sivanesan (1984).

This is a parasitic species, with asci cylindrical, ascospores 13-15 × 3-4 µm. Additional material (New Zealand, Auckland, Ianapathi, on *E. delegatensis*, Baker, IV 1977, IMI no. 228671) is the isotype of the anamorph which it indeed contains.

Mycosphaerella nuristanica Petr., Sydowia 7: 90. 1953.— Fig. 599.

Type — Afghanistan: Nuristan, Kamdesch. On dead stems of *Lonicera* (Adoxaceae). Zilli, VI 1950 (W, holotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores $15-18 \times 4.5-5.5 \mu\text{m}$.

Mycosphaerella nyssicola (Cooke) F.A. Wolf, Mycologia 32: 333. 1940 [as "*nyssaecola*"] = *Sphaerella nyssicola* Cooke, Hedwigia 17: 40. 1878 [as "*nyssoccola*"].— Fig. 600.

Type — USA: South Carolina, Aiken. On lower surface of dead leaves of *Nyssa multiflora* (Cornaceae). Ravenel (K, holotype), also distributed in Fungi Americani Exsiccati no. 96 (BPI, isotype).

Anamorph: *Phyllosticta nyssae* Cooke fide Wolf (op. cit.).

The types are totally postmature, even overgrown with hyphae of basidiomycetes, but the ascomata suggest that is is morphologically indistinguishable from *M. punctiformis*. Additional material studied (USA, North Carolina, Durham, on *N. sylvatica*, Wolf, distributed in Reliquiae Farlowianae no. 931, III 1939, B) is *M. punctiformis*, with asci cylindrical, ascospores $6-8 \times 2-2.5 \mu\text{m}$.

Sphaerella oblivia Cooke, J. Bot. 4: 246. 1866 = *Sphaerella maculiformis* var. *oblivia* (Cooke) Auersw., in Gonn. & Rabenh., Mycol. Europaea 5-6: fig. 7. 1869 [as "*maculaeformis* var."].— Fig. 601.

Type — United Kingdom: Kent, Darenth. On lower surface of dead leaves of *Castanea vesca* (Fagaceae). Cooke, Fungi Britannici Exsiccati, no. 693 (K, 10 isotypes).

Anamorph: *Septoria castanicola* Desm. fide Cooke (op. cit.).

This belongs to section *Caterva* and is morphologically indistinguishable from *M. subradians*, with asci cylindrical, ascospores $12-15 \times 2.5-3.5 \mu\text{m}$. Additional material examined (Cooke, Fungi Britannici Exsiccati, ed. 2, no. 262 (B, IMI no. 16777) is *M. punctiformis*.

Mycosphaerella occulta Bubák, Ann. Mycol. 13: 26. 1915 = *Sphaerella occulta* (Bubák) Trotter, Syll. Fung. 24: 861. 1928.

Type — Czech Republic: *Rhododendron ponticum* (Ericaceae).

Anamorph: Associated with *Phyllosticta occulta* Bubák fide Bubák (op. cit.).

No material was studied as the type was not found in BPI and is therefore probably lost.

Mycosphaerella octopetalae (Oudem.) Jacz., Opredelitel' gribov 2: 613. 1917 = *Sphaerella octopetalae* Oudem., Verslagen Meded. Afd. Natuurk. Kon. Akad. Wetensch., ser. 3, 2: 159. 1885.

Type — Russia: On upper and lower surface of dead leaves and on dead bracts of *Dryas octopetala* (Rosaceae).

Synonymised with *Davidiella allicina* (as *Mycosphaerella tassiana*) by von Arx (1949), cited as synonymous with *Mycosphaerella dryadis* by Tomilin (1979). The type could not be found in L, and is probably lost. Material seen (United Kingdom, Scotland, Sutherland, Loch Eriboll, Ard Neakie, Cannon, V 1985, IMI no. 312720) contains various ascomycetes, including a *Sporormia*, but no *Mycosphaerella*.

Mycosphaerella ["*Mycosphaerium*"] *octopetalae* var. *major* ["*majus*"] Clem., Cryptogamae Formationum Coloradensium no. 223. 1906, nomen herbariorum (not validly published, Article 32).

Authentic material — USA: Colorado, Bottomless Pit. On upper and lower surface of dead leaves of *Dryas octopetala* (Rosaceae). Clements, VII 1906, Cryptogamae Formationum Coloradensium no. 223 (BPI).

The material is postmature.

Mycosphaerella oculata Syd. & P. Syd., Ann. Mycol. 11: 403. 1913 = *Sphaerella oculata* (Syd. & P. Syd.) Trotter, Syll. Fung. 24: 892. 1928.— Fig. 602.

Type — Philippines: Luzon, Prov. Laguna, Los Baños. On white spots on upper surface of living leaves of *Premna odorata* (Verbenaceae). Reyes no. 1532, VIII 1913 (S, holotype).

This is morphologically indistinguishable from *Davidiella clandestina*, although the material is badly preserved and asci were not observed, but characteristic of this species are the large and wide ascospores $22-27 \times 4-6.5 \mu\text{m}$.

Mycosphaerella oedema (Fr.) J. Schröt., in Cohn, Kryptog.-Fl. Schlesien 3(2): 335. 1894 ["1893"] = *Sphaeria oedema* Fr., in Duby, in DC., Botan. Gallicum, ed. 2, 2: 696. 1830 = *Sphaerella oedema* (Fr.) Fuckel, Jahrb. Nassauischen Vereins Naturk. 23-24: 104. 1870.— Fig. 603.

Type — France: Vosges. On lower surface of dead leaves of *Ulmus campestris* (Ulmaceae). Mougeot & Nestler no. 880 (L, isotype), also distributed in Roumeguère, Fungi Gallici Exsiccati no. 361 (L, isotype).

This is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores $8-10 \times 2.5-3 \mu\text{m}$.

Sphaerella oenanthicola Fuckel, Jahrb. Nassauischen Vereins Naturk. 23-24: 102. 1870 = *Laestadia oenanthicola* (Fuckel) Sacc., Syll. Fung. 1: 431. 1882.

Type — Germany: Oestrich, Altersand. On dead stems of *Oenanthe* ("*Phellandrium*") *aquatica* (Apiaceae). Fuckel, Fungi Rhenani Exsiccati no. 1574 (G, holotype; G, isotype [sub "*perpusilla*"]).

This belongs to section *Caterva* and is morphologically indistinguishable from *M. subradians*, with asci cylindrical, ascospores immature.

Mycosphaerella oenotherae (Ellis & Everh.) Lindau, in Engl. & Prantl, Natürlichen Pflanzenf. 1(1): 424. 1897 = *Sphaerella oenotherae* Ellis & Everh., J. Mycol. 1: 151. 1885.— Fig. 604.

Type — USA: New Jersey, Newfield. On dead stems and fruits of *Oenothera biennis* (Onagraceae). Ellis, VII 1885 (NY, holotype), also distributed in Ellis & Everhart, North American Fungi no. 1681 (L, NY, 3 isotypes).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. superflua*, with asci cylindrical, ascospores $14-16 \times 3.5-4.5 \mu\text{m}$.

Mycosphaerella oerteliana (Sacc.) Migula, in Thomé, Fl. Deutschl. Österr. Schweiz. X, 1. Kryptog.-Fl., III, 3(1): 285.

1912 [“1913”] = *Sphaerella oerteliana* Sacc., Ann. Mycol. 2: 528. 1904.— Fig. 605.

Type — Germany: Thüringen, Sondershausen. On dead stems of *Coronilla montana* (Fabaceae). Oertel, VI 1904, distributed in Sydow, Mycotheca Germanica no. 237 (L, isotype).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradicans*, with asci cylindrical, ascospores 10-12 × 2.5-3 µm.

Mycosphaerella ohnowa Crous & M.J. Wingf. in Crous *et al.*, Stud. Mycol. 50: 206. 2004.

Type — South Africa: Mpumalanga, Hazy View, on leaves of *Eucalyptus grandis* (Myrtaceae). Wingfield, III 1995 (PREM 51912, holotype); cultures ex-type CPC 1004 = CBS 112896, CPC 1005 = CBS 112973, CPC 1006 = CBS 110949.

No material was studied of this recently described species.

Mycosphaerella oleandri S. Ahmad, Biologia (Lahore) 17: 83. 1971.

Type — Pakistan: *Nerium odorum* (Apocynaceae).

No type material was preserved in ZT.

Mycosphaerella oleina (Cooke) J.H. Miller, Mycologia 33: 80. 1941 = *Sphaerella oleina* Cooke, J. Bot. 21: 107. 1883.— Fig. 606.

Type — USA: Georgia, Darien. On white spots with black margins on upper surface of living leaves of *Olea americana* (Oleaceae). Ravenel, III 1881 (K, 5 isotypes).

Anamorph: *Phyllostica oleina* Cooke *fide* Cooke (op. cit.).

This is a parasitic species, with asci cylindrical, ascospores 11-14 × 3-4 µm.

Sphaerella olenjana Sacc., Syll. Fung. 9: 619. 1891, nomen novum (Article 58) for *Sphaerella saxifragae* P. Karst., Meddeland. Soc. Fauna Fl. Fenn. 14: 87. March 1887, later homonym (illegitimate, Article 53).

Type — Finland: *Saxifraga rivularis* (Saxifragaceae).

No material was studied as the type was not included in a loan from H. Synonymised with *Davidiella allicina* (as *Mycosphaerella tassiana*) by von Arx (1949).

Mycosphaerella olindensis Petr., Sydowia 7: 391. 1953.— Fig. 231.

Type — Hawaii: Maui, Olinda Pipe Line. On dead rhachis of *Pteridium* [“*Pteris*”] *excelsa* (Polypodiaceae). Shear & Stevens, XII 1927 (BPI, isotype).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. filicum*, with asci cylindrical, ascospores 11.5-14 × 3-3.5 µm.

Mycosphaerella omalanthi Syd. & P. Syd., Ann. Mycol. 18: 100. 1920 [as “*homalanthi*”] = *Sphaerella omalanthi* (Syd. & P. Syd.) Trotter, Syll. Fung. 24: 863. 1928 [as “*homalanthi*”].

Type — Philippines: *Omalanthus* [“*Homalanthus*”] *alpinus* (Asteraceae).

No material was studied as the type was not included in a loan from S.

Mycosphaerella omphalosporoides Petr., Ann. Mycol. 23: 71. 1925.— Fig. 607.

Type — Czech Republic: Weißkirchen, Podhorn. On upper and lower surface of dead leaves of *Melica uniflora* (Poaceae). Petrak (W, holotype), also distributed in Flora Bohemiae et Moraviae Exsiccata no. 2007, V 1924 (W, isotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. disseminata*, with asci pyriform, ascospores 10-11 × 3-3.5 µm.

Mycosphaerella onobrychidis (Hollós) Tomilin, Novosti Sist. Nizsh. Rast. 1968: 166. 1968 = *Sphaerella onobrychidis* Hollós, Növényt. Közlem. 6: 2. 1907.

Type — Hungary: *Onobrychis sativa* (Fabaceae).

The type is not in BP and may have been destroyed during the war.

Sphaerella ononidis Kirschst., nomen herbariorum (not validly published, Article 32).— Fig. 608. Authentic material — Germany: Heilbrunn. On dead stems of *Ononis spinosa* (Fabaceae). Kirschstein, VI 1939 (B).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradicans*, with ascomata large, asci cylindrical, ascospores 10-12 × 2.5-3.5 µm.

Mycosphaerella ontariensis R. Stone, Phytopathology 5: 6. 1915 = *Sphaerella ontariensis* (R. Stone) Trotter, Syll. Fung. 24: 873. 1928.— Fig. 609.

Type — Canada: Ontario, Guelph. On dead stems and fruit pods of *Lathyrus sativus* (Fabaceae). Stone, X 1912 (NY, holotype); also XI 1912 (NY, 2 topotypes).

Anamorph: *Ascochyta ontariensis* R. Stone *fide* Stone (op. cit.).

According to Tomilin (1979), this is no *Mycosphaerella*. However, the type shows that this belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradicans*, with ascomata large, asci cylindrical, ascospores 11-13 × 3-4.5 µm.

Mycosphaerella ootheca (Sacc.) Magnus, in Dalla Torre & Sarnth., Fl. Tirol, Vorarlberg, Liechtenstein, III, Pilze: 465. 1905 = *Sphaerella ootheca* Sacc., Michelia 2: 160. 1880.— Fig. 610.

Type — USA: Alaska. On upper surface of dead leaves of *Dryas octopetala* (Rosaceae). Harrington, Trelease, 1871-1872 (PAD, holotype).

This belongs to *Wettsteinina*, and is morphologically indistinguishable from *W. dryadis* (Rostr.) Petr., for which it would provide an earlier epithet, but it is probably immature, with ascospores 20-23 × 8-11 µm. Previously cited as synonymous with the heterotypic *Mycosphaerella dryadis* by Tomilin (1979).

Mycosphaerella operculata (Sacc.) J.C. Gilman & Wadley, Mycologia 44: 219. 1952 = *Sphaerella operculata* Sacc., Nuovo Giorn. Bot. Ital., ser. 2, 27: 76. 1920.— Fig. 611.

Type — USA: Oregon, Grants Pass. On small pale spots with brown margins on upper and lower surface of living leaves of *Quercus chrysolepis* (Fagaceae). Weir no. 10007, IX 1916 (BPI, lectotype, here designated); also Weir no.

10048, IX 1916 (BPI, paratype); also Weir, IX 1919 (BPI, topotype).

This is a species of *Planistromella*, with ascomata embedded in massive pseudosromata, lifting off parts of the (stiff) epidermis (obviously a character induced by the host-interaction), asci cylindrical, surrounded by 4-12 µm large cells, ascospores 11-12.5 × 2.5-3 µm. Therefore the following combination is proposed: **Planistromella operculata** (Sacc.) Aptroot comb. nov., **MB 500374**. **Basionym:** *Sphaerella operculata* Sacc., Nuovo Giorn. Bot. Ital., ser. 2, 27: 76. 1920.

Sphaerella ophiopogonis Sacc., Atti Soc. Veneto-Trentina Sci. Nat. 2: 142. 1873 ≡ *Leptosphaeria ophiopogonis* (Sacc.) Sacc., Syll. Fung. 2: 68. 1883.

Type — Italy: *Ophiopogon japonicus* (Asparagaceae).

No material was studied as the type was not found in any of the herbaria consulted.

Mycosphaerella opuntiae (Ellis & Everh.) Dearn., in Dearn. & House, New York State Mus. Bull. 205-206: 55. 1919 ≡ *Sphaerella opuntiae* Ellis & Everh., J. Mycol. 4: 97. 1888 ≡ *Monographella opuntiae* (Ellis & Everh.) Arx, Trans. Brit. Mycol. Soc. 83: 374. 1984.— Fig. 612.

Type — USA: Louisiana, St. Martinsville. On bark of dead stems of *Opuntia* (Cactaceae). Langlois no. 1261, IV 1888 (NY, holotype).

Anamorph: *Microdochium lunatum* (Ellis & Everh.) Arx *vide* von Arx (op. cit.).

This belongs to section *Longispora*, and is morphologically indistinguishable from *M. latebrosa*, with asci cylindrical, ascospores 20-27 × 3-3.5 µm, with obtuse ends. Additional material (USA, Texas, Perdinales Falls State Park, Sutton, IV 1980, IMI no. 293691) agrees.

Mycosphaerella orbicularis (Peck) House, New York State Mus. Bull. 233-234: 28. 1921 ≡ *Sphaerella orbicularis* Peck, Annual Rep. New York State Mus. 28: 81. 1876 ≡ *Venturia orbicularis* (Peck) M. Morelet, Soc. Franç. Phytopath., Travaux dédiés à G. Viennot-Bourgien: 260. 1977 ≡ *Phaeosphaerella orbicularis* (Peck) Tomilin, Oprelitel' gribov roda *Mycosphaerella* Johans.: 285. 1979.— Fig. 613, 986.

Type — USA: New York, Albany Co., Center. On upper surface of dead leaves of *Populus grandidentata* (Salicaceae). Peck no. 2722, V 1874 (BPI, isotype).

Anamorph: *Pollaccia radiosus* (Lib.) E. Bald. & Cif. *vide* Sivanesan (1984) (= *Fusicladium radiosum* (Lib.) Lind).

Cited as synonymous with *Venturia macularis* (Fr. : Fr.) E. Müll. & Arx by Sivanesan (1977). However, the type shows that this is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores 7.5-9 × 2-3 µm.

Mycosphaerella orchidearum (P. Karst.) Tomilin, Novosti Sist. Nizsh. Rast. 1967: 189. 1967 ≡ *Sphaerella orchidearum* P. Karst., Acta Soc. Fauna Fl. Fenn. 2(6): 68. 1885.

Type — Finland: *Gymnadenia conopsea* (Orchidaceae).

No material was studied as the type was not included in a loan from H.

Sphaerella oreoselini Auersw., in Gonn. & Rabenh., Mycol. Europaea 5-6: 15. 1869, nomen nudum (illegitimate, Article 32).

Authentic material — Germany: *Oreoselinum* (Apiaceae).

No material was studied as the type was not found in B and might be lost. Cited as synonymous with *Sphaerella umbelliferarum* Rabenh. by Auerswald (op. cit.).

Sphaerella ornithogali (Grognot) Roum., Fungi Selecti Exsiccati no. 4443. 1888 ≡ *Sphaeria ornithogali* Grognot, Plantes cryptog. cellulaires du département de Saône-et-Loire, avec descriptions de plusieurs espèces nouvelles. 1863.

Type — France: *Ornithogalum* (Alliaceae).

No material was studied as the type was not included in a loan from PC.

Sphaerella ornithogali f. *funkiae* Roum., Fungi Selecti Exsiccati no. 4443. 1888.

Type — Belgium: Brussels, Meise, Botanical Garden. On dead branches of *Funkia lancifolia* (Alliaceae). Marchal, distributed in Roumeguère, Fungi Selecti Exsiccati no. 4443 (NY, isotype).

The isotype contains only a coelomycete.

Mycosphaerella orobi (Hazsl.) Lindau, Hilfsb. Sammeln Ascomyceten: 62. 1903 ≡ *Sphaerella orobi* Hazsl., Math. Természettud. Közlem. 25: 105. 1892.

Type — Hungary: *Lathyrus* ["*Orobus*"] *vernus* (Fabaceae).

The type is not in BP and may have been destroyed during the war.

Mycosphaerella orobi Karakulin. See *Mycosphaerella karakulinii* Tomilin.

Sphaerella orontii Ellis & Everh., J. Mycol. 1: 151. 1885.

Type — USA: On *Orontium aquaticum* (Araceae).

The type was not found in NY or BPI, and should be considered lost.

Mycosphaerella ["*Mycosphaerium*"] *orthospora* ["*orthosporum*"] Clem., Cryptogamae Formationum Coloradensium no. 423. 1908, nomen herbariorum (not validly published, Article 32).

Authentic material — USA: Colorado, Sulphur Springs. On dead stems of *Pentstemon procerus* (Plantaginaceae). Clements, VII 1907, Cryptogamae Formationum Coloradensium no. 423 (BPI).

The material is postmature.

Mycosphaerella oryzae (Garov. & Catt.) Jacz., Oprelitel' gribov 2: 615. 1917 ≡ *Pleospora oryzae* Garov. & Catt., Rendiconti Reale Ist. Lombardo Sci., ser. 2, 7: 155. 1874 ≡ *Sphaerella oryzae* (Garov. & Catt.) Sacc., Syll. Fung. 1: 527. 1882.— Fig. 614.

Type — Italy: Pavese. On upper and lower surface of dead leaves of *Oryza sativa* (Poaceae). Cattaneo, distributed in Rabenhorst, Fungi Europaei Exsiccati no. 2450 (L, isotype).

Anamorphs: *Ascochyta* and *Phoma* *vide* Tomilin (1979).

This is morphologically indistinguishable from *Glomerella cingulata* (Stoneman) Spauld. & H. Schrenk, with asci

clavate, dextrinoid, ascospores not septate, 12-15 × 4-5 µm, for which it would present an older epithet.

Mycosphaerella oryzopsidis (Ellis & Everh.) Coons, Rep. Michigan Acad. Sci. 14: 236. 1912 [as “*oryzopsidis*”] ≡ *Sphaerella oryzopsidis* Ellis & Everh., Proc. Acad. Nat. Sci. Philadelphia 45: 134. 1893 [as “*oryzopsis*”].— Fig. 615.

Type — USA: Michigan, Lansing. On dead leaves, mostly immersed between upper and lower cortex, of *Oryzopsis asperifolia* (Poaceae). Hicks no. 573, V 1892 (NY, holotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. disseminata*, with asci pyriform, ascospores 14-18 × 3-3.5 µm.

Mycosphaerella osborniae D. Hawksw. & Sivan., Trans. Brit. Mycol. Soc. 67: 42. 1976.— Fig. 616.

Type — United Kingdom: Devon, Slapton Ley Nature Reserve. On dead stems of *Artemisia vulgaris* (Asteraceae). Hawksworth no. 3566, VIII 1973 (IMI no. 178281a, holotype).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradians*, with asci cylindrical, ascospores 10-12 × 2-3 µm.

Mycosphaerella osmundicola (Kirschst.) M. Morelet, Ann. Soc. Sci. Nat. Archéol. Toulon & Var 20: 106. 1968 ≡ *Sphaerella osmundicola* Kirschst., Hedwigia 81: 194. 1944.— Fig. 617.

Type — Germany: Westfalen, Kreis Olpe, Silberg. On upper surface of dead fronds of *Osmunda regalis* (Osmundaceae). Ludwig, VI 1940 (B, lectotype, designated here; BPI (2×), isolectotypes).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. filicum*, with ascomata tiny (only 60-90 µm and with just 5-10 asci in one ascoma), asci cylindrical, ascospores 11-12.5 × 2.5-4 µm.

Sphaerella ostruthii (Fr. : Fr.) Cooke, Handb. Brit. Fungi 2: 922. 1871 ≡ *Sphaeria ostruthii* Fr., Observ. Mycol. Fl. Suecicam 1: 174. 1815, sanctioned by Fr., Systema Mycol. 2: 526. 1823.

Type — Sweden. On upper and lower surface of dead leaves of *Imperatoria ostruthium* (Apiaceae). Fries, Scleromyceti Suecici no. 4 (UPS, holotype; B, isotype).

The types studied contain only a coelomycete.

Sphaerella othonnopsidis Tassi, Bull. Lab. Orto Bot. Reale Univ. Siena 1899: 27. 1899.— Fig. 618.

Type — Italy: Siena, Botanical Garden. On upper and lower surface of dead leaves of *Othonnopsis cheirifolia* (Asteraceae).

Anamorph: probably *Phoma sordida* Durieu & Mont. *fide* Tassi (op. cit.).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores 17-22 × 4.5-6.5 µm.

Mycosphaerella oxalidis (Rabenh.) Magnus, Oesterr. Bot. Z. 44: 203. 1894 ≡ *Carlia oxalidis* Rabenh., in Klotzsch, Herbarium Vivum Mycologicum, ed. nov., no. 567. 1857 ≡

Laestadia oxalidis (Rabenh.) Sacc., Syll. Fung. 1: 429. 1882 ≡ *Sphaerella carlii* Fuckel, Jahrb. Nassauischen Vereins Naturk. 23-24: 103. 1870, superfluous (illegitimate, Article 52) nomen novum (Article 58).— Fig. 619.

Type — Germany: Bohemia, Schluckenau. On white to brownish spots with brown margins on upper and lower surface of living leaves of *Oxalis acetosella* (Oxalidaceae). Karl, distributed in Rabenhorst, in Klotzsch, Herbarium Vivum Mycologicum, ed. nov., no. 567 (B, L, isotypes).

This is the type species of *Carlia* Rabenh. The isotype in B, as well as additional material studied (Germany, Oberbayern, Oberammergau, Mereschkowitz, Allescher & Schnabl, Fungi Bavarici no. 338, VI 1894, L) show that this is a parasitic species, and is morphologically indistinguishable from *M. depazeaeformis*, with asci cylindrical, ascospores 10-14 × 2.5-3.5 µm. The presence of a brown epispore as mentioned in the protologue could not be confirmed.

Mycosphaerella oxalidis Sawada, J. Taihoku Soc. Agric. 7: 126. 1942, lacking Latin description (not validly published, Article 36) and later homonym (illegitimate, Article 53).— Fig. 620.

Authentic material — Taiwan: Taipei, Urai. On white spots with brown margins on upper surface of living leaves of *Oxalis repens* (Oxalidaceae). Sawada, IV 1915 (BPI).

This is a parasitic species, and is morphologically indistinguishable from *M. depazeaeformis*, with asci cylindrical, ascospores 10-14 × 2.5-3.5 µm.

Mycosphaerella oxyacanthae Jaap, Abh. Bot. Vereins Prov. Brandenburg 49: 15. 1907 ≡ *Sphaerella oxyacanthae* (Jaap) Sacc. & Trotter, Syll. Fung. 22: 129. 1913 ≡ *Carlia oxyacanthae* (Jaap) Höhn., Hedwigia 62: 57. 1920.— Fig. 621.

Type — Germany: Brandenburg, Prignitz, Triglitz. On upper and lower surface of dead leaves of *Crataegus oxyacantha* (Rosaceae). Jaap, Fungi Selecti Exsiccati no. 188, VIII 1906 (L, 2 isotypes).

Anamorph: *Phloeospora oxyacanthae* (G. Kunze & Schmidt) Wallr. *fide* Sivanesan (1984).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. superflua*, with asci cylindrical, ascospores 15-20 × 3-4 µm.

Mycosphaerella oxycocci Dearn. & House, New York State Mus. Bull. 243-244: 76. 1923 [“1921”].— Fig. 622.

Type — USA: New York, Essex Co., Newcomb. On lower and upper surface of dead leaves of *Vaccinium macrocarpum* [“*Oxycoccus macrophyllus*”] (Ericaceae). House, VII 1920 (BPI, isotype).

Anamorph: *Phyllosticta elongata* Weid. *fide* Sivanesan (1984).

Already cited as synonym of *Guignardia vaccinii* Shear (as *Botryosphaeria vaccinii* (Shear) M.E. Barr) by Sivanesan (1984), with which the isotype agrees well, with asci clavate, stipitate, not surrounded by hamathecium filaments, ascospores simple, 12-14 × 4-5 µm.

Sphaerella oxydendri F.A. Wolf. See *Mycosphaerella caroliniana* (F.A. Wolf) J.H. Miller.

Mycosphaerella oxyriae Savile, in Savile & Parmelee, Canad. J. Bot. 42: 715. 1964.

Type — Canada: Ellesmere Island, Hazen Camp. On upper surface of dead leaves of *Oxyria digyna* (Polygonaceae). Savile no. 4629, VII 1962 (IMI no. 200600, isotype, slide only).

Anamorph: *Cercoseptoria oxyriae* (Trail) Gjaerum fide Sivanesan (1984) (= *Pseudocercospora oxyriae* (Trail) U. Braun).

Cited as synonymous with *Davidiella allicina* (as *Mycosphaerella allicina*) by Tomilin (1979), with which the isotype slide agrees well, with asci pyriform. Additional material (Franklin, Somerset Island, Savile no. 3567B, VII 1958, IMI no. 200601) agrees as well.

Mycosphaerella pachyasca (Rostr.) Vestergr., Bih. Kongl. Svenska Vetensk.-Akad. Handl. 26(3, 12): 7. 1900 = *Sphaerella pachyasca* Rostr., Meddel. Grønland 3: 552. 1888.— Fig. 623.

Type — Greenland: Ilivnek. On upper and lower surface of dead leaves of *Arabis holboellii* (Brassicaceae). Warming & Hertz, VII 1884 (C, lectotype, designated here).

Previously synonymised with *Davidiella allicina* (as *Mycosphaerella tassiana*) by von Arx (1949), but the lectotype indicates that this is morphologically indistinguishable from *D. clandestina*, with asci pyriform, ascospores (18)22-26 × 5.5-8 µm. The lectotype was selected among 19 syntypes in C (all from Greenland, all belonging to the same species but from different localities and hosts), as this was the specimen from which Rostrup made a drawing of the asci and ascospores. Additional material studied (USA, California, Mariposa Co., Bagby, on *Eriogonum tripodum* (Polygonaceae), Howell no. 29887, V 1954, B) agrees.

Mycosphaerella pachyasca var. *alpina* (Ferraris) Moesz, Balkán-Kutat. Tud. Eredm. 3: 137. 1926 = *Sphaerella pachyasca* var. *alpina* Ferraris, Malpighia 16: 450. 1902.

Type — Italy: *Armeria alpina* (Plumbaginaceae).

No material was studied as the type was not found in any of the herbaria consulted.

Sphaerella pachyasca var. *ribicola* Gonz. Frag., Bol. Soc. Esp. Hist. Nat. 18: 368. 1918.— Fig. 624.

Type — Spain: Segovia, S. Rafael. On dead bark of living branches of *Ribes uva-crispa* (Grossulariaceae). Bolivar no. 3189, VI 1918 (MA, holotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores 17-20 × 5-6 µm.

Mycosphaerella pachypleuri (Fuckel) Jacz., Opredelitel' gribov 2: 615. 1917 = *Sphaerella pachypleuri* Fuckel, in Heuglin, Reisen nach dem Nordpolarmeer 3: 319. 1874.

Type — Novaya Zemlya: Kostin Schar. On dead stems of *Pachypleurum alpinum* (Apiaceae). (G, holotype).

The holotype contains only overmature fungi, mostly *Heteropatella lacera* Fuckel.

Mycosphaerella pachysandrae (Hemmi) Katum. & Harada, Trans. Mycol. Soc. Japan 28: 31. 1987 [as "*Mycosphaerella*"]

= *Cyclodothis pachysandrae* Hemmi, Bot. Mag. (Tokyo) 29: 415. 1915.

Type — Japan: *Pachysandra terminalis* (Buxaceae).

No material was studied as the type was not included in a loan from TNS.

Mycosphaerella pachystimae Dearn. See *Mycosphaerella paxystimae* Dearn.

Mycosphaerella pachythechia (Speg.) Tomilin, Novosti Sist. Nizsh. Rast. 14: 127. 1977 = *Sphaerella pachythechia* Speg., Rev. Fac. Agron. Univ. Nac. La Plata, ep. 2, 6: 56. 1910.

Type — Chile: *Cryptocarya peumus* (Lauraceae).

No material was studied as the type was not included in a loan from LPS.

Mycosphaerella padina (P. Karst.) Jørst., Meld. Stat. Plantepatol. Inst. 1: 87. 1945 = *Sphaerella cerasina* subsp. *padina* P. Karst., Hedwigia 23: 2. 1884.

Type — Finland: *Tavastia australis*, Tammela, Ikkris. *Prunus padus* (Rosaceae). Karsten no. 4576, V 1865 (H, holotype).

Anamorph: *Septoria* fide Jørstad (op. cit.).

This is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores immature.

Mycosphaerella paepalanthi Rehm, Hedwigia 40: 110. 1901 = *Sphaerella paepalanthi* (Rehm) Sacc., Syll. Fung. 16: 474. 1902.— Fig. 625.

Type — Brazil: Serra Geral. On upper surface of dead leaves of *Paepalanthus* (Eriocaulaceae). Ule no. 1748, II 1891 (S, holotype).

This is morphologically indistinguishable from *Davidiella clandestina*, with asci pyriform, ascospores 25-30 × 6-8 µm.

Mycosphaerella paleicola (Henn.) Tomilin, Novosti Sist. Nizsh. Rast. 1966: 173. 1966 = *Sphaerella paleicola* Henn., Deutsche Südpolar-Exped. 1901-1903, Bot., Pilze 8: 9. 1907 ["1906"].

Type — Kerguelen Island: *Festuca erecta* (Poaceae).

No material was studied as the type was not found in SP or B and is probably lost.

Cited as synonymous with *M. recutita*, which is morphologically indistinguishable from *Davidiella disseminata*, by Tomilin (1979).

Mycosphaerella pales (Sacc.) Tomilin, Novosti Sist. Nizsh. Rast. 1967: 189. 1967 = *Sphaerella pales* Sacc., Nuovo Giorn. Bot. Ital. 7: 305. 1875.

Type — Italy. On dead stems of *Hemerocallis fulva* (Xanthorrhoeaceae). Saccardo (PAD, holotype).

The type contains only a coelomycete. Cited as synonymous with *Davidiella allicina* (as *Mycosphaerella allicina*) by Tomilin (1979).

Mycosphaerella palmae Miles, Trans. Illinois State Acad. Sci. 10: 252. 1917 = *Sphaerella palmae* (Miles) Trotter, Syll. Fung. 24: 881. 1928.— Fig. 626.

Type — Puerto Rico: Guanica. On upper and lower surface of dead leaves of *Arecaceae*. Stevens no. 9107, VII 1915 (K, NY, isotypes).

This belongs to *Davidiella* and is morphologically indistinguishable from *D. ammophilae*, with asci pyriform, ascospores $12-15 \times 4-5.5 \mu\text{m}$.

Mycosphaerella palmicola Chaudhury & P.N. Rao, Mycopathol. Mycol. Appl. 22: 221. 1964.

Type — India: *Cocos nucifera* (Arecaceae).

No material was studied as the type was not found in LWG or IMI.

Sphaerella paludosa Ellis & Everh., North American Fungi no. 2357. 1890, nomen nudum (not validly published, Article 32).— Fig. 627.

Authentic material — Canada: London. On white spots on upper and lower surface of living leaves of *Nuphar advena* (Nymphaeaceae). Dearness, VIII 1889, distributed in Ellis & Everhart, North American Fungi no. 2357 (L, NY).

This belongs to *Davidiella*, of which it represents a parasitic species, with asci pyriform, ascospores $14-18 \times 3-4.5 \mu\text{m}$. Already cited as morphologically indistinguishable from *Davidiella pontederiae* (as *Mycosphaerella pontederiae*) by Barr (1972).

Sphaerella palustris (Fr.) Cooke, Grevillea 18: 86. 1890 = *Sphaeria palustris* Fr., in Duby, in DC., Botan. Gallicum, ed. 2, 2: 710. 1830 = *Laestadia palustris* (Fr.) Sacc., Syll. Fung. 9: 577. 1891.

Type — France: *Caltha palustris* (Ranunculaceae).

No material was studied as the species was already redisposed.

Sphaerella pampini Thüm., Pilze des Weinstockes: 27. 1878 = *Leptosphaeria pampini* (Thüm.) Sacc., Syll. Fung. 2: 31. 1883.

Type — Italy: *Vitis vinifera* (Vitaceae).

No material was studied as the type was not found in any of the herbaria consulted.

Mycosphaerella panacis Bunkina, in Bunkina & Bondartsev, Bot. Mater. Otd. Sporov. Rast. Bot. Inst. Komarova Akad. Nauk SSSR 13: 131. 1960 = *Mycosphaerella panacis-ginseng* Tomilin, Novosti Sist. Niz Rast. 7: 204. 1970 [as “*panacis-ginsengi*”], superfluous (illegitimate, Article 52) nomen novum (Article 58).

Type — Russia: *Panax ginseng* (Araliaceae).

No material was studied as the type was not included in loans from LE or LEP. Cited as synonymous with *D. allicina* (as *Mycosphaerella allicina*) by Tomilin (1979).

Mycosphaerella panacis (Cooke) Tomilin, Novosti Sist. Niz Rast. 7: 204. 1970, later homonym (illegitimate, Article 53) = *Sphaerella panacis* Cooke, J. Bot. 21: 138. 1883.— Fig. 629.

Type — New Zealand: South Island, Bealy Gorge. On lower surface of dead leaves of *Panax crassifolium* (Araliaceae). Kirk no. 89 (K, holotype; K, NY, isotypes).

This is morphologically indistinguishable from *Glomerella cingulata*, with ascospores simple, $18-22 \times 6-8 \mu\text{m}$, for which it would represent an older epithet.

Mycosphaerella panacis-ginseng Tomilin. See *Mycosphaerella panacis* Bunkina.

Mycosphaerella pandani Tilak, in T.S. Viswan. & Tilak, Sydowia 14: 311. 1960.

Type — India: *Pandanus fascicularis* (Pandanaeae).

No material was studied as the type was not found in LWG or IMI.

Sphaerella pandurata Ellis & Everh., Bull. Torrey Bot. Club 10: 117. 1883.— Fig. 630.

Type — USA: New Jersey, Plainfield. On dead leaves, mostly immersed between the upper and lower epidermis, of *Quercus* (Fagaceae). Meschutt, IX 1883 (NY, holotype).

This is morphologically indistinguishable from *Leiosphaerella succinea* (Roberge) E. Müll., with ascomata only visible by a papillate ostiole, asci clavate, paraphyses simple, $1 \mu\text{m}$ wide, ascospores $11-13 \times 3-4 \mu\text{m}$.

Mycosphaerella panicicola Henn., Hedwigia 41: 109. 1902 = *Sphaerella panicicola* (Henn.) Sacc. & D. Sacc., Syll. Fung. 17: 646. 1905.— Fig. 628.

Type — Brazil: São Paulo, Botanical Garden. On upper and lower surface of dead leaves of *Panicum* (Poaceae). Puttemans 150, IV 1900 (SP, lectotype, designated here because the holotype in B is destroyed; BPI, isolectotype).

The type is badly preserved, but shows that this belongs to section *Fusispora*, with asci cylindrical, ascospores fusiform, $17-20 \times 4-5 \mu\text{m}$.

Sphaerella panicum Cooke, Grevillea 5: 153. 1877 = *Metasphaeria panicum* (Cooke) Sacc., Syll. Fung. 2: 176. 1883 [as “*panicorum*”].

Type — USA: *Panicum* (Poaceae).

No material was studied as the type was not found in NY or K.

Sphaerella pantacanthae Speg., Anales Mus. Nac. Hist. Nat. Buenos Aires 19 (ser. 3, 12): 354. 1909.

Type — Argentina: *Pantacantha ameghinoi* (Solanaceae).

No material was studied as the type was not included in a loan from LPS.

Mycosphaerella papaveris (Fuckel) Jacz., Opredelitel' gribov 2: 615. 1917 = *Sphaerella papaveris* Fuckel, in Heuglin, Reisen nach dem Nordpolarmeer 3: 319. 1874.

Type — Russia: *Papaver nudicaule* (Papaveraceae).

No material was studied as the type was not included in a loan from G or found in any other herbarium. Synonymised with *Davidiella allicina* (as *Mycosphaerella tassiana*) by von Arx (1949), probably without examination of the type.

Mycosphaerella papuana Sivan., Trans. Brit. Mycol. Soc. 85: 743. 1985.— Fig. 631.

Type — Papua-New Guinea: Nembi Valley. On white spots with brown margins on upper surface of living leaves of *Dioscorea* (Dioscoreaceae). Waller no. 2144, IX 1984 (IMI no. 293049, holotype).

Anamorph: associated with *Cercospora dioscoreae-pyrifoliae* J.M. Yen *vide* Sivanesan (1984).

This is a parasitic species, with asci cylindrical, ascospores $10-12 \times 2-2.5 \mu\text{m}$, showing much less variation than mentioned in the protologue.

Mycosphaerella papyrifera (Pass.) Miles, Trans. Illinois State Acad. Sci. 10: 250. 1918 \equiv *Sphaerella papyrifera* Pass., Atti Reale Accad. Lincei, Mem. Cl. Sci. Fis., ser. 4, 3: 6. 1887.

Type — Italy: *Aralia papyrifera* (Araliaceae).

No material was studied as the type was not found in any of the herbaria consulted.

Sphaerella parallela P. Karst., Bidrag Kännedom Finlands Natur Folk 23: 184. 1873 \equiv *Apiospora parallela* (P. Karst.) Sacc., Syll. Fung. 1: 540. 1882.

Type — Finland: *Calamagrostis* (Poaceae).

No material was studied as the species was already redisposed.

Mycosphaerella parallelogramma (Rehm) Lindau, Hilfsb. Sammeln Ascomyceten: 50. 1903 \equiv *Sphaerella parallelogramma* Rehm, Hedwigia 24: 239. 1885.— Fig. 632.

Type — Austria: Tyrol, Ortler, Sulden Gletscher. On dead culms of Poaceae. Rehm, Ascomyceten no. 834, VII 1884 (S, holotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. disseminata*, with asci clavate, ascospores $12.5-14.5 \times 3-4 \mu\text{m}$.

Sphaerella paraneura Auersw., in Gonn. & Rabenh., Mycol. Europaea 5-6: 4. 1869, nomen nudum (illegitimate, Article 32).

Authentic material — Germany: *Acer campestre* (Sapindaceae).

No material was studied as the type was not found in B and might be lost. Cited as synonymous with *M. punctiformis* by Tomilin (1979).

Sphaerella parasita Fautrey, in Roum., Rev. Mycol. (Toulouse) 12: 64. 1890 \equiv *Sphaerella pucciniophila* Sacc. & P. Syd., Syll. Fung. 14: 533. 1899, superfluous (illegitimate, Article 52) nomen novum (Article 58).— Fig. 633.

Type — France: Côte d'Or, Noidan. On white spots on upper surface of living leaves of *Alcea rosea* (Malvaceae). Fautrey, distributed in Roumeguère, Fungi Selecti Exsiccati no. 5237 (PC, holotype).

This is a *Didymella*, with asci pyriform, ascospores $17-19 \times 6-8 \mu\text{m}$, with an *Ascochyta* anamorph. The spots are caused by *Puccinia malvacearum* (Basidiomycota, Uredinales), which is sporulating on the lower surface of the spots. The ascomycete is not in close contact with the rust and can therefore not be regarded as mycoparasitic, although both epithets have been coined based on this assumption.

Mycosphaerella parasitica (G. Winter) Lindau, in Engl. & Prantl, Natürlichen Pflanzenf. 1(1): 426. 1897 \equiv *Sphaerella parasitica* G. Winter, Hedwigia 25: 19. 1886 \equiv *Epicymatia winteri* J. Kunze, Fungi Selecti Exsiccati no. 65 \equiv *Epicymatia winteri* f. *cenangii-fuliginosi* J. Kunze, Fungi Selecti Exsiccati no. 65.— Fig. 634.

Type — Germany: Sachsen, Mansfeld. On dead stromata and apothecia of *Godronia* [*Cenangium*] *fuliginosa* (Ascomycota, Leotiaceae). Kunze, Fungi Selecti Exsiccati no. 65, VI 1875 (L, holotype).

This is a species of *Stigmatidium*, with ascomata with periphyses, asci pyriform, ascospores $11-15 \times 3-3.5 \mu\text{m}$. The genus *Stigmatidium* so far contains only parasites of lichenized ascomycetes, but this species is certainly congeneric. Therefore the following combination is proposed here: ***Stigmatidium parasiticum*** (G. Winter) Aptroot comb. nov. **MB 500378**. **Basionym:** *Sphaerella parasitica* G. Winter, Hedwigia 25: 19. 1886.

Sphaerella parasitica var. *tapesiae* Feltgen, Recueil Mém. Trav. Soc. Bot. Grand-Duché Luxembourg 1901: 183. 1901.

Type — Luxembourg: *Tapesia rosae* (Ascomycota, Dermateaceae).

No material was studied as this is certainly no *Mycosphaerella*.

Mycosphaerella pardalota (Cooke & Ellis) Miles, Mycologia 18: 165. 1926 \equiv *Sphaerella pardalota* Cooke & Ellis, Grevillea 6: 16. 1877.— Fig. 635.

Type — USA: New Jersey, Newfield. On lower surface of dead leaves of *Myrica cerifera* (Myricaceae). Ellis no. 2576, V 1877 (NY, holotype); also distributed in Ellis & Everh., North American Fungi no. 2136, VI 1888 (L, topotype).

The holotype is immature, but the topotype shows that this belongs to section *Longispora*, and is morphologically indistinguishable from *M. millegrana*, with asci cylindrical, ascospores $14-18 \times 2-3 \mu\text{m}$.

Mycosphaerella parjumanica Petr., Sydowia 16: 339. 1963 [“1962”].— Fig. 636.

Type — Afghanistan: Parjuman, Kuh Tscheling Safed. On dead stems of *Euphorbia* (Euphorbiaceae). Reehinger, distributed in Reliquiae Petrakianae no. 2253, VIII 1962 (H, L, isotypes).

This is morphologically indistinguishable from *Davidiella allicina*, with asci pyriform, ascospores $17-20 \times 5-6 \mu\text{m}$.

Mycosphaerella parkii Crous, M.J. Wingf., Ferreira & Alfenas, Mycol. Res. 97: 582. 1993.

Type — Brazil: Aracruz. On pale spots with brown margins on upper and lower surface of living leaves of *Eucalyptus grandis* (Myrtaceae). Wingfield, II 1990 (PREM 50668, holotype, not seen).

Anamorph: *Stenella parkii* Crous & Alfenas *vide* Crous (1998).

Material studied (Indonesia, Sumatra, Alfenas, XI 1993, IMI no. 361580 and no. 361581) contains sterile dried cultures.

Mycosphaerella parnassiae (Rostr.) Larsen, in Rosenvinge & Warming, Botany of Iceland 2(3), 9. Fungi of Iceland: 487. 1932 \equiv *Sphaerella parnassiae* Rostr., Bot. Tidsskr. 25: 302. 1903.— Fig. 637.

Type — Iceland: Hof. On dead flower stem of *Parnassia palustris* (Parnassiaceae). Davidsson, VII 1901 (C, holotype).

This belongs to section *Caterva*, with ascomata small, asci cylindrical, ascospores $11-15 \times 2-3 \mu\text{m}$. Additional material studied (Italy, Dolomites, Gröden, between Plau and Sellajoch, on living leaves, Bornmüller, VII 1907, B) is immature, but undoubtedly different.

Sphaerella paronychia Cooke, J. Bot. 21: 138. 1883 = *Laestadia paronychia* (Cooke) Sacc., Syll. Fung. 2: XXXIII. 1883.— Fig. 638.

Type — France: *Paronychia serpyllifolia* (Caryophyllaceae).

No material was studied as the type was not found in K.

Mycosphaerella paronychia M. Morelet, Ann. Soc. Sci. Nat. Archéol. Toulon & Var 20: 106. 1968, nomen novum (Article 58) for *Sphaerella paronychia* Unamuno, Bol. Soc. Esp. Hist. Nat. 34: 140. 1934, later homonym (illegitimate, Article 53).

Type — Spain: León, Ponferrada. On upper and lower surface of dead leaves of *Paronychia argentea* (Caryophyllaceae). Rothmaler no. 10288, IV 1933 (MA, holotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores $16-19 \times 5-6 \mu\text{m}$.

Mycosphaerella parva R.F. Park & Keane, Trans. Brit. Mycol. Soc. 79: 99. 1982.— Fig. 639.

Type — Australia: Victoria, Nowa Nowa. On pale spots with brown margins occupying most of the upper and lower surface of living leaves of *Eucalyptus globulus* (Myrtaceae). Park, VII 1981 (IMI no. 263258, isotype); also same locality, on *E. grandis*. Carnegie, XI 1990 (IMI no. 353729a, topotype).

This is a parasitic species, with asci cylindrical, ascospores $9-11 \times 2-3 \mu\text{m}$.

Sphaerella parvimacula Pass., Nuovo Giorn. Bot. Ital. 7: 256. 1875.

Type — Italy: *Fagus sylvatica* (Fagaceae).

No material was studied as the type was not found in any of the herbaria consulted.

Mycosphaerella pascuorum (Fautrey) Dias, Mem. Soc. Brot. 21: 70. 1970 = *Sphaerella pascuorum* Fautrey, Rev. Mycol. (Toulouse) 18: 71. 1896.— Fig. 640.

Type — France: On dead stems of *Leucanthemum vulgare* (Asteraceae). Fautrey, distributed in Roumeguère, Fungi Selecti Exsiccati no. 6992, X 1895 (PC, holotype).

Anamorph: associated with *Coniothyrium artemisiae* (Gonz. Frag.) Petr. & Syd. *fide* Dias (op. cit.).

Cited as synonymous with *Mycosphaerella parallelogramma* by Tomilin (1979). The holotype and additional material studied (Czech Republic, Weißkirchen, on lower surface of dead leaves of *Tanacetum vulgare*, Petrak, Flora Bohemiae et Moraviae exsiccata no. 1248, VIII 1916, L) belongs to *Davidiella* and shows that it is morphologically indistinguishable from *D. ammophilae*, with asci pyriform, ascospores $10-12 \times 2.5-3.5 \mu\text{m}$.

Sphaerella pascuorum Kirschst. See *Mycosphaerella kirschsteinii* Tomilin.

Mycosphaerella pashkiensis Petr., Sydowia 16: 340. 1963 [“1962”].— Fig. 641.

Type — Afghanistan: Nuristan, Pashki. On dead stems and on upper and lower surface of dead leaves of *Draba* (Brassicaceae). Edelberg, II 1948 (W, holotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores $15-18 \times 3.5-5 \mu\text{m}$.

Mycosphaerella passeriniana (Sacc.) Johanson ex Oudem., Rév. Champ. Pays-Bas 2: 211. 1897 = *Sphaerella passeriniana* Sacc., Syll. Fung. 2: IXI. 1883, intended as nomen novum for *Sphaerella cruciferarum* sensu Pass. non (Fr.) Sacc. (1881).— Fig. 642.

Type — Italy: Parma, Vigheffio. On dead stems of *Rapistrum rugosum* (Brassicaceae). Passerini, distributed in Rabenhorst-Winter, Fungi Europaei Exsiccati no. 3447 (L, isotype).

Already synonymised with *Davidiella allicina* (as *Mycosphaerella tassiana*) by von Arx (1949), with which the isotype and additional material studied (Netherlands, Goes, on *Sisymbrium officinale* (Brassicaceae), van den Bosch, L) agree well, with asci pyriform, ascospores $14-18 \times 5-7 \mu\text{m}$.

Mycosphaerella passiflorae Rehm, Hedwigia 40: 110. 1901 = *Sphaerella passiflorae* (Rehm) Sacc. & P. Syd., Syll. Fung. 16: 470. 1902.

Type — Brazil: *Passiflora* (Passifloraceae).

The type was not found in S or in any of the other herbaria consulted. Material studied (Trinidad, intercepted at New York, on mature fruits, Isakson, V 1979, BPI) is immature.

Mycosphaerella passiflorae J.F. Lue & P.K. Chi, J. S. China Agric. Coll. 13: 93. 1992, later homonym (illegitimate, Article 53).

Type — China: *Passiflora edulis* (Passifloraceae).

No material was studied as the location of the type was not known.

Mycosphaerella passiflorae var. *bignoniae* Rehm, Hedwigia 44: 4. 1905.— Fig. 643.

Type — Brazil: São Francisco. On white spots on upper and lower surface of living leaves of *Bignonia* (Bignoniaceae). Ule no. 287 (S, holotype).

This is a parasitic species, with asci cylindrical, ascospores $13-15 \times 3-4 \mu\text{m}$.

Sphaerella patagonica Speg., Bol. Acad. Nac. Ci. 11: 39. 1888 [“1887”].

Type — Argentina: *Bolax glebaria* (Apiaceae).

No material was studied as the type was not included in a loan from LPS.

Mycosphaerella pataguae (Speg.) Cash, Syll. Fung. 26: 348. 1972 = *Sphaerella pataguae* Speg., Bol. Acad. Nac. Ci. 25: 61. 1921.

Type — Chile: *Crinodendron patagua* (Elaeocarpaceae).

No material was studied as the type was not included in a loan from LPS.

Mycosphaerella patouillardii (Sacc.) Maire & Werner, Mém. Soc. Sci. Nat. Maroc 45: 27. 1937 [as “*patouillardii*”] = *Sphaerella patouillardii* Sacc., in Berl. & Voglino, Syll. Fung., Addit. ad vol. 1-4: 407. 1886 [as “*patouillardii*”].— Fig. 644.

Type — Italy: Roma, Villa Borghese. On white spots on upper and lower surface of living leaves of *Buxus sempervirens* (Buxaceae). Saccardo, Mycotheca Italica no. 849, IV 1902 (L, topotype).

Cited as synonymous with *Mycosphaerella buxicola* by Tomilin (1979). This belongs to section *Longispora*, and is morphologically indistinguishable from *M. latebrosa*, with asci cylindrical, surrounded by many immature asci which are resembling paraphyses, ascospores $25\text{--}30 \times 3\text{--}4 \mu\text{m}$.

Sphaerella patouillardii subsp. *briardi* (Sacc.) Sacc. See *Sphaerella briardi* Sacc.

Mycosphaerella patriniae Petr., Hedwigia 68: 209. 1928.

Type — Russia: *Patrinia intermedia* (Caprifoliaceae).

No material was studied as the type was not included in a loan from W.

Mycosphaerella paulowniae Shirai & Hara, Bot. Mag. (Tokyo) 25: 70. 1911 = *Sphaerella paulowniae* (Shirai & Hara) Trotter, Syll. Fung. 24: 886. 1928.

Type — Japan: *Paulownia tomentosa* (Paulowniaceae).

No material was studied as the type was not included in a loan from TNS.

Mycosphaerella paulula (Cooke) Tomilin, Novosti Sist. Nizsh. Rast. 1966: 173. 1966 = *Sphaerella paulula* Cooke, Grevillea 6: 146. 1878.

Type — USA: *Zea mays* (Poaceae).

No material was studied as the type was not found in NY or K.

Mycosphaerella pavonina Petr. & Cif., Ann. Mycol. 30: 214. 1932.— Fig. 645.

Type — Dominican Republic: Santiago, Hato del Yaque, Hatillo. On ill-defined spots on upper surface of living leaves of *Pavonia* (Malvaceae). Ciferri no. 4133, II 1931 (BPI, holotype).

This is morphologically indistinguishable from *Didymella bryoniae* (Auersw.) Rehm, with asci clavate to pyriform, paraphysoids septate, ca. $1.5\text{--}2 \mu\text{m}$ wide, ascospores $15\text{--}16.5 \times 3.5\text{--}5.5 \mu\text{m}$. Additional material (Sudan, Gilo, on *P. urens* var. *hirsuta*, Tarr, XI 1954 (IMI no. 59730 and 59731) agrees.

Mycosphaerella paxystimae Dearn., Mycologia 18: 246. 1926 [as “*pachystimae*”].— Fig. 646.

Type — USA: Idaho, Bonner Co., Coolin. On white spots with brown margins on upper and lower surface of living leaves of *Paxystima* [“*Pachystima*”] *myrsinites* (Celastraceae). Boyce no. 1287, VII 1926 (BPI, holotype).

This is a parasitic species, with asci cylindrical, ascospores $18.5\text{--}20 \times 3.5\text{--}4.5 \mu\text{m}$.

Mycosphaerella peckii Lindau, in Engl. & Prantl, Natürlichen Pflanzenf. 1(1): 425. 1897, nomen novum (Article 58) for *Sphaerella peckii* Sacc., Syll. Fung. 9: 649. 1891, later homonym (illegitimate, Article 53), nomen novum (Article 58) for *Sphaerella conigena* Peck, Annual Rep. New York State Mus. 38: 104. 1885, later homonym (illegitimate, Article 53) = *Delphinella peckii* (Lindau) M.E. Barr, in M.E. Barr, C.T. Rogerson, S.J. Sm. & Haines, New York State Mus. Bull. 459: 16. 1986.— Fig. 647.

Type — USA: New York, Albany Co., Helderberg Mts. On scales of dead cones of *Tsuga* [“*Abies*”] *canadensis* (Pinaceae). Peck, V (NY, isotype).

Accepted as *Delphinella peckii* (Lindau) M.E. Barr by Barr, Rogerson, Sm. & Haines (op. cit.), with which the isotype agrees well.

Sphaerella peckii Speg. See *Mycosphaerella spegazzinii* Tomilin.

Mycosphaerella pectinis (Pass.) Tomilin, Novosti Sist. Nizsh. Rast. 7: 204. 1970 = *Sphaerella pectinis* Pass., Atti Reale Accad. Lincei, Mem. Cl. Sci. Fis., ser. 4, 4: 57. 1888 [as “*pectin*”].

Type — Italy: *Scandix pecten-veneris* (Apiaceae).

No material was studied as the type was not found in any of the herbaria consulted.

Mycosphaerella pedicularidis (P. Karst.) Lind, Danish Fungi: 209. 1913 [as “*pedicularis*”] = *Sphaerella pedicularidis* P. Karst., Öfvers. Förh. Kongl. Svenska Vetensk.-Akad. 1872(2): 107. 1872 [as “*pedicularis*”].— Fig. 648.

Type — Svalbard: On upper and lower surface of dead leaves of *Pedicularis hirsuta* (Orobanchaceae).

Anamorphs: *Phyllosticta* and *Ramularia obducens* Thüm. *vide* Tomilin (1979).

The type was not found in H or in any of the herbaria consulted. Scandinavian material studied (Sweden, Lappmark, Jukkasjärvi, Nuolja, Nannfeldt, VII 1927, BPI) belongs to section *Caterva* and is morphologically indistinguishable from *M. subradians*, with asci cylindrical, ascospores $9\text{--}10.5 \times 2\text{--}2.5 \mu\text{m}$. Material from the Alps (Germany, Schwaben, Allgäu, Kr. Sonthofen, Oberstdorf, on *P. foliosa*, Starcs no. 9578, VI 1948, B) contains a immature ascomycete, probably *Pleospora herbarum* (Pers. : Fr.) Rabenh.

Mycosphaerella pellucida (Bubák & Dearn.) Tomilin, Novosti Sist. Nizsh. Rast. 1967: 190. 1967 = *Sphaerella pellucida* Bubák & Dearn., in Bubák, Hedwigia 58: 15. 1916.

Type — Canada: London. On white spots with brown margins on upper surface of living leaves of *Smilax herbacea* (Smilacaceae). Dearness, VIII-IX 1910 (BPI, isotype).

The isotype only contains various coelomycetes, e.g. one with apiosporous conidia 1-2-septate, $15\text{--}20 \times 5\text{--}6 \mu\text{m}$, that fit the ascospores mentioned in the protologue.

Mycosphaerella pelvetiae G.K. Sutherl., New Phytologist 24: 34. 1915 = *Sphaerella pelvetiae* (G.K. Sutherl.) Trotter, Syll. Fung. 24: 849. 1928.

Type — United Kingdom: In thallus of *Pelvetia canaliculata* (Phaeophyta, Fucaceae).

Already cited as synonymous with *Mycosphaerella ascophylli* by Tomilin (1979) and as synonymous with *Mycophycias ascophylli* by Kohlmeyer & Volkmann-Kohlmeyer (1998). It is accepted here as *Stigmidium ascophylli* (Cotton) Aptroot (see above).

The location of the type is unknown. Materials studied (United Kingdom, Channel Islands, Herm, Rhodes no. 4986 and 4956, XI 1931, B; also Devon, Portlemouth Down, Hawksworth no. 4199, VIII 1975, IMI no. 196222) agree well.

Mycosphaerella penstemonis Earle, in Greene, Plantae Bakerianae 2(1): 19. 1901 [as “*pentastemonis*”] = *Sphaerella penstemonis* (Earle) Sacc. & P. Syd., Syll. Fung. 16: 1133. 1902 [as “*pentastemonis*”].— Fig. 649.

Type — USA: Colorado, Pagosa Peak. On dead stems of *Penstemon* [“*Pentastemon*”] (Plantaginaceae). Baker no. 125, VIII 1899 (NY, isotype), also distributed in Earle, Plants of Southern Colorado no. 38 (NY, isotype).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradians*, with asci cylindrical, ascospores 10-12 × 3-4 µm.

Sphaerella perconferta Speg., Bol. Acad. Nac. Ci. 23: 470. 1919.

Type — Brazil: *Drimys* (Winteraceae).

Anamorph: Associated with *Phyllosticta perconferta* Speg. *fide* Spegazzini (op. cit.).

No material was studied as the type was not included in a loan from LPS.

Mycosphaerella peregrina (Cooke) Lindau, in Engl. & Prantl, Natürlichen Pflanzenf. 1(1): 424. 1897 = *Sphaerella peregrina* Cooke, Grevillea 7: 88. 1879.— Fig. 650.

Type — United Kingdom: Hereford, Symonds' Yat. On dead stems and upper and lower surface of dead leaves of *Rubia peregrina* (Rubiaceae). Ross, 1878, distributed in Cooke, Fungi Britannici Exsiccati, ed. 2 no. 700 (B, IMI, K (2×), isotypes).

This belongs to section *Caterva* and is morphologically indistinguishable from *M. subradians*, with asci cylindrical, ascospores 13-16 × 3-4 µm.

Mycosphaerella perexigua (P. Karst.) Johanson, Öfvers. Förh. Kongl. Svenska Vetensk.-Akad. 41(9): 166. 1884 = *Sphaerella perexigua* P. Karst., Öfvers. Förh. Kongl. Svenska Vetensk.-Akad. 1872(2): 107. 1872, non *Sphaeria perexigua* Lév., non *Mycosphaerella punctiformis* var. *perexigua* (Lév.) Rabenh.— Fig. 651.

Type — Svalbard: On dead stems of *Juncus biglumis* (Juncaceae).

No type material was found. Material studied (Sweden, Torne Lappmark, Jukkasjärvi, Björkman, VII 1920, CBS) belongs to *Davidiella*, and is possibly morphologically indistinguishable from *D. allicina*, with ascomata tiny, asci pyriform, ascospores 18-21 × 5-6 µm.

Mycosphaerella perexigua Starbäck. See *Sphaerella starbäckii* Sacc. & Traverso.

Sphaerella perexigua Fuckel ex Thüm., Fungi Austriaci Exsiccati, cent. 9 no. 869. 1873, nomen nudum (not validly published, Article 32) and later homonym (illegitimate, Article 53).— Fig. 652.

Authentic material — Germany: On lower surface of dead leaves of *Quercus robur* (Fagaceae). Thümen, Fungi Austriaci Exsiccati no. 869 (L).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. ammophilae*, with asci pyriform, ascospores 9-11 × 3-3.5 µm.

Mycosphaerella perexigua var. *minima* Johanson, Öfvers. Förh. Kongl. Svenska Vetensk.-Akad. 41(9): 166. 1884 = *Sphaerella perexigua* var. *minima* (Johanson) Berl. & Voglino, Syll. Fung., Addit. ad vol. 1-4: 83. 1886.

Type — Sweden: *Scirpus caespitosus* (Cyperaceae).

No material was studied as the type was not included in a loan from S. Cited as synonymous with *Mycosphaerella perexigua* (P. Karst.) Johanson by Barr (1972).

Sphaerella pereziae Speg., Anales Mus. Nac. Hist. Nat. Buenos Aires 19 (ser. 3, 12): 354. 1909.

Type — Argentina: *Perezia patagonica* (Asteraceae).

No material was studied as the type was not included in a loan from LPS.

Mycosphaerella perforans (Roberge) Lind, Danish Fungi: 205. 1913 = *Sphaeria perforans* Roberge, in Desm., Ann. Sci. Nat. Bot., ser. 2, 19: 23. 1843 = *Sphaerella perforans* (Roberge) Ces. & De Not., Comment. Soc. Crittog. Ital. 1(4): 238. 1863 = *Paradidymella perforans* (Roberge) Munk, Dansk Bot. Ark 17(1): 179. 1957.— Fig. 653.

Type — France: On lower surface of dead, inrolled leaves of *Ammophila* [“*Calamagrostis*”] *arenaria* (Poaceae). Desmazières, Fungi Exsiccati no. 788 (L, isotype).

The type shows that this is an older name for the well-known species *Montagnula spartii* (Castagne) Aptroot, which is accepted by Aptroot (1995b), with ascospores red-brown, 19-23 × 10-12 µm. Therefore the following new combination is proposed here: **Montagnula perforans** (Roberge) Aptroot comb. nov., **MB 500370**. **Basionym:** *Sphaeria perforans* Roberge, in Desm., Ann. Sci. Nat. Bot., ser. 2, 19: 23. 1843. See for description, illustration and additional synonymy Aptroot (1995b). Additional material seen (Netherlands, Hollands duin, Oudemans, L) is immature.

Mycosphaerella pericampyli Syd. & P. Syd., Philipp. J. Sci. 8: 270. 1913 = *Sphaerella pericampyli* (Syd. & P. Syd.) Trotter, Syll. Fung. 24: 878. 1928.— Fig. 654.

Type — Philippines: Luzon, Prov. Sorsogon. On white spots on lower surface of living leaves of *Pericampylus incanus* (Menispermaceae). Elmer, Philippine Island Plants no. 15086, III-VIII 1916 (L, 2 isotypes).

This is a parasitic species, with asci cylindrical, ascospores 13-15 × 3-3.5 µm.

Mycosphaerella pericopsidis Henn., Hedwigia 41: 109. 1902 ≡ *Sphaerella pericopsidis* (Henn.) Sacc. & D. Sacc., Syll. Fung. 17: 636. 1905.— Fig. 655.

Type — Brazil: São Paulo, Botanical Garden. On dead spots, especially on leaf margins, on upper and lower surface of living leaves of *Pericopsis mooniana* (Fabaceae). Puttemans 296, VII 1900 (SP, lectotype, designated here, as the holotype in B has been destroyed).

This is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores ca. $7 \times 2 \mu\text{m}$.

Sphaerella perimens Kirschst., Hedwigia 80: 123. 1941.— Fig. 656.

Type — Germany: Westfalen, Siegen, Olpe, Dornbruch. On dead culms of *Juncus filiformis* (Juncaceae). Ludwig, VII 1937 (B, holotype; B, isotype); also VIII 1921 (NY, topotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. disseminata*, with asci pyriform, ascospores $12-16 \times 3.5-4.5 \mu\text{m}$. Additional material (Stegakopf, Ludwig, VII 1924, B) agrees.

Mycosphaerella periplocae (Pass.) Tomilin, Opredelitel' gribov roda *Mycosphaerella* Johans.: 58. 1979 ≡ *Sphaerella periplocae* Pass., Atti Reale Accad. Lincei, Mem. Cl. Sci. Fis., ser. 4, 6: 457. 1890.

Type — Italy: *Periploca graeca* (Periplocaceae).

No material was studied as the type was not found in any of the herbaria consulted.

Mycosphaerella pernettyae (Speg.) Syd., Ann. Mycol. 26: 106. 1928 ≡ *Sphaerella pernettyae* Speg., Bol. Acad. Nac. Córdoba 11: 207. 1887 [as “*pernettiae*”].— Fig. 657.

Type — Argentina: On upper surface of dead tips of living leaves of *Pernettya* [“*Pernettia*”] *mucronata* (Ericaceae).

The type was not included in a loan from LPS. Material studied (Chile, Chiloé Island, Castro, Werdermann no. 1738, II 1924, B, IMI no. 16781) belongs to section *Caterva* and suggested that this is morphologically indistinguishable from *M. subradicans*, with asci cylindrical, ascospores $10-13 \times 2.5-3.5 \mu\text{m}$.

Mycosphaerella perparva House, New York State Mus. Bull. 233-234: 29. 1921, nomen novum (Article 58) for *Sphaerella perparva* Sacc., Syll. Fung. 11: 300. 1895, later homonym (illegitimate, Article 53), nomen novum (Article 58) for *Sphaerella minutissima* G. Winter, Internationale Polarforschung 1882-1883, Deutsche Expedition, 2: 93. 1890, later homonym (illegitimate, Article 53) ≡ *Mycosphaerella minutissima* Tomilin, Opredelitel' gribov roda *Mycosphaerella* Johans.: 208. 1979, superfluous (illegitimate, Article 52) nomen novum (Article 58).

Type — Greenland: *Thelypteris* [“*Lastrea*”] *fragrans* (Polypodiaceae).

No material was studied as the type was not found in B and might be lost. Cited as synonymous with *Mycosphaerella harthensis* by Tomilin (1979).

Mycosphaerella perparva (Pass. & Beltrani) Tomilin, Novosti Sist. Nizsh. Rast. 1967: 190. 1967, later homonym (illegitimate, Article 53) ≡ *Sphaerella perparva* Pass. &

Beltrani, Atti Reale Accad. Lincei, Mem. Cl. Sci. Fis., ser. 3, 7: 3. 1882.

Type — Italy: *Juncus acutus* (Juncaceae).

No material was studied as the type was not found in any of the herbaria consulted.

Sphaerella perpusilla (Desm.) Ces. & De Not., Comm. Soc. Crittog. Ital. 1(4): 237. 1863 ≡ *Sphaeria perpusilla* Desm., Ann. Sci. Nat. Bot., ser. 3, 6: 80. 1846 ≡ *Laestadia perpusilla* (Desm.) Sacc., Syll. Fung. 1: 423. 1882.— Fig. 658.

Type — France: On upper and lower surface of dead leaves of *Phragmites australis* [“*Arundo phragmites*”] (Poaceae).

Material studied (Germany, Rollsdorf, Halle am See, Winter, distributed in Rabenhorst, Fungi Europaei Exsiccati no. 1729, VIII 1873, L) is morphologically indistinguishable from *Davidiella allicina*, with asci pyriform, ascospores $16-24 \times 5-6.5 \mu\text{m}$.

Sphaeria perpusilla var. *typhae* (Lasch) Auersw. See *Mycosphaerella typhae* (Lasch) Lindau.

Mycosphaerella perseae Miles, Trans. Illinois State Acad. Sci. 10: 251. 1917 [as “*persiae*”] ≡ *Sphaerella perseae* (Miles) Trotter, Syll. Fung. 24: 871. 1928.— Fig. 659.

Type — Puerto Rico: Maricao. On white spots with black margins on upper surface of living leaves of *Persea americana* (Lauraceae). Stevens no. 4486, XI 1913 (BPI, isotype).

This is a parasitic species, with asci cylindrical, ascospores $8-11 \times 2.5-3.5 \mu\text{m}$. Additional material (Mexico, Huauchinango, Puebla, Quintero, III 1969, IMI no. 145133) agrees.

Mycosphaerella persica Syd. & P. Syd., Ann. Mycol. 6: 529. 1908 ≡ *Sphaerella persica* (Syd. & P. Syd.) Sacc. & Trotter, Syll. Fung. 22: 121. 1913.— Fig. 660.

Type — Iran: *Moriera stenoptera* (Brassicaceae).

Synonymised with *Davidiella allicina* (as *Mycosphaerella tassiana*) by von Arx (1949), with which material studied (Lybia, on dead stems of *Tarsetia aegyptica* (Brassicaceae), Pamparini, IV 1913, PAD) agrees, with asci pyriform, ascospores $17-20 \times 5-7.5 \mu\text{m}$.

Mycosphaerella persicae B.B. Higgins & F.A. Wolf. See *Mycosphaerella pruni-persicae* Deighton.

Mycosphaerella personata B.B. Higgins, Amer. J. Bot. 16: 295. 1929.

Type — USA: Georgia, Experiment. On lower surface of dead leaves of *Vitis vinifera* (Vitaceae). Higgins no. 713, V 1914 (NY, isotype).

Anamorph: *Pseudocercospora vitis* (Lév.) Speg. *vide* Sivanesan (1984).

The type material is in poor shape, but shows that this is morphologically indistinguishable from *M. punctiformis*.

Mycosphaerella persooniae Henn., Hedwigia 42: 81. 1903 ≡ *Sphaerella persooniae* (Henn.) Sacc. & D. Sacc., Syll. Fung. 17: 639. 1905 ≡ *Anthracostroma persooniae* (Henn.) Petr., Sydowia 8: 97. 1954.

Type — Australia: *Persoonia salicina* (Proteaceae).
Anamorph: *Camarosporula persooniae* (Henn.) Petr. *vide* Petrak (op. cit.).

Accepted as *Anthracostroma persooniae* (Henn.) Petr. by Petrak (op. cit.), with which material studied (New South Wales, Sydney, on upper and lower surface of living leaves of *Persoonia laevis*, Walker no. DAR 13514, VII 1964, L) may agree, as it shows immature, superficial stromata.

Mycosphaerella peruviana (Speg.) Tomilin, Oprelidelitel' gribov roda *Mycosphaerella* Johans.: 88. 1979 = *Sphaerella peruviana* Speg., Anales Soc. Ci. Argent. 12: 115. 1881.

Type — Peru: *Salicornia peruviana* (Amaranthaceae). Cited by J. & E. Kohlmeyer (1979) as synonymous with *M. salicorniae*, which is morphologically indistinguishable from *Davidiella ammophilae*. Material studied (U.S.A., California, San Benito County, Hollister alkaline flats, on *S. subterminalis*, Rose, VIII 1933, B) agrees.

Mycosphaerella petasitidis (Rabenh.) Feltgen, Recueil Mém. Trav. Soc. Bot. Grand-Duché Luxembourg 14: 287. 1899 = *Sphaeria petasitidis* Rabenh., in Klotzsch, Herbarium Vivum Mycologicum, cent. 10 no. 977. 1846 = *Sphaerella petasitidis* (Rabenh.) Fuckel, Jahrb. Nassauischen Vereins Naturk. 23-24: 99. 1870 (non *Stigmatea petasitidis* Fuckel, Jahrb. Nassauischen Vereins Naturk. 23-24: 96. 1870 = *Venturia petasitidis* (Fuckel) Sacc., Syll. Fung. 1: 592. 1882 = *Epipolaeum petasitidis* (Fuckel) E. Müll., in E. Müll. & Arx, Beitr. Kryptogamenfl. Schweiz 11(2): 186. 1962).

Type — Germany: Sonnewalde. On upper and lower surface of dead leaves of *Petasites officinalis* (Asteraceae). Kretzschmar, distributed in Klotzsch, Herbarium Vivum Mycologicum, cent. 10 no. 977 (B, L, isotypes), also distributed in Rabenhorst, Herbarium Mycologicum. ed. 2, no. 733 (L, 2 isotypes).

The isotypes studied are immature.

Sphaerella petasitidis f. *adenostylidis* Sacc., Syll. Fung. 1: 538. 1882.

Type — Italy: Belluno, Agordo. On upper surface of dead leaves of *Adenostyles alpina* (Asteraceae). Saccardo, IX 1905, Mycotheca Italiana no. 1658 (BPI, L, topotypes).

The topotypes studied are immature, with black stromata.

Sphaeria petasitidis var. *digitalidis* Fuckel, Enumeratio Fungi Nassauiae no. 530b [as “*digitalis*”].

Type — Germany: Zange. On spots on upper surface of living leaves of *Digitalis purpurea* (Plantaginaceae). Fuckel, Enumeratio Fungi Nassauiae no. 530b (L, isotype).

The isotype studied is immature.

Mycosphaerella petchii M. Morelet, Ann. Soc. Sci. Nat. Archéol. Toulon & Var 20: 106. 1968, nomen novum (Article 58) for *Sphaerella lobeliae* Petch, Ann. Roy. Bot. Gard. (Peradeniya) 7: 304. 1922 = *Mycosphaerella zeylanica* Cash, Syll. Fung. 26: 359. 1972, superfluous (illegitimate, Article 52) nomen novum (Article 58).

Type — Sri Lanka: Nuwara Eliya, Haputale. On white spots with black margins on upper surface of living leaves of

Lobelia nicotianaefolia (Lobeliaceae). Petch no. 5625, XII 1917 (K, isotype).

The isotype contains three coelomycetes, viz. an *Ascochyta*, with conidia 1-septate, 16-19 × 4.5-5.5 µm, a *Pestalotiopsis* and a *Phoma*, of which the *Ascochyta* agrees to some extent with the protologue and may have led to the description of the *Sphaerella*.

Mycosphaerella petiolicola (Desm.) Migula, in Thomé, Fl. Deutschl. Österr., Schweiz X, 1. Kryptog.-Fl. III, 3(1): 292. 1912 [“1913”] = *Sphaeria petiolicola* Desm., Ann. Sci. Nat. Bot., ser. 3, 11: 354. 1849 = *Sphaerella petiolicola* (Desm.) Auersw., in Gonn. & Rabenh., Mycol. Europaea 5-6: 8. 1869 [as “*Sphaeria*”].— Fig. 661.

Type — France: On dead petioles of *Fraxinus excelsior* (Oleaceae). Desmazières, Plantes Cryptogames de France no. 2077 (NY, isotype).

Cited as synonymous with *M. punctiformis* by Tomilin (1979), with which immature isotype and additional material studied (USA, New Jersey, Newfield, on dead petioles of *Juglans regia*, Ellis, distributed in Ellis & Everh., North American Fungi no. 795b, VI 1881, L) agrees well, with asci cylindrical, ascospores 10-12 × 2-2.5 µm.

Sphaerella petiolicola f. *robiniae* Sacc., Syll. Fung. 1: 490. 1882.— Fig. 662.

Type — France: On dead petioles of *Robinia pseudacacia* (Fabaceae).

Material studied (USA, New Jersey, Newfield, Ellis, distributed in Ellis & Everhart, North American Fungi no. 795a, VI 1881, L) is *M. punctiformis*, with asci cylindrical, ascospores 10-12 × 2-2.5 µm.

Mycosphaerella petrakii Tomilin. See *Mycosphaerella calamagrostidis* Petr.

Mycosphaerella phacae-frigidae E. Müll. & Wehm., Sydowia 8: 190. 1954.— Fig. 663.

Type — Switzerland: Graubünden, St. Moritz, Coroiglia. On upper and lower surface of dead leaves of *Phaca frigida* (Fabaceae). Müller, VII 1953 (ZT, holotype).

This belongs to section *Caterva* and is morphologically indistinguishable from *M. subradicans*, with asci cylindrical, ascospores 11-13 × 3-3.5 µm.

Mycosphaerella phaceliiphila (Speg.) Cash, Syll. Fung. 26: 349. 1972 = *Sphaerella phaceliiphila* Speg., Bol. Acad. Nac. Ci. 25: 59. 1921.— Fig. 664.

Type — Chile: Los Perales. On dead stems of *Phacelia circinata* (Boraginaceae). Spegazzini no. 6150, 1917 (LPS, holotype).

This is morphologically indistinguishable from *Davidiella allicina*, with asci pyriform, ascospores 12-15 × 5-6 µm.

Mycosphaerella phalaridis (Gonz. Frag.) Maire & Werner, Mém. Soc. Sci. Nat. Maroc 45: 27. 1937 = *Sphaerella phalaridis* Gonz. Frag., Bol. Soc. Esp. Hist. Nat. 25: 103. 1925.

Type — Mauritania: Larache. On dead leaves of *Phalaris bulbosa* (Poaceae). Caballero no. 6531 and 6532, VI 1923 (MA, holotype).

The holotype collections contains nothing identifiable.

Mycosphaerella phaseoli Chona & Munjal, Indian Phytopathol. 9: 53. 1956.

Type — India: *Phaseolus aureus* (Fabaceae).

Anamorph: *Cercospora kikuchii* T. Matsumoto & Tomoy. *vide* Chona & Munjal (op. cit.).

No material was studied as the type was not found in LWG or IMI.

Mycosphaerella phaseolicola (Roberge) Siemaszko, Mater. Mikol. Fitopatol. Rossii 3: 26. 1915 \equiv *Depazea phaseolicola* Roberge, in Desm., Ann. Sci. Nat. Bot., ser. 3, 11: 358. 1849 \equiv *Sphaerella phaseolicola* (Roberge) Sacc., Syll. Fung. 1: 503. 1882.— Fig. 665.

Type — France: On dead parts, not in clearly defined spots, of living leaves of *Phaseolus* (Fabaceae). Desmazières, Plantes Cryptogames de France no. 359 (B-Desmazières, IMI no. 243623 (slide only), isotypes).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores immature, 10-15 \times 5-7 μ m.

Mycosphaerella phaseolorum Siemaszko, Mater. Mikol. Fitopatol. Rossii 3: 25. 1915 \equiv *Sphaerella phaseolorum* (Siemaszko) Trotter, Syll. Fung. 24: 872. 1928.

Type — Georgia: *Phaseolus mungo* (Fabaceae).

No material was studied as the type was not found in BPI.

Mycosphaerella pheidasca J. Schröt., in Cohn, Kryptog.-Fl. Schlesien 3(2): 342. 1894 [“1893”] \equiv *Phaeosphaerella pheidasca* (J. Schröt.) Sacc., Syll. Fung. 11: 312. 1895.— Fig. 666, 971.

Type — Germany: On dead culms of *Juncus effusus* [“*leersii*”] (Juncaceae).

Cited as synonym of *Monascostroma innumerosa* (Desm.) Höhn. by Müller & von Arx (1962), with which material studied (Oberpfalz, Haidhof, Rehm, distributed in Ascomyceten no. 1990, BPI) agrees well, with asci pyriform, ascospores 20-23 \times 5-6 μ m, thick-walled, immature, hyaline, brown when mature, ornamented with warts and surrounded with a 1 μ m thick gelatinous sheath.

Sphaerella phellos (Schwein.) Cooke, J. Bot. 21: 106. 1883 \equiv *Sphaeria phellos* Schwein., Trans. Amer. Philos. Soc., ser. 2, 4: 225. 1832.

Type — USA: On lower surface of dead leaves of *Quercus phellos* (Fagaceae). Bartrams (PH, holotype; K, PH, isotype).

Already cited as synonymous with *M. punctiformis* by Tomilin (1979). The type specimens contain only the corresponding *Asteromella* spermatial state.

Mycosphaerella philochorta (Cooke) Tomilin, Novosti Sist. Nizsh. Rast. 1967: 190. 1967 \equiv *Sphaerella philochorta* Cooke, J. Bot. 21: 137. 1883.— Fig. 667.

Type — USA: Maine: Scarborough. On upper and lower surface of dead leaves of Poaceae. Ellis no. 116 (K, holotype).

This is a species of the Microthyriaceae, with conical ascomata with meandering cells, without hamathecium, ascospores 15-18 \times 5-6 μ m.

Mycosphaerella philodendronis (Pat. & Gaillard) Lindau, in Engl. & Prantl, Natürlichen Pflanzenf. 1(1): 426. 1897 [as “*philodendri*”] \equiv *Sphaerella philodendronis* Pat. & Gaillard, Bull. Soc. Mycol. France 4: 112. 1888 [as “*phylloendronis*”].— Fig. 668.

Type — Venezuela: Puerto Zamuro. On white spots on upper and lower surface of living leaves of *Philodendron* [“*Phylloendron*”] (Araceae). No. 1, V 1887 (FH-Patouillard, holotype).

The type and additional material studied (Surinam, Paramaribo, on *Philodendron acutatum*, Kramer & Hekking, Flora Surinamensis no. 2068/A, XI 1960, L) both belong to section *Plaga*, with asci broad-cylindrical, ascospores 11-13 \times 2.5-3.5 μ m in the type but 14-17 \times 3.5-4.5 μ m in the specimen from Surinam.

Mycosphaerella phlogina (Ellis & Everh.) Earle, in Greene, Plantae Bakerianae 2: 19. 1901 \equiv *Sphaerella phlogina* Ellis & Everh., J. Mycol. 4: 65. 1888 \equiv *Phaeosphaerella phlogina* (Ellis & Everh.) Tomilin, Opredelitel' gribov roda *Mycosphaerella* Johans.: 285. 1979.— Fig. 669.

Type — USA: Montana, Belt Mts. On upper and lower surface of dead leaves of *Phlox longifolia* (Polemoniaceae). Anderson no. 142, VI 1885 (NY, holotype; NY, isotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. clandestina*, with asci pyriform, ascospores 23-27 \times 7-9 μ m. The types contain also *Pleospora herbarum*, with ascospores brown, muriform, 24-27 \times 10-13 μ m.

Mycosphaerella phlomidicola Tomilin, Novosti Sist. Nizsh. Rast. 1966: 150. 1966.

Type — Russia: *Phlomis* (Lamiaceae).

No material was studied as the type was not included in loans from LE or LEP. Cited as synonymous with *Mycosphaerella denigrans* by Tomilin (1979).

Mycosphaerella phlomidis Lebedeva, Mater. Mikol. Fitopatol. Rossii 5(3): 2. 1921 \equiv *Mycosphaerella lebedevae* Tomilin, Novosti Sist. Nizsh. Rast. 1966: 151. 1966, superfluous (illegitimate, Article 52) nomen novum (Article 58).— Fig. 670.

Type — Russia: On dead stems and upper and lower surface of dead leaves of *Phlomis pungens* (Lamiaceae). (LE 35018, lectotype, here designated, LE 35019, isotype).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. superflua*, with large ascomata, asci cylindrical, 17-21 \times 3-4 μ m.

Mycosphaerella phlomidis (Bubák & Gonz. Frag.) Tomilin, Novosti Sist. Nizsh. Rast. 1966: 150. 1966, later homonym (illegitimate, Article 53) \equiv *Sphaerella phlomidis* Bubák & Gonz. Frag., Hedwigia 57: 4. 1915 [“1916”].— Fig. 671.

Type — Spain: Sevilla, Pedrozo de la Sierra. On dead calyces and pedicels of *Phlomis purpurea* (Lamiaceae). Bubák and Fragoso no. 155, I 1914 (MA, holotype).

Already synonymised with *Davidiella allicina* (as *Mycosphaerella tassiana*) by von Arx (1949), with which the type agrees well, with asci pyriform, ascospores 17-23 × 5-7 µm.

Sphaerella phoenicis Ces., in Rabenh., Fungi Europaei Exsiccati, ed. nov. ser. 2 no. 2531. 1881, nomen nudum (not validly published, Article 32) ≡ *Phoma phoenicis* Sacc., Syll. Fung. 11: 493. 1895 ≡ *Phomopsis cesatii* Gonz. Frag., Asoc. Esp. Progr. Ci. 6: 39. 1921, superfluous (illegitimate, Article 52) nomen novum (Article 58).

Type — Italy: Napoli. On dead bracts of *Phoenix dactylifera* (Arecaceae). (NY, isotype).

According to Saccardo (op. cit.), this is a coelomycete, with which the isotype agrees.

Mycosphaerella phragmitis (Ellis & Everh.) Lavrov, Trudy Toms. Gosud. Univ. 100: 141. 1951 ≡ *Sphaerella phragmitis* Ellis & Everh., Proc. Acad. Nat. Sci. Philadelphia 45: 446. 1893.— Fig. 672.

Type — USA: Indiana, Pine. On dead leaves, immersed between upper and lower epidermis, of *Phragmites australis* ["*communis*"] (Poaceae). Harper no. 4, IX 1892 ["1893"] (NY, holotype; NY, 2 isotypes).

Cited as synonymous with *M. polygramma* by Tomilin (1979). The types show that this belongs to *Davidiella*, and is morphologically indistinguishable from *D. disseminata*, with asci pyriform, ascospores 13-16 × 4.5-6 µm.

Mycosphaerella phyllachoroides (Sacc.) Tomilin, Novosti Sist. Nizsh. Rast. 1966: 173. 1966 ≡ *Sphaerella phyllachoroides* Sacc., Nuovo Giorn. Bot. Ital. 7: 304. 1875.— Fig. 673.

Type — Italy On dead leaves of *Agrostis* (Poaceae). Saccardo (PAD, holotype).

This is a *Didymella* species, with paraphysoids present, ascospores 10-12 × 3.5-4 µm.

Mycosphaerella phyllanthi Dennis, Kew Bull. 26: 45. 1971.— Fig. 674.

Type — Kenya: Nyeri, Aberdare Moorland, Magura River. On lower surface of living leaves of *Phyllanthus* (Phyllanthaceae). Verdcourt no. 4001A, III 1964 (K, holotype).

This is a parasitic species, with ascomata aggregated with 5-20 together in dense stromata, asci cylindrical, ascospores 12-16 × 2-3 µm.

Mycosphaerella phyllitis Petr., Bot. Jahrb. Syst. 62, Beibl. 142: 124. 1928.— Fig. 675.

Type — Madeira: Ribeiro Frio. On upper and lower surface of dead leaves of *Phyllis nobla* (Rubiaceae). Ade, VI 1926 (W no. 10055, holotype; W no. 26835, isotype).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. superflua*, with asci cylindrical, ascospores 15-18 × 3-4 µm.

Mycosphaerella phyllostachydicola Tomilin. See *Mycosphaerella shibataeae* I. Miyake & Hara.

Mycosphaerella phyllostachydis (Hara) Hara, in Hino & Katum., Bull. Fac. Agric. Yamagata Univ. 9: 901. 1958 ≡ *Mycosphaerella bambusae* var. *phyllostachydis* Hara, Bot. Mag. (Tokyo) 27: 249. 1913 [as "*phyllostachydos*"].

Type — Japan: *Phyllostachys bambusoides* (Poaceae).

No material was studied as the type was not included in a loan from TNS.

Mycosphaerella physostegiae W.A. Jenkins, Phytopathology 35: 329. 1945.

Type — USA: Virginia, Chatham. On upper and lower surface of dead leaves of *Physostegia virginiana* (Lamiaceae). Jenkins, VI 1944 (BPI, holotype).

Anamorph: *Cercospora physostegiae* W.A. Jenkins *vide* Jenkins (op. cit.). The type and numerous other specimens collected by Jenkins were all postmature, and their identity remains unknown.

Mycosphaerella phyteumatis (Jacz.) Lindau, Hilfsb. Sammeln Ascomyceten: 76. 1903 ≡ *Sphaerella phyteumatis* Jacz., Bull. Soc. Mycol. France 12: 113. 1896.— Fig. 676.

Type — Switzerland: *Phyteuma spicatum* (Campanulaceae). The type was not included in loans from LE or LEP. Material studied (Latvia, Vidzeme, Vestiena, Starcs no. 1541, V 1934, B, 2×) belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradians*, with asci cylindrical, ascospores 12-14 × 2.5-3.5 µm.

Sphaerella picconii De Not., Comment. Soc. Crittog. Ital. 2(3): 457. 1867 ≡ *Didymella picconii* (De Not.) Sacc., Syll. Fung. 1: 548. 1882.— Fig. 677.

Type — Italy: Albissola marina. On living young twig of *Picea abies* (Pinaceae). Piccone, 1864 (RO, holotype).

This is a species of *Arthopyrenia*, with pseudoparaphyses simple, cellular, with cells 5-9 × 1 µm, asci cylindrical with small ocular chamber, ascospores ellipsoid with rounded ends, 12-15 × 2-3 µm. The species does not fit any *Arthopyrenia* species treated in the recent literature or otherwise known to the author. Therefore the following new combination is proposed: **Arthopyrenia picconii** (De Not.) Aptroot comb. nov., **MB 374198**. **Basionym:** *Sphaerella picconii* De Not., Comment. Soc. Crittog. Ital. 2(3): 457. 1867.

Sphaerella pieris Sacc., Nuovo Giorn. Bot. Ital. 7: 302. 1875.— Fig. 678.

Type — Italy: Cavolo. On pale spots with black margins on upper surface of living leaves of *Tussilago farfara* (Asteraceae). Saccardo (PAD, holotype).

Anamorph: *Phyllosticta farfarae* Sacc. *vide* Saccardo (op. cit.).

This is a *Didymella* species, with paraphysoids present, ascospores 15-19 × 5-7.5 µm. It was however cited as synonymous with *Davidiella allicina* (as *Mycosphaerella allicina*) by Tomilin (1979).

Mycosphaerella piliostigmatis T. Kobay. & E.D. Guzmán, Bull. Forestry Forest Prod. Res. Inst., Ushiku 351: 157. 1988.

Type — Philippines: *Piliostigma malavaricum* var. *acidum* (Fabaceae).

Anamorph: *Cercospora bauhiniae* Syd. & P. Syd. *vide* Kobayashi & de Guzman (op. cit.) (= *Pseudocercospora bauhiniae* (Syd. & P. Syd.) Deighton).

No material was studied as the type was not included in a loan from TNS.

Mycosphaerella pimpinellae Petr., Hedwigia 74: 33. 1934.— Fig. 679.

Type — Russia: Siberia, Tomsk. On dead stems of *Pimpinella saxifraga* (Apiaceae). Ziling no. 122, VI 1927 (W, holotype).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradians*, with asci cylindrical, ascospores 11-13 × 2.5-3.5 µm.

Sphaerella pinastri (DC.) Cooke, J. Bot. 4: 248. 1866 ≡ *Sphaeria pinastri* DC., Fl. Française 5(6): 133. 1815 ≡ *Laestadia pinastri* (DC.) Sacc., Syll. Fung. 1: 426. 1882.

Type — France: *Pinus* (Pinaceae).

No material was studied as the species was already redisposed.

Mycosphaerella pini Rostr., in Munk, Dansk Bot. Ark. 17(1): 312. 1957 ≡ *Eruptio pini* (Rostr.) M.E. Barr, Mycotaxon 60: 438. 1996.— Fig. 680.

Type — Denmark: Jutland, Tvorup. On dead needles of *Pinus montana* (“*silvestris*” in publication) (Pinaceae). Rostrup, X (C, holotype).

Accepted as *Eruptio pini* (Rostr.) M.E. Barr by Barr (op. cit.). The holotype shows that this is indeed a species of *Mycosphaerella*, and is morphologically indistinguishable from *Mycosphaerella juniperina*, with ascomata thick-walled, breaking through the epidermis, asci pyriform, ascospores 10-12 × 3-3.5 µm.

Mycosphaerella pini (A. Funk & A.K. Parker) Arx, Proc. Kon. Ned. Akad. Wetensch. C 86: 33. 1983, later homonym (illegitimate, Article 53) ≡ *Scirrhia pini* A. Funk & A.K. Parker, Canad. J. Bot. 44: 1171. 1966.

Type — Canada: *Pinus* (Pinaceae).

Anamorph: *Dothistroma septospora* (Dorog.) M. Morelet *vide* Evans (1984).

No material was studied as the species is thought to belong indeed to *Scirrhia*.

Mycosphaerella pinicola (Fautrey) Naumov, Zap. Ural'sk Obshch. Lyubit. Estestv. 35: 21. 1915 ≡ *Sphaerella pinicola* Fautrey, in Roum., Rev. Mycol. (Toulouse) 13: 125. 1891.

Type — France: Côte-d'Or, Epoisses. On dead needles of *Pinus austriaca* (Pinaceae). Fautrey, distributed in Roumeguère, Fungi Selecti Exsiccati no. 5719, IX 1890 (PC, holotype).

The holotype contains only empty ascomata.

Sphaerella pinicola f. *conorum* Naumov ex Trotter, Syll. Fung. 24: 858. 1928.

Type — Russia: *Pinus* (Pinaceae).

No material was studied as the type was not included in loans from LE or LEP, but is certainly no *Mycosphaerella*.

Mycosphaerella pinifolia (Ducomet) Tomilin, Novosti Sist. Nizsh. Rast. 1966: 173. 1966 ≡ *Sphaerella pinifolia* Ducomet, Ann. École Nat. Agric. Rennes 2: 23. 1908.— Fig. 681.

Type — France: On dead, but still attached, needles *Pinus maritima* (Pinaceae).

The location of the type is unknown. Material studied (India, Burrihat, State Forest Service, on *P. kesiya*, Mehrotra, VIII 1981, IMI no. 261599 and no. 261601b) is *Mycosphaerella juniperi*, with asci pyriform, ascospores 6-7 × 2.5-3 µm.

Mycosphaerella pini-patulae M. Morelet, Bull. Soc. Sci. Nat. Archéol. Toulon & Var 196: 7. 1971.

Type — Madagascar: *Pinus patula* (Pinaceae).

No material was studied as the location of the type is unknown.

Mycosphaerella pinodes (Berk. & A. Bloxam) Vestergr., Bih. Kongl. Svenska Vetensk.-Akad. Handl. 22 (3, 6): 15. 1896 ≡ *Sphaeria pinodes* Berk. & A. Bloxam, in Berk. & Broome, Ann. Mag. Nat. Hist., ser. 3, 7: 454. 1861 ≡ *Sphaerella pinodes* (Berk. & A. Bloxam) Niessl, in Rabenh., Fungi Europaei Exsiccati, cent. 20 no. 1947. 1875 ≡ *Didymellina pinodes* (Berk. & A. Bloxam) Höhn., Ann. Mycol. 16: 67. 1918 ≡ *Didymella pinodes* (Berk. & A. Bloxam) Petr., Ann. Mycol. 22: 18. 1924.— Fig. 682.

Type — United Kingdom: On dead stems of *Pisum sativum* (Fabaceae). (K, holotype [sub “*pinoides*”]).

Anamorph: *Ascochyta pinodes* L.K. Jones *vide* Sivanesan (1984).

Accepted as *Didymella pinodes* (Berk. & A. Bloxam) Petr. by Corbaz (1956), with which the holotype agrees well, with ascomata pyriform, hamathecium present (but rudimentary due to bad preservation), asci thick-walled, clavate, ascospores 14-18 × 5-7.5 µm. Additional material studied (Austria, Voitsberg, Niessl, distributed in Rabenhorst, Fungi Europaei Exsiccati no. 1947, L) is morphologically indistinguishable from *Davidiella allicina*, with asci pyriform, ascospores 14-17 × 6-7 µm.

Mycosphaerella pinsapo (Thüm.) Lindau, Hilfsb. Sammeln Ascomyceten: 2. 1903 ≡ *Sphaerella pinsapo* Thüm., in Bolle & Thüm., Boll. Soc. Adriat. Sci. Nat. Trieste 6. 1880.

Type — Yugoslavia: *Abies pinsapo* (Pinaceae).

No material was studied as the type was not found in any of the herbaria consulted.

Mycosphaerella piperis Sawada ex Aptroot, spec. nov. **500504**. J. Taihoku Soc. Agric. 7: 126. 1943, lacking Latin description (not validly published, Article 36). Latin description: *Mycosphaerella piperis infestans ascosporis* 14-15 × 3-3.5 µm.— Fig. 683.

Type — Taiwan: Taipei, Urai. On white spots with black margins on upper surface of living leaves of *Piper futokadsura* (Piperaceae). Sawada, IV 1915 (BPI, isotype).

This is a parasitic species, with asci cylindrical, ascospores 14-15 × 3-3.5 µm. The name is formally validated here, so that it can be used.

Mycosphaerella pirina (Ellis & Everh.) J.H. Mill. See *Mycosphaerella pirina* (Ellis & Everh.) J.H. Mill.

Mycosphaerella pirolae (Rostr.) Lind. See *Mycosphaerella pyrolae* (Rostr.) Lind.

Mycosphaerella pirolina Kirschst. See *Mycosphaerella pyrolina* Kirschst.

Mycosphaerella pistaciae (Cooke) Tomilin, Novosti Sist. Nizsh. Rast. 6: 121. 1970 [“1969”] ≡ *Sphaerella pistaciae* Cooke, J. Bot. 21: 109. 1883.— Fig. 684.

Type — France: Bouches du Rhône, Marseille. On upper surface of dead leaves of *Pistacia vera* (Anacardiaceae). Roux 207, II 1872 (K, lectotype, designated here; K, isotype).

This is morphologically indistinguishable from *Davidiella allicina*, with asci pyriform, ascospores 17-23 × 6-7.5 μm.

Mycosphaerella pistaciarum Chitzan., Ann. Inst. Phytopathol. Benaki 10: 42. 1956.

Type — Greece: *Pistacia vera* (Anacardiaceae). Anamorph: *Septoria pistaciarum* Caracc. fide Chitzanidis (op. cit.).

No material was studied as the location of the type is unknown.

Mycosphaerella pistacina Chitzan., Ann. Inst. Phytopathol. Benaki 10: 42. 1956.

Type — Greece: *Pistacia vera* (Anacardiaceae). Anamorph: *Septoria pistacina* Allesch. fide Chitzanidis (op. cit.).

No material was studied as the location of the type is unknown.

Mycosphaerella pithecolobiicola (Speg.) Cash, Syll. Fung. 26: 350. 1972 [as “*pithecolobiicola*”] ≡ *Sphaerella pithecolobiicola* Speg., Anales Mus. Nac. Hist. Nat. Buenos Aires 31: 410. 1922 [as “*pithecolobiicola*”].

Type — Paraguay: Asunción. On upper and lower surface of dead leaves of *Pithecolobium scalare* (Fabaceae). Spegazzini no. 6299, VII 1919 (LPS, holotype).

The holotype contains only empty ascomata.

Mycosphaerella pittierii Syd., Ann. Mycol. 28: 77. 1930.— Fig. 685.

Type — Venezuela: Caguita. On upper and lower surface of dead leaves of *Ficus* (Moraceae). Sydow, XII 1927, Fungi Exotici Exsiccati no. 811 (L, 2 isotypes), also distributed in Reliquiae Petrakianae no. 657 (H, L, isotypes).

This is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores 8-10 × 2.5-3 μm.

Mycosphaerella pittospori (Cooke) A. Weiss, Index Plant Diseases United States: 861. 1950 ≡ *Sphaerella pittospori* Cooke, Grevillea 7: 53. 1878.

Type — USA: South Carolina, Aiken. On upper and lower surface of dead leaves of *Pittosporum* (Pittosporaceae). Ravenel no. 2589 (K, holotype), also distributed in Fungi Americani Exsiccati no. 382 (NY, 2 isotypes).

In the types only a coelomycete could be found.

Mycosphaerella plantaginicola (Schwein.) Dearn., in Dearn. & House, New York State Mus. Bull. 233-234: 38. 1921 ≡ *Sphaeria plantaginicola* Schwein., Trans. Amer. Philos. Soc., ser. 2, 4: 226. 1832 ≡ *Sphaerella plantaginicola* (Schwein.) Cooke, J. Bot. 21: 139. 1883.— Fig. 687.

Type — USA: Philadelphia, Bethlehem. On lower surface of dead leaves of *Plantago lanceolata* (Plantaginaceae). Schweinitz, 1829 (PH, holotype; PH, isotype).

Both type specimens are immature, but show that this is morphologically indistinguishable from *M. punctiformis*, with cylindrical asci.

Mycosphaerella plantaginicola (Pat.) F. Stevens, Illinois Biol. Monographs 11: 199. 1927, later homonym (illegitimate, Article 53) and based on an illegitimate name ≡ *Sphaerella plantaginicola* Pat., in Pat. & Lagerh., Bull. Soc. Mycol. France 9: 153. 1893, later homonym (illegitimate, Article 53) ≡ *Carlina plantaginicola* Höhn., Mitt. Bot. Lab. TH Wien, ser. 2, 3: 88. 1925, nomen novum (Article 58).— Fig. 686.

Type — Ecuador: Quito, Pancillo. On upper and lower surface of dead leaves of *Plantago* (Plantaginaceae). Lagerheim, II 1892 (FH-Patouillard, holotype).

The holotype and additional material studied (Ecuador, Prov. Tungurahua, Baños, on not delineated brown spots on upper and lower surface of living leaves of *Plantago hirtella*, Sydow, Fungi Exotici Exsiccati no. 1192, XII 1937, B, L) are morphologically indistinguishable from *Wettsteinina kashmirensis* Shoemaker & C.E. Bab., for which it represents an earlier epithet, with asci pyriform, ascospores with one median septum and without or with 2 additional pseudosepta, 25-40 × 7-10 μm. Therefore the following new combination is proposed: **Wettsteinina plantaginicola** (Höhn.) Aptroot comb. nov., **MB 500379**. **Basionym:** *Carlina plantaginicola* Höhn., Mitt. Bot. Lab. TH Wien, ser. 2, 3: 88. 1925. The holotype also contains *Mycosphaerella ammophilae*, which does not fit the protologue.

Mycosphaerella plantaginis (Sollm.) Vesterg., Bih. Kongl. Svenska Vetensk.-Akad. Handl. 22(3, 6): 15. 1896 ≡ *Sphaerella plantaginis* Sollm., Bot. Zeitung (Berlin) 22: 281. 1864.— Fig. 688.

Type — Germany: Coburg. On dead stems and flowers of *Plantago lanceolata* (Plantaginaceae). Sollmann, distributed in Rabenhorst, Fungi Europaei Exsiccati no. 763 (L, isotype).

Anamorph: *Septoria plantaginis* (Ces.) Sacc. fide Saccardo (1882).

This is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores 10-12 × 2-3 μm.

Mycosphaerella plantaginis (Ellis) Theiss. See *Mycosphaerella theissenii* Tomilin.

Mycosphaerella platani (Ellis & G. Martin) Tomilin, Novosti Sist. Nizsh. Rast. 1968: 167. 1968 ≡ *Sphaerella*

platani Ellis & G. Martin, Amer. Naturalist 19: 77. 1885.— Fig. 689.

Type — USA: On lower surface of dead leaves of *Platanus occidentalis* (Platanaceae).

Anamorph: Associated with *Phyllosticta fide* Ellis & Martin (op. cit.).

The type was not found in NY and is probably lost. Material studied (North Carolina, Durham, Wolf, 1936, B, IMI no. 227777) is *M. punctiformis*, with asci cylindrical, ascospores $7-8 \times 2-2.5 \mu\text{m}$.

Mycosphaerella platanifolia (Cooke) F.A. Wolf, Mycologia 30: 62. 1938 \equiv *Sphaerella platanifolia* Cooke, J. Bot. 21: 106. 1883.— Fig. 690.

Type — USA: Georgia, Darien. On lower surface of dead leaves of *Platanus occidentalis* (Platanaceae). Ravenel, III 1881, distributed in Fungi Americani Exsiccati no. 756 (K (5 \times), NY, isoelectotypes, designated by Crous & Corlett, 1998).

Anamorph: *Cercospora platanicola* Ellis & Everh. fide Wolf (op. cit.) (= *Pseudocercospora platanicola* (Ellis & Everh.) U. Braun), but not fide Crous & Corlett (1998).

Reported to be morphologically indistinguishable from *Mycosphaerella albocrustata* (Schwein.) Crous & Corlett, which probably is morphologically indistinguishable from *M. punctiformis*, by Crous & Corlett (1998). The isoelectotypes contain mostly postmature ascomata that may represent *M. punctiformis*, but also a *Didymella* with asci clavate, pseudoparaphyses anastomosing, ascospores $13-15 \times 3-4 \mu\text{m}$.

Mycosphaerella platylobii Sivan. & R.G. Shivas, Mycol. Res. 106: 360. 2002.

Type — Australia: *Platylobium fromosum* (Fabaceae).

No material was studied of this recently described species.

Mycosphaerella platythea (P. Karst.) Tomilin, Novosti Sist. Nizsh. Rast. 6: 121. 1970 [“1969”] \equiv *Sphaerella platythea* P. Karst., Hedwigia 22: 179. 1883.

Type — Finland: *Tilia platyphylla* (Malvaceae).

No material was studied as the type was not included in a loan from H.

Mycosphaerella plectranthi Doidge, Bothalia 4: 862. 1948.

Type — South Africa: *Plectranthus fruticosus* (Lamiaceae).

No material was studied as the type was not found in any of the herbaria consulted.

Sphaerella plegmariae Ces. & Becc., in Ces., Atti Accad. Sci. Fis. 8: 23. 1879 \equiv *Metasphaeria plegmariae* (Ces. & Becc.) Sacc., Syll. Fung. 2: 183. 1883.

Type — Sri Lanka: *Lycopodium plegmaria* (Lycopodiaceae).

No material was studied as the type was not found in any of the herbaria consulted

Sphaerella pleuronervia (De Not.) Ces. & De Not., Comment. Soc. Crittog. Ital. 1(4): 237. 1863 \equiv *Sphaeria pleuronervia* De Not., Mem. Reale Accad. Sci. Torino, ser. 2, 16: 457. 1857.— Fig. 691.

Type — Italy. Oregina, Margherita. On upper and lower surface of dead leaves of *Quercus*. De Notaris 21, I 1842 (RO, holotype; B, isotype).

The holotype, the isotype and two paratypes (Genua, V 1842, RO and Mediola, 1832, RO) show that this is morphologically indistinguishable from *Plagiostoma pustula* (Pers. : Fr.) Arx, with asci dehiscent, ascospores pale yellowish, 1-septate, $20-25 \times 3.5-4.5 \mu\text{m}$, with polar gelatinous appendages.

Mycosphaerella pneumatophorae Kohlm., Ber. Deutsch. Bot. Ges. 79: 32. 1966.— Fig. 692, 976, 977.

Type — Liberia: Upper Buchanan, Benson River. On or immersed in bark of living and dead areal roots of *Avicennia nitida* (Acanthaceae). Kohlmeyer no. 1783, III 1965 (B, holotype; NY, isotype).

This species shows close connections to a genus with several well-known maritime species, viz. *Collemopsidium* (syn. *Pyrenocollema*) in the Xanthopyreniaceae. It is characterized by ascomata only carbonized above, wall small-cellular with lumina ca. $3 \mu\text{m}$, asci clavate, without ocular chamber, pseudoparaphyses anastomosing, ca. $2 \mu\text{m}$ wide, ascospores rather thick-walled, $15-21 \times 6-9 \mu\text{m}$, somewhat ornamented when old. Additional material studied (USA, Florida, Pirate Grove Key, Kohlmeyer no. 1710, I 1964, B) agrees. Therefore the following combination is proposed:

Collemopsidium pneumatophorae (Kohlm.) Aptroot comb. nov., **MB 500337**. **Basionym:** *Mycosphaerella pneumatophorae* Kohlm., Ber. Deutsch. Bot. Ges. 79: 32. 1966. This species is the first non-lichenized member of the genus, but the known species already vary considerably in phycobiont presence, from a dominant cyanobacterium in the type species to nearly absent (and often various different cyanobacteria or even a green alga present) in most species.

Mycosphaerella podagrariae (Roth : Fr.) Petr., Ann. Mycol. 19: 203. 1921 \equiv *Sphaeria podagrariae* Roth, Catalecta Botanica, fasc. 1 no. 230. 1797 \equiv *Dothidea podagrariae* (Roth) Fr., Systema Mycol. 2: 556. 1823 \equiv *Phyllachora podagrariae* (Roth : Fr.) P. Karst., Bidrag Kännedom Finlands Natur Folk 23: 228. 1873 \equiv *Carlia podagrariae* (Roth : Fr.) Höhn., Centralbl. Bakteriolog., 2. Abth., 60: 2. 1923 \equiv *Oligostroma podagrariae* (Roth : Fr.) Höhn. ex Petr., Ann. Mycol. 19: 203. 1921.

Type — Sweden: On spots on upper and lower surface of living leaves of *Aegopodium podagraria* (Apiaceae).

Anamorph: *Septoria podagrariae* Lasch fide von Arx (1983).

The type was not found in any of the herbaria studied. Materials studied (Switzerland, Zürich, Schinz, distributed in Cryptogamae Vindobonenses no. 3357, L; also United Kingdom, Scotland, Ivernesshire, Beaully, Cluanie Deer Farm Park, Cannon, VIII 1990, IMI no. 345065) contain only the anamorph and immature ascomata in superficial stromata.

Mycosphaerella podocarpus (Cooke) Lindau, in Engl. & Prantl, Natürliches Pflanzenf. 1(1): 425. 1897 \equiv *Sphaerella podocarpus* Cooke, J. Bot. 21: 106. 1883.— Fig. 693.

Type — Indonesia: Java, Bogor, Botanical Garden. On upper and lower surface of dead “leaves” of *Podocarpus* (Podocarpaceae). “KWB” no. 549 (K, 2 isotypes).

The isotypes studied probably represent a species of *Dothidea*, with large parenchymatous cells between the asci, asci clavate, thick-walled, ascospores immature, 13-16 × 5-6 μm.

Additional material studied (Japan, Prov. Mino, Kawaue-mura, on upper and lower surface of dead needles of *Podocarpus macrophyllus*, Hara, distributed in Sydow, Fungi exotici exsiccati no. 175, VII 1912, L) has ascomata crowded aggregated in stromata, asci pyriform, ascospores 12-16 × 3.5-4.5 μm.

Mycosphaerella podocarpicola Henn., Verh. Bot. Vereins Prov. Brandenburg 40: 156. 1898 ≡ *Sphaerella podocarpicola* (Henn.) Sacc. & P. Syd., Syll. Fung. 16: 473. 1902.

Type — Germany: *Podocarpus chinensis* (Podocarpaceae). No material was studied as the type was not found in SP or B and is probably lost. Cited as synonymous with *Mycosphaerella podocarpi* by Tomilin (1979).

Mycosphaerella podperae Picb., in Baudyš & Picb., Práce Morav. Přír. Společn. 1: 294. 1924.

Type — Czech Republic: *Scorzonera austriaca* (Asteraceae).

No material was studied as the type was not found in any of the herbaria consulted. Synonymised with *Davidiella allicina* (as *Mycosphaerella tassiana*) by von Arx (1949), probably without examination of type material.

Mycosphaerella pogostemonis Khokhr., Bot. Mater. Otd. Sporov. Rast. Bot. Inst. Komarova Akad. Nauk SSSR 7: 143. 1951.— Fig. 694.

Type — Russia: On white spots on upper surface of living leaves of *Plectranthus graveolens* [“*Pogostemon patchouly*”] (Lamiaceae). (LEP, holotype).

Anamorph: associated with *Phyllosticta pogostemonis* Khokhr. *fide* Khokhrjakov (op. cit.).

This belongs to *Davidiella*, of which it represents a parasitic species, with asci pyriform, ascospores 16-19 × 5-8 μm. Therefore the following new combination is made:

Davidiella pogostemonis (Khokhr.) Aptroot comb. nov., **MB 500358**. **Basionym:** *Mycosphaerella pogostemonis* Khokhr., Bot. Mater. Otd. Sporov. Rast. Bot. Inst. Komarova Akad. Nauk SSSR 7: 143. 1951. The anamorph, described as *Phyllosticta pogostemonis*, is morphologically indistinguishable from *Phoma exigua* Desm.

Mycosphaerella polaris (P. Karst.) Lindau, in Engl. & Prantl, Natürlichen Pflanzenf. 1(1): 424. 1897 ≡ *Sphaerella genuflexa* var. *polaris* P. Karst., Öfvers. Förh. Kongl. Svenska Vetensk.-Akad. 1872(2): 104. 1872 ≡ *Sphaerella polaris* (P. Karst.) Sacc., Syll. Fung. 1: 486. 1882.

Type — Svalbard: *Salix polaris* (Salicaceae).

No material was studied as the type was not included in a loan from H.

Mycosphaerella polemonii Lind, Biol. Meddel. Kongel. Danske Vidensk. Selsk. 11(2): 71. 1934.

Type — Denmark: *Polemonium humilis* (Polemoniaceae).

No material was studied as the type was not found in C. Cited as synonymous with *Mycosphaerella pachypleuri* by Tomilin (1979).

Mycosphaerella polia Petr., in Syd., Ann. Mycol. 22: 261. 1924 ≡ *Sphaerella polia* (Petr.) Kirschst., Kryptog.-Fl. Mark Brandenburg 7(3): 358. 1938.— Fig. 695.

Type — Germany: Schlesien, Krummhübel, Riesengebirge. On dead stems of *Epilobium dodonaei* (Onagraceae). Sydow, Mycotheca Germanica no. 2114, VIII 1922 (L, isotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. ammophilae*, with asci pyriform, ascospores 9-12 × 3-3.5 μm.

Mycosphaerella polifoliae (Ellis & Everh.) Bubák, Ann. Mycol. 4: 109. 1906 [as “*poliifolia*”] ≡ *Sphaerella polifoliae* Ellis & Everh., Proc. Acad. Nat. Sci. Philadelphia 42: 231. 1891 [“1890”, as “*polifolia*”].— Fig. 696.

Type — Canada: London. On upper surface of dead parts of living leaves of *Andromeda polifolia* (Ericaceae). Dearness, VIII 1889 (NY, holotype).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradians*, with asci cylindrical, ascospores 9-12 × 3-4 μm.

Mycosphaerella polycarpa (Kirschst.) Tomilin, Novosti Sist. Nizsh. Rast. 10: 111. 1973 ≡ *Sphaerella polycarpa* Kirschst., Kryptog.-Fl. Mark Brandenburg 7: 387. 1938.— Fig. 697.

Type — Germany: Krossen, Baudach. On upper and lower surface of dead leaves of *Syringa vulgaris* (Oleaceae). Kirschstein, IV 1908 (B, holotype).

This is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores 8-11 × 2.5-3 μm.

Mycosphaerella polygalina Petr., Hedwigia 68: 209. 1928.— Fig. 698.

Type — Russia: Siberia, Tsherepanovo-Tshumysk. On dead stems of *Polygala* (Polygalaceae). Murashinsky no. 413, VI 1925 (W, holotype).

Cited as synonymous with *Mycosphaerella hypsicola* by Tomilin (1979). This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradians*, with asci cylindrical, ascospores 9-12 × 2.5-3 μm.

Sphaerella polygonati (Schwein.) Cooke, J. Bot. 21: 70. 1883 ≡ *Sphaeria polygonati* Schwein., Trans. Amer. Philos. Soc., ser. 2, 4: 225. 1832 ≡ *Laestadia polygonati* (Schwein.) Sacc., Syll. Fung. 2: XXXII. 1883 ≡ *Guignardia polygonati* (Schwein.) Lindau, in Engl. & Prantl, Natürlichen Pflanzenf. 1(1): 423. 1897.

Type — USA: Bethlehem. On upper and lower surface of dead leaves of *Polygonatum* (Asparagaceae). Schweinitz (PH, holotype; PH, isotype).

The type specimens are immature, but show that this belongs to section *Caterva* and is morphologically indistinguishable from *M. subradians*, with asci cylindrical.

Mycosphaerella polygoni-cuspidati Hara, J. Pl. Protect. 5: 617. 1918 [as "*polygonii cuspidatii*"].

Type — Japan: *Polygonum cuspidatum* (Polygonaceae).

No material was studied as the type was not included in a loan from TNS.

Mycosphaerella polygonorum (Crié) Lind, Skr. Vidensk.-Selsk. Christiania, Math.-Naturvidensk. Kl. 1909(9): 6. 1910 ["1909", as "*polygonum*"] = *Depazea polygonorum* Crié, Ann. Sci. Nat. Bot., ser. 6, 7: 45. 1878 = *Sphaerella polygonorum* (Crié) Sacc., Syll. Fung. 1: 512. 1882, later homonym (illegitimate, Article 53).

Type — France: *Polygonum* (Polygonaceae).

Anamorphs: Associated with *Diplodia*, *Hendersonia* and *Septoria* fide Crié (op. cit.).

No material was studied as the type was not included in a loan from PC.

Sphaerella polygonorum Auersw., in Marcucci, Unio Itineraria Exsiccati no. 3. 1867 = *Laestadia polygonorum* (Auersw.) Sacc., Syll. Fung. 2: XXXII. 1883.— Fig. 699.

Type — Italy: Sardinia, Guspini. On dead stems of *Polygonum equisetiforme* (Polygonaceae). Marcucci, Unio Itineraria Exsiccati no. 3, 1866 (L, 2 isotypes).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. ammophilae*, with asci pyriform, ascospores immature, ca. $11 \times 3 \mu\text{m}$.

Mycosphaerella polygramma (Fr.) Starbäck, Bih. Kongl. Svenska Vetensk.-Akad. Handl. 15(3, 2): 9. 1889 = *Sphaeria polygramma* Fr., Systema Mycol. 2: 432. 1832 = *Sphaeropsis polygramma* (Fr.) Fr., Summa Veget. Scand., sect. post.: 419. 1849 = *Sphaerella polygramma* (Fr.) Niessl, Oesterr. Bot. Z. 25: 87. 1875.

Type — Sweden: On dead stems of *Ballota nigra* (Lamiaceae). Fries, Scleromyceti Suecici no. 196 (UPS, holotype; B, isotype).

Anamorph: *Phoma polygramma* Sacc. fide Grove (1935).

The types studied only contain an immature ascomycete and a *Phoma* anamorph. Additional material studied (Germany, Brandenburg, Buckow, Sydow, Mycotheca Germanica no. 238, V 1904, L) contains only a coelomycete.

Sphaerella polygramma f. *betonicae* Fautrey, Rev. Mycol. (Toulouse) 13: 8. 1891.— Fig. 700.

Type — France: Forêt de Clamercy. On dead stems of *Betonica officinalis* (Lamiaceae). Fautrey, distributed in Roumeuguère, Fungi Selecti Exsiccati no. 5544, V 1890 (PC, holotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. ammophilae*, with asci pyriform, ascospores $11-13 \times 3-3.5 \mu\text{m}$.

Mycosphaerella polygramma f. *origani* (Gonz. Frag.) Maire & Werner, Mém. Soc. Sci. Nat. Maroc 45: 27. 1938 ["1937"]: *Sphaerella polygramma* f. *origani* Gonz. Frag., Bol. Soc. Esp. Hist. Nat. 25: 103. 1925.— Fig. 701.

Type — Mauritania: Samoa. On dead stems of *Origanum compactum* (Lamiaceae). Pau no. 7098, VIII 1922 (MA, holotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. ammophilae*, with asci pyriform, ascospores $9-11 \times 3-3.5 \mu\text{m}$.

Sphaerella polygramma var. *caruanae* Sacc., Nouvo Giorn. Bot. Ital., n. ser., 22: 35. 1915.

Type — Malta: *Teucrium flavum* (Lamiaceae).

No material was studied as the type was not found in any of the herbaria consulted.

Mycosphaerella polymorpha D.J. Sm. & C.O. Sm. See *Mycosphaerella stigmia-platani* F.A. Wolf.

Mycosphaerella polypodii (Rabenh.) Johanson ex Oudem., Rév. Champ. Pays-Bas 2: 205. 1897 = *Sphaeria polypodii* Rabenh., Herb. Vivum Mycol., ed. nov. no. 533. 1857 = *Sphaerella polypodii* (Rabenh.) Ces. & De Not., Comment. Soc. Crittog. Ital. 1(4): 238. 1863 = *Leptosphaeria polypodii* (Rabenh.) Auersw., in Gonn. & Rabenh., Mycol. Europaea 5-6: 20. 1869 = *Laestadia polypodii* (Rabenh.) Magnus & Sacc., Atti Reale Ist. Veneto Sci., ser. 6, 3: 737. 1885 = *Physospora polypodii* (Rabenh.) Höhn., Ann. Mycol. 16: 55. 1918 = *Plectosphaera polypodii* (Rabenh.) Arx & E. Müll., Beitr. Kryptogamenfl. Schweiz 11(1): 208. 1954.

Type — Switzerland: On lower surface of dead fronds of *Polypodium* (Polypodiaceae). Rabenhorst, Herb. Vivum Mycol., ed. nov. no. 533, 1856 (L, 2 isotypes).

Accepted as *Plectosphaera polypodii* (Rabenh.) Arx & E. Müll. by von Arx & Müller (op. cit.). The isotypes and additional material studied (Netherlands, Oudemans no. 434, L) contain only coelomycetes.

Sphaeria polypodii f. *aspidii* Fuckel, Fungi Rhenani Exsiccati no. 854. 1864 = *Sphaerella polypodii* f. *aspidii* (Fuckel) Fuckel, Jahrb. Nassauischen Vereins Naturk. 23-24: 102. 1870.— Fig. 702.

Type — Germany: Oestrich. On upper surface of dead fronds of *Dryopteris* ["*Aspidium*"] *filix-mas* (Polypodiaceae). Fuckel, Fungi Rhenani Exsiccati no. 854 (L, isotype).

This is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores $8-10 \times 2-2.5 \mu\text{m}$.

Sphaeria polypodii f. *asplenii* Fuckel, Fungi Rhenani Exsiccati no. 1775. 1866 = *Sphaerella polypodii* f. *asplenii* (Fuckel) Fuckel, Jahrb. Nassauischen Vereins Naturk. 23-24: 102. 1870.— Fig. 703.

Type — Germany: Kiedrich. On upper surface of dead leaf parts of living fronds of *Asplenium trichomanes* (Polypodiaceae). Fuckel, Fungi Rhenani Exsiccati no. 1775 (L, isotype).

This belongs to section *Caterva*, and is morphologically indistinguishable from *Mycosphaerella asplenii*, with asci cylindrical, ascospores $12-16 \times 3.5-4.5 \mu\text{m}$.

Sphaeria polypodii f. *polypodii-vulgaris* Fuckel, Fungi Rhenani Exsiccati no. 2019. 1867 = *Sphaerella polypodii* f. *polypodii-vulgaris* (Fuckel) Fuckel, Jahrb. Nassauischen Vereins Naturk. 23-24: 102. 1870.

Type — Germany: Oestrich. On upper surface of dead leaf parts of living fronds of *Polypodium vulgare* (Polypodiaceae). Fuckel, Fungi Rhenani Exsiccati no. 2019 (L, isotype).

The isotype studied is immature.

Sphaeria polypodii f. *pteridinis* Fuckel, Fungi Rhenani Exsiccati no. 853. 1864 = *Sphaerella polypodii* f. *pteridinis* (Fuckel) Fuckel, Jahrb. Nassauischen Vereins Naturk. 23-24: 102. 1870 [as “*pteridis*”].

Type — Germany: Oestrich. On upper and lower surface of dead leaf parts of living fronds of *Pteridium aquilinum* (Polypodiaceae). Fuckel, Fungi Rhenani Exsiccati no. 853 (L, isotype).

The isotype studied is immature.

Sphaeria polypodii f. *spermogonifera* Marcucci, Unio Itineraria Exsiccati no. 53. 1867 [as “*Sphaeriae*”].

Type — Italy: On lower surface of dead leaf parts of living fronds of *Polypodium* (Polypodiaceae). Marcucci, Unio Itineraria Exsiccati no. 53, 1866 (L, 3 isotypes).

The isotypes studied contain only an *Asteromella* spermatial state.

Mycosphaerella polyspora Johanson, Öfvers. Förh. Kongl. Svenska Vetensk.-Akad. 41: 164. 1884 = *Sphaerella polyspora* (Johanson) Berl. & Voglino, Syll. Fung., Addit. ad vol. 1-4: 82. 1886 = *Diplosphaerella polyspora* (Johanson) Grove, J. Bot. 50: 91. 1912 = *Delphinella polyspora* (Johanson) E. Müll., in E. Müll. & Arx, Beitr. Kryptogamenfl. Schweiz 11(2): 27. 1962.

Type — Sweden: *Rhododendron* [“*Azalea*”] *procumbens* (Ericaceae).

Accepted as *Delphinella polyspora* (Johanson) E. Müll. by Müller & von Arx (op. cit.) and therefore not studied.

Sphaerella polystigma Ellis & Everh., Bull. Torrey Bot. Club 10: 117. 1883 = *Laestadia polystigma* (Ellis & Everh.) Sacc., Syll. Fung. 9: 588. 1891 = *Anisostomula polystigma* (Ellis & Everh.) Höhn., Ann. Mycol. 16: 49. 1918 = *Pseudomassaria polystigma* (Ellis & Everh.) Arx, Ber. Schweiz. Bot. Ges. 62: 352. 1952.— Fig. 704.

Type — USA: Ohio, Fairfield Co. On lower and upper surface of dead leaves of *Quercus coccinea* (Fagaceae). Kellerman no. 257, V 1883 (NY, holotype), also distributed in Ellis & Everhart, North American Fungi no. 1353 (L, NY (4×), isotypes).

Accepted as *Pseudomassaria polystigma* (Ellis & Everh.) Arx by Müller & von Arx (1962), with which the types studied agree well, with asci clavate, ascospores asymmetrically septate, 10-12 × 3.5-5 µm.

Mycosphaerella pomacearum (Crié) Oudem., Verh. Kon. Ned. Akad. Wetensch., Afd. Natuurk., Tweede Sect. 11: 249. 1905 [“1904”] = *Depazea pomacearum* Crié, Ann. Sci. Nat. Bot., ser. 6, 7: 44. 1878 = *Sphaerella pomacearum* (Crié) Sacc., Syll. Fung. 1: 482. 1882.

Type — France: Rosaceae.

Anamorphs: *Septoria* and *Diplodia* fide Saccardo (1882).

The type was not included in a loan from PC. Material studied (Netherlands, Nunspeet, on upper and lower surface

of dead leaves of *Cydonia japonica*, Beins, IX 1899, L) is immature.

Mycosphaerella pomi (Pass.) Lindau, in Engl. & Prantl, Natürlichen Pflanzenf. 1(1): 424. 1897 = *Sphaerella pomi* Pass., in Rabenh., Fungi Europaei Exsiccati, ed. nov., ser. 2 no. 2443. 1876.— Fig. 705.

Type — Italy: Parma. On upper and lower surface of dead leaves of *Malus* (Rosaceae). Passerini, distributed in Rabenhorst, Fungi Europaei Exsiccati, ed. nov., ser. 2 no. 2443 (L, isotype).

Anamorph: *Cylindrosporium pomi* C. Brooks fide Farr et al. (1989).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores 17-22 × 5-7 µm.

Mycosphaerella pongamiae (Racib.) Sivan., Trans. Brit. Mycol. Soc. 84: 363. 1985 = *Stigmatea pongamiae* Racib., Parasitische Algen und Pilze Javas 3: 36. 1900.

Type — Indonesia: *Pongamia glabra* (Fabaceae).

Anamorph: *Asperisporium pongamiae* (Syd.) Deighton ex M.B. Ellis fide Sivanesan (op. cit.).

No material was studied as the location of the type is unknown.

Mycosphaerella pontederiae (Peck) House, New York State Mus. Bull. 233-234: 29. 1921 [as “*pontedereae*”] = *Sphaerella pontederiae* Peck, Annual Rep. New York State Mus. 40: 67. 1887.— Fig. 706.

Type — USA: New York, Washington Co., Whitehall. On pale spots on upper surface of living leaves of *Pontederia cordata* (Pontederiaceae). Peck (BPI, isotype).

This belongs to *Davidiella*, of which it represents a parasitic species, with asci pyriform, ascospores 16-19 × 3.5-4.5 µm. Therefore the following new combination is made: **Davidiella pontederiae** (Peck) Aptroot comb. nov., **MB 500359**. **Basionym:** *Sphaerella pontederiae* Peck, Annual Rep. New York State Mus. 40: 67. 1887. Additional material (Canada, Ontario, Frontenac Co., Silver Lake, on *Nymphaea advena*, Cain, VIII 1941, IMI no. 73322) is immature.

Mycosphaerella poonensis T.S. Viswan., Mycopathol. Mycol. Appl. 10: 222. 1959.

Type — India: *Ehretia aspera* (Boraginaceae).

Anamorph: *Cercospora poonensis* T.S. Viswan. fide Viswanathan (op. cit.).

No material was studied as the type was not found in LWG or IMI.

Mycosphaerella populi (Auersw.) J. Schröt., in Cohn, Kryptog.- Fl. Schlesien 3(2): 336. 1894 [“1893”] = *Sphaerella populi* Auersw., in Gonn. & Rabenh., Mycol. Europaea 5-6: 11. 1869.— Fig. 707.

Type — Germany: Sachsen, Eisleben. On upper and lower surface of dead leaves of *Populus dilatata* (Salicaceae). Kunze, distributed in Rabenhorst, Fungi Europaei Exsiccati no. 2541, (L, isotype).

Anamorph: *Septoria populi* Desm. fide Sivanesan (1984).

The isotype studied is immature. Additional material studied (Königstein, on upper and lower surface of dead leaves of *Populus berolinensis*, Krieger, Fungi Saxonici no. 2066, IV 1908, L; also The Netherlands, Limburg, Valkenburg, Rick, 1900, L) belongs to section *Longispora*, with asci cylindrical, ascospores $40-45 \times 4.5-5.5 \mu\text{m}$.

Sphaerella populi Fuckel, Jahrb. Nassauischen Vereins Naturk. 27-28: 20. 1873, later homonym (illegitimate, Article 53) \equiv *Sphaerella populi* var. *fuckelii* Sacc., Syll. Fung. 1: 488. 1882.

Type — Germany: Eltville. On upper surface of dead leaves of *Populus nigra* (Salicaceae). Fuckel, Fungi Rhenani no. 2647 (L, isotype).

The isotype studied contains only empty ascomata.

Sphaerella populi f. *populi-dilatatae* J. Kunze, Fungi Selecti Exsiccati no. 247. 1879.— Fig. 708.

Type — Germany: Sachsen, Eisleben. On upper and lower surface of dead leaves of *Populus dilatata* (Salicaceae). Kunze, Fungi Selecti Exsiccati no. 247, VI 1879 (L, holotype).

This is identical to the nominal form and belongs to section *Longispora*, with asci cylindrical, ascospores $40-48 \times 3.5-4.5 \mu\text{m}$.

Sphaerella populi f. *populi-nigrae* Thüm., Fungi Austriaci no. 349. 1871 [as “*populina*”].— Fig. 709.

Type — Austria: Krems. On upper and lower surface of dead leaves of *Populus nigra* (Salicaceae). Thümen, Fungi Austriaci no. 349 (B, isotype).

This belongs to section *Longispora* and is morphologically indistinguishable from a small-spored form of *M. populi*, with asci cylindrical, ascospores ca. $25 \times 5 \mu\text{m}$. Additional material (Germany, Leipzig, Winter, distributed in Thümen, Mycotheca Universalis no. 349, 1874, L) agrees with the nominal form, with ascospores $45-60 \times 3.5-4.5 \mu\text{m}$.

Sphaerella populi var. *fuckelii* Sacc. See *Sphaerella populi* Fuckel.

Mycosphaerella populi-albae Tomilin. See *Sphaerella subcrassa* Sacc. & P. Syd.

Mycosphaerella populicola G.E. Thomps., Phytopathology 31: 251. 1941.— Fig. 710, 985.

Type — Canada: Ontario, Lake Temagami distr., Bear Island. On upper and lower surface of dead leaves of *Populus tacamahaca* (Salicaceae). Thompson, V 1933 (BPI, isotype).

Anamorph: *Septoria populicola* Peck fide Thompson (op. cit.).

This belongs to section *Longispora*, and is morphologically indistinguishable from *M. latebrosa*, with asci cylindrical, ascospores $21.5-23 \times 3-4.5 \mu\text{m}$. Additional material (same locality, but on *P. balsamifera*, Thompson, VIII 1931, IMI no. 226963, slide only, topotype) contains only the anamorph.

Mycosphaerella populifolia (Cooke) House, New York State Mus. Bull. 233-234: 29. 1921 \equiv *Sphaerella populifolia* Cooke, J. Bot. 21: 107. 1883.— Fig. 711.

Type — USA: South Carolina, Seaboard. On lower surface of dead leaves of *Populus angulata* (Salicaceae). Ravenel no. 3240 (K, holotype), also distributed in Fungi Americani Exsiccati no. 689 (K (9x), NY, isotypes).

This belongs to section *Longispora*, and is morphologically indistinguishable from *M. millegrana*, with asci cylindrical, ascospores $15-18 \times 3-4 \mu\text{m}$.

Sphaerella populina Auersw. See *Sphaerella populi* f. *populi-nigrae* Thüm.

Sphaerella populina f. *populi-nigrae* Thüm. See *Sphaerella populi* f. *populi-nigrae* Thüm.

Mycosphaerella populnea (Sacc.) House, New York State Mus. Bull. 233-234: 29. 1921 \equiv *Sphaerella populnea* Sacc., Ann. Mycol. 13: 115. 1915.— Fig. 712.

Type — USA: New York, Tupper Lake. On white spots with black margins on upper surface of living leaves of *Populus balsamifera* (Salicaceae). House no. 1456, VIII 1913 (PAD, holotype).

Anamorph: Probably *Septoria populifolia* Peck fide Saccardo (op. cit.).

This belongs to section *Longispora*, and is morphologically indistinguishable from *M. millegrana*, with asci cylindrical, ascosporesca. $12 \times 2 \mu\text{m}$. Additional material studied (India, Assam, Burnihat, on *P. deltoides*, Mehrotra, VI 1982, IMI no. 2684585) is immature.

Mycosphaerella populorum G.E. Thomps., Phytopathology 31: 246. 1941.— Fig. 713, 978.

Type — USA: New York, Ithaca. On lower and upper surface of dead leaves of *Populus balsamifera* (Salicaceae). Thompson, V 1937 (BPI, isotype).

Anamorph: *Septoria musiva* Peck fide Thompson (op. cit.).

This belongs to *Davidiella*, with asci pyriform, ascospores $12.5-13.5 \times 3-3.5 \mu\text{m}$, surrounded by an even, ca. $1 \mu\text{m}$ thick gelatinous sheath. Therefore the following new combination is made: **Davidiella populorum** (G.E. Thomps.) Aptroot comb. nov., **MB 500360**. **Basionym:** *Mycosphaerella populorum* G.E. Thomps., Phytopathology 31: 246. 1941. Additional material (Canada, Ontario, Ottawa, Arboretum, on *P. deltoides*, Bier, 1936, IMI no. 215783, slide only) contains only the anamorph.

Mycosphaerella poraqueibae Bat. & Cavalc., in Batista et al., Publ. Inst. Micol. Univ. Fed. Pernambuco 309: 23. 1961.

Type — Brazil: *Poraqueiba* (Icacinaeae).

No material was studied as the type was not included in a loan from URM.

Mycosphaerella porocyphi (Stein) J. Schröt., in Cohn, Kryptog.-Fl. Schlesien 3(2): 342. 1894 [“1893”] \equiv *Arthopyrenia porocyphi* Stein, in Cohn, Kryptog.-Fl. Schlesien 2(2): 343. 1879 \equiv *Pharcidia porocyphi* (Stein) G. Winter, in Rabenh., Kryptog.-Fl. Deutschl., Österr. Schweiz, ed. 2, 1(2): 347. 1885.

Type — Switzerland: *Porocyphus cataractarum* (Ascomycota, Pyrenopsidaceae).

Accepted as *Pharcidia porocyphi* (Stein) G. Winter by Clauzade, Diederich & Roux (1989) and therefore not studied.

Mycosphaerella potentillae (Oudem.) Jacz., *Opredelitel' gribov* 2: 616. 1917 ≡ *Sphaerella potentillae* Oudem., *Verslagen Meded. Afd. Natuurk. Kon. Akad. Wetensch.*, ser. 3, 2: 15. 1885 ≡ *Mycosphaerella weberi* Oudem., nomen herbariorum (not validly published, Article 32).— Fig. 714.

Type — Novaya Zemlya: on upper and lower surface of dead leaves of *Potentilla fragiformis* (Rosaceae). (L-holotype [sub "*Mycosphaerella weberi*"]).

Already synonymised with *Davidiella allicina* (as *Mycosphaerella tassiana*) by von Arx (1949), but the type indicates that this is morphologically indistinguishable from *D. clandestina*, with asci pyriform, ascospores 18-26 × 4.5-6.5 µm.

Mycosphaerella potentillae (E. Müll.) E. Müll. See *Mycosphaerella muelleriana* M. Morelet.

Mycosphaerella potentillae-stipularis Tomilin, *Novosti Sist. Nizsh. Rast.* 20: 129. 1983.

Type — Russia: *Potentilla stipularis* (Rosaceae).

No material was studied as the type was not included in loans from LE or LEP.

Mycosphaerella pourthiaeae Hara, *List of Japanese fungi hitherto known*: 402. 1954.

Type — Japan: *Pourthiaea villosa* (Rosaceae).

No material was studied as the type was not included in a loan from TNS.

Mycosphaerella pouzolsiae Sawada, *Rep. Gov. Res. Inst. Formosa* 87: 25. 1944, lacking Latin description (not validly published, Article 36).

Type — Taiwan: *Pouzolsia zeylandica* var. *alienata* (Urticaceae).

No material was studied as the type was not found in BPI.

Mycosphaerella praecox (Pass.) Lindau, in *Engl. & Prantl, Natürlichen Pflanzenf.* 1(1): 424. 1897 ≡ *Sphaerella praecox* Pass., *Erbario Crittogamico Italiano*, no. 987. 1880.— Fig. 715.

Type — Italy: Parma, Collecchio. On dead stems and inflorescences of *Lactuca saligna* (Asteraceae). Passerini, *Erbario Crittogamico Italiano*, no. 987, X 1880 (B, isotype). Cited as synonymous with *Mycosphaerella lactucae* by Tomilin (1979). The isotype studied contains only various coelomycetes. Additional materials studied (Malaysia, Sabah, North Borneo, Keningau, on ill-defined green spots on the upper surface of living leaves of *L. indica*, Johnston, IX 1959, IMI no. 79326; also Malaysia, Serdang, on ill-defined green spots on the upper surface of living leaves of *L. sativa*, Johnston, VIII 1952, IMI no. 51290) belong to a species of *Didymella*, with asci cylindrical, pseudoparaphyses < 1 µm wide, ascospores 10-12 × 3.5-4.5 µm.

Mycosphaerella praeparva (Pass. & Beltrani) Tomilin, *Novosti Sist. Nizsh. Rast.* 1967: 190. 1967 ≡ *Sphaerella praeparva* Pass. & Beltrani, *Atti Reale Accad. Lincei, Mem. Cl. Sci. Fis.*, ser. 3, 7: 3. 1882.

Type — Italy: *Juncus acutus* (Juncaceae).

No material was studied as the type was not found in any of the herbaria consulted.

Mycosphaerella prasii (Pat.) Maire & Werner, *Mém. Soc. Sci. Nat. Maroc* 45: 27. 1937 ≡ *Sphaerella prasii* Pat. ex Pit., *Contr. Fl. Maroc*: 63. 1931.

Type — Morocco: *Prasium* (Lamiaceae).

No type material was preserved in FH.

Mycosphaerella prenanthicola Höhn., in *Strasser, Verh. Zool.-Bot. Ges. Wien* 69: 361. 1919 ≡ *Sphaerella prenanthicola* (Höhn.) Trotter, *Syll. Fung.* 24: 857. 1928.— Fig. 716.

Type — Austria: Niederösterreich, Sonntagsberg. On upper and lower surface of *Prenanthes purpurea* (Asteraceae). *Strasser, IV 1916* (FH-Höhnel, holotype).

This is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores 8-10 × 2-3 µm.

Mycosphaerella prenanthis (Ellis & Everh.) M.E. Barr, *Contr. Univ. Michigan Herb.* 9: 593. 1972 ≡ *Laestadia prenanthis* Ellis & Everh., *J. Mycol.* 8: 66. 1902.— Fig. 717.

Type — USA: Alabama, Tuskegee. On ill-defined, pale brown spots on lower surface of living leaves of *Prenanthes crepidinea* (Asteraceae). Carver no. 925, VIII 1901 (NY, holotype).

This is a parasitic species, with asci cylindrical, ascospores 9-11 × 3-3.5 µm.

Sphaerella prignitzensis Kirschst. See *Mycosphaerella hieracii* (Sacc. & Briard) Jaap.

Mycosphaerella primulae (Auersw. & Heufl.) J. Schröt., in *Cohn, Kryptog.-Fl. Schlesien* 3(2): 338. 1894 ["1893"] ≡ *Stigmatea primulae* Auersw. & Heufl., in *Auersw., Oesterr. Bot. Z.* 18: 277. 1868 ≡ *Sphaerella primulae* (Auersw. & Heufl.) G. Winter, *Hedwigia* 19: 166. 1880.— Fig. 718.

Type — Austria: On upper and lower surface of dead leaves of *Primula minima* (Primulaceae).

The type was not found in B and might be lost. The only specimen found in Berlin (Switzerland, Sankt Gallen, on *P. veris*, no. 395, III 1871, B) is immature. Additional material studied (on *P. auricula*, Müller, VI 1949, CBS) belongs to section *Caterva*, with asci cylindrical, ascospores 20-24 × 3.5-5 µm.

Sphaerella primulae var. *macrospora* Keissl., *Akad. Wiss. Wien Sitzungsber., Math.-Naturwiss. Kl., Abt. 1*, 61(2): 12. 1924.— Fig. 719.

Type — China: Yunnan, Piepun, Chungtien. On upper surface of dead leaves of *Primula dryadiifolia* (Primulaceae). Handel-Mazzetti no. 4707, VIII 1914 (W, holotype).

This belongs to the genus *Wettsteinina*, and is morphologically indistinguishable from and a new, heterotypic synonym of *W. macrospora* (Wehm.) Petr., with ascospores distoseptate with 1 septum and two additional constrictions, 55-60 × 15-17 µm.

Mycosphaerella ["*Mycosphaerium*"] *primulae* var. *major* ["*majus*"] Clem., Cryptogamae Formationum Coloradensium no. 225. 1906, nomen herbariorum (not validly published, Article 32).— Fig. 720, 980.

Authentic material — USA: Colorado, Bottomless Pit. On upper and lower surface of dead leaves of *Primula parryi* (Primulaceae). Clements, VIII 1906, Cryptogamae Formationum Coloradensium no. 225 (BPI).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. clandestina*, with asci pyriform, ascospores thick-walled, 22-27 × 7-8.5 µm.

Sphaerella primulaecola G. Winter, Hedwigia 19: 166. 1880 = *Leptosphaeria primulaecola* (G. Winter) Sacc., Syll. Fung. 2: 43. 1883 [as "*primulicola*"] = *Leptosphaerulina primulaecola* (G. Winter) Crivelli, Diss. Eidgen. Techn. Hochschule 7318: 134. 1983.— Fig. 721.

Type — Switzerland: Rhätische Alpen, Albulapas. On upper and lower surface of dead leaves of *Primula latifolia* (Primulaceae). Winter, distributed in Rabenhorst-Winter, Fungi Europaei Exsiccati no. 2849, VIII 1882 (L, topotype). Accepted as *Leptosphaerulina primulaecola* (G. Winter) Crivelli by Crivelli (op. cit.), with which the topotype material agrees well, with ascospores olivaceous brown, 1-5-septate, becoming submuriform, 18-24 × 8-10 µm.

Sphaerella prini Cooke, J. Bot. 21: 106. 1883.— Fig. 722.

Type — USA: South Carolina, Pinopolis. On upper and lower surface of dead leaves of *Prinos glaber* (Aquifoliaceae). Ravenel no. 2895 (K, holotype), also distributed in Fungi Americani Exsiccati no. 753 (K (9x), NY, isotypes).

This is a species of *Guignardia* s.s., with asci clavate, pseudoparaphyses moniliform, ca. 3-4 µm wide, ascospores simple, yellowish, 14-17 × 6-8 µm, both ends with white gelatinous appendages. The isotype in NY is overmature.

Mycosphaerella prinsepiae Padwick & Mehr, Mycol. Pap. 7: 2. 1943.

Type — India: Simla. On small white spots with broad black margins on upper and lower surface of living leaves of *Prinsepia utilis* (Rosaceae). Padwick, IX 1941 (IMI no. 16790 and IMI no. 4013, isotypes); also Sharma, X 1962 (IMI no. 99045, topotype).

The types and additional materials studied (India, Jammu, Kud, Pandotra, IX 1963, IMI no. 114549; also Nepal, Kakani, Tuladhar, VI 1970, IMI no. 152750) are all a *Didymella*, with asci clavate, pseudoparaphyses anastomosing, ca. 1 µm wide, ascospores 13-20 × 3-5.5 µm.

Mycosphaerella prominula (Speg.) Lindau, in Engl. & Prantl, Natürlichen Pflanzenf. 1(1): 426. 1897 = *Sphaerella prominula* Speg., Michelia 1: 456. 1879 = *Didymella prominula* (Speg.) Piroz. & Morgan-Jones, Trans. Brit. Mycol. Soc. 51: 198. 1968.— Fig. 723.

Type — Italy: Conegliano. On upper surface of dead fronds of *Pteridium* ["*Pteris*"] *aquilinum* (Polypodiaceae). Spegazzini, 1879, distributed in Decades Mycologiae Italicae no. 44 (BPI, isotype).

Although not in accordance with the protologue, the isotype shows that this is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores 6-8 × 2-2.5 µm.

Mycosphaerella proteae (Syd. & P. Syd.) Arx, in von Arx & E. Müll., Beitr. Kryptogamenfl. Schweiz 11(2): 357. 1962 = *Oligostroma proteae* Syd. & P. Syd., Ann. Mycol. 12: 265. 1914.— Fig. 724.

Type — South Africa: Natal, Kentani. On spots on upper and lower surface of dead but still green leaves of *Protea flanaganii* (Proteaceae). Pegler no. 5163, VII 1912 (S, holotype [sub "*Oligostroma Protea*"]).

The type and some additional material studied (Pegler no. 1899, 1915, B, topotype; Kirstenbosch, on *Protea*, Örtendahl, III 1931, B) agree and show that this is a peculiar ascomycete, with ascomata immersed in groups, only carbonized in the upper half, asci cylindrical, ascospores 22-32 × 5-6.5 µm, thick-walled and surrounded by a gelatinous sheath. Synonymised with *Teratosphaeria maculiformis* (G. Winter) Joa.E. Taylor & Crous in IMI descriptions of fungi 1344 (1998). Its attribution to *Teratosphaeria* is morphologically probably justified, but was not supported by recent DNA studies. Other material (Cape Province, on *Protea repens*, van Wijk, 1972, BS) is a parasitic species, with asci cylindrical, ascospores 10-12 × 2-3 µm. When accepted in *Mycosphaerella*, the epithet *proteae* should be used, as the older epithet *maculiformis* G. Winter is predated in *Mycosphaerella* by *maculiformis* Pers.

Mycosphaerella proteae-arboreae (Wyk, Marasas & Knox-Dav.) Joa.E. Taylor & Crous, Mycol. Res. 107: 657. 2003 = *Teratosphaeria proteae-arboreae* Wyk, Marasas & Knox-Dav., J. S. African Bot. 41: 232. 1975.

Type — South Africa: *Protea arborea* (Proteaceae).

No material was studied of this recently described species.

Sphaerella proustiae Speg., Anales Mus. Nac. Hist. Nat. Buenos Aires 19 (or ser. 3, 12): 352. 1909.

Type — Argentina: On upper and lower surface of dead leaves of *Proustia ilicifolia* (Asteraceae).

The type was not included in a loan from LPS. Material studied (Las Heras, Mendoza, Villavicencio, on *Proustia*, Ruiz Leal no. 8635, I 1944, CBS) is immature.

Sphaerella proximella P. Karst., Bidrag Kännedom Finlands Natur Folk 23: 177. 1873 = *Didymella proximella* (P. Karst.) Sacc., Syll. Fung. 1: 558. 1882 = *Didymosphaeria proximella* (P. Karst.) G. Winter, in Rabenh., Kryptog.-Fl. Deutschl., Österr. Schweiz, ed. 2, 2(1): 423. 1885.— Fig. 725.

Type — Finland: Mustiala. On dead leaves of *Carex acuta* (Cyperaceae). Karsten, V, distributed in Finland Fungi no. 897 (NY, isotype).

Anamorph: *Stagonospora* fide Corlett & J.D. Sm. (1978).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores $19-22 \times 5-7.5 \mu\text{m}$.

Mycosphaerella pruni-persicae Deighton, Trans. Brit. Mycol. Soc. 50: 328. 1967, nomen novum (Article 58) for *Mycosphaerella persicae* B.B. Higgins & F.A. Wolf, Phytopathology 27: 695. 1937, later homonym (illegitimate, Article 53) [as “*persica*”] = *Mycosphaerella pruni-persicae* Tomilin, Novosti Sist. Nizsh. Rast. 1968: 167. 1968, superfluous (illegitimate, Article 52) nomen novum (Article 58) and later homonym (illegitimate, Article 53).— Fig. 726.

Type — USA: North Carolina, Durham. On upper and lower surface of dead leaves of *Prunus persica* (Rosaceae). Wolf, IV 1938 (BPI, isotype).

Anamorph: *Miuraea persicae* (Sacc.) Hara *fide* Deighton (op. cit.).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradicans*, with asci cylindrical, ascospores $(8.5-10)-12 \times 2.5-3 \mu\text{m}$.

Mycosphaerella pruni-persicae Tomilin. See *Mycosphaerella pruni-persicae* Deighton.

Mycosphaerella psammae (Rostr.) Lind, Danish Fungi: 205. 1913 = *Sphaerella psammae* Rostr., Bot. Tidsskr. 22: 275. 1899.— Fig. 727.

Type — Denmark: Tisvilde. On the outside of in-rolled dead leaves of *Ammophila* [“*Psamma*” in publication; “*Calamagrostis*” on label] *arenaria* (Poaceae). Rostrup, VII 1898 (C, holotype).

This belongs to *Davidiella* and is morphologically indistinguishable from *D. disseminata*, with asci pyriform, ascospores $14-16 \times 2.5-3.5 \mu\text{m}$.

Mycosphaerella psammae var. *elymifloris* Munk. See *Mycosphaerella munkii* Tomilin.

Mycosphaerella psammae var. *stromatica* Munk. See *Mycosphaerella stromatica* (Munk) Tomilin.

Sphaerella psammisiae Cooke, Grevillea 9: 99. 1881.— Fig. 728.

Type — Venezuela: Caracas. On white spots with black margins on upper surface of dead leaves of *Psammisia penduliflora* (Ericaceae). Ernst, IX 1880 (K, holotype; K, isotype).

This is a *Guignardia* species, with ascomata without apparent hamathecium, ascospores simple, $15-17 \times 6-8 \mu\text{m}$.

Mycosphaerella pseudacaciae (Auersw.) Lindau, in Engl. & Prantl, Natürlichen Pflanzenf. 1(1): 424. 1897 = *Sphaerella pseudacaciae* Auersw., in Gonn. & Rabenh., Mycol. Europaea 5-6: 8. 1869.

Type — Germany: Barnim. On bark of dead branches of *Robinia pseudacacia* (Fabaceae). Auerswald no. 81 (B, holotype, sub “*Sphaeria*”).

This contains only a coelomycete. However, there is some doubt whether this is really the holotype of the *Sphaerella*,

as the substrate is mentioned to be petioles (not bark) in the protologue.

Sphaerella pseudacori Kirschst., Kryptog.-Fl. Mark Brandenburg 7(3): 412. 1938.— Fig. 729.

Type — Germany: Warthewiesen, Tamsel. On upper and lower surface of dead margins of living leaves of *Iris pseudacorus* (Iridaceae). Vogel, IX 1912 (B, lectotype, here designated); also Zossen, Rangsdorf, Syd., IX 1907 (B, paratype).

Both original syntypes are identical and show that this belongs to *Davidiella* and is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores $12-15 \times 5-5.5 \mu\text{m}$. It was previously synonymised with *Mycosphaerella iridis* by von Arx (1949), probably mostly on host connection, without actually studying type material.

Mycosphaerella pseudomaculiformis (Desm.) J. Schröt., in Cohn, Kryptog. Fl. Schlesien 3(2): 337. 1894 [“1893”, as “*pseudomaculaeformis*”] = *Sphaeria pseudomaculiformis* Desm., Ann. Sci. Nat. Bot., ser. 3, 6: 83. 1846 [as “*pseudomaculaeformis*”] = *Sphaerella pseudomaculiformis* (Desm.) Auersw., in Rabenh., Fungi Europaea Exsiccati no. 1158. 1869 [as “*pseudomaculaeformis*”].— Fig. 730.

Type — France: On upper and lower surface of dead leaves of *Sanguisorba officinalis* [“*Poterium sanguisorba*”] (Rosaceae). Desmazières, Plantes Cryptogames de France no. 1798 (NY, isotype).

Anamorph: *Ovularia bulbiger* (Fuckel) Sacc. *fide* Sivanesan (1984) (= *Phacellium bulbigerum* (Fuckel) U. Braun).

This is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores $7-9 \times 2-3 \mu\text{m}$. Additional material studied (Czech Republic, Weißkirchen, on dead stems of *Sanguisorba minor*, Petrak, distributed in Reliquiae Petrakianae no. 69, VI 1924, L) belongs to *Davidiella*, and is morphologically indistinguishable from *D. ammophilae*, with asci pyriform, ascospores $7-9 \times 3-3.5 \mu\text{m}$.

Mycosphaerella pseudoplatani Zerova, Bot. Zhurn. (Kiev) 9: 37. 1952.

Type — Ukraine: *Acer negundo* (Sapindaceae).

Anamorph: *Cylindrosporium pseudoplatani* (Roberge & Desm.) Diedicke *fide* Tomilin (1979).

No material was studied as the location of the type is unknown.

Mycosphaerella pseudopsammae Munk, Dansk Bot. Arkiv 17(1): 317. 1957.— Fig. 731.

Type — Denmark: Jutland, Blokhus, Grenå. On the outside of in-rolled dead leaves of *Ammophila arenaria* (Poaceae). Lind, VII 1929 (C, holotype).

This belongs to *Davidiella* and is morphologically indistinguishable from *D. disseminata*, with asci pyriform, ascospores $13-18 \times 3-4 \mu\text{m}$.

Mycosphaerella pseudoseptorioides Tomilin. See *Sphaerella thalicticola* Sacc. & P. Syd.

Mycosphaerella pseudosphaerioides Petr., Hedwigia 65(1): 232. 1925.— Fig. 732.

Type — Poland. On dead stems of *Althaea pallida* (Malvaceae). Wolczyniec no. 1645, VII 1918 (W, holotype).

Already cited as synonymous with *M. subradians* (as *M. asteroma*) by Tomilin (1979), with which the type agrees well, with asci cylindrical, ascospores $9-11 \times 2.5-3.5 \mu\text{m}$.

Mycosphaerella psilospora J.C. Gilman & Wadley, Mycologia 44: 219. 1952.

Type — USA: *Quercus* (Fagaceae).

Anamorph: *Septoria querceti* Thüm. *fide* Gilman & Wadley (op. cit.).

No material was studied as the type was not found in NY or BPI.

Sphaerella psorae Anzi, Atti Soc. Ital. Sci. Nat. 11: 27. 1868 \equiv *Metasphaeria psorae* (Anzi) Sacc., Syll. Fung. 2: 183. 1883 \equiv *Arthopyrenia psorae* (Anzi) Müll. Arg., Rev. Mycol. (Toulouse) 6: 15. 1884 \equiv *Pharcidia psorae* (Anzi) G. Winter, in Rabenh., Kryptog.-Fl. Deutschl., Österr. Schweiz, ed. 2, 1(2): 345. 1885 \equiv *Endococcus psorae* (Anzi) H. Olivier, Bull. Acad. Int. Géogr. Bot. 17: 127. 1907.

Type — Italy: *Psora decipiens* (Ascomycota, Lecideaceae). Cited as synonymous with *Stigmatidium allogenum* (Nyl.) D. Hawksw. by Clauzade, Diederich & Roux (1989) and therefore not studied.

Mycosphaerella ptarmicae (P. Karst. & Starbäck) Petr. & Syd., Ann. Mycol. 22: 358. 1924 \equiv *Laestadia ptarmicae* P. Karst. & Starbäck, in P. Karst., Hedwigia 26: 125. 1887.— Fig. 733.

Type — Finland: Tavastia australis, Tammela, Mustiala. On upper and lower surface of dead leaves of *Achillea ptarmica* (Asteraceae). Starbäck, Karsten no. 4153, VI 1887 (H, holotype).

This is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores $9-11 \times 2.5-3 \mu\text{m}$.

Mycosphaerella pteridicola Dearn. & House, Circ. New York State Mus. 24: 33. 1940, lacking Latin description (not validly published, Article 36).— Fig. 734.

Type — USA: New York, Essex, Newcomb. On upper surface of dead fronds of *Pteridium* [*“Pteris”*] *aquilinum* [*“latiusculum”*] (Polypodiaceae). House no. 815, VII 1925 (NY, isotype).

The type and additional material studied (Canada, Ontario, Dufferin Co., Cain no. 32442, VI 1955, L) belong to section *Caterva*, and this is morphologically indistinguishable from *M. filicum*, with asci cylindrical, ascospores $15-17 \times 2.5-3.5 \mu\text{m}$.

Mycosphaerella pteridis (Desm.) J. Schröt., in Cohn, Kryptog.-Fl. Schlesien 3(2): 341. 1894 [*“1893”*] \equiv *Sphaeria punctiformis* var. *pteridis* Fr., Scleromyceti Sueciae no. 86. 1819, sanctioned by Fr., Systema Mycol. 2: 525. 1823 \equiv *Sphaeria pteridis* Desm., Ann. Sci. Nat. Bot., ser. 2, 19: 359. 1843 \equiv *Sphaerella pteridis* (Desm.) De Not., Sferiacei Italici 1: 87. 1863.— Fig. 735.

Type — France: On upper surface of dead fronds of *Pteridium aquilinum* (Polypodiaceae). Desmazières, Fungi Exsiccati no. 795 (L, isotype).

The isotype studied is immature, and no ascospores were mentioned in the original description. Additional material studied (Germany, Brandenburg, Lebus, Dahmsdorf, Syd., Mycotheca Germanica no. 3311, V 1939, L) belongs to section *Longispora*, with asci cylindrical, ascospores $36-40 \times 4.5-5.5 \mu\text{m}$.

Mycosphaerella pterocarpi Kranz, Nova Hedwigia 18: 238. 1969.

Type — Guinea: Kindia. On white spots with black margins on upper surface of living leaves of *Pterocarpus santalinus* [*“Millettia”* on the original label] (Fabaceae). Kranz no. 298, IX 1962 (IMI no. 98555, holotype).

In the holotype material, only a coelomycete could still be found.

Mycosphaerella pterophila (Pass.) Tomilin, Novosti Sist. Nizsh. Rast. 7: 205. 1970 \equiv *Sphaerella pterophila* Pass., Atti Reale Accad. Lincei, Mem. Cl. Sci. Fis., ser. 4, 3: 7. 1887.

Type — Italy: *Fraxinus ornus* (Oleaceae).

No material was studied as the type was not found in any of the herbaria consulted.

Sphaerella pucciniophila Sacc. & P. Syd. See *Sphaerella parasita* Fautrey.

Mycosphaerella puerariae Sawada, Rep. Gov. Res. Inst. Formosa 86: 12. 1943, lacking Latin description (not validly published, Article 36).— Fig. 737.

Type — Taiwan: Tai Chung. On white spots with brown margins on upper surface of living leaves of *Pueraria thunbergiana* (Fabaceae). Sawada, VIII 1944 (BPI, topotype).

This is a *Phaeosphaeria*, with asci cylindrical, pseudoparaphyses simple, ca. $2 \mu\text{m}$ wide, ascospores brown, 3-septate, $17-19 \times 4-4.5 \mu\text{m}$, mostly postmature.

Mycosphaerella puerariae (Keissl.) Petr. See *Mycosphaerella keissleri* Tomilin.

Mycosphaerella puerariicola Weimer & Luttr., Phytopathology 38: 350. 1948.

Type — USA: Georgia, Experiment. On upper and lower surface of dead leaves of *Pueraria thunbergiana* (Fabaceae). Weimer & Luttrell, 1947 (BPI, IMI no. 226280 (slide only), isotypes), also Weimer no. 71526, XI 1946 (B, topotype).

Anamorph: *Pseudocercospora puerariicola* (W. Yamam.) Deighton *fide* Sivanesan (1984).

The types contain various immature or postmature ascomycetes and hyphomycetes, none of which are identifiable.

Sphaerella puiggarii Speg., Bol. Acad. Nac. Ci. 11: 518. 1889.

Type — Brazil: *Buddleja* [as *“Bouddleia”*] (Scrophulariaceae).

No material was studied as the type was not included in a loan from LPS.

Mycosphaerella pulchella (Syd. & P. Syd.) Arx, in E. Müll. & Arx, Beitr. Kryptogamenfl. Schweiz 11(2): 359. 1962 ≡ *Cyclodothis pulchella* Syd. & P. Syd., Ann. Mycol. 11: 266. 1913.

Type — Philippines: Mindanao, Davao, Todaya. On margins of brown spots on lower surface of living leaves of *Piper celtidiforme* [“*carylistachyon*” on label] (Piperaceae). Elmer, Philippine Islands Plants no. 11163, VII 1909 (S, holotype).

The type contains only a coelomycete with circular stromata.

Sphaerella pulchra G. Winter, Hedwigia 11: 145. 1872 ≡ *Leptosphaeria pulchra* (G. Winter) Sacc., Syll. Fung. 2: 53. 1883 ≡ *Leptosphaerulina pulchra* (G. Winter) M.E. Barr, Contr. Inst. Bot. Univ. Montréal 73: 7. 1959.

Type — Austria: *Potentilla caulescens* (Rosaceae).

No material was studied as the type was not found in B and might be lost.

Mycosphaerella pulmonariae Fakirova, Mycotaxon 64: 465. 1997.

Type — Bulgaria: Vitosha Mts. On overwintered leaves of *Pulmonaria rubra* (Boraginaceae). Fakirova, IV 1995 (SOM 21574-M, holotype, not seen).

No material was studied from this recently described species. According to the description, habitat (overwintered leaves) and illustration, this recently described species is probably just another synonym of *M. punctiformis*.

Mycosphaerella pulsatillae (Lasch) Johanson, Öfvers. Förh. Kongl. Svenska Vetensk.-Akad. 41(9): 165. 1884 ≡ *Sphaeria pulsatillae* Lasch, in Klotzsch, Herbarium Vivum Mycologicum no. 759. 1845 ≡ *Sphaerella pulsatillae* (Lasch) Auersw., in Gonn. & Rabenh., Mycol. Europaea 5-6: 12. 1869.

Type — Germany: Driesen. On upper and lower surface of dead leaves of *Pulsatilla vulgaris* [“*pratensis*”] (Ranunculaceae). (B, isotype).

The type contains only a coelomycete. Additional material studied (Budenheim, Fuckel, Fungi Rhenani Exciccati no. 840, L) contains *Pleospora herbarum* (Pers. : Fr.) Rabenh. and a *Microsphaeropsis* anamorph.

Mycosphaerella pulsatillae var. *major* (Riofrio) Cash, Syll. Fung. 26: 351. 1972 ≡ *Sphaerella pulsatillae* var. *major* Riofrio, Mem. Real Soc. Esp. Hist. Nat. 15: 385. 1929.

Type — Spain: *Anemone narcissiflora* (Ranunculaceae).

No material was studied as the type was not found in MA and might be lost.

Sphaerella pulsatillae var. *minor* Sacc., Syll. Fung. 1: 500. 1882.

Type — Germany: *Pulsatilla pratensis* (Ranunculaceae).

No material was studied as the type was not found in any of the herbaria consulted.

Mycosphaerella pulviscula (Cocc. & Morini) Maire & Werner, Mém. Soc. Sci. Nat. Maroc 45: 27. 1937 [as

“*pulvisculus*”] ≡ *Sphaerella pulviscula* Cocc. & Morini, Mem. Reale Accad. Sci. Ist. Bologna, ser. 4, 6: 395. 1885.

Type — Italy: *Dianthus brachyanthus* (Caryophyllaceae).

No material was studied as the location of the type is unknown. Cited as synonymous with *Mycosphaerella desmazieri*, which is morphologically indistinguishable from *Mycosphaerella iridis*, by Tomilin (1979).

Mycosphaerella punctata Dearn. & House ex M.E. Barr, Contr. Univ. Michigan Herb. 9: 591. 1972.

Type — USA: *Thalictrum polygamum* (Ranunculaceae).

No material was studied as the type was not found in NY or BPI.

Mycosphaerella punctiformis (Pers. : Fr.) Starbäck, Bih. Kongl. Svenska Vetensk.-Akad. Handl. 15(3, 2): 9. 1889 ≡ *Sphaeria punctiformis* Pers., Ann. Bot. (Usteri) 11: 26. 1794; Tent. Disposit. Method. Fungi: 51. 1797, sanctioned by Fr., Systema Mycol. 2: 525. 1823 ≡ *Sphaerella punctiformis* (Pers. : Fr.) Rabenh., Herb. Vivum Mycol., ed. nov., cent. 3 no. 264. 1856.

Anamorph: *Ramularia fide* Barr (1972) (= *Ramularia endophylla* Verkley & U. Braun).— Fig. 738.

Type — Netherlands: On lower surface of dead leaves of *Quercus* (Fagaceae). Persoon (L-Persoon, lectotype, designated by Verkley *et al.* 2004).

The species has traditionally been separated from *M. maculiformis* by the simple (not aggregated) ascomata. However, this character is very inconstant, even on one leaf, and both species have been synonymised, e.g. by Barr (1972). Both names have been published simultaneously. The earlier name *Sphaeria corylea* is not available for this well-known species, as a later name based on this, *Sphaeria maculiformis*, was sanctioned by Fr. The lectotype is the only material under this name in the Persoon herbarium that was not classified in another (often invalid) variety by himself (see below for the respective entries). It is typical for the species, with asci cylindrical, ascospores 8-10 × 2-3 µm.

Sphaerella punctiformis Cooke. See *Sphaerella cookeana* Auersw.

Mycosphaerella punctiformis f. *coryli* Kleb., Haupt- und Nebenfruchtformen der Askomyzeten: 100. 1918.

Type — Germany: *Corylus avellana* (Betulaceae).

No material was studied as the Klebahn types were not kept.

Mycosphaerella punctiformis f. *quercus* Kleb., Haupt- und Nebenfruchtformen der Askomyzeten: 95. 1918.

Type — Germany: *Quercus* (Fagaceae).

No material was studied as the Klebahn types were not kept.

Sphaerella punctiformis f. *salicis-auritae* Rabenh., Fungi Europaei Exsiccati, ed. nov., ser. 2 no. 942. 1866.

Type — Germany: Bohemia, Reichenberg. *Salix aurita* (Salicaceae). Siegmund, distributed in Rabenhorst, Fungi Europaei Exsiccati, ed. nov., ser. 2 no. 942 (B, isotype).

The isotype studied contains nothing identifiable.

Mycosphaerella punctiformis f. *tiliae* Kleb., Haupt- und Nebenfruchtformen der Askomyzeten: 84. 1918.

Type — Germany: *Tilia* (Malvaceae).

No material was studied as the Klebahn types were not kept.

Sphaeria punctiformis var. *ambigua* Pers., Synopsis Meth. Fung.: 91. 1801.

Type — Netherlands: On upper surface of dead leaves of *Quercus* (Fagaceae). (L-Persoon, holotype).

This is morphologically indistinguishable from *Phacidium dentatum* Kunze & Schwein.

Sphaeria punctiformis var. *angeliaria* Pers., nomen herbariorum (not validly published, Article 32).

Authentic material — France: On lower surface of dead leaves of *Angelica sylvestris* (Apiaceae). Chaillet no. 806 (L-Persoon).

This is immature, but morphologically indistinguishable from *M. punctiformis*.

Mycosphaerella punctiformis var. *clematidis* Jaap. See *Sphaerella clematidis* (Jaap) Kirschst.

Sphaeria punctiformis var. *corylaria* Pers., nomen herbariorum (not validly published, Article 32).

Authentic material — France: On lower surface of dead leaves of *Corylus avellana* (Betulaceae). Chaillet no. 206a (L-Persoon).

This is immature, but morphologically indistinguishable from *M. punctiformis*.

Sphaeria punctiformis var. *fagi* Pers., nomen herbariorum (not validly published, Article 32).— Fig. 739.

Authentic material — Netherlands: On upper surface of dead leaves of *Fagus sylvatica* (Fagaceae). Persoon (L-Persoon).

This is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores $8-10 \times 2-2.5 \mu\text{m}$.

Sphaeria punctiformis var. *graminaria* DC., Fl. Française, suppl. 1805.

Type — France: On upper and lower surface of dead leaves of Poaceae. Desmazières no. 337 (L, isotype).

This is immature.

Sphaeria punctiformis var. *hederae* Pers., Synopsis Meth. Fung.: 90. 1801.

Type — Netherlands: On spots on upper and lower surface of living leaves of *Hedera helix* (Araliaceae). (L-Persoon, holotype).

The holotype contains only a *Colletotrichum* anamorph. Additional material studied (Funck no. 604, L-Persoon) contains only a *Septoria* anamorph.

Sphaerella punctiformis var. *perexigua* (Lév.) Rabenh., Herb. Vivum Mycol., ed. nov., cent. 3 no. 264. 1856 \equiv *Sphaeria perexigua* Lév., Ann. Sci. Nat. Bot., ser. 3, 9: 144.

Type — France: On lower surface of dead leaves of *Quercus* (Fagaceae).

Type was not included in a loan from PC. Cited as synonymous with *M. punctiformis* by Tomilin (1979), with which material studied (Rabenhorst, Herb. Vivum Mycol., ed. nov., cent. 3 no. 264, L), although immature, seems to agree.

Sphaeria punctiformis var. *peritheciis-minutissimis* Desm., Plantes Cryptogames de France no. 284. 1828.

Authentic material — France: On lower surface of dead leaves of *Quercus robur* (Fagaceae). Desmazières, Plantes Cryptogames de France no. 284 (L-isotype).

This is immature, but morphologically indistinguishable from *M. punctiformis*.

Sphaerella punctoidea Cooke, J. Bot. 4: 247. 1866 \equiv *Laestadia punctoidea* (Cooke) Auersw., Hedwigia 8: 177. 1869 \equiv *Guignardia punctoidea* (Cooke) Schoeter, in Cohn, Kryptog.-Fl. Schlesien 3(2): 329. 1894 [“1893”] \equiv *Phyllachora punctoidea* (Cooke) Höhn., Ann. Mycol. 16: 47. 1918 \equiv *Physalospora punctoidea* (Cooke) Petr., Ann. Mycol. 22: 52. 1924.— Fig. 741.

Type — United Kingdom: On upper and lower surface of dead leaves of *Quercus* (Fagaceae).

Accepted as *Guignardia punctoidea* (Cooke) J. Schröt. by Eriksson (1992). Material studied (Czech Republic, Weißkirchen, Petrak, V 1923, B, 2 \times) agrees well, with ascospores simple, rhomboidal, $11-13 \times 4-5 \mu\text{m}$.

Mycosphaerella pusilla (Auersw.) Johanson, Öfvers. Förh. Kongl. Svenska Vetensk.-Akad. 41(9): 166. 1884 \equiv *Sphaerella pusilla* Auersw., in Gonn. & Rabenh., Mycol. Europaea 5-6: 17. 1869 \equiv *Sphaerella tassiana* var. *pusilla* (Auersw.) P. Karst., Bidrag Kännedom Finlands Natur Folk 23: 170. 1873.

Type — Germany: *Carex brachystachys* [“*tenuis*”] (Cyperaceae).

The type was not found in B and might be lost. Material studied (Finland, Ostrobothnia australis, Siippy, on upper and lower surface of dead leaves of *Festuca arenaria*, Roivainen, Liro, Mycotheca Fennica no. 877, X 1951, L) contains only a coelomycete.

Sphaerella pusilla f. *tritici-monococci* Thüm., Herb. Mycol. Oecon. no. 601. 1877.— Fig. 742.

Type — Italy: Parma. On upper and lower surface of dead leaves of *Triticum monococcum* (Poaceae). Pass., distributed in Thümen, Herbarium Mycologicum Oeconomicum no. 601, 1877 (L, isotype), also distributed in Rabenhorst, Fungi Europaei Exsiccati no. 2440 (L, isotype). Already synonymised with *Davidiella allicina* (as *Mycosphaerella tassiana*) by von Arx (1949), with which the isotype agrees well, with asci pyriform, ascospores $16-20 \times 6-7 \mu\text{m}$.

Sphaerella pusilla f. *tritici-vulgaris* Thüm., Mycotheca Universalis no. 1355. 1877.— Fig. 743.

Type — Italy: Parma. On upper and lower surface of dead leaves of *Triticum vulgare* (Poaceae). Pass., distributed in Thümen, Mycotheca Universalis no. 1355, III 1877 (L, isotype).

Synonymised with *Davidiella allicina* (as *Mycosphaerella tassiana*) by von Arx (1949), with which the isotype agrees well, with asci pyriform, ascospores 16-18 × 5-6 µm.

Mycosphaerella putoriae (Unamano) Maire & Werner, Mém. Soc. Sci. Nat. Maroc 45: 27. 1937 ≡ *Sphaerella putoriae* Unamano, Bol. Soc. Esp. Hist. Nat. 33: 38. 1933.— Fig. 744.

Type — Spain: Melilla, Barranco del Nano. On upper and lower surface of dead leaves of *Putoria brevifolia* (Rubiaceae). Sennen & Mauricio no. 9698, VI 1931 (MA, holotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. ammophilae*, with asci pyriform, ascospores 7-9 × 2-3 µm.

Mycosphaerella puttemansii Henn., Hedwigia 41: 301. 1902 ≡ *Sphaerella puttemansii* (Henn.) Sacc. & D. Sacc., Syll. Fung. 17: 640. 1905.

Type — Brazil: São Paulo, Botanical Garden. On spots on upper and lower surface of living leaves of *Plantago tomatova* (Plantaginaceae). Puttemans 256, V 1901 (SP, lectotype [holotype lost in B]).

The type contains only an *Asteromella* spermatial state.

Sphaerella puyae Speg., Revista Fac. Agron. Univ. Nac. La Plata, ep. 2, 6: 57. 1910.

Type — Chile: *Puya chilensis* (Bromeliaceae).

No material was studied as the type was not included in a loan from LPS.

Mycosphaerella pyrenaica (Speg.) Arx, Sydowia 3: 64. 1949 ≡ *Sphaerella pyrenaica* Speg., Rev. Mycol. (Toulouse) 4: 78. 1882.— Fig. 745.

Type — France: On upper and lower surface of dead leaves of *Petrocallis pyrenaica* (Brassicaceae).

The type was not included in a loan from LPS. Material studied (Austria, Niederösterreich, Schneeberg, Petrak, distributed in Reliquiae Petrakianae no. 1643, VII 1950, L) belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores 14-19 × 4-5 µm.

Mycosphaerella pyri (Auersw.) Boerema, Netherlands J. Pl. Pathol. 76: 166. 1970 ≡ *Sphaerella pyri* Auersw., in Gonn. & Rabenh., Mycol. Europaea 5-6: 11. 1869.— Fig. 747.

Type — Germany: *Pyrus* (Rosaceae).

Anamorph: *Septoria pyricola* (Desm.) Desm. *fide* Boerema *et al.* (1993).

No material was studied as the type was not found in B and might be lost.

Sphaerella pyri [as “*pyrina*”] f. *pyri-communis* Thüm., Fungi Austriaci no. 158. 1871.— Fig. 748, 980.

Type — Austria: Krems. On upper and lower surface of dead leaves of *Pyrus communis* (Rosaceae). Thümen, Fungi Austriaci no. 158, IV 1871 (B, BPI, isotypes).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores thick-walled, 16-18 × 6-7.5 µm.

Mycosphaerella pyrina (Ellis & Everh.) J.H. Mill., Mycologia 33: 80. 1941 ≡ *Sphaerella pyrina* Ellis & Everh., N. Amer. Pyrenomyc.: 275. 1892 [also as “*pyrina*”].— Fig. 746.

Type — USA: New Jersey, Newfield. On upper and lower surface of dead leaves of *Pyrus communis* (Rosaceae). Ellis, VII 1880, distributed in Ellis & Everhart, North American Fungi no. 597 [sub “*Sphaerella sentina*”] (NY, 2 isotypes), also VI 1895, distributed in Ellis & Everhart, Fungi Columbiani no. 934 [as “*pyrina*”] (NY, topotype).

This is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores 8-10 × 2-3 µm.

Mycosphaerella pyrolae (Rostr.) Lind, Rep. Sci. Results Norw. Exped. Novaya Zemlya 1921, 19: 12. 1924 [as “*pirolae*”] ≡ *Sphaerella pyrolae* Rostr., Meddel. Grønland 3(2): 551. 1888.— Fig. 749.

Type — Greenland: Kirutasat. On dead, blackend parts on upper surface of leaves of *Pyrola grandiflora* (Ericaceae). Warming & Holm, VII 1884 (C, holotype).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradians*, with asci cylindrical, ascospores 15-18 × 3-4 µm. Additional material studied (Russia, Tambow, on upper and lower surface of dead leaves of *Orthilia secunda*, Schiragensky, V 1911, L) is immature.

Mycosphaerella pyrolae (Ehrenberg : Fr.) B. Erikss., Svensk Bot. Tidskr. 68: 216. 1974, later homonym (illegitimate, Article 53) ≡ *Sphaeria pyrolae* Ehrenberg, Sylvae Mycol. Berolinenses: 29. 1818, sanctioned by Fr., Systema Mycol. 2: 528. 1823.— Fig. 750.

Type — Germany: *Chimaphila* [“*Pyrola*”] *umbellata* (Ericaceae).

The type was not found in B and might be lost. Cited as synonym of *Mycosphaerella chimaphilae* (Ellis & Everh.) Höhn. by Eriksson (1992). Material studied (Czech Republic, Weißkirchen, Hrabuvka, on *Orthilia secunda* [as “*Ramischia*”], Petrak, distributed in Reliquiae Petrakianae no. 2479, VI 1921, B) is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores 7-9 × 2-2.5 µm.

Mycosphaerella pyrolina Kirschst., Ann. Mycol. 33: 209. 1935 [as “*pirolina*”] ≡ *Sphaerella pyrolina* (Kirschst.) Kirschst., Kryptog.-Fl. Mark Brandenburg 7(3): 349. 1938 [as “*pirolina*”].— Fig. 751.

Type — Germany: Niederbayern, Eisenstein, Steinhütte. On dead inflorescences of *Orthilia secunda* [as “*Pirola und Ramischia*”] (Ericaceae). Kirschstein, VII 1933 (B, holotype); also Neuhütte VII 1935 (B, topotype).

The types and additional material studied (Oberbarnim, Falkenberg, Sydow, Mycotheca Germanica no. 3508, VII 1938, L) belong to *Davidiella*, and show that this is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores 12-15 × 4-5.5 µm.

Mycosphaerella quadrangulata (Ellis & Everh.) Tomilin. See *Mycosphaerella sapindi* (Ellis & Everh.) Lindau.

Mycosphaerella queenslandica Sivan. & R.G. Shivas, Mycol. Res. 106: 361. 2002.

Type — Australia: *Dianella* (Xanthorrhoeaceae).

No material was studied of this recently described species.

Sphaerella quercicola (Desm.) Roum., Fungi Gallici Exsiccati no. 1085 = *Sphaeria quercicola* Desm., Ann. Sci. Nat. Hist., ser. 3, 8: 25. 1847.— Fig. 752.

Type — France: On upper and lower surface of dead leaves of *Quercus robur* (Fagaceae).

The type was not included in a loan from PC. Material studied (France, Lyon, Therry, distributed in Roumeguère, Fungi Gallici Exsiccati no. 1085, IX 1879, L) is *Sphaerulina myriadea* (DC. : Fr.) Sacc., with asci cylindrical, ascospores 3-septate, 30-34 × 2.5-3 µm.

Mycosphaerella quercifolia (Gonz. Frag.) M. Morelet, Bull. Soc. Sci. Nat. Archéol. Toulon & Var 177: 9. 1968 = *Sphaerella quercifolia* Gonz. Frag., Bol. Soc. Brot., ser. 2, 2: 27. 1924.

Type — Portugal: S. Gens, Póvoa de Lanhoso. On upper and lower surface of dead leaves of *Quercus pedunculata* (Fagaceae). Sampaio no. 5849, XI 1922 (MA, holotype).

In the holotype material nothing identifiable was found.

Mycosphaerella quercina (Jacq.) Lindau, Hilfsb. Sammeln Ascomyceten: 93. 1903 = *Sphaerella quercina* Jacq., Bull. Soc. Mycol. France 12: 104. 1896.

Type — Switzerland: *Quercus* (Fagaceae).

Anamorph: *Aposphaeria quercina* Jacq. fide Jaczewski (op. cit.).

No material was studied as the type was not found in any of the herbaria consulted.

Sphaerella rabenhorstii Ces. & De Not., Comment. Soc. Crittog. Ital. 1(4): 238. 1863, nomen novum (Article 58) for *Sphaeria herbarum* f. *euphrasiae* Rabenh., Fungi Europaei Exsiccati, ed. nov., ser. 2 no. 258. 1860, non *Sphaerella euphrasiae* Pass. (1887) = *Laestadia rabenhorstii* (Ces. & De Not.) Sacc., Syll. Fung. 1: 431. 1882.

Type — Germany: *Euphrasia* (Orobanchaceae).

No material was studied as the species was already redisposed.

Mycosphaerella rabiei Kovatsch. ex Gruyter, in Boerema *et al.*, Phoma identification manual: 398. 2004 = *Mycosphaerella rabiei* Kovatsch., The blight of the chickpea (*Cicer arietinum*). *Mycosphaerella rabiei* n. sp., Minist. Agric. Nat. Dom. Sofia: 70. 1936, lacking Latin diagnosis (not validly published, Article 36) = *Didymella rabiei* (Kovatsch.) Arx, in E. Müll. & Arx, Beitr. Kryptogamenfl. Schweiz 11(2): 364. 1962, lacking Latin diagnosis (not validated, Article 36).

Type — Bulgaria: *Cicer arietinum* (Fabaceae).

Anamorph: *Ascochyta rabiei* (Pass.) Labrousse fide von Arx (op. cit.) (= *Phoma rabiei* (Pass.) Khune ex Gruyter).

No material was studied as the species was already redisposed.

Mycosphaerella radiata (Ranoj.) Tomilin, Novosti Sist. Nizsh. Rast. 8: 151. 1971 = *Sphaerella radiata* Ranoj., Ann. Univ. Grenoble 30: 371. 1918.

Type — France: *Centranthus angustifolius* (Caprifoliaceae). No material was studied as the type was not found in any of the herbaria consulted.

Sphaerella rajaniae Ellis & Everh., in Ellis, Annual Rep. Missouri Bot. Gard. 9: 118. 1898.— Fig. 753.

Type — Bahamas: Nassau. On ill-defined, pale brown spots on upper and lower surface of living leaves of *Rajania microphylla* ["*hastata*"] (Polygonaceae). Hitchcock, XI 1890 (NY, holotype).

This is most probably a species of *Discostroma*, with asci clavate, paraphyses simple, 2-3 µm wide, surrounded by a gelatinous substance, ascospores thick-walled, 18-23 × 5-6.5 µm.

Mycosphaerella ramulorum (Pass.) Tomilin, Novosti Sist. Nizsh. Rast. 8: 151. 1971 = *Sphaerella ramulorum* Pass., Atti Reale Accad. Lincei, Mem. Cl. Sci. Fis., ser. 4, 3: 7. 1887.

Type — Italy: *Lonicera caprifolium* (Adoxaceae).

No material was studied as the type was not found in any of the herbaria consulted. Cited as synonymous with *Mycosphaerella collina* by Tomilin (1979).

Mycosphaerella ranunculi (Fr.) Lind, Meddel. Grønland 71: 167. 1926 = *Stigmatea ranunculi* Fr. = *Sphaerella ranunculi* (Fr.) P. Karst., Öfvers. Förh. Kongl. Svenska Vetensk.-Akad. 1872(2): 105. 1872.— Fig. 754.

Type — *Ranunculus repens* (Ranunculaceae).

Anamorph: *Cercospora fide* Savile & Parmelee (1964).

No material was studied as the type was not included in a loan from UPS. Material seen (Canada, Franklin, Somerset Island, on *R. nivalis*, Savile no. 3657A, VII 1958, IMI 200596, slide only; also Finland, Mustiala, Karsten no. 4577, IX 1871, H) belongs to *Davidiella*, morphologically morphologically indistinguishable from *D. allicina*, with asci rather slender, but pyriform, ascospores 17-19 × 4-5 µm.

Sphaerella rapanae Keissl., Akad. Wiss. Wien Sitzungsber., Math.-Naturwiss. Kl., Abt. 1, 60(11): 76. 1924 ["1923"] = *Melanops rapanae* (Keissl.) Petr., Sydowia 1: 362. 1947.— Fig. 755.

Type — China: Guizhou, Zhenning. On lower surface of living leaves of *Rapanea* (Myrsinaceae). Handel-Mazzetti no. 10375, VI 1917 (K, isotype).

This is a species of *Guignardia*, with asci clavate, thick-walled, pseudoparaphyses moniliform, ascospores simple, hyaline, 14-16 × 6-8 µm.

Mycosphaerella rauwolfiae T.S. Ramakr. & K. Ramakr., Proc. Indian Acad. Sci., sect. B, 32: 206. 1950 [as "*rauwolfiae*"].

Type — India: On dark spots on lower surface of living leaves of *Rauwolfia serpentina* (Apocynaceae).

The location of the type is unknown and its preservation uncertain. Material studied, including topotypes (Kerala, Vellankkara, Varma, IX 1990, IMI no. 342691; also

Madhya Pradesh, Amarkantak, Sharma, X 1992, IMI no. 356621; also Assam, Gauhati, Sarma, X 1980, IMI no. 252718; also Jabalpur, Poli-pather Nursery, Mehrotra & Deo, X 1973, IMI no. 180288; also Assam, Gauguly, IV 1972, IMI no. 165553) all contains only coelomycetes.

Mycosphaerella ravenelii (Cooke) Tomilin, Novosti Sist. Nizsh. Rast. 9: 116. 1972 ≡ *Sphaerella ravenelii* Cooke, Grevillea 7: 53. 1878.— Fig. 756.

Type — USA: South Carolina, Aiken. On lower surface of dead leaves of *Quercus* (Fagaceae). Ravenel no. 2596 (K (9x), NY (2x), isotypes) [as “*raveneli*”].

All isotypes studies are mostly overmature, but the few ascospores with ascospores show that this is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores 9-11 × 2.5-3 µm.

Mycosphaerella readeriellophora Crous & J.P. Mansilla in Crous *et al.*, Stud. Mycol. 50: 207. 2004.

Type — Spain: Pontevedra, Lourizán, Areeiro, on leaves of *Eucalyptus globulus* (Myrtaceae). Mansilla, 2003 (CBS 9901, holotype); culture ex-type CBS 114240 = CPC 10375. Anamorph: *Readeriella readeriellophora* Crous & J.P. Mansilla *vide* Crous *et al.* (op. cit.).

No material was studied of this recently described species.

Mycosphaerella recutita (Fr.) Johanson, Öfvers. Förh. Kongl. Svenska Vetensk.-Akad. 41(9): 166. 1884 ≡ *Sphaeria recutita* Fr., Systema Mycol. 2: 524. 1823 ≡ *Sphaerella recutita* (Fr.) Rabenh., Herb. Vivum Mycol. no. 659. 1858 ≡ *Carlia recutita* (Fr.) Höhn., Centralbl. Bakteriol., 2. Abth., 60: 6. 1923.— Fig. 757.

Type — Sweden: Skåne, Lund. Poaceae. Fries, F-09808 (UPS-FRIES, holotype).

Anamorphs: *Scolicotrichum graminis* Fuckel (= *Passalora graminis* (Fuckel) Höhn.) and *Septoria graminum* Desm. *vide* Cooke (1871).

According to Eriksson (1992) the type contains no *Mycosphaerella*. The oldest available name for *Mycosphaerella recutita* sensu von Arx is *Davidiella disseminata*, not *Mycosphaerella wichuriana*, as suggested by Eriksson (1992). Material studied (Germany, Oestrich, on upper and lower surface of dead leaves of *Dactylis glomerata*, Fuckel, Fungi Rhenani no. 2434, L) belongs to *Davidiella*, and is morphologically indistinguishable from *D. disseminata*, with asci pyriform, ascospores 13-15 × 3-3.5 µm.

Sphaerella recutita f. *dispersa* Rehm, Hedwigia 24: 239. 1885.

Type — Austria: Tyrol, Ortler, Sulden-gletscher. On dead culms of *Dactylis glomerata* (Poaceae). Rehm no. 835, VII 1884 (S, holotype).

The holotype is in bad shape, but shows that it is morphologically indistinguishable from *Davidiella disseminata*.

Sphaerella recutita f. *molinae* Roum., Fungi Gallici Exsiccati no. 2294.

Type — France: Forêt de Fontainebleau. On upper and lower surface of dead leaves of *Molinia coerulea* (Poaceae).

Feuilleaubois, distributed in Roumégère, Fungi Gallici Exsiccati no. 2294, XII [“188”]. (L, isotype).

The isotype studied is immature.

Sphaerella recutita f. *poae* Roum., Fungi Gallici Exsiccati no. 3032 [as “*Sphaerella*”].

Type — France: Seine-Inférieure, Parc du Grand-Quevilly. On upper and lower surface of dead leaves of *Poa pratensis* (Poaceae). Letendre, distributed in Roumégère, Fungi Gallici Exsiccati no. 3032, VII 1883 (L, isotype).

The isotype studied contains only empty ascospores.

Mycosphaerella rehmana Jaap, Verh. Bot. Vereins Prov. Brandenburg 56: 81. 1914 ≡ *Sphaerella rehmana* (Jaap) Trotter, Syll. Fung. 24: 865. 1928.— Fig. 758.

Type — Italy: Lago di Garda, Gardone. On upper and lower surface of dead leaves of *Adiantum capillus-veneris* (Polyodiaceae). Jaap, Fungi Selecti Exsiccati no. 563, V 1912 (L, 2 isotypes).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. ammophilae*, with asci pyriform, ascospores 9-11 × 3-3.5 µm. The isotypes also contain a *Sphaerulina*, probably *Sphaerulina myriadea* (DC. : Fr.) Sacc., with asci cylindrical, ascospores 3-septate, ca. 30 × 3 µm.

Mycosphaerella resedae (Pass.) Woron., Trudy Tiflissk. Bot. Sada, ser 2, 3: 139. 1923 ≡ *Sphaerella resedae* Pass., Atti Reale Accad. Lincei, Mem. Cl. Sci. Fis., ser. 4, 4: 56. 1888.

Type — Italy: *Reseda luteola* (Resedaceae).

No material was studied as the type was not found in any of the herbaria consulted.

Mycosphaerella resedicola Ade ex Petr., Bot. Jahrb. Syst. 62, Beibl. 142: 126. 1928.— Fig. 759.

Type — Canary Islands: Tenerife, Guimar. On dead stems of *Reseda scoparia* (Resedaceae). Ade, V 1926 (W no. 10070, holotype; W no. 26837, isotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. ammophilae*, with asci pyriform, ascospores 7-10 × 3-4 µm.

Mycosphaerella retinisporae (Berl. & Bres.) Magnus, in Dalla Torre & Sarnth., Fl. Tirol, Vorarlberg, Liechtenstein, III, Pilze: 466. 1905 [as “*retinisporae*”] ≡ *Sphaerella retinisporae* Berl. & Bres., Annuario Soc. Alpinisti Tridentini 14: 27. 1889 [as “*retinisporae*”].

Type — Italy: *Retinispora squarrosa* (Cupressaceae).

No material was studied as the type was not found in any of the herbaria consulted.

Mycosphaerella reyesi Syd. & P. Syd., Ann. Mycol. 12: 200. 1914 ≡ *Sphaerella reyesi* (Syd. & P. Syd.) Trotter, Syll. Fung. 24: 885. 1928.

Type — Philippines: Luzon, Prov. Rizal. On upper and lower surface (not in spots) of living leaves of *Sapindus saponaria* (Sapindaceae). Ramos, Bureau of Science no. 21868, VIII 1913 (S, holotype; BPI, 2 isotypes).

The holotype and isotypes are immature, but show that it is a Phyllachoraceae with ascomata large-cellular, asci globose, ascospores simple.

Sphaerella rhabdophila Kirschst. See *Sphaerella rhabdospora* Kirschst.

Sphaerella rhabdospora Kirschst., Ann. Mycol. 37: 104. 1939.

Type — Germany: Rathenow, Körgraben. On dead twigs of *Salix viminalis* (Salicaceae). Kirschstein, VI 1905 (B, holotype [on the label as “*rhabdophila*”]).

The type is overmature, but the ascoma structure suggests that this is morphologically indistinguishable from *Cryptodiaporthe salicella* (Fr. : Fr.) Petr.

Mycosphaerella raphithamni (Keissl.) Petr., Ann. Mycol. 38: 221. 1940 = *Didymellina raphithamni* Keissl., in Skottsberg (ed.), Nat. Hist. Juan Fernandez and Easter Island 2: 480. 1927.— Fig. 760.

Type — Juan Fernandez: Masatierra, Villagra. On white spots with brown margins at the margins of the upper and lower surface of living leaves (but still saprobic) of *Raphithamnus longiflorus* (Verbenaceae). C. & J. Skottsberg, I 1917 (W, holotype).

This is a new synonym of *Lophiostoma rubi* (Fuckel) E.C.Y. Liew, Aptroot & K.D. Hyde (Liew *et al.* 2002), with simple hamathecium filaments of ca. 2 µm wide, ascospores 1-3-septate, 17-22 × 4-5 µm.

Mycosphaerella rhea (Fautrey) Koshk., in Koshkelova, Frolov & Dzhuraeva, Mikoflora Badkhyza, Karabilya i Yuzhnoi Chasti Murgabskogo Oazisa: 101. 1970, full reference to basionym not given (not validly published, Article 33) = *Sphaerella rhea* Fautrey, Rev. Mycol. (Toulouse) 16: 162. 1894.

Type — France: *Ruta graveolens* (Rutaceae).

Anamorph: Associated with *Phyllosticta rhea* Fautrey and *Septoria rhea* Fautrey *vide* Fautrey (op. cit.).

No material was studied as the type was not included in a loan from PC.

Sphaerella rhododendri De Not., Sferiacei Italici 1: 86. 1863 = *Laestadia rhododendri* (De Not.) Sacc., Syll. Fung. 1: 428. 1882 = *Carlina rhododendri* (De Not.) Kuntze & Bonord. ex Oudem., Rév. Champ. Pays-Bas 2: 197. 1897 = *Physalospora rhododendri* (De Not.) Rehm, Ann. Mycol. 5: 536. 1907 = *Hyponectria rhododendri* (De Not.) Rehm, Ann. Mycol. 5: 544. 1907.— Fig. 763.

Type — Italy: Riva. On upper surface of dead leaves of *Rhododendron ferrugineum* (Ericaceae). Carestia no. 215, VIII 1861 (RO, holotype).

Although the holotype was annotated by Traverso to be empty, it was found to belong to *Physalospora*, in which it was accepted by von Arx & Müller (1954), with paraphyses simple, branching above the asci, ascospores simple, 37-42 × 7-9 µm. Also studied: Valle Vegazzo, 1862 (B, RO, paratypes).

Mycosphaerella rhododendri Feltgen, Recueil Mém. Trav. Soc. Bot. Grand-Duché Luxembourg: 181. 1901, nomen

novum (Article 58) for *Sphaerella rhododendri* Cooke, J. Bot. 21: 108. 1883, later homonym (illegitimate, Article 53) = *Sphaerella clintoniana* House, New York State Mus. Bull. 205-206: 40. 1919 [“1918”], nomen novum (Article 58) = *Mycosphaerella clintoniana* (House) House, New York State Mus. Bull. 233-234: 26. 1921.— Fig. 200, 763.

Type — United Kingdom: Jordan. On upper and lower surface of dead leaves of *Rhododendron* (Ericaceae). Vize, X 1879 (K (4×), IMI no. 81280 (slide only), isotypes).

This is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores 8-11 × 2-2.5 µm.

Mycosphaerella rhodophila (Pass.) Cruchet, Bull. Soc. Vaud. Sci. Nat. 55: 45. 1923 = *Sphaerella rhodophila* Pass., Atti Reale Accad. Lincei, Mem. Cl. Sci. Fis., ser. 4, 3: 6. 1887.

Type — Italy: *Rosa* (Rosaceae).

Anamorph: Associated with *Discosia artocreas* Tode : Fr. *vide* Saccardo (1891).

No material was studied as the type was not found in any of the herbaria consulted.

Sphaerella rhodora Cooke, Grevillea 13: 99. 1885 = *Laestadia rhodora* (Cooke) Berl. & Voglino, Syll. Fung., Addit. ad vol. 1-4: 62. 1886 = *Botryosphaeria rhodora* (Cooke) M.E. Barr, Mycologia 62: 381. 1970.

Type — United Kingdom: Surrey, Kew, Botanical Garden. On large brown spots with black margins on upper surface of leaves of *Rhododendron* (Ericaceae). Cooke, IV 1885 (K, holotype). Accepted as *Botryosphaeria rhodora* (Cooke) M.E. Barr by Barr (op. cit.), with which the type agrees, with clavate, thick-walled asci, although no ascospores were found.

Mycosphaerella rhodostacheos (Speg.) Cash, Syll. Fung. 26: 351. 1972 = *Sphaerella rhodostacheos* Speg., Bol. Acad. Nac. Ci. 25: 61. 1921.

Type — Chile: *Rhodostachys litoralis* (Bromeliaceae).

No material was studied as the type was not included in a loan from LPS.

Mycosphaerella rhoina (Sacc.) Zerova, Bot. Zhurn. (Kiev) 8: 66. 1951 = *Sphaerella rhoina* Sacc., Ann. Mycol. 6: 561. 1908.— Fig. 761.

Type — Germany: Brandenburg, Tamsel. On lower surface of dead leaves of *Rhus toxicodendron* (Anacardiaceae). Vogel, distributed in Sydow, Mycotheca Germanica no. 680, V 1908 (L, isotype).

Anamorph: *Cercospora* *vide* Joly (1965).

Cited as synonymous with *Mycosphaerella fagi*, which is morphologically indistinguishable from *M. punctiformis*, by Tomilin (1979), with which the isotype studied agrees well, with asci cylindrical, ascospores 8-10 × 2-2.5 µm.

Mycosphaerella rhois (Sawada & Katsuki) C.C. Chen, Bot. Bull. Acad. Sin. 8: 140. 1967, based on an anamorph (illegitimate, Article 59) = *Cercospora rhois* Sawada & Katsuki (1959) non Castell. (1942).

Type — Taiwan: *Rhus semialata* (Anacardiaceae).

No material was studied as the type was not found in BPI.

Mycosphaerella rhois (Sawada) Sivan., *Biblioth. Mycol.* 59: 117. 1977, lacking Latin description (not validated, Article 36), later homonym (illegitimate, Article 53) = *Venturia rhois* Sawada, *Taiwan Univ. Coll. Agric. Special Publ.* 8: 73. 1959, lacking Latin description (not validly published, Article 36).— Fig. 762.

Type — Taiwan: Nantou, Hsinyi. On upper surface of dead leaves of *Rhus javanica* var. *roxburghii* ["*roxburghii*"] (Anacardiaceae). Sawada, VIII 1944 (IMI no. 174953, isotype).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradicans*, with asci cylindrical, ascospores 11-13 × 2.5-3 µm.

Mycosphaerella rhynchosporae Petr., *Kryptog. Forsch.* 2: 168. 1931.

Type — Germany: Oberbayern, Haselmoor near Kachel. On dead leaves and stems of *Rhynchospora alba* (Cyperaceae). Ade, VII 1930 (W, holotype; NY, 2 isotypes).

The holotype and isotypes contain only a coelomycete.

Sphaerella rhytismoides (Bab.) De Not., *Comment. Crittog. Ital.* 2(3): 488. 1867 = *Sphaeria rhytismoides* Bab., *Ann. Mag. Nat. Hist.* 6: 361. 1841 = *Isothea rhytismoides* (Bab.) Fr., *Summa Veg. Scand.*: 421. 1849 = *Laestadia rhytismoides* (Bab.) Sacc., *Syll. Fung.* 1: 424. 1882 = *Hypopsila rhytismoides* (Bab.) Niessl, in G. Winter, in Rabenh., *Fungi Europaei Exsiccati*, ed. nov., ser. 2 no. 3263. 1885.— Fig. 765.

Type — United Kingdom: On upper surface of dead leaves of *Dryas octopetala* (Rosaceae).

Accepted as *Isothea rhytismoides* (Bab.) Fr. by Müller & von Arx (1954), with which material studied (Italy, Valsesia, Riva, Carestia, 1869, distributed in Rabenhorst, *Fungi Europaei Exsiccati* no. 1343, CBS) agrees well, with ascospores simple, 11-14 × 5-7 µm.

Mycosphaerella ribis (Sacc.) Feltgen, *Recueil Mém. Trav. Soc. Bot. Grand-Duché Luxembourg* 14: 285. 1899 = *Sphaerella ribis* Sacc., *Syll. Fung.* 1: 486. 1882, nomen novum (Article 58) for *Sphaeria ribis* Fuckel, *Jahrb. Nassauischen Vereins Naturk.* 23-24: 108. 1870, later homonym (illegitimate, Article 53), non Tode : Fr. (1823).— Fig. 766.

Type — Germany: On upper and lower surface of dead leaves of *Ribes rubrum* (Grossulariaceae).

Anamorphs: *Septoria aurea* Ellis & Everh. and *Septoria ribis* Desm. *vide* Eriksson (1992).

Cited as synonymous with *Mycosphaerella aurea* R. Stone by Eriksson (1992), but the name *Mycosphaerella ribis* has priority. Material studied (Germany, Brandenburg, Baumschulen zu Tamsel, Vogel, distributed in Sydow, *Mycotheca Germanica* no. 2929, VI 1934, L) belongs to section *Longispora*, and is morphologically indistinguishable from *M. latebrosa*, with asci cylindrical, ascospores 23-26 × 3-3.5 µm.

Mycosphaerella ricciae Pavgi & L. Singh, in L. Singh & Pavgi, *Beih. Nova Hedwigia* 63: 19. 1979.— Fig. 767.

Type — India: Utar Pradesh, Varanasi. On upper surface of thallus of *Riccia himalayensis* (Hepaticae, Ricciaceae). Singh, X 1971 (IMI no. 190603, holotype).

This is most probably a species of *Didymella*, with ascomata ca. 0.5 mm diam, thick-walled, asci cylindrical, pseudoparaphyses < 1 µm wide, ascospores variable in shape, 1-3-septate, 15-21 × 6-8 µm.

Mycosphaerella richeae Petr., *Sydowia* 8: 201. 1954.— Fig. 768.

Type — Australia: New South Wales, Mt. Kosciusko. On upper and lower surface of dead leaves of *Richea continens* (Ericaceae). Gauba, distributed in *Reliquiae Petrakianae* no. 862, IV 1953 (H, L, isotypes).

This belongs to section *Longispora*, and is morphologically indistinguishable from *M. millegrana*, with asci cylindrical, ascospores 16-20 × 3-3.5 µm.

Mycosphaerella ricinicola (Speg.) Hemmi & Matuo, in Matuo, *Res. Rep. Fac. Textile Seric. Shinshu Univ.* 3: 8. 1953 = *Sphaerella ricinicola* Speg., *Anales Mus. Nac. Hist. Nat. Buenos Aires* 19: 355. 1909.

Type — Argentina: *Ricinus communis* (Euphorbiaceae).

Anamorph: *Ascochyta ricinella* Sacc. & Scalia *vide* Hemmi & Matuo (op. cit.).

No material was studied as the type was not included in a loan from LPS.

Mycosphaerella ritro (Pass.) Woronow, *Trudy Tiflissk. Bot. Sada*, ser. 2, 3: 139. 1923 = *Sphaerella ritro* Pass., *Atti Reale Accad. Lincei, Mem. Cl. Sci. Fis.*, ser. 4, 3: 7. 1887.

Type — Italy: *Echinops ritro* (Asteraceae).

No material was studied as the type was not found in any of the herbaria consulted.

Mycosphaerella robiniae Siemaszko, *Trudy Byuro Prikl. Bot.* 6: 712. 1913 = *Sphaerella robiniae* (Siemaszko) Trotter, *Syll. Fung.* 24: 874. 1928.

Type — Ukraine: *Robinia pseudacacia* (Fabaceae).

Anamorph: Associated with *Septoria robiniae* Desm. *vide* Siemaszko (op. cit.).

No material was studied as the type was not found in BPI.

Sphaerella rosae Auersw., in Gonn. & Rabenh., *Mycol. Europaea* 5-6: tab. 7, fig. 96. 1869., nomen nudum (not validly published, Article 36) = *Laestadia rosae* Auersw., *Hedwigia* 8: 178. 1869 = *Guignardia rosae* (Auersw.) Petr., *Ann. Mycol.* 18: 109. 1920 = *Discosphaerina rosae* (Auersw.) Petr., *Ann. Mycol.* 22: 36. 1924.

Type — Germany: *Rosa canina* (Rosaceae).

Accepted as *Guignardia rosae* (Auersw.) Petr. by von Arx & Müller (1954) and therefore not studied.

Mycosphaerella rosae Sawada, *Rep. Gov. Res. Inst. Formosa* 87: 26. 1944, lacking Latin description (not validly published, Article 36.1).

Type — Taiwan: *Rosa taiwanensis* (Rosaceae).

No material was studied as the type was not found in BPI.

Mycosphaerella rosicola (Pass.) B.H. Davis, *Mycologia* 30: 296. 1938, based on an anamorph (illegitimate, Article 59) =

Cercospora rosicola Pass., Bot. Jahresber. (Just) 3: 276. 1877 [as “*rosaecola*”] = *Phaeosphaerella rosicola* (Pass.) Tomilin, Opredelitel’ gribov roda *Mycosphaerella* Johans.: 285. 1979.

Type — Italy: Parma. On upper surface of living leaves of *Rosa* (Rosaceae). Passerini, 1874, distributed in Thümen, Mycotheca Universalis no. 1086 (NY, isotype).

The isotype contains only the anamorph (= *Passalora rosicola* (Pass.) U. Braun), as is to be expected.

Mycosphaerella rosigena (Ellis & Everh.) McMurrin ex F.C. Stewart, New York Agric. Exp. Sta. Bull. 328: 389. 1910 = *Sphaerella rosigena* Ellis & Everh., J. Mycol. 3: 45. 1887.— Fig. 769.

Type — USA: Louisiana. On brown spots with purple margins on upper surface of living leaves of *Rosa* (Rosaceae). Langlois no. 689, VI 1886 (NY, holotype).

This belongs to *Davidiella*, of which it represents a parasitic species, with asci pyriform, ascospores 10-12 × 3.5-5 µm.

Therefore the following new combination is made: **Davidiella rosigena** (Ellis & Everh.) Aptroot comb. nov., **MB 500361**. **Basionym:** *Sphaerella rosigena* Ellis & Everh., J. Mycol. 3: 45. 1887.

Sphaerella rosigena var. *comoriensis* Bouriquet & Augé, Agron. Trop. (Nogent-sur-Marne) 12: 310. 1957, lacking Latin description (not validly published, Article 36).

Type — Comores: *Rosa* (Rosaceae).

No material was studied as the type was not included in a loan from PC.

Mycosphaerella rosigena var. *madagascariensis* (Bouriquet) Wallace, Rep. (Annual) Dept. Agric. Tanganyika 1950: 171. 1952, lacking Latin description (not validated, Article 36) = *Sphaerella rosigena* var. *madagascariensis* Bouriquet, Encycl. Mycol. 12: 506. 1946, lacking Latin description (not validly published, Article 36).— Fig. 770.

Type — Madagascar: On white spots with black margins on upper surface of living leaves of *Rosa* (Rosaceae).

The location of the type is unknown and its preservation uncertain. Material studied (Tanzania, Moshi, Lyamungu, V 1950, IMI no. 150328; also China, Hong Kong, Sheun Shui Mountain Garden, Johnston, VI 1962, IMI no. 94801b; also Mauritius, Felix, XI 1958, IMI no. 75374) belong to section *Plaga*, with asci cylindrical, ascospores 13-16 × 3-4 µm.

Mycosphaerella rostkoviae (Speg.) Lindau, in Engl. & Prantl, Natürlichem Pflanzenf. 1(1): 426. 1897 = *Sphaerella rostkoviae* Speg., Bol. Acad. Nac. Ci. 11: 207. 1888 [“1887”].

Type — Argentina: *Rostkovia grandiflora* (Juncaceae).

No material was studied as the type was not included in a loan from LPS.

Mycosphaerella rottboelliae Kranz, Nova Hedwigia 18: 237. 1969.— Fig. 771.

Type — Guinea: Kindia. On white spots with black margins on upper surface of living leaves of *Rottboellia exaltata* (Poaceae). Kranz, X 1963 (IMI no. 104989, holotype).

This belongs to *Davidiella*, of which it represents a parasitic species, and is morphologically indistinguishable from *D. spilota* (see below), with asci pyriform, ascospores 17-22 × 4-5.5 µm.

Sphaerella rottlerae (Berk. & Broome) Sacc., Syll. Fung. 1: 536. 1882 = *Sphaeria rottlerae* Berk. & Broome, J. Linn. Soc., Bot. 14: 130. 1875.

Type — Sri Lanka: On white spots with brown margins on upper surface of living leaves of *Tetragastris* [“*Rottlera*”] *quadricocca* (Burseraceae). Broome no. 1228 (K, holotype; K, isotype).

The type contains only coelomycetes, which is in agreement with the original drawing on the packet.

Mycosphaerella roureae Syd. & P. Syd., Philipp. J. Sci. 8: 270. 1913 = *Sphaerella roureae* (Syd. & P. Syd.) Trotter, Syll. Fung. 24: 858. 1928.— Fig. 772.

Type — Philippines: Luzon, Prov. Bataan. On spots on upper surface of living leaves of *Rourea erecta* (Connaraceae). Graff, distributed in Sydow, Fungi Exotici Exsiccati no. 128, XI 1912 (L, isotype).

This is a parasitic species, with asci cylindrical, ascospores 15-17 × 3-3.5 µm, associated with a *Septoria* anamorph.

Mycosphaerella rousseliana (Desm.) Rehm, Oesterr. Bot. Z. 56: 295. 1906 = *Sphaeria rousseliana* Desm., Ann. Sci. Nat. Bot., ser. 3, 11: 355. 1849 = *Sphaerella rousseliana* (Desm.) Auersw., in Rabenh., Fungi Europaei Exsiccati, ed. nov., ser. 2 no. 1345. 1870.

Type — France: *Phleum phleoides* [“*boehmeri*”] (Poaceae). Material studied (Geramny, Stralsund, on dead culms and upper and lower surface of dead leaves of *Phleum pratense*, Fischer, distributed in Rabenhorst, Fungi Europaei Exsiccati, ed. nov., ser. 2 no. 1345, CBS) contains *Pleospora herbarum* (Pers. : Fr.) Rabenh. and a *Phaeosphaeria*.

Sphaerella rousseliana f. *poae* Roum., Fungi Gallici Exsiccati no. 2454.— Fig. 773.

Type — France: Toulouse. On dead culms of *Poa* (Poaceae). Roumeguère, Fungi Gallici Exsiccati no. 2454, 1882 (L, isotype).

This is a *Leptosphaeria*, with ascospores brown, 17-20 × 3-4 µm.

Sphaerella rouxii (Mont.) Ces. & De Not., Comment. Soc. Crittog. Ital. 1(4): 237. 1863 = *Sphaeria rouxii* Mont., Ann. Sci. Nat. Bot., ser. 4, 12: 184. 1858 = *Laestadia rouxii* (Mont.) Sacc., Syll. Fung. 1: 428. 1882.

Type — France: *Erica multiflora* (Ericaceae).

No material was studied as the species was already redisposed.

Mycosphaerella rubefaciens B. Erikss., Svensk Bot. Tidskr. 68: 217. 1974.

Type — Finland: On upper and lower surface of reddish but are still attached dead leaves of *Vaccinium vitis-idaea* (Ericaceae).

The type of this recently described species was not studied. Material seen (Canada, Yukon, Dempster, Ginns no. 5713 & Cady, VIII 1980, BPI) is immature.

Mycosphaerella rubella (Niessl) Magnus, in Dalla Torre & Sarnth., Fl. Tirol, Vorarlberg, Liechtenstein, III, Pilze: 465. 1905 = *Sphaerella rubella* Niessl, in Rabenh., Fungi Europaei Exsiccati, cent. 22 no. 2131. 1876 = *Haplotheciella rubella* (Niessl) Höhn., Mitt. Bot. Lab. TH Wien 2: 92. 1925.— Fig. 774.

Type — Germany: Baden, Rastatt. On dead stems of *Angelica sylvestris* (Apiaceae). Schröter, distributed in Rabenhorst, Fungi Europaei Exsiccati no. 2131, IV-V 1876 (L, isotype).

This belongs to section *Caterva*, with mycelium turning the substratum red, asci cylindrical, ascospores 13-17 × 3-4.5 µm. Additional material studied (Netherlands, Baarn, Arx, IV 1950, L) agrees well.

Mycosphaerella rubi Roark, Phytopathology 11: 329. 1921.

Type — USA: Wisconsin, Madison. On white spots on upper surface of living leaves of *Rubus occidentalis* (Rosaceae). Honey, X 1944 (B, topotype).

Anamorph: *Septoria rubi* (Duby) Westend. *vide* Roark (op. cit.).

The isotype studied contains only the anamorph.

Mycosphaerella rubiae (Brunaud) Tomilin, Novosti Sist. Nizsh. Rast. 8: 151. 1971 = *Sphaerella rubiae* Brunaud, Ann. Soc. Sci. Nat. La Rochelle 1892: 105. 1892.

Type — France: *Rubia tinctoria* (Rubiaceae).

No material was studied as the type was not included in a loan from PC.

Mycosphaerella rubicola (McAlpine) Tomilin, Novosti Sist. Nizsh. Rast. 1968: 167. 1968 = *Sphaerella rubicola* McAlpine, Proc. Linn. Soc. New South Wales 29: 126. 1904.

Type — Australia: *Rubus parvifolius* (Rosaceae).

No material was studied as the type was not found in any of the herbaria consulted. This is no *Mycosphaerella* according to Tomilin (1979).

Mycosphaerella rubiginosa (Cooke) Lindau, in Engl. & Prantl, Natürlichen Pflanzenf. 1(1): 425. 1897 = *Sphaerella rubiginosa* Cooke, Grevillea 14: 90. 1886.— Fig. 775.

Type — Australia: Queensland, Johnston River. On upper surface of dead leaves of *Pittosporum rubiginosum* (Pittosporaceae). Müller (K, holotype).

This is a *Guignardia* species, without hamathecium, ascospores simple, 14-16 × 5-6 µm. On the type also an *Aulaxina* species is present.

Mycosphaerella rubina (Peck) House, New York State Mus. Bull. 233-234: 30. 1921 = *Sphaerella rubina* Peck, Annual Rep. New York State Mus. 48: 114. 1897 [“1894”].— Fig. 776.

Type — USA: New York, Albany Co., Menands. On dead stems of *Rubus idaeus* (Rosaceae). Peck, IV (NY, isotype).

Anamorph: *Phoma* *vide* Corbaz (1957).

Cited by Corbaz (1957) as morphologically indistinguishable from *Didymella applanata* (Niessl) Sacc., with which the isotype agrees well, with ascomata applanate, pseudoparaphyses anastomosing, 1-2 µm wide, asci clavate, ascospores 14-17 × 4-6 µm.

Mycosphaerella [“*Mycosphaerium*”] *rudbeckiae* Clem., Cryptogamae Formationum Coloradensium no. 226. 1906, nomen herbariorum (not validly published, Article 32).

Authentic material — USA: Colorado, Ruxton Dell. On dead stems of *Rudbeckia flava* (Asteraceae). Clements, VII 1906, Cryptogamae Formationum Coloradensium no. 226 (BPI).

The material is postmature.

Mycosphaerella rumicis (Desm.) Cruchet, Bull. Soc. Vaud. Sci. Nat. 55: 43. 1923 = *Sphaeria rumicis* Desm., Ann. Sci. Nat. Bot., ser. 2, 19: 361. 1843 = *Sphaerella rumicis* (Desm.) Cooke, J. Bot. 4: 251. 1866 = *Venturia rumicis* (Desm.) G. Winter, in Rabenh., Kryptog.-Fl. Deutschl., Österr. Schweiz, ed. 2, 1(2): 435. 1885 = *Stigmatea rumicis* (Desm.) J. Schröt., in Cohn, Kryptog.-Fl. Schlesien 3(2): 332. 1894 [“1893”] = *Spilosticta rumicis* (Desm.) Syd., Ann. Mycol. 21: 172. 1923.— Fig. 777.

Type — France: On upper surface of living leaves of *Rumex nemolapathus* (Polygonaceae). Desmazières, Fungi Exsiccati no. 798 (L, isotype).

Anamorph: *Ovularia obovata* (Fuckel) Sacc. *vide* Saccardo (1882) (= *Ramularia rubella* (Bonord.) Nannf.).

Accepted as *Venturia rumicis* (Desm.) G. Winter by Sivanesan (1977), with which the isotype and additional material studied (The Netherlands, Haarlem, on *R. obtusifolius*, Oudemans no. 574, 1870, L) agree well, with asci cylindrical, surrounded by paraphysoids, ascospores pale brown, asymmetrically septate, 14-17 × 5-7 µm.

Mycosphaerella rumicis f. *caulicola* Grove, J. Bot. 71: 253. 1933.

Type — United Kingdom: *Rumex* (Polygonaceae).

No material was studied as the type was not found in K and might be lost.

Mycosphaerella rusci D. Gupta, Padhi & P.N. Chowdhry, Indian Phytopathol. 33: 502. 1981 [“1980”].

Type — India: *Ruscus aculeatus* (Asparagaceae).

No material was studied as the type was not found in LWG or IMI.

Sphaerella rusci (Fr.) Ces. & De Not., Comment. Soc. Crittog. Ital. 1(4): 237. 1863 = *Sphaeria atrovirens* var. *rusci* Fr., Systema Mycol. 2: 501. 1823 = *Sphaeria rusci* (Fr.) Wallr., Fl. Cryptog. Germ. 2: 776. 1833 = *Leptosphaeria rusci* (Fr.) Sacc., Syll. Fung. 2: 74. 1883.— Fig. 778.

Type — Sweden: On upper and lower surface of dead phyllocladia of *Ruscus aculeatus* (Asparagaceae).

Material studied (France, Pau, Roumeguère, Fungi Selecti Galliae no. 290, VII 1878, L) belongs to *Paraphaeosphaeria glaucopunctata* (Greville) Shoemaker & C.E. Babc., with asci surrounded by hamathecial filaments, ascospores consistently 4-septate, brown, 18-26 ×

4.5-6 μm , which agrees with the description by Saccardo (op. cit.).

Mycosphaerella ruscicola A. Pande, Current Sci. 49: 523. 1980.

Type — India: Pune. On upper and lower surface of dead phyllocladia of *Ruscus aculeatus* (Asparagaceae). Bansude no. 4367, VII 1978 (LWG, holotype).

Anamorph: *Cercospora ruscicola* V.G. Rao & A.S. Patil *vide* Pande (op. cit.).

The holotype contains only empty ascomata.

Mycosphaerella ruthenica Petr., Hedwigia 65(1): 233. 1925.— Fig. 779.

Type — Poland: Stanislaw, Podluzie. On dead stems of *Astragalus glycyphyllos* (Fabaceae). Petrak, Fungi Polonici Exsiccati no. 583, III 1918 (W, holotype; W, 2 isotypes).

Cited as synonymous with *Mycosphaerella elodis* by Tomilin (1979). Examination of the types show that this belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradicans*, with ascomata very variable in size, asci cylindrical, ascospores 12-14 \times 3-4 μm .

Sphaerella sabaligena Ellis & Everh., J. Mycol. 2: 101. 1886.— Fig. 780.

Type — USA: Louisiana, Point à la Hache. On upper and lower surface of dead leaves of *Sabal palmetto* (Arecaceae). Langlois no. 426, I [in original letter; erroneously given as “IV” in the publication] 1886 (NY, holotype); also distributed in Ellis & Everh., North American Fungi no. 1800, V 1886 (L, topotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. disseminata*, with asci pyriform, ascospores 13-15 \times 3-4 μm ; the holotype now contains only coelomycetes.

Mycosphaerella sabalis (Sousa da Câmara) Cash, Syll. Fung. 26: 351. 1972 [as “*sabalidis*”] \equiv *Sphaerella sabalis* Sousa da Câmara, Bol. Agric. 2: 20. 1936 [as “*sabalidis*”].

Type — Portugal: *Sabal palmetto* (Arecaceae).

Anamorph: *Coniothyrium sabalis* Sousa da Câmara *vide* Sousa da Câmara (op. cit.).

No material was studied as the type was not found in LISE.

Mycosphaerella sabinae Feltgen, Recueil Mém. Trav. Soc. Bot. Grand-Duché Luxembourg 15: 258. 1903 \equiv *Sphaerella sabinae* (Feltgen) Sacc. & D. Sacc., Syll. Fung. 17: 642. 1905.

Type — Belgium: *Juniperus sabina* (Cupressaceae).

No material was studied as this is certainly no *Mycosphaerella*. Cited as synonymous with *Mycosphaerella juniperina* by Tomilin (1979).

Mycosphaerella saccardoana Jaap, Ann. Mycol. 14: 12. 1916 \equiv *Sphaerella saccardoana* (Jaap) Trotter, Syll. Fung. 24: 870. 1928.

Type — Yugoslavia: *Juglans regia* (Juglandaceae).

Anamorph: Associated with *Septoria letendreaana* Sacc. *vide* Trotter (op. cit.).

No material was studied as the type was not found in any of the herbaria consulted. Cited as synonymous with *Mycosphaerella terebinthi* by Tomilin (1979).

Mycosphaerella sacchari (Speg.) Seaver & Chardón, Sci. Surv. Porto Rico & Virgin Islands 8: 62. 1926 \equiv *Sphaerella sacchari* Speg., Revista Fac. Agron. Univ. Nac. La Plata, 1896: 230. 1896.

Type — Argentina: *Saccharum officinarum* (Poaceae).

No material was studied as the type was not included in a loan from LPS.

Sphaerella sacchari Wakker. See *Mycosphaerella wakkeri* (Sacc. & P. Syd.) Tomilin.

Mycosphaerella saccharoides (Peck) Tomilin, Novosti Sist. Nizsh. Rast. 1967: 190. 1967 \equiv *Sphaerella saccharoides* Peck, New York State Mus. Bull. 167: 48. 1913.

Type — Cuba: *Saccharum officinarum* (Poaceae).

No material was studied as the type was not found in NY or BPI.

Mycosphaerella sagedioides (G. Winter ex Sacc.) Lindau, in Engl. & Prantl, Natürlichen Pflanzenf. 1(1): 424. 1897 \equiv *Sphaerella sagedioides* G. Winter ex Sacc., Syll. Fung. 1: 517. 1882 \equiv *Sphaerella sagedioides* f. *dipsaci-sylvestris* G. Winter, in Kunze, Fungi Selecti Exsiccati, cent. 4 no. 324. 1880, superfluous (illegitimate, Article 52) nomen novum (Article 58).— Fig. 781.

Type — Switzerland: Zürich. On dead stems of *Dipsacus sylvestris* (Dipsacaceae). Winter, distributed in Kunze, Fungi Selecti Exsiccati, cent. 4 no. 324, VIII 1878 (L, lectotype, here designated).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. ammophilae*, with asci slender pyriform, ascospores 10-12 \times 3-3.5 μm .

Sphaerella sagedioides f. *dauci-carotae* G. Winter, in J. Kunze, Fungi Selecti Exsiccati, cent. 4 no. 323. 1880.— Fig. 782.

Type — Switzerland: Zürich. On dead stems of *Daucus carota* (Apiaceae). Winter, distributed in Kunze, Fungi Selecti Exsiccati no. 323, VIII 1878 (L, holotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. ammophilae*, with asci slender pyriform, ascospores 10-12 \times 3-3.5 μm .

Sphaerella sagedioides f. *dipsaci-sylvestris* G. Winter see *Mycosphaerella sagedioides* (G. Winter ex Sacc.) Lindau.

Mycosphaerella saginae Urries, Anales Inst. Bot. Cavanilles 14: 161. 1956.

Type — Canary Islands: La Palma, Baranco de Gallegos. On dead parts of living leaves of *Sagina* [“*procumbens* var.”] *apetala* (Caryophyllaceae). Urries, IV 1954 (MA, holotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with ascomata in dense groups, asci pyriform, ascospores immature.

Mycosphaerella sagittariae (Tassi) Tomilin, Novosti Sist. Nizsh. Rast. 1966: 173. 1966 = *Sphaerella sagittariae* Tassi, Bull. Lab. Orto Bot. Reale Univ. Siena 3: 118. 1900.— Fig. 783.

Type — Italy: Siena, Botanical Garden. On upper and lower surface of dead leaves of *Sagittaria montevidensis* (Alismataceae). Tassi, X 1900 (SIENA, holotype).

Already cited as synonymous with *Davidiella allicina* (as *Mycosphaerella allicina*) by Tomilin (1979), with which the holotype agrees well, with asci pyriform, ascospores 17-22 × 4.5-6 µm.

Mycosphaerella sajanyca Petr., Hedwigia 68: 210. 1928 [“1929”].— Fig. 784.

Type — Russia: Siberia, Sajany. On dead stems of *Rhaponicum* [“*Leuzea*”] *carthamoides* (Asteraceae). Murashkinsky no. a27, VII 1927 (W, lectotype, here designated).

Additional material studied: Same data, nos a52 & o66 (W, paratypes). This belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores 18-20 × 4.5-5.5 µm.

Mycosphaerella salicicola (Fr.) Johanson ex Oudem., Rév. Champ. Pays-Bas 2: 210. 1897 = *Depazea salicicola* Fr., Systema Mycol. 2: 530. 1823 = *Sphaerella salicicola* (Fr.) Fuckel, Jahrb. Nassauischen Vereins Naturk. 23-24: 106. 1870 = *Sphaeria salicicola* (Fr.) Rabenh., in Fuckel, Fungi Rhenani no. 836.

Type — Sweden: *Salix caprea* (Salicaceae).

Anamorph: *Septoria salicicola* (Fr.) Sacc. *fide* Grove (1935).

The type was not included in a loan from UPS. Already cited as synonymous with *M. punctiformis* by Tomilin (1979), with which material studied (Germany, On upper and lower surface of dead leaves of *Salix triandra*, Fuckel, Fungi Rhenani no. 836, L; also Leipzig, on lower surface of dead leaves of *S. amygdalina*, Auerswald, B), which are both immature, probably agree.

Mycosphaerella salicicola f. *amygdalinae* Jaap, Verh. Bot. Vereins Prov. Brandenburg 47: 85. 1905 = *Sphaerella salicicola* f. *amygdalinae* (Jaap) Sacc. & Trotter, Syll. Fung. 22: 140. 1913.— Fig. 785.

Type — Germany: Brandenburg, Triglitz in der Prignitz. On lower surface of dead leaves of *Salix amygdalina* (Salicaceae). Jaap, Fungi Selecti Exsiccati no. 79c, V 1904 (L, isotype).

Anamorph: *Ramularia rosea* (Fuckel) Sacc. *fide* Saccardo & Trotter (op. cit.).

This is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores 8-10 × 2-2.5 µm.

Sphaerella salicicola f. *salicis-fragilis* Thüm., Fungi Austriaci no. 161. 1871.

Type — Austria: Niederösterreich, Krems. On small white spots with black margins on upper surface of living leaves of *Salix fragilis* (Salicaceae). Thümen, Fungi Austriaci no. 161, IX 1871 (B, BPI, isotypes).

Anamorph: *Septoria salicis* Westend. *fide* Thümen (op. cit.).

The isotypes contain only the anamorph, and another coelomycete, which is in accordance with the exsiccate label, which mentions “Fungus spermogonium”.

Sphaerella salicicola f. *salicis-triandrae* J. Kunze, Fungi Selecti Exsiccati no. 243. 1879.— Fig. 786.

Type — Germany: Sachsen, Eisleben. On upper and lower surface of dead leaves of *Salix triandra* (Salicaceae). Kunze, Fungi Selecti Exsiccati no. 243, V 1879 (L, holotype).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradicans*, with asci cylindrical, ascospores 11-13 × 2-2.5 µm.

Sphaerella salicicola var. *borealis* P. Karst., Hedwigia 23: 3. 1884.

Type — Finland: *Salix phylicifolia* (Salicaceae).

No material was studied as the type was not included in a loan from H.

Mycosphaerella salicina (Ellis & Everh.) Tomilin, Novosti Sist. Nizsh. Rast. 1967: 190. 1967 = *Sphaerella salicina* Ellis & Everh., J. Mycol. 9: 166. 1903.— Fig. 787.

Type — USA: Kansas, Rooks Co. On dead branches of *Salix cordata* (Salicaceae), Bartholomew no. 2949, V 1902 (BPI, isotype).

No type material was preserved in NY, so the isotype in BPI may represent the only existing material. It shows that this is morphologically indistinguishable from *Cryptodiaporthe salicella* (Fr.) Petr., with ascomata gregarious, ostioles fused, asci fusiform, loose in the ascomata when mature, ascospores 14.5-19 × 4.5-5 µm. Its anamorph *Discella carbonacea* (Fr.) Berk. & Broome is also present.

Sphaerella salicinearum Pass., in Rabenh., Fungi Europaei Exsiccati, ed. nov., ser. 2, cent. 25 no. 2445. 1876 = *Sphaerella salicinearum* f. *salicis-albae* Pass., in Rabenh., Fungi Europaei Exsiccati, ed. nov., ser. 2, cent. 25 no. 2445. 1876, superfluous (illegitimate, Article 52) nomen novum (Article 58) = *Leptosphaeria salicinearum* (Pass.) Sacc., Syll. Fung. 2: 50. 1883.

Type — Italy: On upper and lower surface of dead leaves of *Salix alba* (Salicaceae). Passerini, distributed in Rabenhorst, Fungi Europaei Exsiccati, ed. nov., ser. 2 no. 2445 (BPI, CBS, isotypes).

The isotype in CBS contains only the coelomycete *Phoma*, the isotype in BPI an immature ascomycete.

Sphaerella salicinearum f. *populi-nigrae* Pass., in Rabenh., Fungi Europaei Exsiccati, ed. nov., ser. 2, cent. 25 no. 2444. 1876.— Fig. 788.

Type — Italy: On white spots on upper and lower surface of living leaves of *Populus nigra* (Salicaceae). Passerini, distributed in Rabenhorst, Fungi Europaei Exsiccati no. 2444 (CBS, isotype).

The isotype studied belongs to *Phaeosphaeria*, with asci surrounded by hamathecial filaments, ascospores consistently 3-septate, brown, 17-22 × 3.5-5 µm, which agrees with the description by Saccardo (1883).

Sphaerella salicinearum f. *salicis-albae* Pass. See *Sphaerella salicinearum* Pass.

Sphaerella salicis Bubák & Vleugel, in Vleugel, Svensk Bot. Tidskr. 11: 310. 1917.— Fig. 789.

Type — Sweden: Luleå. On lower surface of dead leaves of *Salix nigricans* (Salicaceae). Vleugel, V 1913 (BPI, holotype).

Anamorph: *Phloeospora salicis* Bubák & Vleugel *vide* Bubák & Vleugel (op. cit.).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. superflua*, with asci clavate to cylindrical, ascospores 14-17 × 4.5-5 µm.

Mycosphaerella salicorniae (Rabenh.) Lindau, Hilfsb. Sammeln Ascomyceten: 103. 1903 = *Pleospora herbarum* f. *salicorniae* Rabenh., Fungi Europaei Exsiccati no. 145 = *Sphaerella salicorniae* (Rabenh.) Auersw., in Gonn. & Rabenh., Mycol. Europaea 5-6: 16. 1869.— Fig. 790.

Type — Germany: Leipzig. On dead stems of *Salicornia herbacea* (Amaranthaceae). Auerswald (B, lectotype).

Material studied (USA, Connecticut, Green Farm, Thaxter, distributed in Ellis & Everhart, North American Fungi no. 2759, V 1890, L) belongs to *Davidiella*, and is morphologically indistinguishable from *D. ammophilae*, with asci pyriform to globose, ascospores 10-13 × 3-4 µm.

Mycosphaerella salvatorensis Jaap, Ann. Mycol. 15: 105. 1917 [as “*salvatorensis*”] = *Sphaerella salvatorensis* (Jaap) Trotter, Syll. Fung. 24: 882. 1928.

Type — Switzerland: *Helleborus viridis* (Ranunculaceae).

No material was studied as the type was not found in any of the herbaria consulted.

Mycosphaerella salviae (Strasser) Tomilin, Novosti Sist. Nizsh. Rast. 7: 205. 1970 = *Sphaerella salviae* Strasser, Verh. K.K. Zool.-Bot. Ges. Wien 57: 311. 1907.— Fig. 791.

Type — Austria: Rosenau. On upper and lower surface of dead leaves of *Salvia glutinosa* (Lamiaceae). Strasser, Kryptogamae Exsiccatae no. 2406 (L, isotype).

Cited as synonymous with *Mycosphaerella isariphora* by Tomilin (1979). The isotype shows that it belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradicans*, with asci cylindrical, ascospores 10-13 × 2.5-3.5 µm.

Mycosphaerella samanae Chardón, Bol. Soc. Venez. Ci. Nat. 40: 362. 1939, lacking Latin description (not validly published, Article 36).— Fig. 792.

Type — Venezuela: Caracas. On white spots with brown margins on lower surface of living leaves of *Samanea saman* (Fabaceae). Muhler no. 2075, XII 1937 (BPI, 2 isotypes).

This is a parasitic species, with asci cylindrical, ascospores 16-19 × 4-5 µm.

Mycosphaerella sanguisorbae Lobik, Bolezni Rast. 17: 164. 1928.— Fig. 793.

Type — Russia: On white spots with purple margins on lower surface of living leaves of *Sanguisorba officinalis* (Rosaceae). (LE 35032, holotype).

Cited as synonymous with *Mycosphaerella pseudomaculiformis* by Tomilin (1979). The type shows that this belongs to *Davidiella*, and is morphologically

indistinguishable from *D. ammophilae*, with asci pyriform, ascospores 10-12 × 2.5-3.5 µm.

Mycosphaerella sapindi (Ellis & Everh.) Lindau, in Engl. & Prantl, Natürlichem Pflanzenf. 1(1): 425. 1897 = *Sphaerella sapindi* Ellis & Everh., J. Mycol. 2: 101. 1886 = *Sphaerella quadrangulatae* Ellis & Everh., N. Amer. Pyrenomyc.: 274. 1892 [as “*quadrangulata*”], superfluous (illegitimate, Article 52) nomen novum (Article 58) = *Mycosphaerella quadrangulatae* (Ellis & Everh.) Tomilin, Novosti Sist. Nizsh. Rast. 7: 205. 1970 [as “*quadrangulata*”], illegitimate combination, because basionym illegitimate.— Fig. 794.

Type — USA: Missouri, Columbia. On white spots with dark margins on upper surface of living leaves of *Fraxinus quadrangulata* (Oleaceae) [initially erroneously identified as “*Sapindus marginatus* (Sapindaceae)” but this does not legitimate a name change of the fungal species]. Galloway, VII 1886 (NY, holotype).

Anamorph: *Alternaria* [“*Macrosporium*”] *vide* Ellis & Everhart (1892).

Cited as synonymous with *Mycosphaerella effigurata* by Tomilin (1979). The type is in bad shape, but shows that this belongs to *Davidiella*, of which it represents a parasitic species, with asci pyriform, ascospores thick-walled, 15-17 × 5.5-7 µm. Therefore the following new combination is made: **Davidiella sapindi** (Ellis & Everh.) Aptroot comb. nov., **MB 500362**. **Basionym:** *Sphaerella sapindi* Ellis & Everh., J. Mycol. 2: 101. 1886.

Mycosphaerella sapindicola Pai, Mycopathol. Mycol. Appl. 35: 127. 1968.— Fig. 795.

Type — India: Solan, Simla Hills. On white spots with brown margins covering the upper surface of the tips of living leaves of *Sapindus mukorosii* (Sapindaceae). Sohi, IV 1963 (IMI no. 132809, probable isotype).

Anamorph: *Phyllosticta sapindi-emarginata* V.G. Rao *vide* Pai (op. cit.).

This is a species of *Didymella*, with asci cylindrical, pseudoparaphyses present, ascospores pointed, variable in size and shape, with an *Ascochyta* anamorph.

Sphaerella sarmentorum Pirota, Arch. Triennale Lab. Bot. Crittog. 2-3: 132. 1879, nomen novum (Article 58) for *Sphaerella vitis* Schulzer, Verh. K.K. Zool.-Bot. Ges. Wien 20: 643. 1870 (autumn), later homonym (illegitimate, Article 53) [cited by Saccardo as “*vitigena*”] = *Leptosphaeria vitigena* Sacc., Syll. Fung. 2: 29. 1883, superfluous (illegitimate, Article 52) nomen novum (Article 58).

Type — Austria: *Vitis vinifera* (Vitaceae).

No material was studied as the species was already redisposed.

Mycosphaerella sarothamni Petr., Ann. Mycol. 22: 57. 1924.— Fig. 797.

Type — Czech Republic: Weißkirchen, Hrabuvka. On dead branches of *Sarothamnus scoparius* (Fabaceae). Petrak, distributed in Reliquiae Petrakianae no. 70, IX 1923 (H, L, isotypes)

This is no *Mycosphaerella*, but possibly an immature *Dothidea*, with asci thick-walled, bitunicate, cylindrical,

surrounded by paraplectenchymatous cells, ascospores 16-20 × 5-6 µm.

Mycosphaerella sarraceniae (Schwein. : Fr.) Henn., Verh. Bot. Vereins Prov. Brandenburg 40: 157. 1898 ≡ *Sphaeria sarraceniae* Schwein., Schriften Naturf. Ges. Leipzig 1: 35. 1822, sanctioned by Fr., Systema Mycol. 2: 516. 1823 ≡ *Sphaerella sarraceniae* (Schwein. : Fr.) Peck, Annual Rep. New York State Mus. 29: 70. 1878.— Fig. 798.

Type — USA: New Jersey, Hope. On upper and lower surface of dead leaves of *Sarracenia* (Sarraceniaceae). (PH, holotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. ammophilae*, with asci pyriform, ascospores 10-12 × 4-5 µm. Additional material seen (Salem, PH) agrees.

Mycosphaerella sarracenia (Sacc. & Roum.) Lindau, Hilfsb. Sammeln Ascomyceten: 112. 1903 [as “sarracenia”] ≡ *Sphaerella sarracenia* Sacc. & Roum., in Roum. & Sacc., Rev. Mycol. (Toulouse) 3: 46. 1881.— Fig. 796.

Type — Belgium: Malmedy. On upper and lower surface of dead leaves of *Senecio fuchsii* [“sarracenicus”] (Asteraceae). Libert, distributed in Roumeguère, Fungi Gallici Exsiccati no. 1607, VII 1881 (L, isotype).

The isotype studied contains only empty ascomata. Additional material studied (Czech Republic, Weißkirchen, Petr., distributed in Reliquiae Petrakianae no. 242, V 1914, L) is *M. punctiformis*, with asci cylindrical, ascospores 8-10 × 2-2.5 µm.

Mycosphaerella sassafras (Ellis & Everh.) Cash, Syll. Fung. 26: 352. 1972 ≡ *Sphaerella sassafras* Ellis & Everh., Bull. Torrey Bot. Club 10: 98. 1883.— Fig. 799.

Type — USA: Ohio, Amanda, Fairfield Co. On upper and lower surface of dead leaves of *Sassafras officinale* (Lauraceae). Kellerman no. 230, V [on label; “VI” in publication] 1883 (NY, holotype), also distributed in Ellis & Everh., North American Fungi no. 1683 (L, isotype), also distributed in Rabenhorst-Winter, Fungi Europaei Exsiccati no. 3448 (L, isotype).

This is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores 7-10 × 2-3 µm.

Mycosphaerella sassafras f. *major* (Nann.) Cash, Syll. Fung. 26: 352. 1972 ≡ *Sphaerella sassafras* f. *major* Nann., Atti Reale Accad. Fisiocrit. Siena, ser. 10, 4: 89. 1929.

Type — Italy: Siena, Botanical Garden. On upper and lower surface of dead leaves of *Sassafras officinale* (Lauraceae). Nannizzi, X 1928 (SIENA, holotype).

The holotype studied is immature.

Mycosphaerella saussureae-alpinae Petr., Ann. Mycol. 34: 447. 1936.— Fig. 800.

Type — Finland: Lapponia, Petsamo, Pummanki. On upper and lower surface of dead leaves of *Saussurea alpina* (Asteraceae). Kari, VII 1931 (W, holotype).

Cited as synonymous with *Mycosphaerella lysimachiae* by Tomilin (1979). The type shows that it belongs to section *Caterva*, and is morphologically indistinguishable from *M.*

subradians, with asci cylindrical, ascospores 12-14 × 2.5-3.5 µm.

Mycosphaerella sawadae Tomilin, Novosti Sist. Nizsh. Rast. 1968: 167. 1968, nomen novum (Article 58) for *Mycosphaerella aquilegiae* Sawada, Bull. Gov. Forest Exp. Sta. 53: 165. 1952, later homonym (illegitimate, Article 53). Type — Japan: *Aquilegia buergeriana* (Ranunculaceae). No material was studied as the type was not found in BPI.

Mycosphaerella saxatilis (J. Schröt.) Tomilin, Novosti Sist. Nizsh. Rast. 1967: 190. 1967 ≡ *Sphaerella saxatilis* J. Schröt., Jahresber. Schles. Ges. Vaterl. Cult. 58: 173. 1880.

Type — Sweden: *Carex saxatilis* (Cyperaceae). No material was studied as the type was not found in B and might be lost. Synonymised with *Davidiella allicina* (as *Mycosphaerella tassiana*) by von Arx (1949).

Mycosphaerella saxifragae (Pass.) Lind, Biol. Meddel. Kongel. Danske Vidensk. Selsk. 11(2): 69. 1934 ≡ *Sphaerella saxifragae* Pass., Atti Reale Accad. Lincei, Mem. Cl. Sci. Fis., ser. 4, 3: 6. January 1887.— Fig. 801.

Type — Italy: *Saxifraga muscoides* (Saxifragaceae). Anamorph: *Cercospora saxifragae* Rostr. fide Sivanesan (1984) (= *Pseudocercospora saxifragae* (Rostr.) U. Braun).

Cited as synonymous with *Davidiella allicina* (as *Mycosphaerella allicina*) by Tomilin (1979). No type material was found in any of the herbaria consulted. Material seen (Canada, Northwest Territories, Keewatin District, Chesterfield Inlet, on *S. nivalis*, Sairle & Watts, VII 1950, IMI no. 200598; also same locality, on *S. cernua*, Sairle & Watts, VIII 1950, IMI no. 200599, both slides only) contains a *Mycosphaerella* with asci cylindrical, ascospores 19-22 × 3-4.5 µm. Its identity is difficult to judge from slides only.

Sphaerella saxifragae P. Karst. See *Sphaerella olenjana* Sacc.

Mycosphaerella scabiosae Tomilin. See *Mycosphaerella sylvatica* (Sacc. & Speg.) Laib.

Mycosphaerella scaevolae F. Stevens & K. Young, in F. Stevens, Bernice P. Bishop Mus. Bull. 19: 104. 1925.— Fig. 802, 989.

Type — Hawaii: Oahu. On brown spots with black margins on upper and lower surface of living leaves of *Scaevola gaudichadiana* (Goodeniaceae). Pryer, IV 1947 (BPI, topotype).

This is a parasitic species, with asci cylindrical, ascospores (15-)17-19 × 4-5 µm.

Sphaerella schaeferi (A. Massal.) Anzi, Atti Soc. Ital. Sci. Nat. (Milano) 11: 180. 1868 ≡ *Sphaeria schaeferi* A. Massal., Nota Sulla Lecidea Hookeri: 8. 1853 ≡ *Stigmidium schaeferi* (A. Massal.) Trevis., Conspect. Verruc.: 17. 1860 ≡ *Verrucaria schaeferi* (A. Massal.) Nyl., Flora 53: 358. 1865 ≡ *Pharcidia schaeferi* (A. Massal.) Arnold, Verh. K.K. Zool.-Bot. Ges. Wien 19: 638. 1869 ≡ *Epicymatia schaeferi* (A. Massal.) Sacc., Syll. Fung. 1: 571. 1882 ≡

Sphaerulina schaeferi (A. Massal.) Sacc., Syll. Fung. 17: 695. 1905.

Type — Italy: *Solorina* (Ascomycota, Peltigeraceae).

Accepted as *Stigmidium schaeferi* (A. Massal.) Trevis. by Clauzade, Diederich & Roux (1989) and therefore not studied.

Mycosphaerella schelkovnikovii Woron., Izv. Kavkazsk. Muz. 9: 9. 1915.— Fig. 803.

Type — Russia: Elisabethpol. On dead parts of living branches of *Berberis vulgaris* (Berberidaceae). Schelkovnikov, XII 1912 (LEP, holotype).

This is a species of *Didymella*, with asci pyriform, surrounded by paraphysoids, ascospores 12-15 × 3-4 µm.

Mycosphaerella schisandrae Mitrosh., Bot. Mater. Otd. Sporov. Rast. Bot. Inst. Akad. Nauk SSSR 6: 82. 1949.

Type — Russia: *Schisandra chinensis* (Schisandraceae).

No material was studied as the location of the type is unknown.

Mycosphaerella schoenocauli Syd., Ann. Mycol. 28: 79. 1930.

Type — Venezuela: *Schoenocaulon officinale* (Alliaceae).

No material was studied as the type was not included in a loan from S.

Mycosphaerella schoenoprasi (Auersw.) J. Schröt., in Cohn, Kryptog.-Fl. Schlesien 3(2): 339. 1894 [“1893”] = *Vermicularia schoenoprasi* Auersw., in Rabenh., Fungi Europaei Exsiccati, ed. nov., ser. 2 no. 346. 1861 = *Gnomonia schoenoprasi* (Auersw.) Ces. & De Not., Comment. Soc. Crittog. Ital. 1(4): 232. 1863 = *Sphaerella schoenoprasi* (Auersw.) Auersw., in Gonn. & Rabenh., Mycol. Europaea 5-6: 19. 1869.— Fig. 804.

Type — Germany: Sachsen, Pankow. On upper and lower surface of dead leaves of *Allium schoenoprasum* (Alliaceae). Auerswald, IV 1861 (L, isotype).

Anamorph: *Colletotrichum* fide Saccardo (1882).

Already synonymised with *Davidiella allicina* (as *Mycosphaerella tassiana*) by von Arx (1949), with which the isotype studied agrees well, with asci pyriform, ascospores 16-21 × 5-7 µm.

Sphaerella schoenoprasi var. *romuleae* Sacc., Ann. Mycol. 2: 14. 1904.

Type — Italy: *Romulea bulbocodium* (Iridaceae).

No material was studied as the type was not found in any of the herbaria consulted.

Sphaerella schroeteriana Kirschst. See *Mycosphaerella comari* (J. Schröt.) Tomilin.

Sphaerella schumacheri E.C. Hansen, Vidensk. Meddel. Dansk Naturhist. Foren. Kjøbenhavn 1876: 52. 1876 = *Rosellinia schumacheri* (E.C. Hansen) Sacc., Syll. Fung. 1: 276. 1882 = *Microascus schumacheri* (E.C. Hansen) Curzi, Boll. Staz. Patol. Veg. Roma 11: 60. 1931 = *Pithoascus schumacheri* (E.C. Hansen) Arx, Proc. Kon. Ned. Akad. Wetensch. C 76: 292. 1973.

Type — Denmark: On dung.

Accepted as *Pithoascus schumacheri* (C.E. Hansen) Arx by von Arx (op. cit.) and therefore not studied.

Mycosphaerella sciadophila (Pass.) Magnus, in Dalla Torre & Sarnth., Fl. Tirol, Vorarlberg, Liechtenstein, III, Pilze: 465. 1905 = *Sphaerella sciadophila* Pass., Rev. Mycol. (Toulouse) 2: 33. 1880.

Type — Italy: *Chaerophyllum temulum* (Apiaceae).

No material was studied as the type was not found in any of the herbaria consulted.

Sphaerella sciadophila var. *chelidonii* Traverso, Fl. Ital. Cryptog. 1(11): 600. 1913.

Type — Italy: Riva-Valdobbia. On dead stems of *Chelidonium majus* (Papaveraceae). Carestia, IV 1899 (PAD, holotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores immature.

Mycosphaerella scirpi-lacustris (Auersw.) Lindau, in Engl. & Prantl, Natürlichen Pflanzenf. 1(1): 425. 1897 = *Sphaerella scirpi-lacustris* Auersw., in Gonn. & Rabenh., Mycol. Europaea 5-6: 18. 1869 = *Paradidymella scirpi-lacustris* (Auersw.) Munk, Dansk Bot. Ark 17(1): 179. 1957.— Fig. 805.

Type — Germany: *Scirpus lacustris* (Cyperaceae).

Anamorph: *Rhabdospora scirpi* (Sacc.) Allesch. fide Allesch. (1901).

Cited as probably synonymous with *Mycosphaerella lineolata* by von Arx (1949). No type material was found in B or any of the other herbaria consulted, and it should be considered lost. Material studied (British Isles, Lynn, Plowright, 1876, distributed in Sphaeriacei Britannici no. 91, B) belongs to *Davidiella*, and is morphologically indistinguishable from *D. clandestina*, with asci pyriform, ascospores 23-26 × 7-9 µm. Another collection of the same material (IMI 132627, slide only) contains a *Lophiostoma*, probably *L. amphibium* (Magnes & Hafellner) Aptroot & K.D. Hyde or *L. arundinaceum* (Sowerby) Aptroot & K.D. Hyde (Hyde *et al.* 2002), with ascomata immersed, asci cylindrical, pseudoparaphyses septate, 1-2 µm wide, ascospores 18-21 × 5-6.5 µm.

Mycosphaerella scirrroides M.E. Barr, Contr. Univ. Michigan Herb. 9: 585. 1972, nomen novum (Article 58) for *Scirrhia lineata* Dearn. & House, Circ. New York State Mus. 24: 31. 1940, lacking Latin description (not validly published, Article 36) non *Mycosphaerella lineata* Clements (1903).

Type — USA: *Carex strictior* (Cyperaceae).

No material was studied as the type nor any additional material was found in NY or BPI.

Mycosphaerella scopulorum (Sacc. & Cavara) Petr., Sydowia 7: 23. 1953 = *Sphaerella scopulorum* Sacc. & Cavara, Nuovo Giorn. Bot. Ital. 7: 281. 1900.

Type — Italy: Vallombrosa. On dead branches of *Spartium junceum* (Fabaceae). Cavara, V 1896 (PAD, holotype).

This is an unidentifiable fungus. No spores were seen in the type, not even by the original authors, according to their annotations on the label.

Mycosphaerella scorzonerae Petr., Ann. Mycol. 42: 77. 1944.— Fig. 806.

Type — Austria: Niederdonau, Baden, Pfaffstätten. On upper and lower surface of dead leaves of *Scorzonera austriaca* (Asteraceae). Petrak, V 1940 (W no. 1871, holotype), also distributed in Mycotheca Generalis no. 1333 (B, W (2×), isotypes).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradians*, with asci cylindrical, ascospores 11-13 × 2.5-3.5 µm. The type also contains a *Phaeosphaeria* species.

Mycosphaerella scrophulariae (Sacc. & Briard) Tomilin, Novosti Sist. Nizsh. Rast. 8: 152. 1971 = *Sphaerella nebulosa* var. *scrophulariae* Sacc. & Briard, in Briard, Rev. Mycol. (Toulouse) 7: 208. 1885 = *Sphaerella scrophulariae* (Sacc. & Briard) Kirschst., Kryptog.-Fl. Mark Brandenburg 7(3): 382. 1938.

Type — France: On dead stems of *Scrophularia aquatica* (Scrophulariaceae).

The type was not found in any of the herbaria consulted. Cited as synonymous with *Mycosphaerella nebulosa* by Tomilin (1979). Material studied (Germany, on dead stems of *Scrophularia nodosa*, Sydow, Mycotheca Germanica no. 1545, L) is immature.

Mycosphaerella securinegae Koshk. & Frolov, Mikoflora Podgornoi Ravniny Kopetdaga i Tsentral'nykh Karakumov: 83. 1973, lacking Latin description (not validly published, Article 36) [as “*securigineae*”].

Type — Turkmenia: *Securinega* [“*Securiginea*”] *suffruticosa* (Euphorbiaceae).

No material was studied as the location of the type is unknown.

Mycosphaerella sedi Naumov, Mater. Mikol. Fitopatol. Rossii 1: 41. 1916, nomen nudum, not validly published (Article 32).

Authentic material — Russia: Petrograd, Luga. On dead stems of *Sedum telephium* (Crassulaceae). Naumov, VI 1915 (LEP).

The authentic material studied is immature.

Mycosphaerella sedicola Petr., Hedwigia 74: 35. 1934.— Fig. 807.

Type — Russia: Siberia, Southern Altaj, Sogornaja. On dead stems of *Sedum hybridum* (Crassulaceae). Ziling no. P9, VII 1926 (W, holotype).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradians*, with asci cylindrical, ascospores 11-12 × 2.5-3.5 µm.

Mycosphaerella selene (Sacc.) Tomilin, Novosti Sist. Nizsh. Rast. 6: 122. 1970 [“1969”] = *Sphaerella selene* Sacc., Nuovo Giorn. Bot. Ital. 7: 301. 1875.

Type — Italy. On white spots with brown margins on upper surface of living leaves of *Oxalis stricta* (Oxalidaceae). Saccardo, VIII 1875 (PAD, holotype).

This is a parasitic species, and is morphologically indistinguishable from *M. depazaeformis*, with asci cylindrical, ascospores immature.

Mycosphaerella semeles Urries, Anales Inst. Bot. Cavanilles 14: 161. 1956 [“1955”, as “*semelei*”].

Type — Canary Islands: *Semele androgyna* (Asparagaceae). No material was studied as the type was not found in MA and might be lost.

Mycosphaerella senecionis (Petch) M. Morelet, Ann. Soc. Sci. Nat. Archéol. Toulon & Var 20: 106. 1968 = *Sphaerella senecionis* Petch, Ann. Roy. Bot. Gard. (Peradeniya) 7: 304. 1922.— Fig. 808.

Type — Sri Lanka: Hakgala. On dead tips and margin on upper surface of living leaves of *Senecio scandens* (Asteraceae). Petch no. 5262, IV 1917 (K, holotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. ammophilae*, with asci long pyriform, ascospores 12-14 × 3-4 µm.

Mycosphaerella sentina (Fr.) J. Schröt., in Cohn, Kryptog.-Fl. Schlesien 3(2): 334. 1894 [“1893”] = *Sphaeria sentina* Fr., Systema Mycol. 2: 520. 1823 = *Sphaerella sentina* (Fr.) Fuckel, Jahrb. Nassauischen Vereins Naturk. 23-24: 104. 1870 = *Phaeosphaerella sentina* (Fr.) Verpl., Bull. Jard. Bot. Bruxelles 10: 343. 1939.— Fig. 809.

Type — Sweden: *Malus sylvestris* [“*Pyrus malus*”] (Rosaceae).

Anamorph: *Septoria pyricola* (Desm.) Desm. *vide* Eriksson (1992).

Cited as synonymous with *Venturia pyrina* Aderh. by Oudemans (1921). The type was reported to contain no asci. Material studied (Germany, on lower surface of dead leaves of *Pyrus communis*, Fuckel, Enumeratio Fungi Nassau. no. 524, L) belongs to section *Longispora*, and is morphologically indistinguishable from *M. latebrosa*, with asci cylindrical, ascospores 27-32 × 3-3.5 µm.

Sphaerella sentina f. *mali* Thüm., Fungi Austriaci no. 465. 1872.— Fig. 810, 972.

Type — Czech Republic: Bohemia, Teplitz. On upper and lower surface of dead leaves of *Malus sylvestris* [“*Pyrus malus*”] (Rosaceae). Thümen, 1872, Fungi Austriaci no. 465, 1872 (B, BPI, isotypes).

This is morphologically indistinguishable from *Venturia inaequalis* (Cooke) G. Winter, with asci cylindrical, pseudoparaphyses septate, simple, ca. 2.5 µm wide, ascospores becoming brownish, unequally septate, 11.5-14.5 × 4-5.5 µm.

Sphaerella sentina f. *pyri-communis* Thüm., Mycotheca Universalis no. 1058. 1878.— Fig. 811.

Type — Germany: Bayern, Bayreuth. On lower surface of dead leaves of *Pyrus communis* (Rosaceae). Thümen, 1874, Mycotheca Universalis no. 1058 (L, isotype).

This belongs to section *Longispora*, and is morphologically indistinguishable from *M. latebrosa*, with asci cylindrical,

ascospores $26-33 \times 3-4 \mu\text{m}$. On the upper surface of the same isotype *M. punctiformis* is present.

Sphaerella sepincolaeformis De Not., Comment. Soc. Crittog. Ital. 2(3): 486. 1867 [as “*saepincolaeformis*”] = *Didymella sepincolaeformis* (De Not.) Sacc., Syll. Fung. 1: 551. 1882 = *Apiosporella sepincolaeformis* (De Not.) Theiss., Ann. Mycol. 15: 275. 1917 = *Pseudomassaria sepincolaeformis* (De Not.) Arx, Ber. Schweiz. Bot. Ges. 62: 350. 1952.— Fig. 812.

Type — Italy: Carestia. On dead branches of *Rosa alpina* (Rosaceae). De Notaris (B, isotype).

Accepted as *Pseudomassaria sepincolaeformis* (De Not.) Arx by Müller & von Arx (1962), with which the isotype agrees well with ascospores septate near the tip (apiosporous), $15-17 \times 6-8 \mu\text{m}$.

Mycosphaerella septorioides (Desm.) Lindau, Hilfsb. Sammeln Ascomyceten: 3. 1903 = *Sphaeria septorioides* Desm., Ann. Sci. Nat. Bot., ser. 3, 6: 81. 1846 = *Sphaerella septorioides* (Desm.) Niessl, in J. Kunze, Fungi Selecti Exsiccati no. 242. 1879 = *Carlina septorioides* (Desm.) Höhn., Hedwigia 62: 73. 1920.— Fig. 813.

Type — France: On upper and lower surface of dead leaves of *Acer campestre* (Sapindaceae). Desmazières, Plantes Cryptogames de France no. 1797 (NY, isotype).

Anamorphs: *Cylindrosporium aceris* Jacz., *Phyllosticta* and *Septoria fide* Tomilin (1979).

The isotype contains only a coelomycete; according to the exsiccatum label, no ascospores were seen by Desmazières either. Additional material studied (Germany, Prov. Hessen-Nassau, Donsbachtale, Ludwig, distributed in Sydow, Mycotheca Germanica no. 3510, V 1941, L) is *M. punctiformis*, with asci cylindrical, ascospores $8-10 \times 2-2.5 \mu\text{m}$.

Sphaerella septorioides Peck. See *Sphaerella thalicticola* Sacc. & P. Syd.

Sphaerella septorioides f. *aceris-campestris* J. Kunze, Fungi Selecti Exsiccati no. 242.— Fig. 814.

Type — Germany: Sachsen, Eisleben, Wolferode. On upper and lower surface of dead leaves of *Acer campestre* (Sapindaceae). Kunze, Fungi Selecti Exsiccati no. 242, V 1879 (L, holotype).

This is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores $8-10 \times 2-2.5 \mu\text{m}$.

Mycosphaerella septorispora (Sacc.) Petr., Hedwigia 62: 286. 1921 = *Sphaerella septorispora* Sacc., Ann. Mycol. 12: 287. 1914.— Fig. 815.

Type — Czech Republic: Weißkirchen, Mariental. On upper and lower surface of dead leaves of *Quercus robur* (Fagaceae). Petrak, Flora Bohemiae et Moraviae Exsiccata no. 1026, V 1914 (L, 2 isotypes), also distributed in Reliquiae Petrakianae no. 71 (L, isotype).

Cited as synonymous with *Sphaerulina myriadea* (DC. : Fr.) Sacc. by Barr (1972), with which the isotype studied agrees well, with ascospores 3-septate, $25-35 \times 2-2.5 \mu\text{m}$.

Mycosphaerella sequoiae Bonar, Mycologia 34: 184. 1942.— Fig. 816.

Type — USA: Humboldt County, Trinidad. On lower surface of dead needles of *Sequoia sempervirens* (Cupressaceae). Parks, California Fungi no. 651, 1932 (H, isotype).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. superflua*, with asci cylindrical, ascospores $15-18 \times 3.5-4.5 \mu\text{m}$.

Sphaerella serograptia (Durieu & Mont.) Auersw., in Gonn. & Rabenh., Mycol. Europaea 5-6: 12. 1869 = *Sphaeria serograptia* Durieu & Mont., Fl. Algérie. Cryptog. 1: 537. 1849 = *Sphaerulina serograptia* (Durieu & Mont.) Sacc., Syll. Fung. 2: 187. 1883.

Type — Algeria: *Quercus coccifera* (Fagaceae).

No material was studied as the type was not included in a loan from PC. Cited as synonymous with *Sphaerulina myriadea* (DC. : Fr.) Rabenh. by Barr (1972).

Mycosphaerella serpylli (Pass.) Tomilin, Oprelidelitel' gribov roda *Mycosphaerella* Johans.: 146. 1979 = *Sphaerella serpylli* Pass., Atti Reale Accad. Lincei, Mem. Cl. Sci. Fis., ser. 4, 3: 5. 1887.

Type — Italy: *Thymus serpyllum* (Lamiaceae).

No material was studied as the type was not found in any of the herbaria consulted.

Mycosphaerella serratulae (Hollós) Tomilin, Oprelidelitel' gribov roda *Mycosphaerella* Johans.: 62. 1979 = *Sphaerella serratulae* Hollós, Ann. Hist.-Nat. Mus. Natl. Hung. 6: 528. 1908.

Type — Hungary: *Serratula tinctoria* (Asteraceae).

The type is not in BP and may have been destroyed during the war.

Mycosphaerella serrulata (Ellis & Everh.) Diehl ex A. Weiss, Pl. Dis. Reporter 26: 296. 1942 = *Sphaerella serrulata* Ellis & Everh., J. Mycol. 3: 45. 1887 = *Laestadia serrulata* (Ellis & Everh.) Sacc., Syll. Fung. 9: 586. 1891 [as “*serrulatae*”].— Fig. 817.

Type — USA: Florida, Lake Co., Eustis. On upper surface of dead leaves of *Sabal serrulata* (Aracaceae). Nash, V-VI 1895, distributed in Plants of Florida no. 1952 (BPI, 2 topotypes).

The holotype was out on loan; the topotypes show that this belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradians*, with asci cylindrical, ascospores $12-13.5 \times 2.5-3.5 \mu\text{m}$, although most ascospores are immature and much smaller.

Mycosphaerella sesami Sivan., Trans. Brit. Mycol. Soc. 85: 397. 1985.— Fig. 818.

Type — Zimbabwe: Kantongo Agricultural Station. On white spots with black margins on upper surface of living leaves of *Sesamum* (Pedaliaceae). Angus no. M1050, III 1961 (IMI no. 89776, holotype).

Anamorph: *Cercospora sesami* Zimm. *fide* Sivanesan (op. cit.) (= *Cercospora apii* s. lat.).

This is a parasitic species, with asci cylindrical, ascospores $14-17 \times 4-5.5 \mu\text{m}$.

Mycosphaerella sesamicola Sivan., Trans. Brit. Mycol. Soc. 85: 397. 1985.— Fig. 819.

Type — Zimbabwe: Kasama, Misambu Experimental Station. On white spots with black margins on upper surface of living leaves of *Sesamum angolense* (Pedaliaceae). Angus no. M758, III 1960 (IMI no. 85701, holotype).

Anamorph: *Cercoseptoria sesami* (Hansf.) Deighton *vide* Sivanesan (op. cit.) (= *Pseudocercospora sesami* (Hansf.) Deighton).

This is a parasitic species, with asci cylindrical, ascospores 11-17 × 3.5-5 µm.

Sphaerella sesbaniae Ellis & Everh., J. Mycol. 4: 98. 1888.— Fig. 820.

Type — USA: Louisiana, St. Martinsville. On dead stems of *Sesbania macrocarpa* (Fabaceae). Langlois no. 1403, VI 1888 (NY, holotype, sub *Botryosphaeria minor*, of which species this specimen is also the holotype).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradians*, with asci cylindrical, ascospores 10-12 × 3.5-4 µm, although most material belongs to coelomycetes.

Mycosphaerella seseli Gucevicz, Bot. Mater. Otd. Sporov. Rast. Bot. Inst. Komarova Akad. Nauk SSSR 13: 190. 1960.

Type — Russia: *Seseli* (Apiaceae).

No material was studied as the type was not included in loans from LE or LEP. Cited as synonymous with *Davidiella allicina* (as *Mycosphaerella allicina*) by Tomilin (1979).

Mycosphaerella setosa Tomilin, Novosti Sist. Nizsh. Rast. 6: 123. 1970 [“1969”].

Type — Russia: *Empetrum nigrum* (Ericaceae).

No material was studied as the type was not included in loans from LE or LEP.

Mycosphaerella shawii Arx & J.L. Bezerra, in Arx, Proc. Kon. Ned. Akad. Wetensch. C 66: 177. 1963.

Type — USA: *Arctostaphylos uva-ursi* var. *coactilis* (Ericaceae).

No material was studied as the type was not found in CBS nor included in a loan from URM.

Mycosphaerella shibataeae I. Miyake & Hara, in Hara, Spec. Dendropath.: 103. 1923 = *Mycosphaerella*

phyllostachydicola Tomilin, Novosti Sist. Nizsh. Rast. 1967: 190. 1967, superfluous nomen novum (Article 58) for *Sphaerella bambusae* (I. Miyake & Hara) Sacc. ex Trotter, Syll. Fung. 24: 867. 1928, later homonym (illegitimate, Article 53) = *Guignardia bambusae* I. Miyake & Hara, Bot. Mag. (Tokyo) 24: 338. 1910 = *Pseudomassaria bambusae* (I. Miyake & Hara) Hino & Katum., J. Jap. Bot. 40: 83. 1965.

Type — Japan: Honshu, Musashi, Komana. On dead parts (e.g. tips) of living leaves of *Shibataea kumasaca* (Poaceae). Hara, V 1909 (TNS 209270, holotype).

Cited as synonymous with *Mycosphaerella malinverniana* by Tomilin (1979).

In the type and additional material seen (Hara, V 1905, TNS 209960, 209961 & 209962) only coelomycetes and immature ascomycetes found.

Mycosphaerella shikaeana Hara, Trans. Mycol. Soc. Japan 2(4): 16. 1960.

Type — Japan: *Damnacanthus angustifolius* (Rubiaceae).

No material was studied as the type was not included in a loan from TNS.

Mycosphaerella shimadae Sawada, Taiwan Univ. Coll. Agr. Special Publ. 8: 64. 1959.

Type — Taiwan: *Oryza sativa* (Poaceae).

No material was studied as the type was not included in a loan from TNS.

Mycosphaerella shiraiana (I. Miyake) Sawada, Taiwan Univ. Coll. Agr. Special Publ. 8: 64. 1959 = *Sphaerella shiraiana* I. Miyake, J. Coll. Agric. Imp. Univ. Tokyo 2: 243. 1910.

Type — Japan: *Oryza sativa* (Poaceae).

No material was studied as the type was not included in a loan from TNS. Cited as synonymous with *Mycosphaerella oryzae* by Tomilin (1979).

Mycosphaerella shomae Hara, J. Pl. Protect. 5: 618. 1918.

Type — Japan.

No material was studied as the type was not included in a loan from TNS.

Mycosphaerella sibirica (Thüm.) Johanson, Öfvers. Förh. Kongl. Svenska Vetensk.-Akad. 41(9): 165. 1884 = *Sphaerella sibirica* Thüm., Bull. Soc. Imp. Naturalistes Moscou 52: 198. 1880.— Fig. 821.

Type — Russia: Siberia. On upper and lower surface of dead leaves and on dead stems of *Dianthus versicolor* (Caryophyllaceae).

The type was not found in any of the herbaria consulted. Synonymised with *Davidiella allicina* (as *Mycosphaerella tassiana*) by von Arx (1949). Material seen (Iceland, Hof, on *Viscaria alpina*, Davisson, IV 1901, BPI) agrees, with asci pyriform, ascospores 19-22 × 5.5-7 µm.

Mycosphaerella sicula (Penz.) Tomilin, Novosti Sist. Nizsh. Rast. 6: 122. 1970 [“1969”] = *Sphaerella sicula* Penz., Michelia 2: 413. 1882.— Fig. 822.

Type — Italy: Sicily, Carini. On upper and lower surface of dead leaves of *Citrus limonum* [“limon”] (Rutaceae). Beltrani, 1878 (BPI, isotype).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradians*, with asci cylindrical, ascospores 10-12 × 3-3.5 µm.

Mycosphaerella sicyicola (Ellis & Everh.) Tomilin, Novosti Sist. Nizsh. Rast. 8: 152. 1971 = *Sphaerella sicyicola* Ellis & Everh., J. Mycol. 3: 45. 1887.

Type — USA: Missouri, Columbia. On living leaves of *Sicyos angulata* (Cucurbitaceae). Galloway no. 51, VII 1886 (NY, holotype), also VIII 1886 (NY, topotype).

Cited by Tomilin (1979) as morphologically indistinguishable from *Didymella bryoniae* (Auersw.)

Rehm, with which the types agree, albeit that they are in poor shape.

Mycosphaerella sidaecola (Ellis & Everh.) Tomilin, Novosti Sist. Nizsh. Rast. 7: 205. 1970 = *Sphaerella sidaecola* Ellis & Everh., Erythea 1: 198. 1893.— Fig. 823.

Type — USA: California, Pasadena. On nearly unchanged spots on upper and lower surface of living leaves of *Sidalcea* ["*Sida*"] *delphinifolia* (Malvaceae). McClatchie no. 191, III 1893 (NY, holotype).

This is morphologically indistinguishable from *Coleroa robertiana*, with asci cylindrical, ascospores greenish, apiosporous, 11-14 × 3-4 µm.

Mycosphaerella sieberiana Sivan., Trans. Brit. Mycol. Soc. 72: 157. 1979.— Fig. 824.

Type — Sierra Leone: Njali. On white spots with black margins on upper surface of living leaves of *Cassia sieberiana* (Fabaceae). Deighton, X 1933 (IMI no. 8182, holotype).

Anamorph: *Pseudocercospora fide* Sivanesan (op. cit.).

This is a parasitic species, with asci cylindrical, ascospores 17-21 × 4.5-5 µm.

Mycosphaerella silenes Höhn., Ann. Mycol. 1: 523. 1903 = *Sphaerella silenes* (Höhn.) Sacc. & D. Sacc., Syll. Fung. 17: 639. 1905.— Fig. 825.

Type — Austria: Tirol, Ötztal. On dead calyces of *Silene inflata* (Caryophyllaceae). Höhnelt, VII 1903 (FH-Höhnelt, holotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. ammophilae*, with asci pyriform, ascospores 11-13 × 3-3.5 µm. A *Septoria* is present on white spots on the upper and lower surface of the living leaves. Additional materials seen (Schneeberg, VIII 1905, FH-Höhnelt; also Kirchhof, III 1870, B) agree.

Mycosphaerella silenes-acaulis (Maire) Lind, Svensk Bot. Tidskr. 22: 64. 1928 = *Sphaerella silenes-acaulis* Maire, Oesterr. Bot. Z. 57: 335. 1907.— Fig. 826.

Type — Austria: On upper and lower surface of dead leaves of *Silene acaulis* (Caryophyllaceae).

Material studied (Austria, Nordliche Kalkalpen, Rax, Petrak, distributed in Reliquiae Petrakianae no. 2255, VII 1940, L) belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradicans*, with asci cylindrical, ascospores 10-13 × 2-3 µm.

Mycosphaerella silenicola Woron. See *Mycosphaerella woronichinii* G. Woron.

Sphaerella silenicola Hollós, Ann. Hist.-Nat. Mus. Natl. Hung. 7: 51. 1909.

Type — Hungary: *Silene otites* (Caryophyllaceae).

The type is not in BP and may have been destroyed during the war.

Mycosphaerella silveirae (Speg.) Cash, Syll. Fung. 26: 353. 1972 = *Sphaerella silveirae* Speg., Anales Soc. Ci. Argent. 93: 114. 1922.

Type — Brazil: *Acacia pedicellata* (Fabaceae).

No material was studied as the type was not included in a loan from LPS.

Sphaerella simulans Cooke, J. Bot. 4: 246. 1866.— Fig. 827.

Type — United Kingdom. On lower surface of dead leaves of *Quercus* (Fagaceae). Cooke (K, holotype; K, isotype).

Already cited as synonymous with *M. maculiformis*, which is morphologically indistinguishable from *M. punctiformis*, by Tomilin (1979), with which the types studied agree well, with ascomata in groups, asci cylindrical and ascospores 7-9 × 2-3 µm.

Sphaerella simulans Berl. & Bres. See *Mycosphaerella berlesiana* (Traverso) Tomilin.

Mycosphaerella singularis (Henn.) Arx, in E. Müll. & Arx, Beitr. Kryptogamenfl. Schweiz 11(2): 357. 1962 = *Lizonia singularis* Henn., Hedwigia 42: 80. 1903 = *Lizoniella singularis* (Henn.) Sacc., Syll. Fung. 17: 662. 1905 = *Haplodothis singularis* (Henn.) Höhn., Sitzungsber. Kaiserl. Akad. Wiss., Math.-Naturwiss. Kl., Abt. 1, 120: 423. 1911.

Type — Australia: *Leucopogon hispidus* (Ericaceae).

No material was studied as the species was already redisposed.

Mycosphaerella sisyrinchiicola (Speg.) Cash, Syll. Fung. 26: 353. 1972 = *Sphaerella sisyrinchiicola* Speg., Bol. Acad. Nac. Ci. 27: 363. 1924.

Type — Argentina: *Sisyrinchium iridifolia* (Iridaceae).

No material was studied as the type was not included in a loan from LPS.

Mycosphaerella skimmiae E. Müll. & S. Ahmad, Biologia (Lahore) 3: 7. 1957.— Fig. 828.

Type — Pakistan: Changla Gali. On upper and lower surface of dead leaves of *Skimmia laureola* (Rutaceae). Ahmad no. 12000, VIII 1954 (ZT, holotype; IMI no. 82002, isotypes), also Ahmad no. 11099 (ZT, isotype), also Ahmad no. 15086 (IMI no. 89505, isotype).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradicans*, with asci cylindrical, ascospores 11-14 × 2.5-3.5 µm. There is no hesitation to assign the specimens with different collection numbers as isotypes, as the remaining collecting details are identical.

Mycosphaerella slaptoniensis D. Hawksw. & Sivan., Trans. Brit. Mycol. Soc. 64: 108. 1975.— Fig. 829.

Type — United Kingdom: Devon, Slapton Ley Nature Reserve. On young but dead twigs of *Crataegus oxyacanthus* (Rosaceae). Hawksworth no. 3411, VII 1973 (IMI no. 178341, holotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. ammophilae*, with ascomata protruding through the bark epidermis, wall only darkened above, asci pyriform, ascospores 10-11 × 2-2.5 µm.

Mycosphaerella smegmatos (Pass.) Johanson ex Oudem., Rév. Champ. Pays-Bas 2: 210. 1897 = *Sphaerella smegmatos* Pass., Nuovo Giorn. Bot. Ital. 7: 256. 1875.

Type — Italy: On upper and lower surface of dead leaves of *Saponaria officinalis* (Caryophyllaceae).

Anamorph: Associated with *Colletotrichum fide* Saccardo (1882).

The type was not found in any of the herbaria consulted. Material studied (Netherlands, Scheveningen, Destrée, X 1888, L) contains *Pleospora herbarum* (Pers. : Fr.) Rabenh.

Mycosphaerella smilacicola (Cooke) Overholts, Mycologia 17: 111. 1925 = *Sphaerella smilacicola* Cooke [as “Schwein.”], Grevillea 6: 146. 1878, nomen novum (Article 58) for *Sphaeria smilacicola* Schwein., Trans. Amer. Philos. Soc., ser. 2, 4: 226. 1832, later homonym of *Sphaeria smilacicola* Schwein. (op. cit.: 196).— Fig. 830.

Type — USA: Philadelphia. On leaves of *Smilax rotundifolia* (Smilacaceae). Schweinitz (BPI, K, isotypes).

The holotype is not preserved in PH. The isotypes studied are sterile. Therefore the following epitype has been selected:

Epitype — USA: South Carolina, Aiken. On brown spots with black margins on upper surface of living leaves of *Smilax* (Smilacaceae). Ravenel, Fungi Americani Exsiccati no. 95 (NY, 2 isoeotypes).

The epitype shows that this is a parasitic species, with asci cylindrical, ascospores 17-20 × 4-5.5 µm. Most other material studied, including Ravenel, Fungi Americani Exsiccati no. 155 (NY, 2×) contains only coelomycetes.

Mycosphaerella smilacifolii Bat. & Peres, in Bat., Peres, Cavalcanti & Heringer, Atas Inst. Micol. 3: 227. 1966.— Fig. 831.

Type — Brazil. On white spots on upper and lower surface of dead leaves of *Smilax* (Smilacaceae). Batista exs. no. 19414 (URM 35945, holotype).

This is morphologically indistinguishable from *Didymella smilacina* Bat. & Peres (of which this specimen is also the type), with paraphyses simple, slightly clavate, up to 2 µm wide, asci clavate and ascospores 14-19 × 3-4.5 µm.

Mycosphaerella smilacina (Ellis & Everh.) Tomilin, Novosti Sist. Nizsh. Rast. 1967: 190. 1967 = *Sphaerella smilacina* Ellis & Everh., J. Mycol. 2: 101. 1886.— Fig. 832.

Type — USA: New Jersey, Newfield. On dead stems of *Smilax* (Smilacaceae). Ellis, V 1886 (NY, holotype; NY, isotype).

Anamorph: associated with *Diplodina smilacis* Ellis & Everh. *fide* Ellis & Everhart (op. cit.).

This is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical to clavate, ascospores 8-10 × 2-2.5 µm.

Sphaerella smilacis Speg., Anales Mus. Nac. Hist. Nat. Buenos Aires 19: 355. 1909.

Type — Argentina: *Smilax campestris* (Smilacaceae).

No material was studied as the type was not included in a loan from LPS.

Mycosphaerella smilacis-glabrae Sawada, Taiwan Univ. Coll. Agric. Special Publ. 8: 64. 1959 [as “*smilacis-glabra*”].

Type — Taiwan: *Smilax glabra* (Smilacaceae).

No material was studied as the type was not found in BPI.

Mycosphaerella sodiroana Petr., Sydowia 4: 504. 1950.— Fig. 833.

Type — Ecuador: Tungurahua, Baños. On upper and lower surface of dead leaves of *Sapium sodiroi* (Euphorbiaceae). Sydow, distributed in Reliquae Petrakianae no. 659, XII 1937 (H, L, isotypes).

This is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores 8-10 × 2-2.5 µm.

Mycosphaerella sojae Hori, in Yamamoto, J. Pl. Protect. 12: 97. 1925.

Type — Japan: *Glycine max* (Fabaceae).

No material was studied as the type was not included in a loan from TNS.

Mycosphaerella solani (Ellis & Everh.) Wollenw., Phytopathology 3: 229. 1913 = *Sphaerella solani* Ellis & Everh., Proc. Acad. Nat. Sci. Philadelphia 45: 134. 1893 = *Didymella solani* (Ellis & Everh.) W. Gams & Gerlagh, Persoonia 5: 177. 1968.— Fig. 834.

Type — USA: Ohio. On small white spots with ochraceous margins on upper surface of living leaves of *Solanum dulcamara* (Solanaceae). Kellerman no. 412, VII 1883 (NY, lectotype, designated here).

This is accepted as *Didymella solani* (Ellis & Everh.) W. Gams & Gerlagh, with which the lectotype agrees well, with asci clavate, pseudoparaphyses present, ascospores 11-14 × 3-3.5 µm. Gams & Gerlagh (op. cit.) also studied a specimen in NY, but did not indicate the collection details, and as several specimens were mentioned in the protologue, it may well have been a different (but identical) specimen.

Sphaerella solidaginea Ellis & Kellerm., J. Mycol. 3: 127. 1887.— Fig. 835.

Type — USA: Kansas, Manhattan. On upper and lower surface of dead leaves of *Solidago canadensis* (Asteraceae). Kellerman no. 1115, X 1887 (NY, holotype).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. superflua*, with asci cylindrical, ascospores 14-17 × 3.5-4.5 µm.

Sphaerella solidaginis (Fr.) P. Karst., Bidrag Kännedom Finlands Natur Folk 23: 180. 1873 = *Xyloma solidaginis* Fr., Observ. Mycol. Fl. Suecicam 1: 199. 1815. = *Sphaeria solidaginis* (Fr.) Fr., Kungl. Svenska Vetenskapsakad. Handl. 1816: 152. 1816 = *Dothidea solidaginis* (Fr.) Fr., Systema Mycol. 2: 562. 1823 = *Ascospora solidaginis* (Fr.) Chevall. ex Fr., Summa Veg. Scand.: 425. 1849 = *Carlia solidaginis* (Fr.) Höhn., Centralbl. Bakteriolog., 2. Abth., 60: 25. 1923.

Type — Sweden: *Solidago virgaurea* (Asteraceae).

No material was studied. It was synonymised with *Asteroma solidaginis* Chevallier by Oudemans (1923).

Mycosphaerella solidaginis Jacz., Opredelitel' gribov 2: 617. 1917, nomen novum (Article 58) for *Sphaerella*

solidaginis Jacz., Bull. Soc. Imp. Naturalistes Moscou, ser. 2, 9: 135. 1895, later homonym (illegitimate, Article 53).

Type — Russia: *Solidago* (Asteraceae).

No material was studied as the type was not included in loans from LE or LEP.

Mycosphaerella sophorae (G. Winter) Tomilin, Opređelitel' gribov roda *Mycosphaerella* Johans.: 123. 1979 = *Sphaerella sophorae* G. Winter, Bol. Soc. Brot. 2: 40. 1884.

Type — Portugal: *Sophora* (Fabaceae).

No material was studied as the type was not found in B and might be lost.

Mycosphaerella sordidula (Speg.) Tomilin, Novosti Sist. Nizsh. Rast. 1968: 167. 1968 = *Sphaerella sordidula* Speg., Anales Soc. Ci. Argent. 18: 280. 1884.

Type — Brazil: Guarapi. On dead, but still attached, fruit pods of *Cassia* (Fabaceae). Balansa, VII 1883, distributed in Roumeguère, Fungi Selecti Exsiccati no. 4136 (NY, isotype).

The isotype contains no identifiable fungus.

Sphaerella sparsa (Wallr.) Auersw., Mycol. Europaea 5-6: 4. 1869 = *Sphaeria sparsa* Wallr., Fl. Cryptog. Germ. 4: 772. 1833.— Fig. 836.

Type — Germany: On upper and lower surface of dead leaves of *Tilia parvifolia* ["*microphylla*"] (Fagaceae).

The type was not found in any of the herbaria consulted. Cited as synonymous with *M. punctiformis* by Tomilin (1979), which, although an older epithet, was mentioned as morphologically indistinguishable from *M. sparsa* by Auerswald (op. cit.), with which material studied (Fuckel, Enum. Fungi Nassau. no. 521, L) agrees well, with asci cylindrical, ascospores 8-10 × 2-2.5 µm.

Sphaerella sparsa f. *aesculi* Thüm., Fungi Austriaci no. 466. 1872.— Fig. 837, 983.

Type — Czech Republic: Bohemia, Teplitz. On upper and lower surface of dead leaves of *Aesculus hippocastanum* (Sapindaceae). Thümen, 1872, distributed in Fungi Austriaci no. 466 (B, BPI, isotypes).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores thick-walled, 14.5-19 × 4-6 µm.

Sphaerella sparsa f. *quercus* Thüm., Fungi Austriaci no. 467. 1872.— Fig. 838.

Type — Czech Republic: Bohemia, Teplitz. On upper and lower surface of dead leaves of *Quercus pedunculata* (Fagaceae). Thümen, 1872, distributed in Fungi Austriaci no. 467 (B, BPI, isotypes).

The isotype in B shows that this is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores 8-10 × 2.5-3 µm. Various other, unidentifiable, ascomycetes were found in the isotype in BPI.

Sphaerella sparsa f. *tiliae* Thüm., Fungi Austriaci no. 245. 1872.— Fig. 839.

Type — Austria: Niederösterreich, Krems. On lower surface of dead leaves of *Tilia* (Malvaceae). Thümen, 1871, distributed in Fungi Austriaci no. 245 (B, isotype).

This is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores 7-9 × 2.5-3 µm.

Sphaerella sparsa f. *tiliae-parvifoliae* Thüm., Mycotheca Universalis no. 348. 1876.— Fig. 840.

Type — Germany: Sachsen, Leipzig. On upper and lower surface of dead leaves of *Tilia parvifolia* (Malvaceae). Winter, 1874, distributed in Thümen, Mycotheca Universalis no. 348 (L, isotype).

This is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores 8-10 × 2-2.5 µm.

Sphaerella sparsa subsp. *corylina* P. Karst. See *Mycosphaerella corylina* (P. Karst.) Tomilin.

Mycosphaerella spartinae (Ellis & Everh.) Tomilin, Novosti Sist. Nizsh. Rast. 1967: 190. 1967 = *Sphaerella spartinae* Ellis & Everh., J. Mycol. 4: 97. 1888 = *Phaeosphaerella spartinae* (Ellis & Everh.) Tomilin, Opređelitel' gribov roda *Mycosphaerella* Johans.: 285. 1979.

Type — USA: Nebraska, Lincoln. On dead leaves of *Spartina cynosuroides* (Poaceae). Webber, X 1887 (BPI, isotype).

The holotype could not be found in NY. The isotype in BPI shows that this is the coelomycete *Ascochyta*, with conidia 1-septate, in accordance with the protologue, where no asci are mentioned.

Mycosphaerella spegazzinii Tomilin, Opređelitel' gribov roda *Mycosphaerella* Johans.: 220. 1979, superfluous (illegitimate, Article 52) nomen novum (Article 58) for *Sphaerella peckii* Speg., in Thümen, Mycotheca Universalis no. 1356. 1879 = *Metasphaeria peckii* (Speg.) Sacc., Syll. Fung. 2: 172. 1883.

Type — USA: New York, Albany. On upper and lower surface of dead leaves of *Amelanchier canadensis* (Rosaceae). Peck, III 1878, distributed in Thümen, Mycotheca Universalis no. 1356 (L, NY, isotypes, sub "*Sphaerella peckii*").

This is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores 8-10 × 2-2.5 µm.

Mycosphaerella spetsbergensis L. Holm & K. Holm, Symb. Bot. Upsal. 31(3): 288. 1996.

Type — Svalbard: *Salix polaris* (Salicaceae). Praestö, VIII 1994 (UPS, holotype, not seen).

No material was studied as the type of this recently described species was not included in a loan from UPS.

Mycosphaerella sphaerelloides (Sacc.) Petr., in Syd. & Petr., Ann. Mycol. 22: 391. 1924 = *Didymella sphaerelloides* Sacc., Nuovo Giorn. Bot. Ital., n. ser., 27: 76. 1920.

Type — USA: *Yucca glauca* (Asparagaceae).

No material was studied as the type was not found in any of the herbaria consulted. Synonymised with *Davidiella allicina* (as *Mycosphaerella tassiana*) by von Arx (1949).

Mycosphaerella sphaerellula (Peck) M.E. Barr, Contr. Univ. Michigan Herb. 9: 604. 1972 = *Sphaeria sphaerellula* Peck, Annual Rep. New York State Mus. 30: 66. 1878 = *Didymella sphaerellula* (Peck) Sacc., Syll. Fung. 1: 547. 1882.

Type — USA: *Acer pensylvanicum* (Sapindaceae).

No material was studied as the type was not found in NY or BPI.

Mycosphaerella sphaerosperma Herter, Florula Uruguayensis Plantae Avasculares 3: 35. 1933, nomen novum (Article 58) for *Sphaerella sphaerosperma* Rostr., nomen herbariorum (not validly published, Article 32).

Type — Uruguay: *Beta vulgaris* (Amaranthaceae).

No material was studied as the type was not found in C and might be lost.

Mycosphaerella sphaerulinae Crous & M.J. Wingf., Sydowia 9: 144. 2003.

Type — Chile. On brown spots on upper surface of living leaves of *Eucalyptus nitens* (Myrtaceae). Wingfield, V 2001 (CBS no. 6597, holotype).

Anamorph: *Pseudocercospora sphaerulinae* Crous & M.J. Wingf. (op. cit.).

This recently described species was not studied further. It deviates from all other species of *Mycosphaerella* by its 3-septate ascospores.

Mycosphaerella spigeliae Medeiros & Nascim., Anais Soc. Bot. Brasil 1964: 376. 1964.

Type — Brazil: *Spigelia anthelmia* (Loganiaceae).

No material was studied as the type was not included in a loan from URM.

Mycosphaerella spilota Syd., in Syd. & P. Syd., Ann. Mycol. 37: 211. 1939.— Fig. 841.

Type — Sierra Leone: York. On brown spots with black margins on upper surface of living leaves of *Andropogon tectorum* (Poaceae). Deighton no. 1338, XII 1936 (IMI no. 16805, isotype).

This belongs to *Davidiella*, of which it represents a parasitic species, with asci pyriform, ascospores 17-21 × 5-7 µm. Therefore the following new combination is made:

Davidiella spilota (Syd.) Aptroot comb. nov., **MB 500363**.

Basionym: *Mycosphaerella spilota* Syd., in Syd. & P. Syd., Ann. Mycol. 37: 211. 1939. Although the protologue mentions rather numerous “paraphyses”, no hamathecial filaments were observed in the material.

Mycosphaerella spinarum (Auersw.) Migula, in Thomé, Fl. Deutschl., Österr. Schweiz. X, 1. Kryptog.-Fl., III, 3(1): 285. 1912 [“1913”] = *Sphaerella spinarum* Auersw., in Gonn. & Rabenh., Mycol Europaea 5-6: 9. 1869.— Fig. 842.

Type — Germany: *Astragalus aristatus* (Fabaceae).

Material studied (Afghanistan, Nuristan, Kamdesch, on upper and lower surface of dead leaves of *Astragalus*

paghmanensis, Gilli, distributed in Reliquiae Petrakianae no. 660, IV 1950, L) belongs to section *Caterva*, and is morphologically indistinguishable from *M. superflua*, with asci cylindrical, ascospores 13-17 × 3-4 µm.

Mycosphaerella spinicola (Pass.) Tomilin, Novosti Sist. Nizsh. Rast. 6: 122. 1970 [“1969”] = *Sphaerella spinicola* Pass., Atti Reale Accad. Lincei, Mem. Cl. Sci. Fis., ser. 4, 3: 6. 1887.

Type — Italy: *Hippophaë rhamnoides* (Elaeagnaceae).

No material was studied as the type was not found in any of the herbaria consulted.

Sphaerella spinicola Ellis & Everh., Proc. Acad. Nat. Sci. Philadelphia 42: 231. 1891 [“1890”], later homonym (illegitimate, Article 53) = *Laestadia spinicola* Sacc., Syll. Fung. 9: 582. 1891, nomen novum (Article 58) = *Guignardia spinicola* (Sacc.) Lindau, in Engl. & Prantl, Natürlichen Pflanzenf. 1(1): 423. 1897.— Fig. 843.

Type — USA: Pennsylvania, West Chester. On dead spines of *Rosa rubiginosa* (Rosaceae). Everhart, VII 1889 (NY, holotype), also distributed in Ellis & Everhart, North American Fungi no. 2537, VII 1890 (L, isotype).

This is morphologically indistinguishable from *Glomerella cingulata* (Stoneman) Spauld. & H. Schrenk, with asci surrounded by simple paraphyses, ascospores not septate, 12-15 × 6-7 µm, for which it would be an older epithet.

Mycosphaerella spiraeae Woron., Trudy Bot. Muz. 21: 122. 1927.

Type — Georgia: *Spiraea hypericifolia* (Rosaceae).

No material was studied as the type was not included in loans from LE or LEP.

Sphaerella spiraeae Westend. ex Roum., Rev. Mycol. (Toulouse) 12: 21. 1890, nomen nudum (not validly published, Article 32).

Authentic material — Belgium: Ardennes, Louette Saint Pierre. On lower surface of dead leaves of *Filipendula* [“*Spiraea*”] *ulmaria* (Rosaceae). Aubert, distributed in Roumeguère, Fungi Selecti Exsiccati no. 5150 (NY).

This is a specimen of *M. punctiformis*, although the material is in bad shape.

Mycosphaerella spissa Syd., Ann. Mycol. 22: 296. 1924.

Type — New Zealand: *Coprosma robusta* (Rubiaceae).

No material was studied as the type was not included in a loan from S.

Mycosphaerella spleniata (Cooke & Peck) House, New York State Mus. Bull. 233-234: 30. 1921 = *Sphaerella spleniata* Cooke & Peck, Annual Rep. New York State Mus. 25: 105. 1873.— Fig. 844.

Type — USA: New York, Albany, Greenbush. On lower surface of dead leaves of *Quercus bicolor* (Fagaceae). Peck no. 29 (K, holotype; NY, isotype).

Additional material studied: USA: Kansas, Rockport. On lower surface of dead leaves of *Quercus macrocarpa*, Bartholomew, distributed in Ellis & Everhart, North American Fungi no. 3110, VII 1894 (L). All material studied belongs to section *Caterva*, and shown that this is

morphologically indistinguishable from *M. superflua*, with asci cylindrical, ascospores 13-15(-18) × 2.5-4 µm.

Mycosphaerella spraguei Tomilin. See *Mycosphaerella zizaniae* (Schwein.) Lindau.

Mycosphaerella staphyleae Miura, Industr. Contr. 27: 165. 1928.

Type — China: *Staphylea bumalda* (Staphyleaceae).

No material was studied as the location of the type is unknown.

Mycosphaerella staphylina (Ellis & Everh.) J.H. Miller, Mycologia 33: 80. 1941 = *Sphaerella staphylina* Ellis & Everh., J. Mycol. 3: 128. 1887.

Type — USA: *Staphylea trifolia* (Staphyleaceae).

Anamorph: Associated with *Alternaria* ["*Macrosporium*"] *vide* Saccardo (1891).

No material was studied as the type was not found in BPI or NY and is probably lost.

Sphaerella starbackii Sacc. & Traverso, Syll. Fung. 20: 828. 1911, nomen novum (Article 58) for *Mycosphaerella perexigua* Starbäck, Ark. Bot. 5: 21. 1905, later homonym (illegitimate, Article 53).— Fig. 845.

Type — Bolivia: Tarija. On dead stems of *Flavaria contragerba* (Asteraceae). Fries, Exped. Suec. in reg. Chaco-Andinis, Fungi no. 268, II 1902 (S, holotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. ammophilae*, with asci pyriform, ascospores 11-13 × 3-4 µm.

Mycosphaerella staticicola (Pat.) Dias, Mem. Soc. Brot. 21: 72. 1970 = *Sphaerella staticicola* Pat., Catalogue raisonné des plantes cellulaires de la Tunisie. Fungi: 104. 1897.— Fig. 846.

Type — Tunisia: Gabès. On dead stems of *Armeria* ["*Staticae*"] (Plumbaginaceae). II 1893 (FH-Patouillard, holotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. ammophilae*, with asci pyriform, ascospores 10-12 × 2.5-3.5 µm.

Mycosphaerella stellariae Magnus. See *Mycosphaerella stellarinearum* (Rabenh.) Johanson.

Sphaerella stellariae f. *holostea* (Auersw.) Thüm. See *Sphaeria stellarinearum* var. *holostea* Auersw.

Mycosphaerella stellarinearum (Rabenh.) Johanson, Öfvers. Förh. Kongl. Svenska Vetensk.-Akad. 41(9): 165. 1884 [also as "*stellarinarum*"] = *Sphaeria stellarinearum* Rabenh., Herb. Vivum Mycol., ed. 1 no. 975. 1846 = *Sphaerella stellarinearum* (Rabenh.) P. Karst., Öfvers. Förh. Kongl. Svenska Vetensk.-Akad. 29: 104. 1872 = *Mycosphaerella stellariae* Magnus, Abh. Naturhist. Ges. Nürnberg 13: 32. 1899, superfluous (illegitimate, Article 52) nomen novum (Article 58) for *Sphaerella stellariae* Fuckel, Jahrb. Nassauischen Vereins Naturk. 23-24: 102. 1870, superfluous (illegitimate, Article 52) nomen novum (Article 58) = *Sphaeria stellarinearum* var. *cerastii* Rabenh., Herb.

Vivum. Mycol., ed. 1 no. 975. 1846, superfluous (illegitimate, Article 52) nomen novum (Article 58) = *Sphaerella cerastii* (Rabenh.) Fuckel, Jahrb. Nassauischen Vereins Naturk. 23-24: 101. 1870, superfluous (illegitimate, Article 52) nomen novum (Article 58) = *Mycosphaerella cerastii* (Rabenh.) Magnus, Abh. Naturhist. Ges. Nürnberg 13: 32. 1900, superfluous (illegitimate, Article 52) nomen novum (Article 58).— Fig. 175, 847.

Type — Germany: Dresden, Driesen. On upper and lower surface of dead leaves of *Cerastium triviale* (Caryophyllaceae). Klotzsch, Herbarium Vivum Mycologicum, ed. 1 no. 975 (L, isotype; B, isotype).

Anamorph: *Isariopsis pusilla* Fresen. *vide* Saccardo (1882).

This is a *Didymella*, with septate, 1-2 µm wide paraphyses, ascospores 18-21 × 6-8 µm. It was cited as synonymous with *Mycosphaerella isariphora* by Tomilin (1979).

Sphaeria stellarinearum var. *cerastii* Rabenh. See *Mycosphaerella stellarinearum* (Rabenh.) Johanson.

Sphaeria stellarinearum var. *holostea* Auersw., in Rabenh., in Klotzsch, Herbarium Vivum Mycologicum, ed. 1 no. 976. 1846 [as "*stellarinarum* var."] = *Sphaerella stellariae* f. *holostea* (Auersw.) Thüm., Mycotheca Universalis no. 163. 1875.— Fig. 848, 849.

Type — Germany: Leipzig. On upper and lower surface of dead leaves of *Stellaria holostea* (Caryophyllaceae). Auerswald, distributed in Rabenhorst, in Klotzsch, Herbarium Vivum Mycologicum no. 976 (L, isotype).

This is morphologically indistinguishable from *M. punctiformis*, with ascomata immersed, asci cylindrical, ascospores 9-11 × 2.5-3 µm.

Mycosphaerella stemmatea (Fr. : Fr.) Romell, Fungi Exsiccati Scandinavici, cent. 1 no. 68. 1890 = *Sphaeria stemmatea* Fr., Kongl. Vetensk. Acad. Handl. 38: 246. 1817, sanctioned by Fr., Systema Mycol. 2: 528. 1823 = *Sphaerella stemmatea* (Fr. : Fr.) Ellis & Everh., N. Amer. Pyrenomyc.: 730. 1892 = *Stigmatea stemmatea* (Fr. : Fr.) J. Schröt., in Cohn, Kryptog.-Fl. Schlesien 3(2): 331. 1894 ["1893"].— Fig. 850.

Type — Sweden: On spots on upper surface of living leaves of *Vaccinium vitis-idaea* (Ericaceae). Fries, Scleromyceti Suecici no. 287 (UPS, holotype).

Spermatial state: *Asteromella stemmatea* (Fr.) Petr. *vide* Petrak (1924).

The holotype is unfortunately immature. Additional material studied (Austria, Tirol, Tuxer Voralpen, Navis, Maas Geesteranus no. 15257, VI 1968, L) is a parasitic species, with asci cylindrical, ascospores 13-15 × 3-3.5 µm.

Sphaeria stemmatea f. *spermogonifera* Rabenh., Fungi Europaei Exsiccati no. 832. 1865.

Type — Poland: Swinoujście ["Swinemünde"], Seebad Heringsdorf. On dark brown spots with black margins on upper surface of living leaves of *Vaccinium vitis-idaea* (Ericaceae). Braun, distributed in Rabenhorst, Fungi Europaei Exsiccati no. 832, IX 1864 (B, isotype).

This contains only a coelomycete, as can be expected from the name, referring to spermatia.

Sphaerella stenospora Ellis & Everh., Bull. Torrey Bot. Club 24: 461. 1897.

Type — USA: *Sphaeralcea* (Malvaceae).

No material was studied as the type was not found in BPI or NY and is probably lost.

Mycosphaerella stephaniae Sawada, Taiwan Univ. Coll. Agric. Special Publ. 8: 65. 1959.

Type — Taiwan: *Stephania sasakii* (Menispermaceae).

No material was studied as the type was not found in BPI.

Mycosphaerella stephaniicola Sawada, Taiwan Univ. Coll. Agric. Special Publ. 8: 65. 1959.

Type — Taiwan: *Stephania sasakii* (Menispermaceae).

No material was studied as the type was not found in BPI.

Mycosphaerella stephanorossiae Duke, Kew Bull. 8: 317. 1926.— Fig. 851.

Type — Kenya: Kisumu, Nairobi. On lower surface of dead but still green leaves of *Stephanorossia* (Apiaceae). Dümmer no. 1749, II 1915 (K, holotype).

This belongs to *Davidiella*, with asci pyriform, with apical thickening, ascospores 26-36 × 5-7 µm. Therefore the following new combination is made: **Davidiella stephanorossiae** (Duke) Aptroot comb. nov., **MB 500364**. **Basionym:** *Mycosphaerella stephanorossiae* Duke, Kew Bull. 8: 317. 1926.

Mycosphaerella stevensii Tomilin, Novosti Sist. Nizsh. Rast. 1968: 167. 1968, nomen novum (Article 58) for *Mycosphaerella erythrinae* F. Stevens, Ann. Mycol. 28: 285. 1930, later homonym (illegitimate, Article 53).— Fig. 852.

Type — Panama: Juan Diaz. On white spots with brown margins on upper and lower surface of living leaves of *Erythrina* (Fabaceae). Stevens no. 1235, VIII 1923 [“1921”] (BPI, isotype).

This is a species of *Phaeosphaeria*, with asci cylindrical, pseudoparaphyses 1 µm wide, surrounded by a gelatinous mass, ascospores 3-septate, second cell from above widest, golden brown, ornamented with warts, 14.5-17 × 3-4 µm.

Mycosphaerella stigmaphyllonis Rangel, Arq. Mus. Nac. Rio de Janeiro 18: 161. 1916 [as “*stigmaphylli*”] = *Sphaerella stigmaphyllonis* (Rangel) Trotter, Syll. Fung. 24: 878. 1928 [as “*stigmaphylli*”].

Type — Brazil: *Stigmaphyllon ciliatum* (Malpighiaceae).

No material was studied as the type was not found in SP nor included in a loan from URM.

Sphaerella stigmatodes Cooke, J. Bot. 21: 68. 1883 = *Laestadia stigmatodes* (Cooke) Sacc., Syll. Fung. 2: XXXI. 1883.

Type — USA: On leaves.

No material was studied as the species has been redisposed.

Mycosphaerella stigmata-platani F.A. Wolf, Mycologia 30: 60. 1938 = *Mycosphaerella polymorpha* D.J. Sm. & C.O. Sm., Hilgardia 14: 206. 1941, superfluous (illegitimate, Article 52) nomen novum (Article 58).— Fig. 853.

Type — USA: North Carolina, Durham. On lower surface of dead leaves of *Platanus occidentalis* (Platanaceae). Wolf, IV 1937 (L, isotype), also distributed in Reliquiae Farlowianae no. 932 (NY, isotype).

Anamorphs: *Stigmata platani* (Fuckel) Sacc. *vide* Sivanesan (1984); *Xenostigmata wolfii* Crous & Corlett *vide* Crous & Corlett (1998).

This is an aberrant species, especially because the unusual anamorph (Crous & Corlett 1998), with asci pyriform, ascospores 11-15 × 3.5-5 µm. As its host, the tree genus *Platanus* has recently been realized to be related to the Proteaceae (Bremer *et al.* 2003), it may turn out to have close relatives in the generally well-studied species on Proteaceae.

Mycosphaerella stipicola Tomilin. See *Sphaerella chlorina* Sacc. & Trab.

Mycosphaerella stipina Petr., Ann. Mycol. 29: 113. 1931.— Fig. 854.

Type — Spain: Valencia, Alicante. On outer surface of dead, inrolled leaves of *Stipa juncea* (Poaceae). Ade, VI 1929 (W, holotype; NY, isotype).

Already cited as synonymous with *Davidiella ammophilae* (as *M. minor*) by Tomilin (1979), with which the type agrees well, with asci pyriform to globose, ascospores 8-10 × 3-4 µm.

Mycosphaerella striatiformans Cobb, Rept. Exp. Station Hawaiian Sugar Planter's Assoc., Bull. 5: 208. 1906 = *Sphaerella striatiformans* (Cobb) Sacc. & Trotter, Syll. Fung. 22: 145. 1913.

Type — Hawaii: *Saccharum officinarum* (Poaceae).

No material was studied as the location of the type is unknown. Cited as synonymous with *Mycosphaerella sacchari* (Speg.) Seaver & Chardón by Tomilin (1979).

Mycosphaerella striola Petr., nomen herbariorum (not validly published, Article 32).— Fig. 855.

Authentic material — Hawaii: Maui, Olinda Pipe Line. On dead rhachis of *Pteridium* [“*Pteris*”] *excelsa* (Polypodiaceae). Shear & Stevens, XII 1927 (B).

This is *Leptopeltis filicina* (Lib.) Höhn., with asci cylindrical, ascospores 3-septate, pale brown, 11-12 × 2.5-3 µm. Material from the same host and locality (but belonging to a different species) has been described as *M. olindensis* Petr.

Mycosphaerella stromatica (Rehm) Arx, in E. Müll. & Arx, Beitr. Kryptogamenfl. Schweiz 11(2): 358. 1962 = *Lizonia stromatica* Rehm, Ann. Mycol. 6: 323. 1908 = *Euryachora stromatica* (Rehm) Höhn., Sitzungsber. Kaiserl. Akad. Wiss., Math.-Naturwiss. Kl., Abt. 1, 120: 421. 1911 = *Lizoniella stromatica* (Rehm) Sacc. & Trotter, Syll. Fung. 22: 167. 1913.

Type — Austria: *Saxifraga* (Saxifragaceae).

No material was studied as the species was redisposed.

Mycosphaerella stromatica (Munk) Tomilin, Opredelitel' gribov roda *Mycosphaerella* Johans.: 187. 1979, later homonym (illegitimate, Article 53) = *Mycosphaerella*

psammae var. *stromatica* Munk, Dansk Bot. Ark. 17(1): 319. 1957.— Fig. 856.

Type — Denmark: Jutland, Tversted. On the outer surface of dead leaves of *Ammophila* [*Psamma*] *arenaria* (Poaceae). Ostenfeld, VII 1905 (C, holotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. disseminata*, with asci pyriform, ascospores 13-15 × 3-3.5 µm. The ascomata are not at all stromatic, in contrast with the original name and description.

Mycosphaerella stromatoidea Dearn., Mycologia 18: 245. 1926.— Fig. 857.

Type — USA: Washington, Clallam Co., Forks. On small brown spots with reddish brown margins on upper surface of living leaves of *Rumex occidentalis* (Polygonaceae). Boyce no 1288, VIII 1924 (BPI, isotype).

Cited as synonym of *Venturia rumicis* (Desm.) G. Winter by Sivanesan (1977), with which the isotype agrees, with pseudoparaphyses numerous, ascospores becoming greenish brown, 15-17 × 6.5-7.5 µm.

Mycosphaerella stromatosa Joa.E. Taylor & Crous, Mycol. Res. 104: 625. 2000.

Type — South Africa: Drakensberg. On spots on living leaves of *Protea* (Proteaceae). Denman, I 1998 (CBS, isotype, living culture).

Anamorph: *Pseudocercospora stromatosa* Joa.E. Taylor & Crous (op. cit.).

The type specimen of this recently described species was not studied; the ex-type strain does not produce ascomata in culture.

Mycosphaerella strychni S.M. Lin & P.K. Chi, in P.K. Chi, Fungal Diseases of Cultivated Medicinal Plants in Guangdong Province: 69. 1994 [as "*strychnoris*"].

Type — China: *Strychnos nux-vomica* (Loganiaceae).

No material was studied as the location of the type is unknown and its preservation uncertain.

Mycosphaerella styracis Starbäck, Bih. Kongl. Svenska Vetensk.-Akad. Handl. 25(3, 1): 57. 1899 = *Sphaerella styracis* (Starbäck) Sacc. & P. Syd., Syll. Fung. 16: 468. 1902.— Fig. 858.

Type — Brazil: Rio Grande do Sul, Cascata de Hermenegilda, Pelatato. *Styrax* (Styracaceae). Malme, 1st Regnellian Exped. no. 109, XII 1892 (S, holotype).

This belongs to section *Longispora*, and is morphologically indistinguishable from *M. latebrosa*, with asci cylindrical, surrounded by some interascal cells, ascospores 25-35 × 3.5-4.5 µm. Additional material studied (Brazil, S. Leopoldo, Rio Grande do Sul, superficial on lower surface of living leaves, but still saprobic, of *Styrax leprosa*, Theissen, Decades fungorum Brasiliensium no. 271, 1907, L) belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with asci pyriform, thick (3 µm)-walled, ascospores 15-20 × 5.5-7.5 µm.

Mycosphaerella suaedae-australis Hansf., Proc. Linn. Soc. New South Wales 79: 12. 1954.

Type — Australia: *Suaeda australis* (Amaranthaceae).

Anamorph: *Septoria suaedae-australis* Hansf. fide Hansford (op. cit.).

No material was studied as the holotype is not in IMI or K.

Sphaerella subalpina Sacc., Michelia 2: 247. 1881.

Type — Italy: Veneto, Cadore. On ochraceous yellow spots with brown margins on upper surface of living leaves of *Senecio cordatus* (Asteraceae). Spegazzini, 1879, distributed in Saccardo, Mycotheca Veneta no. 1490 (NY, isotype).

The type specimen contains only empty ascomata. No ascospores were found by the original author either.

Mycosphaerella subantarctica (Speg.) Cash, Syll. Fung. 26: 354. 1972 = *Sphaerella subantarctica* Speg., Bol. Acad. Nac. Ci. 27: 363. 1924.

Type — Argentina: *Nothofagus pumilio* (Nothofagaceae).

No material was studied as the type was not included in a loan from LPS.

Mycosphaerella subastoma F. Stevens & Dalbey, Mycologia 11: 8. 1919 = *Sphaerella subastoma* (F. Stevens & Dalbey) Trotter, Syll. Fung. 24: 865. 1928.

Type — Puerto Rico: *Anemia adiantifolia* (Polypodiaceae). No material was studied as the type was not found in NY or BPI.

Mycosphaerella subcongregata (Ellis & Everh.) Tomilin, Novosti Sist. Nizsh. Rast. 1967: 190. 1967 = *Sphaerella subcongregata* Ellis & Everh., J. Mycol. 2: 101. 1886 = *Phaeosphaerella subcongregata* (Ellis & Everh.) Tomilin, Opređelitel' gribov roda *Mycosphaerella* Johans.: 285. 1979.— Fig. 859.

Type — USA: Washington, Mt. Paddo. On dead stems of *Erigeron salsuginosus* (Asteraceae). Suksdorf no. 234, VIII 1885 (NY, holotype).

Anamorph: *Diplodina smilacis* Ellis & Everh. fide Ellis & Everhart (op. cit.).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores thick-walled and rough when mature, 19-24 × 5.5-7 µm. *Pleospora permunda* Cooke is also present on the holotype specimen.

Sphaerella subcrassa Sacc. & P. Syd., Syll. Fung. 14: 533. 1899, nomen novum (Article 58) for *Sphaerella maculans* Pass., Atti Reale Accad. Lincei, Mem. Cl. Sci. Fis., ser. 4, 4: 57. 1888, later homonym (illegitimate, Article 53) = *Mycosphaerella populi-albae* Tomilin, Novosti Sist. Nizsh. Rast. 1967: 183. 1967, superfluous (illegitimate, Article 52) nomen novum (Article 58).

Type — Italy: *Populus alba* (Salicaceae).

No material was studied as the type was not found in any of the herbaria consulted. Cited as synonymous with *Mycosphaerella alni* (Fuckel) Tomilin by Tomilin (1979).

Mycosphaerella suberosa Crous, Ferreira, Alfenas & M.J. Wingf., Mycologia 85: 707. 1993.

Type — Brazil: Espirito Santo, Santa Catarina. On leaves of *Eucalyptus dunnii* (Myrtaceae). Wingfield, VIII 1992 (PREM 51082, holotype, not seen).

No material of this recently described species was studied.

Mycosphaerella subgregaria Petr., Ann. Mycol. 25: 285. 1927.— Fig. 860.

Type — Czech Republic: Weißkirchen, Bartelsdorf. On dead stems of *Impatiens* (Balsaminaceae). Petrak, V 1923 (W no. 11883, holotype; W, 3 isotypes).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradicans*, with asci cylindrical, ascospores 10-13 × 3-4 µm.

Mycosphaerella sublibera Petr., Sydowia 4: 505. 1950.— Fig. 861.

Type — Ecuador: Tungurahua, Baños. On white spots on lower surface of living leaves of *Pilea melastomoides* (Urticaceae). Sydow, XII 1937 (BPI, isotype), also distributed in Reliquiae Petrakianae no. 661 (H, L, isotypes).

This is a parasitic species, with asci cylindrical, ascospores 17-19 × 3-4 µm.

Mycosphaerella subnivalis (Rehm) Lindau, Hilfsb. Sammeln Ascomyceten: 28. 1903 = *Sphaerella subnivalis* Rehm, Hedwigia 24: 237. 1885.— Fig. 862.

Type — Austria: Tyrol, Ortler, Sulden-gletscher. On upper and lower surface of dead leaves of *Cerastium latifolium* (Caryophyllaceae). Rehm no. 832, VII 1884 (S, holotype).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. superflua*, with asci cylindrical, ascospores 16-19 × 3.5-4.5 µm.

Mycosphaerella subostiolica (Aggéry) M. Morelet, Bull. Soc. Sci. Nat. Archéol. Toulon & Var 177: 9. 1968, lacking Latin description (not validated, Article 36) = *Sphaerella subostiolica* Aggéry, Bull. Soc. Hist. Nat. Toulouse 68: 11. 1935, lacking Latin description (not validly published, Article 36).

Type — France: *Polypodium* (Polypodiaceae).

No material was studied as the location of the type is unknown.

Mycosphaerella subradicans (Fr. : Fr.) J. Schröt., in Cohn, Kryptog.-Fl. Schlesien 3(2): 339. 1894 [“1893”] = *Sphaeria subradicans* Fr., Kongl. Vetensk. Acad. Handl. 38: 248. 1817, sanctioned by Fr., Systema Mycol. 2: 525. 1823 = *Sphaerella subradicans* (Fr. : Fr.) Auersw., in Gonn. & Rabenh., Mycol. Europaea 5-6: 19. 1869.

Type — Sweden. On upper and lower surface of dead leaves of *Convallaria majalis* (Asparagaceae). Fries, Scleromyceti Suecici no. 204 (UPS, holotype).

Cited as synonym of *M. asteroma* by Eriksson (1992), but the name *M. subradicans* is accepted here, as its basionym is older. It differs from the related *M. superflua* by the shorter and more asymmetrical (septum closer to the upper end) ascospores with often rounded ends. It grows more on dead leaves, while *M. superflua* occurs predominantly on dead stems.

Sphaerella subradicans f. *convallariae-majalis* Thüm., Fungi Austriaci no. 470, 1872.— Fig. 863.

Type — Czech Republic: Bohemia, Kaplitz. On upper and lower surface of dead leaves of *Convallaria majalis* (Asparagaceae). Kirchner, distributed in Thümen, Fungi Austriaci no. 470, 1872 (B, isotype).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradicans*, with asci cylindrical, ascospores 9-11 × 2.5-3 µm. It is identical to the nominal variety and in fact described from the type host.

Mycosphaerella subsequens Munk, in E. Müll., Fungi of the Faeröes 2: 60. 1958.

Type — Faeröes: *Armeria vulgaris* (Plumbaginaceae).

No material was studied as the type was not found in C and might be lost. Cited as synonymous with *Mycosphaerella kirschsteinii* by Tomilin (1979).

Mycosphaerella succedanea (Pass.) Tomilin, Novosti Sist. Nizsh. Rast. 6: 122. 1970 [“1969”] = *Sphaerella succedanea* Pass., Atti Reale Accad. Lincei, Mem. Cl. Sci. Fis., ser. 4, 3: 6. 1887.

Type — Italy: *Vitis vinifera* (Vitaceae).

Anamorph: Associated with *Phoma succedanea* Pass. *vide* Pass. (op. cit.).

No material was studied as the type was not found in any of the herbaria consulted. Cited as synonymous with *Mycosphaerella curvulata* by Tomilin (1979).

Sphaerella succinea (Roberge) Cooke, J. Bot. 21: 106. 1883 = *Sphaeria succinea* Roberge, in Desm., Ann. Sci. Nat. Bot., ser. 3, 10: 354. 1848 = *Charonectria succinea* (Roberge) Sacc., Syll. Fung. 2: LXVIII. 1883 = *Nectriella succinea* (Roberge) Weese, Ann. Mycol. 12: 130. 1914 = *Paradidymella succinea* (Roberge) Petr., Ann. Mycol. 25: 239. 1927 = *Leiosphaerella succinea* (Roberge) E. Müll., in E. Müll. & Arx, Beitr. Kryptogamenfl. Schweiz 11(2): 673. 1962.

Type — France: *Quercus* (Fagaceae).

Accepted as *Leiosphaerella succinea* (Roberge) E. Müll. by Müller & von Arx (op. cit.) and therefore not studied.

Mycosphaerella sumacis (Madej) M. Morelet, Bull. Soc. Sci. Nat. Archéol. Toulon & Var 176: 7. 1968 = *Sphaerella sumacis* Madej, Zesz. Nauk. Wyszszej Szkoły Roln. Szczecinie 21: 146. 1966.

Type — Poland: *Rhus typhina* (Anacardiaceae).

No material was studied as the location of the type is unknown. Cited as synonymous with *Mycosphaerella fagi*, which is morphologically indistinguishable from *M. punctiformis*, by Tomilin (1979).

Mycosphaerella superflua (Auersw.) Petr., Ann. Mycol. 38: 235. 1940 = *Sphaerella superflua* Auersw., in Gonn. & Rabenh., Mycol. Europaea 5-6: 14. 1869 = *Sphaeria superflua* Auersw., in Fuckel, Fungi Rhenani Exsiccati no. 884. 1863, nomen nudum (not validly published, Article 32) = *Didymosphaeria superflua* (Auersw.) Niessl, in J. Kunze, Fungi Selecti Exsiccati no. 258. 1875 = *Didymella superflua* (Auersw.) Sacc., Michelia 2: 316. 1881.— Fig. 864.

Type — Germany: Östrich. On dead stems of *Urtica dioica* (Urticaceae). Fuckel, 1863, Fungi Rhenani Exsiccati no. 884 (BPI, isotype).

Anamorph: *Ramularia urticae* Ces. *vide* Sivanesan (1984).
The type and additional material studied (e.g. Netherlands, Lelystad, Jansen no. 88-106, VI 1988, L) belongs to section *Caterva*, with asci cylindrical, ascospores $12.5-16(-19) \times 3-5 \mu\text{m}$. It differs from *M. subradians* by the longer and symmetrical ascospores, often with pointed ends, and its predominant occurrence on dead stems.

Sphaerella superflua f. *sisymbrii* Anzi, Erbario Crittogamico Italiano, ser. 2, 1868.— Fig. 865.

Type — Italy, Bormio, Valtellini. On dead stems of *Sisymbrium strictissimum* (Brassicaceae). Anzi, Erbario Crittogamico Italiano, ser. 2 (BPI, isotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores $12.5-14 \times 4.5-6 \mu\text{m}$.

Didymosphaeria superflua f. *umbelliferarum* Niessl, nomen herbariorum (not validly published, Article 32).

Authentic material — Austria: Salzburg, on dead stems of *Seseli* (Apiaceae). Niessl, VI 1911 (M).

This contains typical material of *Mycosphaerella superflua*.

Didymosphaeria superflua f. *urticae-dioicae* J. Kunze, Fungi Selecti Exsiccati 258. 1879.

Type — Germany: Eisleben, Neckendorfer Thal, on dead stems of *Urtica dioica* (Urticaceae). Kunze, IV 1879, distributed in Fungi Selecti Exsiccati no. 258 (M, NY (2 ×), isotypes).

The isotypes contain typical material of *Mycosphaerella superflua*.

Didymella superflua ssp. *humuli* P. Karsten, Symb. Mycol. Fenn. 16: 150. 1884.

Type — Finland: Tavestia, Mustiala, on dead stems of *Humulus lupulus* (Urticaceae). Karsten no. 3705, XII 1865 (H, isotype).

The type material studied is very poor and contains only a coelomycete. Additional material (Czech Republic, Brno (“Brünn”), Niessl, VIII 1873, M) contains typical *Mycosphaerella superflua*.

Sphaerella suspecta Auersw., in Rabenh., Fungi Europaei Exsiccati, ed. nov., ser. 2, cent. 8 no. 833. 1865 ≡ *Plagiostoma suspecta* (Auersw.) Fuckel, Jahrb. Nassauischen Vereins Naturk. 23-24: 119. 1870 ≡ *Gnomonia suspecta* (Auersw.) Sacc., Syll. Fung. 1: 566. 1882.— Fig. 866.

Type — Germany: Coburg. On upper and lower surface of dead leaves of *Quercus robur* (Fagaceae). Sollmann, distributed in Rabenhorst, Fungi Europaei Exsiccati no. 833, 1863 (L, isotype).

This is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores $8-10 \times 2-2.5 \mu\text{m}$.

Mycosphaerella suttonii Crous & M.J. Wingf., Canad. J. Bot. 75: 783. 1997 [as the feminine “*suttoniae*”].

Type — Indonesia: Sumatra, Lake Toba. On leaves of *Eucalyptus* (Myrtaceae). Wingfield, III 1996 (PREM 54963a, holotype, not seen).

Anamorphs: *Phaeophleospora epicoccoides* (Cooke & Masee) Crous, Ferreira & B. Sutton (with many synonyms) and *Cercostigmia* *vide* Crous (1998).

No material of this recently described species was studied.

Mycosphaerella swartii R.F. Park & Keane, Trans. Brit. Mycol. Soc. 83: 99. 1984.— Fig. 867.

Type — Australia: Southern Australia, Mt. Gambier. On pale spots with brown margins on upper and lower surface of living leaves of *Eucalyptus leucoxylon* (Myrtaceae). Park, XII 1982 (IMI no. 280474, isotype).

Anamorph: *Sonderhenia eucalyptorum* (Hansf.) H.J. Swart & J. Walker *vide* Crous (1998).

This is a parasitic species, with asci cylindrical, ascospores $18-23 \times 4-6 \mu\text{m}$.

Mycosphaerella sydowii Tomilin, Novosti Sist. Nizsh. Rast. 11: 249. 1974.

Type — Germany: *Tragopogon pratensis* (Asteraceae).

No material was studied as the type was not included in loans from LE or LEP.

Mycosphaerella sylvatica (Sacc. & Speg.) Laib., Centralbl. Bakteriol., 1. Abt., Originale 53: 552. 1921 [as “*silvatica*”] ≡ *Sphaerella sylvatica* Sacc. & Speg., in Sacc., Michelia 1: 380. 1878 ≡ *Mycosphaerella scabiosae* Tomilin, Novosti Sist. Nizsh. Rast. 8: 151. 1971, superfluous (illegitimate, Article 52) nomen novum (Article 58).— Fig. 868.

Type — Italy: On upper and lower surface of dead leaves of *Scabiosa sylvatica* (Dipsacaceae). Spegazzini (PAD, holotype).

Anamorph: *Ramularia knautiae* (C. Massal.) Bubák *vide* von Arx (1983) (= *Ramularia tricherae* Lindr.).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradians*, with asci cylindrical, ascospores $10-13 \times 3.5-4.5 \mu\text{m}$.

Mycosphaerella sylvatica (Sacc.) Petr. & Syd., Ann. Mycol. 23: 281. 1925 [as “*silvatica*”], later homonym (illegitimate, article 64) ≡ *Stigmatea sylvatica* Sacc., Michelia 2: 604. 1882.

Type — France: *Juniperus communis* (Cupressaceae).

No material was studied as the type was not found in any of the herbaria consulted. Cited as synonymous with *Sphaerella fructinex* by Tomilin (1979).

Sphaerella symphoricarpi Pass., Atti Reale Accad. Lincei, Mem. Cl. Sci. Fis., ser. 4, 6: 457. 1890 [“1889”].

Type — Italy: *Symphoricarpos racemosus* (Adoxaceae).

No material was studied as the type was not found in any of the herbaria consulted.

Mycosphaerella symphyostemonis (Speg.) Cash, Syll. Fung. 26: 354. 1972 [as “*symphyostemi*”] ≡ *Sphaerella symphyostemonis* Speg., Bol. Acad. Nac. Ci. 27: 364. 1924 [as “*symphyostemi*”].

Type — Argentina: *Symphyostemon biflorus* (Iridaceae).

No material was studied as the type was not included in a loan from LPS.

Mycosphaerella syringae Cruchet. See *Mycosphaerella cruchetii* M. Morelet.

Mycosphaerella syringicola (G.H. Otth) Lindau, Hilfsb. Sammeln Ascomyceten: 118. 1903 = *Sphaerella syringicola* G.H. Otth, Mitth. Naturf. Ges. Bern 1870: 110. 1871.

Type — Switzerland: *Syringa vulgaris* (Oleaceae).
No material was studied as the type was not found in any of the herbaria consulted.

Mycosphaerella syzygii Crous, Mycol. Res. 103: 611. 1999.
Type — India: Bangalore. On spots on living leaves of *Syzygium cumini* (Myrtaceae). Seshadri & Channamma, VII 1968 (BPI 436209, holotype, not seen).

Anamorph: *Cercostigmina punctata* (Wakef.) Crous *vide* Crous (op. cit.) (= *Pseudocercospora punctata* (Wakef.) B. Sutton).

No material of this recently described species was studied.

Mycosphaerella tabaci (Maubl.) Miles, Trans. Illinois State Acad. Sci. 10: 250. 1917 [as “*tabaca*”] = *Sphaerella tabaci* Maubl., Bull. Soc. Mycol. France 23: 142. 1908 [“1907”].

Type — France: *Nicotiana tabacum* (Solanaceae).
No material was studied as the type was not included in a loan from PC.

Mycosphaerella tabebuiae Miles, Trans. Illinois State Acad. Sci. 10: 249. 1917 = *Sphaerella tabebuiae* (Miles) Trotter, Syll. Fung. 24: 854. 1928.— Fig. 869.

Type — Puerto Rico: Vega Baja. On white spots with purple margins on upper surface of living leaves of *Tabebuia haemantha* (Bignoniaceae). Stevens no. 2021, V 1913 (K, NY, isotypes).

This is a parasitic species, with asci cylindrical, ascospores 14-19 × 3-4 µm.

Mycosphaerella tabifica (Prill. & Delacr.) Lind, Danish Fungi: 203. 1913 = *Sphaerella tabifica* Prill. & Delacr., Bull. Soc. Mycol. France 7: 24. 1891.

Type — France. On up to 5 cm large pale spots on upper and lower surface of living leaves of *Beta vulgaris* (Amaranthaceae). Delacroix (PC, holotype).

Anamorph: Associated with *Phyllosticta tabifica* Prill. *vide* Saccardo (1882).

The type contains only an *Ascochyta* anamorph.

Mycosphaerella tabularis Syd., Ann. Mycol. 26: 107. 1928.— Fig. 870.

Type — Chile: Chiloë, Piquina, Castro. On lower surface of dead fronds of *Lomaria magellanica* [“*Blechnum tabulare*”] (Polypodiaceae). Werdermann no. 1739, II 1924 (S, holotype; B, isotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. ammophilae*, with asci pyriform, ascospores 11-13 × 3-3.5 µm.

Sphaerella taccari Speg., Anales Mus. Nac. Hist. Nat. Buenos Aires 23: 52. 1912.

Type — Argentina: *Taccarum hasslerianum* (Araceae).
No material was studied as the type was not included in a loan from LPS.

Mycosphaerella taediosa (Pass.) Cruchet, Bull. Soc. Vaud. Sci. Nat. 55: 42. 1923 = *Sphaerella taediosa* Pass., in Pass. & Brunaud, Rev. Mycol. (Toulouse) 8: 205. 1886.

Type — France: *Sparganium ramosum* (Sparganiaceae).
No material was studied as the type was not found in any of the herbaria consulted. Cited as synonymous with *Mycosphaerella microscopica* by Tomilin (1979).

Mycosphaerella taeniographa Petr., Ann. Mycol. 38: 230. 1940.— Fig. 871.

Type — Czech Republic: Mähren, Skalička. On dead stems of *Conium maculatum* (Apiaceae). Petrak, distributed in Reliquiae Petrakianae no. 73, V 1934 (H, L, isotypes), also Cryptogamae Exsiccatae no. 3268 (L, isotype).

This belongs to section *Longispora*, and is morphologically indistinguishable from *M. latebrosa*, with asci cylindrical to pyriform, ascospores in one bundle, 20-25 × 2-2.5 µm.

Mycosphaerella taeniographoides Petr., Sydowia 1: 142. 1947.— Fig. 872.

Type — Austria: Niederdonau, Pfaffstätten bei Baden. On dead stems of *Siler trilobum* (Apiaceae). Petrak, Mycotheca Generalis no. 1842, V 1940 (W, holotype; W, isotype).

This belongs to section *Longispora*, and is morphologically indistinguishable from *M. topographica*, with ascomata in superficial black stromata, asci cylindrical, ascospores 22-26 × 4-5 µm.

Mycosphaerella tahitensis (Sacc.) Lindau, in Engl. & Prantl, Natürlichen Pflanzenf. 1(1): 425. 1897 = *Sphaerella tahitensis* Sacc., Atti Reale Ist. Veneto Sci., ser. 6, 2: 461. 1884.— Fig. 873.

Type — Tahiti. On upper and lower surface of dead leaves of *Mangifera indica* (Anacardiaceae). (PAD, holotype).

The type contains a *Pestalotiopsis* anamorph and a *Guignardia* species, with ascospores simple, 17-20 × 4-5 µm, with gelatinous appendages.

Mycosphaerella tajmyrensis Tomilin, Novosti Sist. Nizsh. Rast. 9: 107. 1972.

Type — Russia: *Trisetum spicatum* (Poaceae).
No material was studied as the type was not included in loans from LE or LEP.

Mycosphaerella tamaricis (Hollós) Kurbans., Mikoflora Kugitangtau: 133. 1975, full reference to basionym not given (not validly published, Article 33) = *Sphaerella tamaricis* Hollós, Ann. Hist.-Nat. Mus. Natl. Hung. 4: 332. 1906 = *Phaeosphaerella tamaricis* (Hollós) Tomilin, Opredelitel' gribov roda *Mycosphaerella* Johans.: 285. 1979.

Type — Hungary: *Tamarix africana* (Tamaricaceae).
The type is not in BP and may have been destroyed during the war.

Mycosphaerella tamarindi Henn., Notizbl. Königl. Bot. Gart. Berlin 3: 240. 1903 = *Sphaerella tamarindi* (Henn.) Sacc. & D. Sacc., Syll. Fung. 17: 637. 1905.

Type — Tanzania: *Tamarindus indica* (Fabaceae).
No material was studied as the type was not found in SP or B and is probably lost.

Sphaerella tamseliensis Kirschst., Kryptog.-Fl. Mark Brandenburg 7(3): 380. 1938.— Fig. 874.

Type — Germany: Tamsel, Baumschulen (= tree nursery). On upper and lower surface of dead leaves of *Actinidia polygama* (Actinidiaceae). Vogel, IV 1906 (B, lectotype, here designated).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradians*, with asci cylindrical, ascospores $10-12 \times 2.5-3.5 \mu\text{m}$. It was previously cited as synonymous with *Mycosphaerella hermione* by Tomilin (1979). The lectotype is the only original specimen present now in B under this name. There most have been more specimens, as the protologue mentions “April bis Mai”.

Mycosphaerella taraxaci (P. Karst.) Dearn., Rep. Canad. Arctic Exped. 1913-1918 4, C: 12. 1923 = *Sphaerella taraxaci* P. Karst., Öfvers. Förh. Kongl. Svenska Vetensk.-Akad. 1872: 106. 1872.— Fig. 876.

Type — Svalbard: *Taraxacum phymatocarpum* (Asteraceae).

Anamorph: *Cercospora augustana* Ferraris fide Sivanesan (1984) (= *Pseudocercospora angustana* (Ferraris) U. Braun).

No type material was found in H or any of the herbaria consulted. Material seen (Canada, Northwest Territories, Franklin Distr., Axel Heiberg Island, Wolf River Delta, on upper and lower surface of dead leaves of *T. arctogenum*, Parmelee no. 2195, VIII 1961, IMI no. 105192; also Parmelee no. 2003, VIII 1961, IMI no. 200597) belongs to *Davidiella*, with asci pyriform, ascospores $17-24 \times 3-4.5 \mu\text{m}$.

Mycosphaerella tardiva Syd., Ann. Mycol. 14: 245. 1916 = *Sphaerella tardiva* (Syd.) Trotter, Syll. Fung. 24: 887. 1928.— Fig. 875.

Type — Germany: Brandenburg, Tamsel. On upper and lower surface of dead leaves of *Scrophularia nodosa* (Scrophulariaceae). Vogel, distributed in Sydow, Mycotheca Germanica no. 1337, V 1914 (L, 2 isotypes).

This is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores $8-10 \times 2-2.5 \mu\text{m}$.

Mycosphaerella tassiana (De Not.) Johanson, Öfvers. Förh. Kongl. Svenska Vetensk.-Akad. 41: 167. 1884 = *Sphaerella tassiana* De Not., Sferiacei Italici 1: 87. 1863 = *Davidiella tassiana* (De Not.) Crous & U. Braun, in U. Braun *et al.*, Mycol. Progress 2: 8. 2003.— Fig. 877.

Type — Italy. On upper and lower surface of dead leaves of *Carex nigra* [“*fusca*”] (Cyperaceae). Tassi no. 862 (RO, holotype).

Anamorph: *Cladosporium herbarum* (Pers. : Fr.) Link fide Sivanesan (1984).

The holotype was studied and found to conform to the current opinion about this species, with asci pyriform, ascospores $17-22 \times 5-7 \mu\text{m}$, seemingly with endospore formation.

As already indicated by von Arx (1949), the name *Mycosphaerella allicina* has been used for a polyphagous but supposedly parasitic species. However, the type and other material of *Mycosphaerella allicina* clearly grows on

dead plant materials. Therefore the species should be referred to as *Davidiella allicina*.

Sphaerella tassiana f. *festucae* Fautrey, Rev. Mycol. (Toulouse) 13: 75. 1891.— Fig. 878.

Type — France: Côte-d'Or, Forêt de Charny. On dead culms of *Festuca heterophylla* (Poaceae). Fautrey, distributed in Roumeguère, Fungi Selecti Exsiccati no. 5623, VI 1890 (PC, holotype).

This is morphologically indistinguishable from *Phaeosphaeria fautreyi* Shoemaker & C.E. Babc., with pale brown, 3-septate ascospores of $18-22 \times 5-6 \mu\text{m}$.

Mycosphaerella tassiana var. *arctica* (Rostr.) M.E. Barr, Contr. Inst. Bot. Univ. Montréal 73: 24. 1959 = *Laestadia arctica* Rostr., Meddel. Grønland 3: 547. 1888 = *Mycosphaerella allicina* var. *arctica* (Rostr.) M. Morelet, Bull. Soc. Sci. Nat. Archéol. Toulon & Var 196: 7. 1971.— Fig. 879.

Type — Greenland, Godhavn. On upper and lower surface of dead leaves and on dead stems of *Honckenya* [“*Halianthus*”] *peploides* (Caryophyllaceae). Warming & Holm, VII 1884 (C, holotype).

Already synonymised with *Davidiella allicina* (as *Mycosphaerella tassiana*) by von Arx (1949), but the holotype indicates that this is morphologically indistinguishable from *D. clandestina*, with asci pyriform, ascospores $27-30 \times 6-7 \mu\text{m}$. It was cited as a synonym of *Mycosphaerella halophila* (E. Bommer, M. Rousseau & Sacc.) O.E. Erikss. by Eriksson (1992).

Mycosphaerella tassiana var. *arthopyrenioides* (Auersw.) M.E. Barr. See *Mycosphaerella arthopyrenioides* (Auersw.) Lindau.

Mycosphaerella [“*Mycosphaerium*”] *tassiana* [“*tassianum*”] var. *macrospora* [“*macrosporum*”] Clem., Cryptogamae Formationum Coloradensium no. 228. 1906, nomen herbariorum (not validly published, Article 32).— Fig. 880. Authentic material — USA: Colorado, Minnehaha. On upper and lower surface of dead leaves of *Calamagrostis purpurascens* (Poaceae). Clements, VIII 1906, Cryptogamae Formationum Coloradensium no. 228 (BPI).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores thick-walled, $17-23 \times 6.5-8 \mu\text{m}$. The ascospores are not even particularly large for the species.

Sphaerella tassiana var. *primulae* Rehm, Ber. Naturhist. Vereins Augsburg 26: 74. 1881.

Type — Germany: *Primula* (Primulaceae).

Sphaerella tassiana var. *pusilla* (Auersw.) P. Karst. See *Mycosphaerella pusilla* (Auersw.) Johanson.

Mycosphaerella [“*Mycosphaerium*”] *tassiana* [“*tassianum*”] var. *rupefortensis* [“*rupefortense*”] (Pass.) Clem., Cryptogamae Formationum Coloradensium no. 229. 1906 = *Sphaerella tassiana* var. *rupefortensis* Pass., in Pass. & Brunaud, Rev. Mycol. (Toulouse) 8: 205. 1886.

Type — France: On dead leaves of *Typha latifolia* (Typhaceae).

The type was not included in a loan from PC, and not found in any of the herbaria consulted. Material studied (USA, Colorado, Clements, VIII 1906, Cryptogamae Formationum Coloradensium no. 229, BPI) is overmature.

Sphaerella tassiana var. *turrata* P. Karst., Acta Soc. Fauna Fl. Fenn. 2(6): 67. 1885.

Type — Finland.

No material was studied as the type was not included in a loan from H.

Mycosphaerella tassiana var. *vagnerae* (Earle) Clem. See *Mycosphaerella vagnerae* Earle.

Mycosphaerella tatarica Miura, Industr. Contr. 27: 167. 1928, based on an anamorph (illegitimate, Article 59) = *Septoria tatarica* Syd., Ann. Mycol. 12: 163. 1914.

Type — *Aster tataricus* (Asteraceae).

No material was studied as the type is an anamorph.

Mycosphaerella taxi (Cooke) Lind, Danish Fungi: 204. 1913 = *Sphaerella taxi* Cooke, Grevillea 6: 128. 1878 = *Metasphaeria taxi* (Cooke) Oudem., Ned. Kruidk. Arch., ser. 3, 2: 170. 1900 = *Sphaerulina taxi* (Cooke) Masee, Diseases Cult. Pl.: 220. 1910.

Type — United Kingdom: Cornwall. On upper and lower surface of dead needles of *Taxus baccata* (Taxaceae). Boscaveau (K, holotype; K, 3 isotypes), also distributed in Fungi Britannici Exciccati no. 697, III 1878 (B, isotype), also Plowright, Sphaeriacei Britannici no. 90 (B, isotype).

Anamorphs: *Cytospora taxifolia* Cooke & Masee and *Dothichiza fide* Barr (1972).

Synonymised with *Dothiora taxicola* (Peck) M.E. Barr by Barr (1972). The holotype and isotypes agree well, but all are somewhat immature.

Mycosphaerella taxodii (Cooke) Lindau, in Engl. & Prantl, Natürlichen Pflanzenf. 1(1): 425. 1897 = *Sphaerella taxodii* Cooke, J. Bot. 21: 106. 1883.— Fig. 881.

Type — USA: California, Seaboard of So. On upper and lower surface of dead needles of *Taxodium distichum* (Taxodiaceae). Ravenel, distributed in Ellis & Everhart, North American Fungi no. 1676, III 1883 (L, isotype).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradians*, with asci cylindrical, ascospores 10-13 × 3-4 µm.

Mycosphaerella tecomae F.A. Wolf, Mycologia 35: 507. 1943.— Fig. 882.

Type — USA: North Carolina, Durham. On upper and lower surface of dead leaves of *Tecoma radicans* (Bignoniaceae). Wolf, IV 1943 (NY, IMI no. 228051 (slide only), isotypes).

Anamorph: *Pseudocercospora sordida* (Sacc.) Deighton *fide* Deighton (1976).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. superflua*, with asci cylindrical, ascospores 14-17 × 3-4 µm.

Mycosphaerella tecomae Viégas. See *Mycosphaerella viegasii* M. Morelet.

Mycosphaerella telopeae M.E. Palm & Crous, in Swart, Crous, Denman & Palm, S. Afr. J. Bot. 64: 141. 1998.

Type — New Zealand: *Telopea* (Proteaceae).

No material was studied of this recently described species.

Mycosphaerella terebinthi (Pass.) Jacz., Trudy Tiflissk. Bot. Sada 11: 143. 1910 [as “*terebinhti*”] = *Sphaerella terebinthi* Pass., Atti Reale Accad. Lincei, Mem. Cl. Sci. Fis., ser. 4, 4: 56. 1888.

Type — Italy: *Pistacia terebinthus* (Anacardiaceae).

No material was studied as the type was not found in any of the herbaria consulted.

Mycosphaerella ternstroemiae Petr. & Cif., Ann. Mycol. 30: 216. 1932.

Type — Dominican Republic: Manielito. On white spots with black margins on upper surface of living leaves of *Ternstroemia obovata* (Pentaphragmaceae). Ciferri no. 3073, II 1930 (W, holotype; NY, isotype).

In the types only a coelomycete was found.

Mycosphaerella tetraspora Seaver, Sci. Surv. Porto Rico & Virgin Islands 8: 62. 1926.— Fig. 883.

Type — Puerto Rico: On small pale spots with brown margins on upper and lower surface of living leaves of *Commelina elegans* (Commelinaceae). Seaver & Chardón no. 413, 1923 (NY, holotype).

This is a species of *Glomerella*, with ascocarp wall mostly pale, only blackened above, paraphyses simple, 1-2 µm wide, asci clavate, ascospores consistently 4 in the ascus, 12-16 × 5-6 µm. No species of *Glomerella* is so far known with these characters. Therefore the following combination is proposed: **Glomerella tetraspora** (Seaver) Aptroot comb. nov., **MB 500367**. **Basionym:** *Mycosphaerella tetraspora* Seaver, Sci. Surv. Porto Rico & Virgin Islands 8: 62. 1926.

Mycosphaerella tetroncii (Speg.) Cash, Syll. Fung. 26: 354. 1972 = *Sphaerella tetroncii* Speg., Bol. Acad. Nac. Ci. 27: 365. 1924.

Type — Argentina: *Tetroncium magellanicum* (Juncaginaceae).

No material was studied as the type was not included in a loan from LPS.

Mycosphaerella teucris (Unamuno) M. Morelet, Ann. Soc. Sci. Nat. Archéol. Toulon & Var 20: 106. 1968 = *Sphaerella teucris* Unamuno, Assoc. Esp. Progr. Ci. 1929: 13. 1929.

Type — Spain: León, Vegariensa. On dead, brown sori of *Puccinia annularis* on living leaves of *Teucrium scorodonia* (Lamiaceae). Agustin Melcón no. 8917, IX 1928 (MA, holotype).

In the type only a coelomycete was found.

Mycosphaerella thailandica Crous, Himaman & M.J. Wingf. in Crous *et al.*, Stud. Mycol. 50: 465. 2004.

Type — Thailand: Chachoengsao Prov., Sanamchaikhet, on leaves of *Acacia mangium* (Fabaceae). Pongpanich, V 2003 (CBS 9875, holotype); cultures ex-type CBS 116367 = CPC 10547–10549.

Anamorph: *Pseudocercospora thailandica* Crous, Himaman & M.J. Wingf. *vide* Crous *et al.* (op. cit.).

No material was studied of this recently described species.

Mycosphaerella thais (Sacc.) Negru & Benec, in Negru, Bull. Soc. Mycol. France 80: 319. 1964 = *Sphaerella thais* Sacc., Nuovo Giorn. Bot. Ital. 7: 305. 1875.— Fig. 884.

Type — Italy: *Scirpus triquetrus* (Cyperaceae). (IMI no. 132624, slide only, possible isotype).

The possible isotype shows that this belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradicans*, with asci cylindrical, ascospores 10–13 × 2–3 µm.

Mycosphaerella thalictri (Ellis & Everh.) Lindau, in Engl. & Prantl, Natürlichem Pflanzenf. 1(1): 424. 1897 = *Sphaerella thalictri* Ellis & Everh., J. Mycol. 1: 44. 1885.— Fig. 885.

Type — USA: New Jersey, Parsippany. On white spots with brown margins on upper and lower surface of living leaves of *Thalictrum dioicum* [“*cornuti*” on label] (Ranunculaceae). Trelease, VIII 1884 (NY, holotype); also Iowa, Decorah. Holway, VIII 1884 (NY, paratype).

Types and additional material (USA, New York, McLain, on *T. polygamum*, Higgins, VI 1912, distributed in Bartholomew, Fungi Columbiani no. 4080, IMI no. 16809) belong to section *Plaga*, with asci cylindrical, ascospores 10–12 × 3.5–5 µm.

Sphaerella thalictri Allesch. See *Mycosphaerella allescheri* (Sacc.) Lindau

Sphaerella thalictrocola Sacc. & P. Syd., Syll. Fung. 14: 534. 1899, nomen novum (Article 58) for *Sphaerella septorioides* Peck, Annual Rep. New York State Mus. 32: 52. 1879, later homonym (illegitimate, Article 53) = *Mycosphaerella pseudoseptorioides* Tomilin, Opredelitel' gribov roda *Mycosphaerella* Johans.: 217. 1979, superfluous (illegitimate, Article 52) nomen novum (Article 58).

Type — USA: *Thalictrum dioicum* (Ranunculaceae).

No material was studied as the type was not found in NY or BPI. Cited as synonymous with *Mycosphaerella thalictri* (Ellis & Everh.) Lindau by Barr (1972).

Mycosphaerella thalictrina Petr., Hedwigia 68: 212. 1928.— Fig. 886.

Type — Russia: Siberia, Southern Altaj. On dead stems of *Cimicifuga foetida* (Ranunculaceae). Antonov no. a124, VII 1925 (W, lectotype, here designated).

Cited as synonymous with *Mycosphaerella thalictri* (Ellis & Everh.) Lindau by Tomilin (1979). The lectotype belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradicans*, with asci cylindrical, ascospores 11–13 × 2.5–3.5 µm. The paratype on *Thalictrum simplex* was not found in W.

Sphaerella thallina Cooke, Grevillea 8: 10. 1879 = *Epicymatia thallina* (Cooke) Sacc., Syll. Fung. 1: 572. 1882 = *Pharcidia thallina* (Cooke) Lettau, Festschr. Preuss. Bot. Ver. 1912: 67. 1912 = *Lichenochora thallina* (Cooke) Hafellner, Nova Hedwigia 48: 363. 1989.

Type — United Kingdom: Eastbourne. On living thallus of *Phaeophyscia orbicularis* [“*Physcia obscura*”] (Ascomycota, Physciaceae). Muller, 1879 (K, holotype).

Accepted as *Lichenochora thallina* (Cooke) Hafellner by Hafellner (op. cit.), with which the type agrees.

Sphaerella thallophila (Cooke) Masee, Grevillea 19: 44. 1890 = *Sphaeria thallophila* Cooke, Handb. Brit. Fungi 2: 872. 1871 = *Epicymatia thallophila* (Cooke) Sacc., Syll. Fung. 1: 572. 1882.

Type — United Kingdom: *Lecanora subfusca* (Ascomycota, Lecanoraceae).

Cited as synonymous with *Anisomeridium bifforme* (Borrer) R.C. Harris by Hawksworth (2003) and therefore not studied.

Mycosphaerella thaspiicola H.C. Greene, Trans. Wisconsin Acad. Sci. 46: 153. 1957.

Type — USA: *Zizia aurea* [as “*Thaspium aureum*”] (Apiaceae).

Anamorph: *Cercospora thaspiicola* Davis *vide* Greene (op. cit.) (= *Passalora ziziae* (Ellis & Everh.) U. Braun & Crous).

No material was studied as the type was not found in NY or BPI.

Mycosphaerella theae Hara, Tea Journal 14: 9. 1919 = *Sphaerella theae* (Hara) Trotter, Syll. Fung. 24: 888. 1928.

Type — Japan. On white spots on upper surface of living leaves of *Camellia* [“*Thea*”] *sinensis* (Theaceae). Hara, VI 1918 (TNS 209272, holotype).

In type and additional material seen (Hara, V 1920, TNS 209271 & 209980) only a coelomycete could be found.

Mycosphaerella theissenii Tomilin, Novosti Sist. Nizsh. Rast. 8: 152. 1971, nomen novum (Article 58) for *Mycosphaerella plantaginis* (Ellis) Theiss., Ann. Mycol. 10: 196. 1912, later homonym (illegitimate, Article 53) = *Asterina plantaginis* Ellis, Bull. Torrey Bot. Club 9: 74. 1882 = *Asterella plantaginis* (Ellis) Sacc., Syll. Fung. 9: 398. 1891 = *Coleroa plantaginis* (Ellis) M.E. Barr, Can. J. Bot. 46: 833. 1968 = *Hormotheca plantaginis* (Ellis) Corlett & M.E. Barr, Mycotaxon 25: 256. 1986.— Fig. 887.

Type — USA: Pennsylvania, near Philadelphia. On ill-defined brown spots on upper surface of living leaves of *Plantago major* (Plantaginaceae). Rex, X 1881 (NY, holotype); also distributed in Ellis & Everhart, North American Fungi no. 791 (NY, 2 isotypes).

It was cited as synonymous with *Mycosphaerella plantaginis* (Sollmann) Vesterg. by Tomilin (1979), but is accepted as *Hormotheca plantaginis* (Ellis) Corlett & M.E. Barr by Corlett & Barr (op. cit.), with which the types agree well, with asci globose to pyriform, ascospores 9–11 × 3–4.5 µm.

Mycosphaerella thelypteridis Syd., Ann. Mycol. 19: 139. 1921.— Fig. 888.

Type — Germany: Brandenburg, Zossen. On lower surface of dead leaves of *Thelypteris palustris* [*Aspidium thelypteris*] (Polypodiaceae). Sydow, Mycotheca Germanica no. 1548, VI 1918 (L, 2 isotypes).

This is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores $8-10 \times 2-2.5 \mu\text{m}$.

Mycosphaerella theobromae (F.C. Faber) Tomilin, Novosti Sist. Nizsh. Rast. 6: 122. 1970 [“1969”] = *Sphaerella theobromae* F.C. Faber, Arbeiten Biol. Reichsanst. Land- - Forstw. 7: 219. 1909.

Type — Africa: On *Phytophthora* (Oomycota, Pythiaceae) on *Theobroma cacao* (Malvaceae).

No material was studied as the location of the type is unknown and its preservation uncertain.

Mycosphaerella theodulina (Unamuno) Tomilin, Novosti Sist. Nizsh. Rast. 1967: 190. 1967 = *Sphaerella theodulina* Unamuno, Bol. Soc. Esp. Hist. Nat. 29: 392. 1929.— Fig. 889.

Type — Spain: Burgos, La Vid. On dead stems of *Carex pendula* (Cyperaceae). Theodulus Asensio no. 8996, III 1928 (MA, holotype).

Cited as synonymous with *Mycosphaerella caricicola* by Tomilin (1979). The type shows that it belongs to *Davidiella*, and is morphologically indistinguishable from *D. disseminata*, with asci pyriform, ascospores $12-14 \times 3-4 \mu\text{m}$.

Mycosphaerella thermopsisidis Kalymb., Bot. Mater. Otd. Sporov. Rast. Bot. Inst. Akad. Nauk SSSR 14: 178. 1961 = *Phaeosphaerella thermopsisidis* (Kalymb.) Tomilin, Opredelitel' gribov roda *Mycosphaerella* Johans.: 285. 1979.

Type — Russia: *Thermopsis alpina* (Fabaceae).

No material was studied as the type was not included in loans from LE or LEP.

Mycosphaerella thesii (J. Schröt.) Tomilin, Novosti Sist. Nizsh. Rast. 1968: 168. 1968 = *Sphaerella thesii* J. Schröt., Hedwigia 29: 59. 1890.

Type — Yugoslavia: *Thesium intermedium* (Santalaceae).

No material was studied as the type was not found in any of the herbaria consulted.

Sphaerella thesii var. *pedemontana* Ferraris, Malpighia 16: 14. 1902.

Type — Italy: *Thesium linophyllum* (Santalaceae).

No material was studied as the type was not found in any of the herbaria consulted.

Mycosphaerella thironi Guyot, Bull. Soc. Mycol. France 62: 83. 1946.

Type — France: *Vicia sativa* (Fabaceae).

No material was studied as the type was not included in a loan from PC. Cited as synonymous with *Davidiella allicina* (as *Mycosphaerella allicina*) by Tomilin (1979).

Mycosphaerella thujae Petr., in Syd. & Petr., Ann. Mycol. 20: 179. 1922.

Type — USA: Idaho, Priest River. On dead scales of *Thuja plicata* (Cupressaceae). Weir & Rhoads, V 1929 (W, holotype).

Cited as synonymous with *Scirrhia conigena* (Peck) M.E. Barr by Barr (1972), with which the type agrees well, although it contains mostly coelomycetes.

Mycosphaerella thujopsisidis Sawada, Rep. Gov. Forest Exp. Sta. 46: 142. 1950.

Type — Japan: *Thujopsis dolabrata* (Cupressaceae).

No material was studied as the type was not found in BPI, nor in any of the other herbaria consulted.

Mycosphaerella thysselini (Kirschst.) Tomilin, Opredelitel' gribov roda *Mycosphaerella* Johans.: 52. 1979 = *Sphaerella thysselini* Kirschst., Kryptog.-Fl. Mark Brandenburg 7(3): 401. 1938.— Fig. 890.

Type — Germany: Rathenow, Gräninger See. On dead stems of *Peucedanum palustre* [*Thysselinum palustris*] (Apiaceae). Kirschstein, IX 1904 (B, holotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. ammophilae*, with asci pyriform, ascospores $8-10 \times 2.5-3 \mu\text{m}$. Additional material studied (Latvia, Vidzeme, Kastrane, Starcs no. 7607a, VIII 1939, B) agrees.

Mycosphaerella tilakii Tomilin. See *Mycosphaerella citricola* Tilak.

Mycosphaerella tiliae Naumov, Trudy Bjuro Prikl. Bot. 6: 201. 1913 = *Sphaerella tiliae* (Naumov) Trotter, Syll. Fung. 24: 889. 1928.

Type — Russia: Tula. On white spots on lower surface of living leaves of *Tilia* (Malvaceae). Naumov, VIII 1911 (LEP, holotype).

In the holotype only immature material is left.

Mycosphaerella tingens (Niessl) Lindau, Hilfsb. Sammeln Ascomyceten: 12. 1903 = *Sphaerella tingens* Niessl, in Rabenh., Fungi Europaei Exsiccati, ed. nov., ser. 2 no. 2848. 1882.— Fig. 891.

Type — Switzerland: Rhätische Alpen, Albulapas. On upper and lower surface of dead leaves of *Arenaria ciliata* (Caryophyllaceae). Winter, distributed in Rabenhorst, Fungi Europaei Exsiccati, ed. nov., ser. 2 no. 2848, VIII 1882 (L, isotype).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. superflua*, with ascomata turning the substratum reddish, asci cylindrical, ascospores $14-16 \times 3-5 \mu\text{m}$.

Sphaerella tingens f. *dianthicola* Gonz. Frag., Mem. Real Soc. Esp. Hist. Nat. 11: 88. 1919.— Fig. 892.

Type — Spain: Gerona, Llivia. On upper and lower surface of dead leaves of *Dianthus deltooides* (Caryophyllaceae). Sennen no. 3631, VII 1918 (MA, holotype).

This is morphologically indistinguishable from *Wettsteinina gigaspora* Höhn., with ascospores thick-walled, with 3 incomplete septa, ca. $70 \times 25 \mu\text{m}$.

Sphaerella tini Arcangeli, Erbario Crittogamico Italiano, ser. 2 no. 822. 1879.

Type — Italy: Firenze (Florence). On white spots on upper surface of living leaves of *Viburnum tinus* (Adoxaceae). Arcangeli, in Erbario Crittogamico Italiano no. 822, XI 1878 (L, isotype).

In the isotype studied only coelomycetes were present.

Mycosphaerella tinosporae Ajrekar ex Mundkur, Sci. Monogr. Council Agric. Res. India 12: 15. 1938.

Type — India: Ahmadabad. On lower surface of living leaves of *Tinospora cordifolia* (Menispermaceae). Ajrekar, V 1934 (NY, isotype).

Anamorph: *Cercospora tinosporae* (Lacy & Thirum.) Deighton *vide* Deighton (1973).

The type and additional material seen (India, Madhya Pradesh, Sagar, University Campus, Gupta, X 1988, IMI no. 330902) contain only the anamorph.

Mycosphaerella tirolensis (Auersw.) Magnus, in Dalla Torre & Sarnth., Fl. Tirol, Vorarlberg, Liechtenstein, III, Pilze: 463. 1905 = *Sphaerella tirolensis* Auersw., in Gonn. & Rabenh., Mycol. Europaea 5-6: 20. 1869 [also as “*tyrolensis*”].

Type — Germany: On upper and lower surface of dead fronds of *Polypodium vulgare* (Polypodiaceae).

Material studied (France, Haute-Garonne, Luchon, Fourcade & Roumeguère, distributed in Roumeguère, Fungi Gallici Exsiccati no. 2935, IV 1884, L) contains only empty ascomata.

Sphaerella tirolensis var. *allosoricola* Carestia, in Bagl., Ces. & De Not., Erbario Crittogamico Italiano, ser. 2, fasc. 10 no. 493. 1871.— Fig. 893.

Type — Italy: Valsesia, Riva. On upper and lower surface of dead fronds of *Allosorus crispus* (Polypodiaceae). Carestia, 1870, distributed in Erbario Crittogamico Italiano, ser. 2 no. 493 (BPI, isotype).

This belongs to section *Longispora*, and is morphologically indistinguishable from *M. millegrana*, with asci cylindrical, ascospores 13-16 × 2-2.5 μm.

Sphaerella tirolensis var. *montellica* Sacc., Michelia 1: 34. 1879 [as “*tyrolensis*”].

Type — Italy: *Polypodium vulgare* (Polypodiaceae).

No material was studied as the type was not found in any of the herbaria consulted.

Mycosphaerella tithymali (Pass.) Tomilin, Novosti Sist. Nizsh. Rast. 6: 123. 1970 [“1969”] = *Sphaerella tithymali* Pass., Atti Reale Accad. Lincei, Mem. Cl. Sci. Fis., ser. 4, 3: 8. 1887.

Type — Italy: *Euphorbia cyparissias* (Euphorbiaceae).

No material was studied as the type was not found in any of the herbaria consulted.

Mycosphaerella tocoyenae Bat. & Peres, Publ. Inst. Micol. Univ. Fed. Pernambuco 268: 12. 1960.

Type — Brazil. On black spots on upper surface of living leaves of *Tocoyena brasiliensis* (Rubiaceae). Batista exs. no. 14022 (URM 19151, holotype).

The holotype material is overmature, too old to identify.

Sphaerella todeae Cooke, Grevillea 12: 85. 1884 = *Sphaerulina todeae* (Cooke) Berl. & Voglino, Syll. Fung., Addit. ad vol. 1-4: 159. 1886.

Type — New Zealand: Karori. On fronds of *Todea hymenophylloides* (Osmundaceae). I 1879 (K, holotype).

In the holotype only a coelomycete could be found.

Mycosphaerella togashiana K. Itô & T. Kobay., Bull. Gov. Forest Exp. Sta. 59: 23. 1953.

Type — Japan: *Populus simonii* (Salicaceae).

Anamorphs: *Pseudocercospora salicina* (Ellis & Everh.) Deighton *vide* Tomilin (1979) and Sivanesan (1984).

No material was studied as the type was not included in a loan from TNS.

Mycosphaerella togniniana (Traverso) Woronow, Trudy Tiflisk. Bot. Sada, ser. 2, 3: 140. 1922 = *Sphaerella togniniana* Traverso, Fl. Ital. Cryptog. 1(11): 601. 1913, nomen novum (Article 58) for *Sphaerella trifolii* var. *umbelliferarum* Tognini, Atti Ist. Bot. Univ. Pavia, n.s. 5: 5. 1895, non *Sphaerella umbelliferarum* Rabenh. (1866).

Type — Italy: *Foeniculum vulgare* (Apiaceae).

No material was studied as the type was not found in any of the herbaria consulted. Cited as synonymous with *Davidiella allicina* (as *Mycosphaerella allicina*) by Tomilin (1979).

Mycosphaerella tokoroii Hara, J. Pl. Protect. 5: 537. 1918.

Type — Japan.

No material was studied as the type was not included in a loan from TNS.

Mycosphaerella toledana Crous & G. Bills in Crous *et al.*, Stud. Mycol. 50: 208. 2004.

Type — Spain: Toledo, on leaves of *Eucalyptus* sp. (Myrtaceae). Crous & Bills, V 2003 (CBS 9896, holotype); culture ex-type CBS 113313.

Anamorph: *Phaeophleospora toledana* Crous & G. Bills *vide* Crous *et al.* (op. cit.).

No material was studied of this recently described species.

Mycosphaerella tomlinii Akhundov, Izv. Akad. Nauk Azerb. SSR, Ser. Biol. Med. Nauk 1971(3): 3. 1971 [as “*tomilinae*”].

Type — Azerbaijan: *Erodium turkmenum* (Geraniaceae).

No material was studied as the type was not included in loans from LE or LEP. This is no *Mycosphaerella* according to Tomilin (1979).

Mycosphaerella topographica (Sacc. & Speg.) Vestergr., Bot. Not. 1897: 267. 1897 = *Sphaerella topographica* Sacc. & Speg., Michelia 1: 380. 1878 = *Carlina topographica* (Sacc. & Speg.) Weese, Eumycetes Selecti Exsiccati no. 405. 1930.— Fig. 894.

Type — Italy: Veneto, Conegliano. On lower and upper surface of dead leaves of *Sorbus torminalis* (Rosaceae).

Saccardo, 1878, distributed in Mycotheca Veneta no. 1368 (BPI, 2 isotypes).

Anamorph: Probably *Septoria hyalospora* (Mont. & Ces.) Sacc. *fide* Saccardo (1882).

The isotypes are overmature. Additional material studied (Czech Republic, Weißkirchen, on *S. aucuparia*, Petrak, distributed in Flora Bohemiae et Moraviae Exsiccata no. 622, I 1913, L) shows that this belongs to section *Longispora*, with asci cylindrical, ascospores 21-26 × 3.5-4.5 µm.

Mycosphaerella tormentillae (Sacc.) Tomilin, Opredeletel' gribov roda *Mycosphaerella* Johans.: 225. 1979 ≡ *Sphaerella ariadna* subsp. *tormentillae* Sacc., Michelia 1: 379. 1878 ≡ *Sphaerella tormentillae* (Sacc.) Traverso, Fl. Ital. Cryptog. 1(11): 552. 1913.

Type — Italy: On pale spots on upper surface of living leaves of *Potentilla* ["*Tormentilla*"] *erecta* (Rosaceae).

The type was not found in any of the herbaria consulted. Material studied (Bayern, Eisenstein, on *Potentilla erecta* ["*Potentilla tormentilla*"], Kirschstein, VIII 1939, B) contains only coelomycetes.

Sphaerella tortulae Bubák & Gonz. Frag., Hedwigia 57: 5. 1915.— Fig. 895.

Type — Spain: Badaios, Villalba de los Barros. On setae of *Tortula laevipila* [wrongly identified as "*pulvinata*"] (Musci, Pottiaceae). Casares no. 116, IV 1914 (MA, holotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. ammophilae*, with asci pyriform, ascospores 9-11 × 3.5-5 µm.

Mycosphaerella tournefortiae Petr. & Cif., Ann. Mycol. 28: 392. 1930.

Type — Dominican Republic: *Tournefortia hirsutissima* (Boraginaceae).

No material was studied as the type was not found in BPI or NY nor included in a loan from W.

Mycosphaerella trachycarpi Rehm, Ann. Mycol. 11: 399. 1913 [as "*trochicarp*"] ≡ *Sphaerella trachycarpi* (Rehm) Trotter, Syll. Fung. 24: 870. 1928 [as "*trochicarp*"].— Fig. 897.

Type — Georgia: Batumi. On brown spots with black margins on lower surface of dead leaves of *Trachycarpus* ["*Trochicarpus*"] (Arecaceae). Nevodovski, II 1912 (BPI, isotype).

Synonymised with *Mycosphaerella lineolata* by von Arx (1949). however, the type shows that this is a parasitic species, with asci cylindrical, ascospores 14.5-18 × 3.5-4.5 µm.

Mycosphaerella tragopogonicola Petr., Hedwigia 68: 213. 1928 ≡ *Sphaerella tragopogonicola* (Petr.) Kirschst., Kryptog.-Fl. Mark Brandenburg 7(3): 402. 1938.— Fig. 896.

Type — Russia: Siberia, Central Altaj, Mujuta. On upper and lower surface of dead leaves of *Tragopogon pratensis* (Asteraceae). Antonov no. A86, VIII 9125 (W, holotype; LEP, W, isotypes).

The types are all immature. Additional material studied (Germany, Brandenburg, Tamsel, on dead stems of *Tragopogon pratensis*, Vogel, distributed in Sydow, Mycotheca Germanica no. 2334, II 1909, L) belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores 16-19 × 5.5 µm.

Mycosphaerella tremulae Aderh., nomen herbariorum (not validly published, Article 32).— Fig. 898.

Authentic material — Germany: Westfalen, Siegen, Walpersdorf. On lower surface of dead leaves of *Populus tremula* (Salicaceae). Ludwig, IV 1924 (B).

This is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores 7-9 × 2.5-3 µm.

Mycosphaerella tremulicola (Fr.) Petr., Ann. Mycol. 38: 248. 1940 ≡ *Sphaeria tremulicola* Fr., Systema Mycol. 2: 529. 1823 [as "*tremulaecola*"].

Type — Sweden: *Populus tremula* (Salicaceae).

No material was studied as the type was not included in a loan from UPS. Cited as synonymous with *M. punctiformis* by Tomilin (1979).

Mycosphaerella tremulina (Mouton) Lindau, Hilfsb. Sammeln Ascomyceten: 86. 1903 ≡ *Sphaerella tremulina* Mouton, Bull. Soc. Roy. Bot. Belgique 11: 40. 1900.— Fig. 899.

Type — Belgium: Beaufay. On upper and lower surface of dead leaves of *Populus tremula* (Salicaceae). VI 1899 (BR, holotype).

Already cited as synonymous with *M. punctiformis* by Tomilin (1979), with which the holotype agrees well, with asci cylindrical, ascospores 7-9 × 2-3 µm.

Mycosphaerella trichomanes (Cooke) Lindau, in Engl. & Prantl, Natürlichen Pflanzenf. 1(1): 426. 1897 [as "*trichomanis*"] ≡ *Sphaerella trichomanes* Cooke, Grevillea 14: 12. 1885.— Fig. 900.

Type — Samoa Islands. On upper surface of living fronds of *Trichomanes* (Hymenophyllaceae). Müller (K, holotype). The holotype contains two lichenized ascomycetes, viz. *Strigula phyllogena* (Müll. Arg.) R.C. Harris, of which this is a later synonym, and *Porina albicera* (Krempel.) van Overeem-de Haas. See Aptroot & Lücking (2001) for a description and more details.

Sphaerella trichophila P. Karst., Bidrag Kännedom Finlands Natur Folk 23: 171. 1873.— Fig. 901.

Type — Finland: On upper surface of dead leaves of *Saussurea alpina* (Asteraceae). (K, isotype).

Synonymised with *Davidiella allicina* (as *Mycosphaerella tassiana*) by von Arx (1949), but material seen (Finland, Lappland, Enontekiö, Porojärvet, Jukassjärvi, Ollila & Roivainen, VII 1955, IMI no. 175292, topotype) belongs to section *Caterva*, and suggests that this is morphologically indistinguishable from *M. subradians*, with asci cylindrical, ascospores 10-12 × 2.5-3.5 µm.

Sphaerella trichophila var. *saxifragae* Dearn., Mycologia 9: 346. 1917.

Type — USA: *Saxifraga nelsoniana* (Saxifragaceae).
No material was studied as the type was not found in BPI or NY.

Mycosphaerella trifolii (P. Karst.) Jacz., Gribnye i Bakterial'nya Bolezni Klevera: 59. 1916 = *Sphaerella trifolii* P. Karst., Bidrag Kännedom Finlands Natur Folk 23: 174. 1873.

Type — Finland: *Trifolium pratense* (Fabaceae).
No material was studied as the type was not included in a loan from H. Cited as synonymous with *Davidiella allicina* (as *Mycosphaerella allicina*) by Tomilin (1979).

Mycosphaerella trifolii f. *trifolii-tomentosi* (Gonz. Frag.) Maire & Werner, Mém. Soc. Sci. Nat. Maroc 45: 28. 1937 = *Sphaerella trifolii* f. *trifolii-tomentosi* Gonz. Frag., Mem. Real Soc. Esp. Hist. Nat. 8: 339. 1916.— Fig. 902.

Type — Morocco: Melilla Gurugia. On upper and lower surface of dead leaves of *Trifolium tomentosum* (Fabaceae). Caballero no. 1418, VI 1915 (MA, holotype).

This is a species of *Wettsteinina*, with ascospores 8/ascus, pale brownish, with 3 complete septa, ca. $37 \times 17 \mu\text{m}$.

Sphaerella trifolii var. *umbelliferarum* Tognini. See *Mycosphaerella togniniana* (Traverso) Woronow.

Mycosphaerella triseti (Speg.) Tomilin, Novosti Sist. Nizsh. Rast. 1967: 190. 1967 = *Sphaerella triseti* Speg., Bol. Acad. Nac. Ci. 11: 208. 1888 ["1887"].

Type — Argentina: *Trisetum* (Poaceae).
No material was studied as the type was not included in a loan from LPS. Cited as synonymous with *Mycosphaerella parallelogramma* by Tomilin (1979).

Sphaerella triseti Speg., Anales Mus. Nac. Hist. Nat. Buenos Aires 8 (or ser. 3, 1): 70. 1902, later homonym (illegitimate, Article 53).

Type — Argentina: *Trisetum* (Poaceae).
No material was studied as the type was not included in a loan from LPS.

Mycosphaerella tristaniae Wakef., Bull. Misc. Inform. 1922: 163. 1922.— Fig. 903.

Type — Malaysia: Pulau Penang, Penang Hill. On reddish spots on lower surface of living leaves of *Tristania griffithii* (Myrtaceae). Chipp no. 4694, VIII 1919 (K, holotype).

Although this was reported not to belong to *Mycosphaerella* by Crous (1998), examination of the type shows that this belongs to *Davidiella*, of which it represents a parasitic species, and is morphologically indistinguishable from *D. myrticola*, with asci pyriform, ascospores $14-17 \times 4-5 \mu\text{m}$.

Mycosphaerella trochicarpi Rehm. See *Mycosphaerella trachycarpi* Rehm.

Mycosphaerella tsugae (House) House, New York State Mus. Bull. 233-234: 31. 1921 = *Sphaerella tsugae* House, New York State Mus. Bull. 205-206: 40. 1919 ["1918"] = *Delphinella tsugae* (House) M.E. Barr, Contr. Univ. Michigan. Herb. 9: 563. 1972.— Fig. 904.

Type — USA: New York, Albany Co. On dead scales of cones of *Tsuga canadensis* (Pinaceae). House, IV 1945 (BPI, topotype).

Already synonymised with *Delphinella peckii* (Lindau) M.E. Barr by Barr *et al.* (1986), with which the topotype and additional material studied (Canada, Ontario, Bolton, Cain no. 34147, V 1955, L) agrees, with ascocarps with schizolytic, linear ostioles, asci polysporous, ascospores 8-9.5 (in topotype) or 11-13 (in Cain no. 34147) \times 3-3.5 μm .

Mycosphaerella tuerckheimii Petr., Sydowia 16: 240. 1963 ["1962", as "*Türkheimii*"].— Fig. 905.

Type — Dominican Republic: Constanza, Vallenueva. On upper and lower surface of dead leaves of *Siphocampylus urbanus* (Campanulaceae). Von Tuerckheim, VI 1910 (BPI, isotype).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradicans*, with asci cylindrical, ascospores $10.5-13 \times 2.5-3 \mu\text{m}$.

Mycosphaerella tulasnei (Jancz.) Lindau, Hilfsb. Sammeln Ascomyceten: 111. 1903 = *Sphaerella tulasnei* Jancz., Bull. Int. Acad. Sci. Cracovie 27: 187. 1894.— Fig. 906.

Type — France: On dead leaves of *Secale cereale* (Poaceae).

Anamorphs: *Cladosporium herbarum* (Pers. : Fr.) Link and *Cladosporium fasciculare* (Pers. : Fr.) Fr. *fide* Janczewski (op. cit.).

Topotype material (Lorraine, Forbach, Stieringen, Wendel, Ludwig, VII 1914, B, topotype) belongs to *Davidiella*, and shows that this is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores $18-20 \times 5.5-7 \mu\text{m}$. Other material (Russia, Smolensk, Gzhatsk, Janczewski, BPI) contains only the anamorphs. Additional material studied (USA, California, Marin County, San Francisco, Tiburon, Howell Botanical Garden, on *Calamagrostis ophitidis*, Howell, V 1966, IMI no. 142274) is postmature.

Mycosphaerella tulipifera (Schwein.) B.B. Higgins, Amer. J. Bot. 23: 598. 1936 = *Sphaeria tulipifera* Schwein., Trans. Amer. Philos. Soc., ser. 2, 4: 226. 1832 = *Sphaerella tulipifera* (Schwein.) Cooke, J. Bot. 21: 108. 1883.

Type — USA: Bethlehem. On lower surface of dead leaves of *Liriodendron tulipifera* (Magnoliaceae). 1828 (PH, holotype; PH, isotype).

Spermatial state and anamorph: *Asteromella fide* Tomilin (1979) and *Phaeoisariopsis liriodendri* (Ellis & Harkn.) Morgan-Jones & Brown *fide* Morgan-Jones & Brown (1976) (= *Passalora liriodendri* (Ellis & Harkn.) U. Braun & Crous).

The types studied contain only coelomycetes, including the *Asteromella*.

Mycosphaerella tungurahua Petr., Sydowia 4: 507. 1950.— Fig. 907.

Type — Ecuador: Tungurahua, Baños. On spots on upper surface of living leaves of *Eupatorium inulaefolium* (Asteraceae). Syd., distributed in Reliquiae Petrakianae no. 662, XII 1937 (H, L, isotypes).

This is a species of *Didymella*, with paraphysoids and thick, short paraphysoids, asci cylindrical, ascospores $13-16 \times 3.5-5 \mu\text{m}$. The isotype in L contains only empty ascospores.

Mycosphaerella tupae (Speg.) Tomilin, Novosti Sist. Nizsh. Rast. 8: 152. 1971 \equiv *Sphaerella tupae* Speg., Revista Fac. Agron. Univ. Nac. La Plata, ser. 2, 6: 58. 1910.

Type — Chile: *Lobelia salicifolia* (Lobeliaceae).

No material was studied as the type was not included in a loan from LPS.

Sphaerella turba Fuckel, Jahrb. Nassauischen Vereins Naturk. 23-24: 103. 1870.

Type — Germany: Oestrich. On lower surface of dead leaves of *Salix aurita* (Salicaceae). Fuckel, Fungi Rhenani Exsiccati no. 1577 (BPI, L, isotypes [as “*Sphaeria*”]).

This is no *Mycosphaerella*, but an immature ascomycete with ascospores superficial, conical, asci pyriform, ascospores multiseptate.

Mycosphaerella turckheimii Petr. See *Mycosphaerella tuerckheimii* Petr.

Mycosphaerella tussilaginis (Rehm) Lindau, in Engl. & Prantl, Natürlichen Pflanzenf. 1(1): 424. 1897 \equiv *Sphaerella tussilaginis* Rehm, Ascomyceten no. 100. 1872.— Fig. 908.

Type — Germany: Franken, Windsheim. On upper surface of dead leaves of *Tussilago farfara* (Asteraceae). Rehm, Ascomyceten no. 100, IV 1872 (L, isotype).

The type and additional material studied (Czech Republic, Weißkirchen, Petrak, distributed in Cryptogamae Vindobonenses no. 3269, V, L) belong to section *Caterva*, and show that this is morphologically indistinguishable from *M. superflua*, with asci cylindrical, ascospores $15-18 \times 4.5-5.5 \mu\text{m}$. Additional material from the Netherlands (Apeldoorn, 1897, L) contains only a coelomycete.

Sphaerella tussilaginis f. *tussilaginis-farfarae* J. Kunze, Fungi Selecti Exsiccati no. 248. 1879.

Type — Germany: Sachsen, Eisleben. On upper surface of dead leaves of *Tussilago farfara* (Asteraceae). Kunze, Fungi Selecti Exsiccati no. 248, VI 1879 (L, holotype).

This is morphologically indistinguishable from *Pleospora herbarum* (Pers. : Fr.) Rabenh.

Mycosphaerella typhae J. Schröt., in Cohn, Kryptog.-Fl. Schlesien 3(2): 342. 1894 [“1893”] \equiv *Monascostroma typhae* (J. Schröt.) Munk, Dansk Bot. Ark. 15(2): 108. 1953, full reference to basionym not given (not validly published, Article 33).— Fig. 909.

Type — Germany: *Typha angustifolia* (Typhaceae).

Accepted as *Monascostroma typhae* (J. Schröt.) Munk by Munk (1957). Material studied (On upper and lower surface of dead leaves of *Typha latifolia*, Lütjeharms, L) agrees, with asci pyriform, ascospores only immature, $12-16 \times 3.5-4.5 \mu\text{m}$.

Mycosphaerella typhae (Lasch) Lindau, in Engl. & Prantl, Natürlichen Pflanzenf. 1(1): 425. 1897, later homonym (illegitimate, Article 53) \equiv *Sphaeria typhae* Lasch, in Rabenh., in Klotzsch, Herbarium Vivum Mycologicum,

cent. 7 no. 660. 1844 \equiv *Sphaeria perpusilla* var. *typhae* (Lasch) Auersw., in Rabenh., Fungi Europaei Exsiccati, ed. 3, no. 831 \equiv *Sphaerella typhae* (Lasch) Auersw., in Gonn. & Rabenh., Mycol. Europaea 5-6: 18. 1869 \equiv *Phaeosphaerella typhae* (Lasch) Sacc., Syll. Fung. 11: 312. 1895.— Fig. 910. Type — Germany: Driesen. On upper and lower surface of dead leaves of *Typha latifolia* (Typhaceae). Lasch, distributed in Rabenhorst, in Klotzsch, Herbarium Vivum Mycologicum no. 660 (L, isotype).

This is morphologically indistinguishable from *Phaeosphaeria eustoma* (Fuckel) L. Holm, with ascospores 3-septate, brown, $16-20 \times 4-5 \mu\text{m}$. Additional material (Dresden, Moritzbürg, Auerswald, distributed in Rabenhorst, Fungi Europaei Exsiccati, ed. 3, no. 831, B) agrees.

Mycosphaerella typhaii Ponnappa. See *Mycosphaerella typhina* Ponnappa.

Sphaerella typharum (Desm.) Rabenh., Fungi Europaei Exsiccati, ed. nov., ser. 2, cent. 11 no. 1040. 1860 \equiv *Sphaeria scirpicola* var. *typharum* Desm., Plantes Cryptogames de France, ed. 2 no. 1428 \equiv *Sphaeria typharum* (Desm.) Rabenh., Klotzschii Herbarium Vivum Mycologicum, ed. nov. no. 731. 1858.

Type — France. On dead leaves of *Typha* (Typhaceae). Desmazières, Plantes Cryptogames de France no. 1428 (B, isotype).

This is morphologically indistinguishable from *Phaeosphaeria eustoma* (Fuckel) L. Holm, with ascospores 3-septate, brown, $15-20 \times 3.5-5 \mu\text{m}$. Additional material studied (Germany, Leipzig, Auerswald, distributed in Rabenhorst, in Klotzsch, Herbarium Vivum Mycologicum no. 731, IV 1857, B) agrees.

Sphaerella typharum f. *sparganii* Niessl, in Rabenh., Fungi Europaei Exsiccati, ed. nov., ser. 2: cent. 11 no. 1040. 1862.

Type — Czech Republic: Brno (Brünn). On upper and lower surface of dead leaves of *Sparganium ramosum* (Sparganiaceae). Niessl, distributed in Rabenhorst, Fungi Europaei Exsiccati ser. 2, no. 1040, VIII 1862 (L, M, isotypes).

In the isotype studied only an *Ascochyta* was present. The original illustration suggests a *Lophiostoma* or *Phaeosphaeria*.

Mycosphaerella typhina Ponnappa, Proc. Indian Acad. Sci. 8: 178. 1969 \equiv *Mycosphaerella typhaii* Ponnappa, nomen herbariorum (not validly published, Article 32).

Type — India: Mysore State, Kengeri. On dead [“living” on label] leaves of *Typha angustata* (Typhaceae). Ponnappa, IX 1966 (IMI no. 131896, isotype, sub “*Mycosphaerella typhaii*”).

The isotype, probably the only material still in existence, contains only a coelomycete.

Mycosphaerella tyrolensis (Auersw.) Magnus. See *Mycosphaerella tirolensis* (Auersw.) Magnus.

Sphaerella uliginosa Phillips & Plowr., Grevillea 10: 74. 1881 \equiv *Leptosphaeria uliginosa* (Phillips & Plowr.) Sacc., Syll. Fung. 2: 47. 1883.

Type — United Kingdom: *Stellaria uliginosa* (Caryophyllaceae). (K, isotype).

Accepted as *Leptosphaeria uliginosa* by Saccardo (op. cit.), and therefore type material not studied in detail.

Mycosphaerella ulmariae Kirschst., Ann. Mycol. 34: 187. 1936 \equiv *Sphaerella ulmariae* (Kirschst.) Kirschst., Kryptog.-Fl. Mark Brandenburg 7(3): 358. 1938.

Type — Germany: Niederbayern, Eisenstein, Arberhütte. On dead stems of *Filipendula ulmaria* [*“Ulmaria pentapetala”*] (Rosaceae). Kirschstein, VII 1936 (B, holotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. ammophilae*, with asci pyriform, ascospores 7-9 \times 2.5-3 μ m. Topotype material studied (Bayerisch Häusl, Kirschstein, VII 1937, B, topotype) agrees.

Mycosphaerella ulmi Kleb., Z. Pflanzenkrankh. 12: 257. 1902 \equiv *Sphaerella ulmi* (Kleb.) Sacc. & D. Sacc., Syll. Fung. 17: 642. 1905.— Fig. 911.

Type — Germany: On upper and lower surface of dead leaves of *Ulmus montana* var. *pendula* (Ulmaceae).

Anamorphs: *Phloeospora ulmi* (Fr.) Wallr. and *Phyllosticta bellunensis* Martelli *vide* Sivanesan (1984). According to Tomilin (1979) also reported as associated with *Cercospora klebahniana* Potebnia (= *Phloeospora ulmi* (Fr.) Wallr.) and *Cylindrosporium ulmicola* Jacz.

Topotype material studied (Hamburg, Ohlsdorf, Klebahn, distributed in Jaap, Fungi Selecti Exsiccati no. 376B, IV 1906, L) belongs to section *Longispora*, and is morphologically indistinguishable from *M. topographica*, with asci cylindrical, ascospores 23-28 \times 3.5-5 μ m.

Mycosphaerella ulmifolia (Pass.) Tomilin, Novosti Sist. Nizsh. Rast. 1968: 168. 1968 [as *“ulmifoliae”*] \equiv *Sphaerella ulmifolia* Pass., in Erbario Crittogamico Italiano, ser. 2 no. 1281. 1883.

Type — Italy: Parma. On small white spots with brown margins on upper surface of living leaves of *Ulmus campestris* (Ulmaceae). Passerini, XI 1882, distributed in Erbario Crittogamico Italiano, ser. 2 no. 1281 (NY, isotype). Anamorph: *Hendersonia ulmifolia* Pass. *vide* Saccardo (1891).

The isotype contains only coelomycetes.

Mycosphaerella umbelliferarum (Rabenh.) Lindau, in Engl. & Prantl, Natürlichen Pflanzenf. 1(1): 424. 1897 \equiv *Sphaerella umbelliferarum* Rabenh., Fungi Europaei Exsiccati, ed. nov., ser. 2 no. 1041. 1862 \equiv *Leptopacidium umbelliferarum* (Rabenh.) Höhn., Sitzungsber. Kaiserl. Akad. Wiss., Math.-Naturwiss. Kl., Abt. 1, 127: 331. 1918 \equiv *Guignardia umbelliferarum* (Rabenh.) Petr., Ann. Mycol. 19: 106. 1921 \equiv *Discosphaerina umbelliferarum* (Rabenh.) Petr., Ann. Mycol. 22: 36. 1924.— Fig. 912.

Type — Germany: Dresden. On dead stems of *Peucedanum oreoselinum* (Apiaceae). Rabenhorst, Fungi Europaei Exsiccati, ser. 2 no. 1041, 1862 (L, 2 isotypes).

Spermatial state: *Asteromella chaerophylli* (C. Massal.) Petr. *vide* Tomilin (1979).

Cited as synonymous with *Guignardia foeniculacea* (Mont.) von Arx & E. Müll. by von Arx & Müller (1954), with which the isotype studied agrees well, with asci clavate, ascospores aseptate, 14-16 \times 4.5-5.5 μ m.

Mycosphaerella umbellulariae (Cooke & Harkn.) Lindau, in Engl. & Prantl, Natürlichen Pflanzenf. 1(1): 425. 1897 \equiv *Sphaerella umbellulariae* Cooke & Harkn., Grevillea 13: 21. 1884.— Fig. 913.

Type — USA: California, Jamalpais. On dark spots on upper surface of living leaves of *Umbellularia californica* (Lauraceae). Harkness, distributed in Ellis & Everhart, North American Fungi no. 1680 (L, isotype).

This is a parasitic species, with asci cylindrical, ascospores 13-16 \times 4-5 μ m.

Sphaerella umbrosa Sacc., Michelia 1: 378. 1878.— Fig. 914.

Type — Italy: On pale spots on upper surface of living leaves of *Galeopsis versicolor* (Lamiaceae). Saccardo (PAD, holotype).

Anamorph: Associated with *Phyllosticta galeopsidis* Sacc. *vide* Saccardo (1882).

This is a parasitic species, with asci cylindrical, ascospores 10-12 \times 4-5 μ m. Cited as synonymous with *Mycosphaerella rubella* by Tomilin (1979), but instead morphologically indistinguishable from *Mycosphaerella caulicola*.

Mycosphaerella unedinis Jaap, Ann. Mycol. 14: 13. 1916 \equiv *Sphaerella unedinis* (Jaap) Trotter, Syll. Fung. 24: 861. 1928.— Fig. 915.

Type — Yugoslavia: *Arbutus unedo* (Ericaceae).

The type was not found in any of the herbaria consulted. Cited as synonymous with *Mycosphaerella arbuticola* by Tomilin (1979). Material studied (Greece, Thessaloniki, Agia, on upper and lower surface of dead leaves of *Arbutus andrachne*, Reching, distributed in Reliquiae Petrakianae no. 2256, V 1961, L) is *M. punctiformis*, with asci cylindrical, ascospores 8-10 \times 2-2.5 μ m.

Mycosphaerella ungnadiae Siemaszko, Bull. Musée du Caucase 12: 20. 1919 \equiv *Sphaerella ungnadiae* (Siemaszko) Trotter, Syll. Fung. 24: 1334. 1928 [as *“unguadiae”*].

Type — Georgia: *Ungnadia speciosa* (Sapindaceae).

No material was studied as the location of the type is unknown.

Sphaerella urticae Losa, Anales Jard. Bot. Madrid 5: 99. 1945, lacking Latin description (not validly published, Article 36).

Type — Spain: *Urtica dioica* (Urticaceae).

No material was studied as the type was not found in MA and is probably lost.

Mycosphaerella urticae-dioicae Tomilin, Novosti Sist. Nizsh. Rast. 10: 97. 1973.

Type — Russia: Leningrad. On dead stems of *Urtica dioica* (Urticaceae). Gramenitzkaja, V 1925 (LEP, holotype).

In the holotype only immature material is left and a *Phoma* anamorph. However, it is morphologically indistinguishable from *M. superflua*.

Mycosphaerella ushuvaiensis (Speg.) Tomilin, Novosti Sist. Nizsh. Rast. 8: 152. 1971 = *Sphaerella ushuvaiensis* Speg., Bol. Acad. Nac. Ci. 11: 208. 1888 ["1887"].

Type — Argentina: *Galium antarcticum* (Rubiaceae).

No material was studied as the type was not included in a loan from LPS.

Mycosphaerella uspenskajae Mashkina & Tomilin, Mikol. Fitopatol. 20: 367. 1986.

Type — Russia: *Glycine max* (Fabaceae).

No material was studied as the type was not included in loans from LE or LEP.

Mycosphaerella usteriana (Speg.) Hara, Diseases of the Rice Plant: 139. 1918 = *Sphaerella usteriana* Speg., Revista Mus. La Plata 15 (or ser. 2, 2): 20. 1908.— Fig. 916.

Type — Brazil: On dead leaves of *Oryza sativa* (Poaceae).

The type was not included in a loan from LPS. Material seen (Korea, intercepted at Los Angeles, Koerber, I 1981, BPI) belongs to *Davidiella*, and is morphologically indistinguishable from *D. disseminata*, with asci pyriform, ascospores 11-13 × 3-4 μm.

Mycosphaerella vaccinii (Cooke) J. Schröt., in Cohn, Kryptog.-Fl. Schlesien 3(2): 335. 1894 ["1893"] = *Sphaerella vaccinii* Cooke, J. Bot. 4: 249. 1866 = *Sphaerella myrtilli* Auersw., in Gonn. & Rabenh., Mycol. Europaea 5-6: 10. 1869, superfluous (illegitimate, Article 52) nomen novum (Article 58) = *Carlia vaccinii* (Cooke) Höhn., Hedwigia 62: 39. 1921.— Fig. 917.

Type — United Kingdom: Surrey, Shere. On upper and lower surface of dead leaves of *Vaccinium myrtilus* (Ericaceae). Cooke, IV 1866 (K, holotype), also distributed in Fungi Britannici Exsiccati no. 176 (B, 2 isotypes; K, isotype).

Type and other material studied (Germany, Brandenburg, Grafenbrück, Sydow, Mycotheca Germanica no. 1549, V 1918, L) belong to section *Longispora*, and show that this species is morphologically indistinguishable from *M. millegrana*, with asci cylindrical, ascospores 15-18 × 2-3 μm.

Sphaerella vaccinii Fuckel, Jahrb. Nassauischen Vereins Naturk. 23-24: 106. 1870, later homonym (illegitimate, Article 53).

Type — Germany. On upper surface of living leaves of *Vaccinium uliginosum* (Ericaceae). Fuckel, Fungi Rhenani no. 1779 (B, isotype, sub "*Sphaeria vaccinii*").

According to Sivanesan (1984) this is a synonym of *Gibbera myrtilli* (Cooke) Petr. The isotype studied is overmature.

Sphaerella vaccinii var. *corymbosi* Sacc., Nuovo Giorn. Bot. Ital. 23: 187. 1916.

Type — USA: *Vaccinium corymbosum* (Ericaceae).

No material was studied as the type was not found in any of the herbaria consulted.

Mycosphaerella vaccinicola Ade, in Petr., Bot. Jahrb. Syst. 62, Beibl. 142: 127. 1928.— Fig. 918.

Type — Madeira: Ribeiro frio. On upper and lower surface of dead leaves of *Vaccinium maderense* (Ericaceae). Ade, VI 1926 (W no. 10047, holotype; W no. 26871, isotype).

This is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores 6-7 × 1.5-2 μm.

Mycosphaerella vagabunda (Desm.) Migula, in Thomé, Fl. Deutschl., Österr. Schweiz X, 1. Kryptog.-Fl., III, 3(1): 280. 1912 ["1913"] = *Sphaeria vagabunda* Desm., Ann. Sci. Nat. Bot., ser. 3, 6: 81. 1846 = *Sphaerella vagabunda* (Desm.) Fuckel, Jahrb. Nassauischen Vereins Naturk. 23-24: 106. 1870.

Type — France: On lower surface of dead leaves of *Clematis vitalba* (Ranunculaceae). Desmazières, Plantes Cryptogames de France no. 1795 p.p. (NY, isolectotype, here designated); also on *Crataegus monogyna* (Rosaceae). Desmazières, Plantes Cryptogames de France no. 1795 p.p. (NY, paratype).

The type contains two different hosts from two different plant families; it is lectotypified here with the material on *Clematis*. The types and additional material studied (Seine-et-Marne, Chailly, Feuilleaubeis, distributed in Roumeguère, Fungi Gallici Exsiccati no. 2575, X 1882, L; also Germany, Fuckel, Fungi Rhenani no. 848, B) are immature, but show that this is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores immature.

Mycosphaerella vagans (Ellis & Everh.) Tomilin, Novosti Sist. Nizsh. Rast. 8: 152. 1971 = *Sphaerella vagans* Ellis & Everh., Proc. Acad. Nat. Sci. Philadelphia 47: 421. 1895.— Fig. 919.

Type — USA: Colorado, Larimer Co., Mt. Richtophen, near Cameron Pass. On dead stems of *Castilleja miniata* (Orobanchaceae). Baker no. 271, VII 1894 (NY, lectotype, here designated); also Baker no. 275 (NY, topotype).

The protologue also mentions type material on a different host, viz. *Valeriana sylvatica* (Caprifoliaceae), but this was not found in NY. The lectotype shows that this belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradians*, with asci cylindrical, ascospores 14-18 × 3.5-5 μm. The topotype contains only a coelomycete.

Sphaerella vaginae (Lasch) Fuckel, Jahrb. Nassauischen Vereins Naturk. 23-24: 100. 1870 = *Sphaeria vaginae* Lasch, in Klotzsch, Herbarium Vivum Mycologicum, cent. 7 no. 663. 1844 = *Laestadia vaginae* (Lasch) Sacc., Syll. Fung. 1: 431. 1882.

Type — Germany: On dead culms of *Phragmites australis* ["*communis*"] (Poaceae). Lasch, distributed in Klotzsch, Herbarium Vivum Mycologicum no. 663 (B, isotype).

In the isotype studied, only overmature fungi were found.

Mycosphaerella vagnerae Earle, Bull. New York Bot. Gard. 3: 292. 1904 = *Sphaerella vagnerae* (Earle) Sacc. & Trotter, Syll. Fung. 2: 142. 1913 [as "*wagnerae*"] = *Mycosphaerella tassiana* var. *vagnerae* (Earle) Clem., Cryptogamae

Formationum Coloradensium no. 230. 1906 [as "*Mycosphaerium tassianum*"].— Fig. 920.

Type — USA: Colorado, Carson, Kings Canon. On dead stems of *Smilax* ["*Vagnera*"] (Smilacaceae). Baker, VII 1902 (NY, holotype).

Already synonymised with *Davidiella allicina* (as *Mycosphaerella tassiana*) by von Arx (1949), with which the type agrees well, with asci pyriform, ascospores 16-18 × 5-6 µm.

Mycosphaerella valeppensis Rehm, Oesterr. Bot. Z. 56: 295. 1906, nomen nudum (not validly published, Article 32).— Fig. 921.

Authentic material — Germany: Alpen, Valepp. On upper and lower surface of dead leaves of *Dryas octopetala* (Rosaceae). Rehm, 1905 (S).

This is material of *Pseudomassaria* (= *Chaetapiospora*) *islandica* (Johanson) M.E. Barr, with ascospores yellowish and strongly apiosporous, 20-24 × 10-12 µm.

Mycosphaerella valida Syd. & P. Syd., Ann. Mycol. 34: 390. 1936.— Fig. 922.

Type — Germany: Brandenburg, Strausberg. On dead stems of *Ranunculus lingua* (Ranunculaceae). Sydow, Mycotheca Germanica no. 2930, VI 1934 (L, isotype).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. superflua*, with asci cylindrical, ascospores 12-15 × 3.5-4.5 µm.

Mycosphaerella variabilis Kill., Ann. Épiphyt. 12: 154. 1926.

Type — France: *Digitalis purpurea* (Plantaginaceae).

No material was studied as the type was not included in a loan from PC.

Sphaerella veneta (Sacc. & Speg.) Cooke, Grevillea 16: 77. 1888 = *Laestadia veneta* Sacc. & Speg., in Sacc., Michelia 1: 351. 1878 = *Apiospora veneta* (Sacc. & Speg.) Saccardo ex Kleb., Z. Pflanzenkrankh. 12: 257. 1902 = *Gnomonia veneta* (Sacc. & Speg.) Kleb., Jahrb. Wiss. Bot. 41: 533. 1905 non Spegazzini 1879 = *Guignardia veneta* (Sacc. & Speg.) Traverso, Flora Ital. Cryptog. 2: 392. 1906 = *Apiosporiopsis veneta* (Sacc. & Speg.) Mariani, Atti Soc. Ital. Sci. Nat. 50: 165. 1911 = *Apiognomonia veneta* (Sacc. & Speg.) Höhn., Ann. Mycol. 16: 51. 1918.

Type — Italy: *Platanus occidentalis* (Platanaceae).

Anamorph: *Discula nervisequa* (Fuckel) M. Morelet (sub *platani* (Peck) Sacc.) *vide* Barr (1978).

Accepted as *Apiognomonia veneta* (Sacc. & Speg.) Höhn. by Barr (1978) and therefore not studied.

Mycosphaerella venezuelensis J.H. Miller & Burton, Mycologia 35: 88. 1943.

Type — Venezuela: *Canavalia ensiformis* (Fabaceae).

No material was studied as the type was not found in NY or BPI.

Sphaerella venziana Sacc., Nuovo Giorn. Bot. Ital. 7: 303. 1875.— Fig. 923.

Type — Italy. On dead stems and on upper and lower surface of dead leaves of *Facchinia lanceolata*

(Caryophyllaceae). Saccardo (PAD, holotype; PAD, isotype).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradicans*, with asci cylindrical, ascospores 10-12 × 3-4 µm.

Mycosphaerella veratri Höhn., Sitzungsber. Kaiserl. Akad. Wiss., Math.-Naturwiss. Kl., Abt. 1, 123: 60. 1914 = *Sphaerella veratri* (Höhn.) Trotter, Syll. Fung. 24: 876. 1928.— Fig. 924.

Type — Austria: Wienerwald, Gumpoldskirchen. On upper and lower surface of dead leaves of *Veratrum nigrum* (Asparagaceae). Höhn., IV 1905 (FH-Höhn., holotype).

This is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores 10-12 × 2.5-3 µm. Additional material studied (Italy, Venice, Piave-Knie, Barbitano, on upper and lower surface of dead leaves of *Veratrum nigrum* (Asparagaceae), Hruby, distributed in Reliquiae Petrakianae no. 2480, IV 1918, L) also belongs to *Davidiella*, and is morphologically indistinguishable from *D. ammophilae*, but with asci nearly cylindrical, ascospores 8-10 × 4.5-5.5 µm.

Mycosphaerella veratri (Losa) M. Morelet. See *Mycosphaerella veratri-lobeliani* Fakirova & Denchev.

Mycosphaerella ["*Mycosphaerium*"] *veratri* Clem., Cryptogamae Formationum Coloradensium no. 426. 1908, nomen herbariorum (not validly published, Article 32).— Fig. 925.

Authentic material — USA: Colorado, Sierra Blanca. On upper and lower surface of dead leaves of *Veratrum speciosum* (Asparagaceae). Clements, VI 1907, Cryptogamae Formationum Coloradensium no. 426 (BPI).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. clandestina*, with asci pyriform, ascospores thick-walled, 21-25 × 6-7.5 µm.

Mycosphaerella veratri-lobeliani Fakirova & Denchev, Mycotaxon 88: 119. 2003 = *Mycosphaerella veratri* (Losa) M. Morelet, Ann. Soc. Sci. Nat. Archéol. Toulon & Var 20: 106. 1968, later homonym (illegitimate, Article 53) = *Sphaerella veratri* Losa, Anales Jard. Bot. Madrid 8: 304. 1948 ["1947"], later homonym (illegitimate, Article 53).

Type — Spain: *Veratrum album* (Asparagaceae).

No material was studied as the type was not found in MA and is probably lost. Fakirova & Denchev (op. cit.) give a new type (from Bulgaria) and a Latin description, but cite the two illegitimate names as synonyms, with full references. Therefore, this is a new name for an already described taxon, the type of which originates from Spain.

Mycosphaerella verbascicola (Schwein.) Fairm., Proc. Rochester Acad. Sci. 4: 176. 1905 = *Sphaeria verbascicola* Schwein., Trans. Amer. Philos. Soc., ser. 2, 4: 221. 1832 = *Sphaerella verbascicola* (Schwein.) Ellis, in Britton, Cat. Plants New Jersey: 522. 1890 ["1889"].— Fig. 926.

Type — USA: Bethlehem. On dead stems of *Verbascum* (Scrophulariaceae). (PH, holotype; PH, 3 isotypes).

The types studied contain only *Pleospora herbarum* (Pers. : Fr.) Rabenh., of which this is a new synonym, with

ascospores muriform, brown, 17-23 × 5-6 µm. Also studied and agreement with the type: Pennsylvania, Chester, Michener (PH). Additional material studied (New Jersey, Newfield, on *Verbascum thapsus* no. 591, L) belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores 12-16 × 4-6.5 µm.

Mycosphaerella verbenae (Unamuno) M. Morelet, Ann. Soc. Sci. Nat. Archéol. Toulon & Var 20: 106. 1968 ≡ *Sphaerella verbenae* Unamuno, Anales Jard. Bot. Madrid 2: 39 1942 ["1941"].

Type — Spain: *Verbena officinalis* (Verbenaceae). No material was studied as the type was not found in MA and is probably lost.

Mycosphaerella verecunda Syd., Ann. Mycol. 37: 373. 1939.— Fig. 927.

Type — Ecuador: Quito, Pichincha. On upper surface of green leaves, not in spots, of *Coriaria thymifolia* (Coriariaceae). Sydow, IX 1937 (NY, isotype), also distributed in Fungi Exotici Exsiccati no. 1191 (L, isotype). This is a parasitic species, with asci cylindrical, ascospores 15-19 × 3.5-5 µm.

Sphaerella verna Sacc. & Speg., in Sacc., Michelia 1: 379. 1878.— Fig. 928.

Type — Italy. On upper and lower surface of dead leaves of *Forsythia viridissima* (Oleaceae). Spegazzini (PAD, holotype). This is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores 8-10 × 2-2.5 µm.

Mycosphaerella vernoniae (Petch) Cash, Syll. Fung. 26: 357. 1972 ≡ *Sphaerella vernoniae* Petch, Ann. Roy. Bot. Gard. (Peradeniya) 7: 303. 1922.— Fig. 929.

Type — Sri Lanka: Peradeniya. On pale spots with dark margins on upper and lower surface of living leaves of *Vernonia hookeriana* (Asteraceae). Petch no. 5399, X 1917 (K, holotype). This is a species of *Guignardia*, with asci pyriform and with apical dome, hamathecium absent, ascospores simple, 18-22 × 7-8 µm.

Sphaerella verrucosa Alcalde, Anales Jard. Bot. Madrid 5: 147. 1945.

Type — Spain: Madrid, Botanical Garden. On upper and lower surface of dead leaves of *Hedera helix* (Araliaceae). Alcalde no. 12563, III 1944 (MA, holotype). Cited as synonymous with *Davidiella allicina* (as *Mycosphaerella allicina*) by Tomilin (1979). However, the holotype and a topotype (Same locality, III 1945, MA) contain only a coelomycete.

Mycosphaerella vesicaria (Pass.) Tomilin, Oprelidelitel' gribov roda *Mycosphaerella* Johans.: 112. 1979 ≡ *Sphaerella vesicaria* Pass., Erbario Crittogamico Italiano, ser. 2 no. 1367. 1884.— Fig. 930.

Type — Italy: Parma. On dead pods of *Colutea arborescens* (Fabaceae). Passerini, distributed in Roumeguère, Fungi Gallici Exsiccati no. 2829, XI 1883 (L, isotype). This is morphologically indistinguishable from *Davidiella allicina*, with asci pyriform, ascospores 18-22 × 6-7 µm.

Mycosphaerella vesicariae-arcticae (Henn.) Tomilin, Oprelidelitel' gribov roda *Mycosphaerella* Johans.: 78. 1979 ≡ *Sphaerella vesicariae-arcticae* Henn., in Allesch. & Henn., Biblioth. Bot. 42: 45. 1897.

Type — Denmark: *Vesicaria arctica* (Brassicaceae). No material was studied as the type was not found in B and is probably lost. Synonymised with *Davidiella allicina* (as *Mycosphaerella tassiana*) by von Arx (1949).

Mycosphaerella vespa Carnegie & Keane, Mycol. Res. 102: 1274. 1998.

Type — Australia: Victoria, Gippsland, Flynn. On leaves of *Eucalyptus globulus* (Myrtaceae). Carnegie, III 1995 (DAR 72449, holotype, not seen). No material was studied of this recently described species.

Mycosphaerella vexans (Masse) Tomilin, Novosti Sist. Nizsh. Rast. 6: 123. 1970 ["1969"] ≡ *Sphaerella vexans* Masse, Bull. Misc. Inform. 4: 158. 1914.

Type — Tanzania: Zanzibar. On white spots on upper and lower surface of living leaves of *Eugenia caryophyllata* (Myrtaceae). McClellan, V 1913 (K, holotype; K, isotype). Anamorph: *Stenella vexans* Crous fide Crous (1999). The types contain only a *Pestalotiopsis*.

Mycosphaerella viburni (Nitschke) J. Schröt., in Cohn, Kryptog.-Fl. Schlesien 3(2): 335. 1894 ["1893"] ≡ *Sphaerella viburni* Nitschke, in Fuckel, Jahrb. Nassauischen Vereins Naturk. 23-24: 106. 1870.— Fig. 931.

Type — Germany: Oestrich. On lower surface of dead leaves of *Viburnum opulus* (Adoxaceae). Fuckel, Fungi Rhenani Exsiccati no. 1785 (BPI, L, isotypes). The isotypes studied are immature. Additional material studied (Germany, Baden, Rastatt, Schröter, distributed in Rabenhorst, Fungi Europaei Exsiccati no. 2130, IV 1876, L) belongs to section *Caterva*, and is morphologically indistinguishable from *M. superflua*, with ascocarps tiny, asci cylindrical, ascospores 14-17 × 3-4.5 µm.

Mycosphaerella viburni var. *intermedia* (Cif.) Cash, Syll. Fung. 26: 357. 1972 ≡ *Sphaerella viburni* var. *intermedia* Cif., Ann. Mycol. 20. 39. 1922.

Type — Italy: *Viburnum lantana* (Adoxaceae). No material was studied as the type was not found in any of the herbaria consulted.

Sphaerella viburni var. *lantanae* (Nitschke) Cif. See *Mycosphaerella lantanae* (Nitschke) Migula.

Mycosphaerella viburnicola (Speg.) Tomilin, Novosti Sist. Nizsh. Rast. 8: 152. 1971 ≡ *Sphaerella viburnicola* Speg., Anales Mus. Nac. Hist. Nat. Buenos Aires 19 (or ser. 3, 12): 356. 1909.

Type — Argentina: *Viburnum tinus* (Adoxaceae).

No material was studied as the type was not included in a loan from LPS. Cited as synonymous with *M. lantanae* by Tomilin (1979).

Mycosphaerella viciae (J. Schröt.) Cruchet, Bull. Soc. Vaud. Sci. Nat. 55: 45. 1923 \equiv *Sphaerella viciae* J. Schröt., Jahresber. Schles. Ges. Vaterl. Cult. 65: 274. 1887.

Type — Norway: *Vicia cracca* (Fabaceae).

No material was studied as the type was not found in any of the herbaria consulted.

Mycosphaerella viciarum Petr., Hedwigia 74: 36. 1934.— Fig. 932.

Type — Russia: Siberia, Distr. Tsheropanovo-Tshumysk. On dead stems of *Vicia* (Fabaceae). Antonov no. N 104, VI 1925 (W, holotype).

Cited as synonymous with *M. montellica* by Tomilin (1979). The type shows that it is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores 7-9 \times 1.5-2.5 μ m.

Mycosphaerella viegasii M. Morelet, Ann. Soc. Sci. Nat. Archéol. Toulon & Var 20: 106. 1968, nomen novum (Article 58) but lacking Latin description (not validly published, Article 36) for *Mycosphaerella tecomae* Viégas, Bragantia 4: 164. 1944, lacking Latin description (not validly published, Article 36).

Type — Brazil: *Tecoma* (Bignoniaceae).

No material was studied as the type was not included in a loan from URM.

Mycosphaerella vincae (Fr.) Feltgen, Recueil Mém. Trav. Soc. Bot. Grand-Duché Luxembourg 14: 287. 1899 \equiv *Sphaeria vincae* Fr., in Wallr., Fl. Crypt. Germ. 2: 776. 1833 \equiv *Sphaerella vincae* (Fr.) Auersw., in Gonn. & Rabenh., Mycol. Europaea 5-6: 10. 1869 \equiv *Metasphaeria vincae* (Fr.) Sacc., Syll. Fung. 2: 171. 1883 \equiv *Sacchettoecium vincae* (Fr.) Kirschst., Kryptog.-Fl. Mark Brandenburg 7: 427. 1938.

Type — France: Vosges. *Vinca minor* (Apocynaceae). Mougeot no. 504 (UPS-FRIES, holotype).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradicans*, with asci cylindrical but immature ascospores. Additional materials studied (Germany, Mappen, Fuckel, Fungi Rhenani no. 841, B; also Sachsen, Ponickau, Auerswald, IV 1869, B) are also immature, but morphologically similar.

Sphaerella vincae f. *vincae-minoris* Thüm., Mycotheca Universalis no. 1148.— Fig. 933.

Type — Germany: Bayern, Bayreuth. On upper and lower surface of dead leaves of *Vinca minor* (Apocynaceae). Thümen, Mycotheca Universalis no. 1148, IV 1875 (L, isotype).

This belongs to section *Longispora*, and is morphologically indistinguishable from *M. millegrana*, with asci cylindrical, ascospores 17-22 \times 3.5-4.5 μ m.

Mycosphaerella vincetoxici (Sacc.) Lindau, in Engl. & Prantl, Natürlichen Pflanzenf. 1(1): 424. 1897 \equiv *Sphaerella vincetoxici* Sacc., Syll. Fung. 1: 516. 1882.— Fig. 934.

Type — Italy: Conegliano. On dead stems of *Vincetoxicum officinale* (Apocynaceae). Saccardo (PAD, holotype).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradicans*, with asci cylindrical, ascospores 10-12 \times 2.5-3.5 μ m. Already cited as synonymous with *M. albescens* (Rabenh.) Lind by Munk (1957).

Sphaerella vincetoxici var. *folliculorum* Sacc., Syll. Fung. 1: 516. 1882.— Fig. 935.

Type — Italy: Conegliano. On dead fruits of *Vincetoxicum officinale* (Apocynaceae). Saccardo (PAD, holotype).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. subradicans*, with asci cylindrical, ascospores 10-12.5 \times 2.5-3.5 μ m.

Mycosphaerella vindobonensis Petr., Sydowia 9: 577. 1955.

Type — Austria: *Globularia wilkommii* (Plantaginaceae).

No material was studied as the type was not included in a loan from W. Cited as synonymous with *M. jenensis* by Tomilin (1979).

Mycosphaerella violae Potebnia, Ann. Mycol. 8: 51. 1910 \equiv *Sphaerella violae* (Potebnia) Sacc. & Traverso, Syll. Fung. 20: 829. 1911.

Type — Russia: Kursk. On *Viola hirta* (Violaceae).

Anamorphs: Probably *Cercospora violae* Sacc., *Ramularia lactea* (Desm.) Sacc. and *Septoria violae* Westend. *vide* Potebnia (op. cit.).

No material was studied as the type was not included in loans from LE or LEP.

Mycosphaerella virgaureae Krieger, Ann. Mycol. 9: 216. 1911.— Fig. 936.

Type — Germany: Schandau, Kirnitzschtal. On upper and lower surface of dead leaves of *Solidago virgaurea* (Asteraceae). Krieger, Fungi Saxonici no. 2112, V 1904 (L, isotype).

This belongs to section *Caterva*, and is morphologically indistinguishable from *M. superflua*, with asci cylindrical, ascospores 12-15 \times 2.5-3.5 μ m.

Mycosphaerella vitalbae (Pass.) Petr., Sydowia 1: 145. 1947 \equiv *Sphaerella vitalbae* Pass., Atti Reale Accad. Lincei, Mem. Cl. Sci. Fis., ser. 4, 3: 5. 1887.— Fig. 973, 974, 975.

Type — Italy: On dead stems of *Clematis vitalba* (Ranunculaceae).

No type material was found in any of the herbaria consulted. Material seen (Montenegro, Mornj, Bubák, IV 1903, BPI) contains immature and old specimens of a *Pleospora* species, with ascospores hyaline and 1-septate to golden brown and muriform, 21-25 \times 7.5-12 μ m.

Mycosphaerella vitalbina (Pass.) Petr., Hedwigia 65(1): 234. 1925 \equiv *Sphaerella vitalbina* Pass., Atti Reale Accad. Lincei, Mem. Cl. Sci. Fis., ser. 4, 4: 56. 1888.— Fig. 937.

Type — Italy: On dead stems of *Clematis vitalba* (Ranunculaceae).

The type was not found in any of the herbaria consulted. Material studied (Germany, Heilbrunn, Kirschstein, VI 1939, B) belongs to *Davidiella*, and is morphologically

indistinguishable from *D. ammophilae*, with asci pyriform, ascospores $7-9 \times 2.5-3 \mu\text{m}$.

Mycosphaerella vitensis (Unamuno) Tomilin, Novosti Sist. Nizsh. Rast. 1967: 190. 1967 \equiv *Sphaerella vitensis* Unamuno, Bol. Soc. Esp. Hist. Nat. 29: 392. 1929.

Type — Spain: Burgos, La Vid, Orillas del Duero. On dead culms of *Scirpus holoschoenus* (Cyperaceae). Unamuno no. 8976, VIII 1929 (MA, holotype).

The type contains only a *Stagonospora* anamorph.

Mycosphaerella viticis Hara, Fungi 3-4: 104. 1931.

Type — Japan: *Vitis trifolia* (Vitaceae).

No material was studied as the type was not included in a loan from TNS.

Mycosphaerella viticola (Hollós) Tomilin, Novosti Sist. Nizsh. Rast. 6: 123. 1970 [“1969”] \equiv *Sphaerella viticola* Hollós, Ann. Hist.-Nat. Mus. Natl. Hung. 5: 454. 1907 \equiv *Phaeosphaerella viticola* (Hollós) Tomilin, Opređelitel' gribov roda *Mycosphaerella* Johans.: 285. 1979.

Type — Hungary: *Vitis vinifera* (Vitaceae).

The type is not in BP and may have been destroyed during the war.

Mycosphaerella viticola Koshk., in Koshk., Frolov & Dzhuraeva, Mikoflora Badkhyza, Karabilya i Yuzhnoi Chasti Murgabskogo Oazisa: 102. 1970, later homonym (illegitimate, Article 53), nomen novum (Article 58) for *Mycosphaerella vitis* Koshk., Bot. Mater. Otd. Sporov. Rast. Bot. Inst. Komarova Akad. Nauk SSSR 14: 126. 1961, later homonym (illegitimate, Article 53).

Type — Turkmenia: *Vitis vinifera* (Vitaceae).

No material was studied as the location of the type is unknown.

Mycosphaerella vitis (Fuckel) J. Schröt., in Cohn, Kryptog.-Fl. Schlesien 3(2): 334. 1894 [“1893”] \equiv *Sphaerella vitis* Fuckel, Jahrb. Nassauischen Vereins Naturk. 23-24: 104. 1870 (summer) \equiv *Sphaeria vitis* Rabenh., Herb. Vivum Mycol. no. 1047. 1846, nomen nudum (not validly published, Article 32) \equiv *Mycosphaerella vitis-viniferae* Tomilin, Opređelitel' gribov roda *Mycosphaerella* Johans.: 268. 1979, superfluous (illegitimate, Article 52) nomen novum (Article 58).

Type — Germany: On upper and lower surface of dead leaves of *Vitis vinifera* (Vitaceae). Fuckel, Fungi Rhenani Exsiccati no. 829 (L, isotype).

Anamorph: *Septosporium fuckelii* Thüm. *vide* Fuckel (op. cit.) (= *Passalora dissiliens* (Duby) U. Braun & Crous).

The isotype studied contains only an *Asteromella* spermatial state. According to printed information on the label (“fungus conidiophorus”), the material has never contained a teleomorph.

Mycosphaerella vitis Koshk. See *Mycosphaerella viticola* Koshk.

Sphaerella vitis Schulzer. See *Sphaerella sarmentorum* Pirota.

Mycosphaerella vitis-viniferae Tomilin. See *Mycosphaerella vitis* (Fuckel) J. Schröt.

Mycosphaerella vivipari (G. Winter) Lind, Biol. Meddel. Kongel. Danske Vidensk. Selsk. 13: 20. 1928 \equiv *Sphaerella vivipari* G. Winter, Internationale Polarforschung 1882-1883. Deutsche Expedition, 2: 94. 1890.

Type — Greenland: *Polygonum viviparum* (Polygonaceae). No material was studied as the type was not found in any of the herbaria consulted.

Mycosphaerella vogelii (Syd.) Tomilin, Novosti Sist. Nizsh. Rast. 6: 123. 1970 [“1969”] \equiv *Sphaerella vogelii* Syd., Mycotheca Germanica, fasc. 14-15 no. 681. 1908 \equiv *Carlia cinerea* (Desm.) Höhn., Mitt. Bot. Lab. TH Wien, ser. 2, 3: 91. 1925, based on anamorph (illegitimate, Article 59).— Fig. 938.

Type — Germany: Brandenburg, Tamsel. On upper and lower surface of dead leaves of *Rhamnus catharticus* (Rhamnaceae). Vogel, distributed in Sydow, Mycotheca Germanica no. 681, V 1908 (L, isotype).

Anamorph: *Phoma cinerea* Desm. \equiv *Carlia cinerea* (Desm.) Höhn. *vide* Höhnel (op. cit.).

This belongs to section *Longispora*, and is morphologically indistinguishable from *M. topographica*, with asci cylindrical, ascospores $22-25 \times 3-4 \mu\text{m}$. In the isotype studied, also *M. punctiformis* is present.

Mycosphaerella vogesiaca (Syd.) Tomilin, Novosti Sist. Nizsh. Rast. 1967: 190. 1967 \equiv *Sphaerella vogesiaca* Syd., Mycotheca Germanica, fasc. 20 no. 979. 1911.— Fig. 939.

Type — France: Dept. Haut-Rhin, Vosges, Hohneck. On dead stems of *Juncus conglomeratus* (Juncaceae). Sydow, Mycotheca Germanica no. 979, VII 1910 (L, isotype).

This belongs to *Davidiella*, and is possibly morphologically indistinguishable from *D. ammophilae*, with ascospores $11-13 \times 3-3.5 \mu\text{m}$. In the isotype studied, also three different coelomycetes are present.

Mycosphaerella volkartiana Petr., Reliquiae Petrakianae no. 2689, nomen herbariorum (not validly published, Article 32).

Authentic material — Croatia: Istria, Smogar Plateau. On brown spots on living leaves of *Doronicum halleri* (Asteraceae). Hruby, distributed in Reliquiae Petrakianae no. 2689, IX 1917 (L).

The authentic material studied is sterile.

Mycosphaerella volkartii Tomilin, Novosti Sist. Nizsh. Rast. 7: 198. 1970, based on a nomen nudum (not validly published, Article 32), nomen novum (Article 58) for *Mycosphaerella calamagrostidis* Volkart, in Rehm, Ann. Mycol. 4: 409. 1906, nomen nudum (not validly published, Article 32) \equiv *Sphaerella calamagrostidis* (Volkart) Sacc. & Trotter, Syll. Fung. 22: 145. 1913, based on a nomen nudum (not validly published, Article 32).— Fig. 137.

Authentic material — Austria: Graubünden, Fürstenalp. On dead leaves of *Calamagrostis varia* (Poaceae). Volkart, VI 1905, distributed in Rehm, Ascomyceten no. 1667 (BPI).

Synonymised with *Davidiella allicina* (as *Mycosphaerella tassiana*) by von Arx (1949), with which additional material

studied (Switzerland, Glarus, Filzbach, on upper and lower surface of dead leaves of *Dactylis glomerata*, Müller, VI 1949 CBS, sub *Mycosphaerella calamagrostidis*) agrees well, with asci pyriform, ascospores $14-16 \times 4.5-5.5 \mu\text{m}$. The isotype studied is immature.

Mycosphaerella vulgaris (P. Karst.) Johanson. See *Mycosphaerella maculiformis* (Pers. : Fr.) J. Schröt.

Sphaerella vulgaris (Fuckel) Masee. See *Sphaerella epicymatia* (Wallr.) Speg.

Mycosphaerella vulnerariae (Fuckel) Lindau, Hilfsb. Sammeln Ascomyceten: 12. 1903 \equiv *Sphaerella vulnerariae* Fuckel, Jahrb. Nassauischen Vereins Naturk. 27-28: 21. 1873.— Fig. 940.

Type — Switzerland: Neuchâtel. On upper and lower surface of dead leaves of *Anthyllis vulneraria* (Fabaceae). Morthier no. 72, III 1872 (G, holotype).

Anamorphs: *Cercospora radiata* Fuckel and *Ascochyta vulnerariae* Fuckel *vide* Fuckel (op. cit.).

Type and additional material studied (Morthier, V 1874, B, topotype; also Morthier, distributed in Rabenhorst-Winter, Fungi Europaei Exsiccati no. 2654, III 1881, L, topotype) belong to section *Caterva*, and show that this is morphologically indistinguishable from *M. superflua*, with asci cylindrical, ascospores $(12-13-15(-16) \times (2.5-)3-3.5(-4) \mu\text{m}$.

Mycosphaerella waimeana Crous, Joa.E. Taylor & M.E. Palm, in J.E. Taylor, Crous & M.E. Palm, Mycotaxon 78: 463. 2001.

Type — Hawaii: *Leucospermum* (Proteaceae).

Anamorph: *Stenella fide* Taylor, Crous & Palm (op. cit.).

No material was studied of this recently described species.

Mycosphaerella wakkeri (Sacc. & P. Syd.) Tomilin, Novosti Sist. Nizsh. Rast. 1967: 191. 1967 \equiv *Sphaerella wakkeri* Sacc. & P. Syd., Syll. Fung. 14: 533. 1899, nomen novum (Article 58) for *Sphaerella sacchari* Wakker, in Wakker & Went, De Ziekten van het Suikerriet op Java: 196. 1898, later homonym (illegitimate, Article 53).

Type — Indonesia: *Saccharum officinarum* (Poaceae).

No material was studied as the type was not found in L and is probably lost. Cited as synonymous with *Mycosphaerella sacchari* (Speg.) Seaver & Chardón by Tomilin (1979).

Mycosphaerella walkeri R.F. Park & Keane, Trans. Brit. Mycol. Soc. 83: 104. 1984.— Fig. 941.

Type — Australia: Skipton. On white spots with black margins on upper surface of living leaves of *Eucalyptus globulus* (Myrtaceae). Park, XII 1982 (IMI no. 280473, isotype).

Anamorph: *Sonderhenia eucalypticola* (A.R. Davis) H.J. Swart & J. Walker *vide* Crous (1998).

This is a parasitic species, with asci cylindrical, ascospores $13-17 \times 3-4 \mu\text{m}$.

Mycosphaerella washingtoniae Rehm, Ann. Mycol. 9: 364. 1911 \equiv *Sphaerella washingtoniae* (Rehm) Trotter, Syll. Fung. 24: 881. 1928.

Type — USA: California, Los Angeles. On dead bracts of *Washingtonia brachypoda* (Arecaceae). Baker & Metz 5388 (S, holotype).

Already synonymised with *Davidiella allicina* (as *Mycosphaerella tassiana*) by von Arx (1949), but the isotype shows that this is morphologically indistinguishable from *D. clandestina*, with asci pyriform, ascospores $(20-)22-25 \times 8-11 \mu\text{m}$.

Mycosphaerella weberi Oudem. See *Sphaerella potentillae* Oudem.

Mycosphaerella websteri Wiehe, Mycol. Pap. 55: 2. 1953.— Fig. 942.

Type — Malawi: Cholo, Tung Station. On white spots with brown margins on upper surface of living leaves of *Aleurites montana* (Euphorbiaceae). Webster no. 605, III 1950 (IMI no. 40977, holotype; K, isotype).

Anamorph: *Cercospora fide* Wiehe (op. cit.).

This is a parasitic species, with asci cylindrical, ascospores $13-17 \times 4-5.5 \mu\text{m}$.

Mycosphaerella weigela Fairm., Ann. Mycol. 8: 326. 1910 [as “*weigeliae*”] \equiv *Sphaerella weigela* (Fairm.) Sacc. & Trotter, Syll. Fung. 22: 137. 1913 [as “*weigeliae*”].

Type — USA: On brown spots on upper surface of living leaves of *Weigela rosea* (Adoxaceae).

No type material was found in any of the herbaria consulted. Material seen (USA, Indiana, Richmond, on *Weigela*, Adams, VI 1930, BPI) contains no fungal fruiting bodies at all.

Sphaerella weinmanniae Cooke, Grevillea 14: 130. 1886.— Fig. 943.

Type — New Zealand. On upper surface of dead leaves of *Weinmannia racemosa* (Cunoniaceae). Colenso no. b215, XII 1885 (K, holotype).

This is morphologically indistinguishable from *Glomerella cingulata*, with immersed, large ascomata, hamathecium absent, ascospores simple, $18-20 \times 4-5.5 \mu\text{m}$, ends pointed.

Sphaerella weiriana Sacc., Nuovo Giorn. Bot. Ital. 27: 76. 1920.— Fig. 944.

Type — USA: California, Gasquet. On pale brown spots with brown margins on upper surface of dead (!) leaves of *Castanopsis chrysophylla* (Fagaceae). Weir no. 10046, IX 1916 (PAD, lectotype, here designated); also Oregon, Siskiyou National Forest, Grants Pass. Weir no. 10049, IX 1916 (PAD, paratype).

Already cited as synonymous with *Dothidella janus* (Berk. & M.A. Curtis) Höhn. (as *Mycosphaerella janus*) by Barr (1972), with with the types agree well, with ascomata breaking through the epidermis, lifting flaps of it, asci cylindrical, surrounded by parenchymatic tissue of $7-15 \mu\text{m}$ wide cells, ascospores $22-27 \times 3-4 \mu\text{m}$.

Mycosphaerella welwitschii (A.L. Sm.) Sivan., in Sivan. & Nair, Trans. Brit. Mycol. Soc. 91: 329. 1988 \equiv *Dothidella welwitschii* A.L. Sm., J. Bot. 36: 179. 1898.— Fig. 945.

Type — Namibia [“*Iter Angolense*” but not Angola]: Swakopmund. On dead leaf tips of *Welwitschia mirabilis*

[“*bainesii*”] (Welwitschiaceae). Welwitsch, IX 1859 (IMI no. 313681, isotype, slide only).

The type and additional material (Swakopmund District, Goanikontes, Buck no. 13216, I 1986, IMI no. 313680) shows that this is a species of *Arthopyrenia*, with ascomata in dense groups, wall thick, composed of compressed cells, ostiole conspicuous, paraphysoids copious, thin, anastomosing, asci clavate, ascospores constricted at the septum, lower cell slightly constricted, 16-20 × 4-5.5 µm, when old becoming pale brown and ornamented with warts, surrounded by a 1 µm thick gelatinous sheath. Therefore the following combination is proposed: **Arthopyrenia welwitschii** (A.L. Sm.) Aptroot comb. nov., **MB 500336**. **Basionym:** *Dothidella welwitschii* A.L. Sm., J. Bot. 36: 179. 1898.

Mycosphaerella wichuriana (J. Schröt.) Johanson, Öfvers. Förh. Kongl. Svenska Vetensk.-Akad. 41(9): 166. 1884 ≡ *Sphaerella wichuriana* J. Schröt., Jahresber. Schles. Ges. Vaterl. Cult. 58: 173. 1880.— Fig. 946.

Type — Sweden: On upper and lower surface of dead leaves of *Carex* (Cyperaceae).

Synonymised with *M. recutita* by von Arx (1949). As the type of *M. recutita* does not contain a *Mycosphaerella*, the name *Mycosphaerella wichuriana* was suggested as the oldest available name for *M. recutita* sensu von Arx by Eriksson (1992). However, the name *Mycosphaerella disseminata* has priority for this taxon. No type material of *M. wichuriana* was found in any of the herbaria consulted. Material seen (Jämtland, Åre, Hamdöl, on *Carex aquatilis*, Nannfeldt, VII 1951, BPI) indeed belongs to *Davidiella*, and is morphologically indistinguishable from *D. disseminata*, with asci pyriform, ascospores 11.5-15 × 4-5 µm.

Mycosphaerella wichuriana var. *andromedae-polifoliae* (Sacc.) Negru & R. Sandor, Izv. Akad. Nauk Arm. SSR, Biol. Nauki 18: 57. 1965 [as “*andromeda-polyfoliae*”] ≡ *Sphaerella wichuriana* var. *andromedae-polifoliae* Sacc., Syll. Fung. 1: 530. 1882.

Type — Sweden: *Andromeda polifolia* (Ericaceae).

No material was studied as the type was not found in any of the herbaria consulted. Cited as synonymous with *M. recutita*, which is morphologically indistinguishable from *Davidiella disseminata*, by Tomilin (1979).

Mycosphaerella wichuriana var. *scirpella* Munk, Dansk Bot. Ark. 17(1): 316. 1957.— Fig. 947.

Type — Denmark: Lystrup. On dead leaves of *Scirpus lacustris* (Cyperaceae). Lind, VI 1909 (C, holotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. clandestina*, with asci pyriform, ascospores 25-30 × 5-6.5 µm.

Mycosphaerella winteri (Pass.) Tomilin, Novosti Sist. Nizsh. Rast. 1968: 168. 1968 ≡ *Stigmatea winteri* Pass., Hedwigia 15: 93. 1876.— Fig. 948.

Type — Italy: Parma. On spots on upper surface of living leaves of *Rubus corylifolius* (Rosaceae). Passerini, distributed in Rabenhorst, Fungi Europaei Exsiccati no. 2154, 1875 (L, isotype).

This is a parasitic species, with asci cylindrical, ascospores 15-19 × 3-4 µm.

Mycosphaerella winteriana (Sacc.) J. Schröt., in Cohn, Kryptog.-Fl. Schlesien 3(2): 338. 1894 [“1893”] ≡ *Sphaerella winteriana* Sacc., Syll. Fung. 1: 516. 1882 ≡ *Didymolepta winteriana* (Sacc.) Munk, Dansk Bot. Ark. 15(2): 110. 1953 ≡ *Didymella winteriana* (Sacc.) Petr. ex Munk, Dansk Bot. Ark. 17(1): 337. 1957.

Type — Germany: *Melampyrum nemorosum* (Orobanchaceae).

Anamorph: *Phoma silvatica* Sacc. fide Tomilin (1979).

Accepted as *Didymella winteriana* (Sacc.) Petr. by Müller & von Arx (1962) and therefore not studied.

Sphaerella winteriana f. *melampyri-nemorosi* J. Kunze, Fungi Selecti Exsiccati no. 64 [as “*winteri* f.”].

Type — Germany: Sachsen, Eisleben. On dead stems of *Melampyrum nemorosum* (Orobanchaceae). Kunze, Fungi Selecti Exsiccati no. 64, III 1875 (L, holotype).

The holotype contains only empty ascomata.

Mycosphaerella wisteriae (Cooke) Tomilin, Novosti Sist. Nizsh. Rast. 10: 114. 1973 [as “*wistariae*”] ≡ *Sphaerella wisteriae* Cooke, Grevillea 7: 54. 1878.— Fig. 949.

Type — USA: South Carolina, Aiken. On lower surface of dead leaves of *Wisteria sinensis* (Fabaceae). Ravenel, Fungi Americani Exsiccati no. 379 (K, 5 isotypes).

This is morphologically indistinguishable from *M. punctiformis*, with asci cylindrical, ascospores 9-11 × 2-3 µm.

Mycosphaerella wladivostokensis Petr., Hedwigia 74: 37. 1934.— Fig. 950.

Type — Russia: Primorsky Krai, Vladivostok, Razdol'noe. On dead stems of *Acanthopanax sessiliflorum* (Araliaceae). Ziling, VI 1928 (W, holotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. ammophilae*, with asci pyriform, ascospores 8-10 × 3-4 µm.

Mycosphaerella wollemiae Sivan. & R.G. Shivas, Mycol. Res. 106: 355. 2002.

Type — Australia: *Wollemia nobilis* (Araucariaceae).

No material was studied of this recently described species.

Mycosphaerella woronichinii G. Woron., Trudy Tiflissk. Bot. Sada, ser. 2, 3: 134. 1923 nom. nov. pro

Mycosphaerella silenicola Woron., Vestn. Tiflissk. Bot. Sada 28: 17. 1913, later homonym (illegitimate, Article 53) ≡ *Sphaerella silenicola* (Woron.) Trotter, Syll. Fung. 24: 856. 1928, later homonym (illegitimate, Article 53).

Type — Georgia: Abchazia, Juguin. On spots on upper surface of living leaves of *Silene compacta* (Caryophyllaceae). Woronichin no. 322, VII 1912 (LEP, holotype).

Although this species was synonymised with *Davidiella allicina* (as *Mycosphaerella tassiana*) by von Arx (1949), examination of the holotype shows that it belongs to *Davidiella*, of which it represents a parasitic species, with asci pyriform, ascospores 14-16 × 4-5 µm. Therefore the

following new combination is made: **Davidiella woronichinii** (G. Woron.) Aptroot comb. nov., **MB 500365**. **Basionym:** *Mycosphaerella woronichinii* G. Woron., Trudy Tiflissk. Bot. Sada, ser. 2, 3: 134. 1923.

Mycosphaerella woronowii Jacz., in Woronow, Trudy Tiflissk. Bot. Sada, 11: 143. 1910 \equiv *Sphaerella woronowii* (Jacz.) Sacc. & Trotter, Syll. Fung. 22: 138. 1913.

Type — Georgia: *Juglans regia* (Juglandaceae).

No material was studied as the type was not included in loans from LE or LEP. Cited as synonymous with *Mycosphaerella terebinthi* by Tomilin (1979).

Mycosphaerella xanthiicola (Cooke & Harkn.) Lindau, in Engl. & Prantl, Natürlichen Pflanzenf. 1(1): 424. 1897 \equiv *Sphaerella xanthiicola* Cooke & Harkn., Grevillea 14: 9. 1885 [as “*xanthicola*”].— Fig. 951.

Type — USA. On dead stems of *Xanthium* (Asteraceae). Harkness no. 2451 (K, holotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores 15-18 \times 4.5-6.5 μ m.

Mycosphaerella xerophylli Syd., in Syd. & Petr., Ann. Mycol. 20: 179. 1922.— Fig. 952.

Type — USA: Idaho, Priest River. On dead stems of *Xerophyllum tenax* (Xanthorrhoeaceae). Shattuck, VII 1917 (BPI, isotype).

Type and additional material studied (USA, Washington, Loft, Hedgcock, VI 1931, B, IMI no. 127145) belong to *Davidiella*, and show that this is morphologically indistinguishable from *D. allicina*, with asci pyriform, ascospores 17-21 \times 5.5-7.5 μ m. the isotype is postmature with few ascospores still present.

Mycosphaerella xylomeli Sivan. & R.G. Shivas, Mycol. Res. 106: 355. 2002.

Type — Australia: *Xylomelum pyriforme* (Proteaceae).

No material was studied of this recently described species.

Mycosphaerella yaku-insularia T. Kobay., Bull. Gov. Forest Exp. Sta. 292: 9. 1977.

Type — Japan: *Ilex buergeri* (Aquifoliaceae).

No material was studied as the type was not included in a loan from TNS.

Mycosphaerella yanagawaensis Togashi, Trans. Sapporo Nat. Hist. Soc. 14: 283. 1936.

Type — Japan: *Pteridium aquilinum* (Polypodiaceae).

No material was studied as the type was not included in a loan from TNS.

Mycosphaerella yuccae (Ellis & Everh.) Tomilin, Novosti Sist. Nizsh. Rast. 1967: 191. 1967 \equiv *Sphaerella yuccae* Ellis & Everh., J. Mycol. 8: 67. 1902.— Fig. 953.

Type — USA: Alabama, Tuskegee. On upper and lower surface of dead leaves of *Yucca filamentosa* (Asparagaceae). Carver no. 80, VII 1900 (NY, holotype).

This is a species of *Guignardia*, with asci clavate, ascus tip thickened, no hamathecium filaments, ascospores simple, 12-19 \times 3.5-5 μ m. Additional material studied (USA, North

Carolina, Scotland County, on white spots with brown margins on upper surface of living leaves of *Yucca*, McInnes, V 1993, IMI no. 358511) is *Planistromella acervata*, with ascomata in stromata, cracking through the epidermis, asci cylindrical, surrounded by a parenchymatous tissue, ascospores 16-20 \times 2.5-3.5 μ m.

Mycosphaerella yuccina Woron., Věstn. Tiflissk. Bot. Sada 28: 16. 1913 \equiv *Sphaerella yuccina* (Woron.) Trotter, Syll. Fung. 24: 877. 1928.

Type — Georgia: Tiflis, Botanical garden. On white spots on upper and lower surface of living leaves of *Yucca gloriosa* (Asparagaceae). Koenig, II 1909 (LEP, holotype).

This is an immature ascomycete, possibly a *Dothidea*.

Mycosphaerella zae (Sacc.) Kirchn., Die Krankheiten und Beschädigungen unserer landwirtschaftlichen Kulturpflanzen, ed. 3: 92. 1923 \equiv *Sphaerella zae* Sacc., Michelia 1: 377. 1878.— Fig. 954.

Type — Italy. On pale spots with brown margins on upper surface of living leaves of *Zea mays* (Poaceae). Saccardo (PAD, holotype).

This is a parasitic species, with asci clavate, ascospores 17-20 \times 5-7 μ m.

Mycosphaerella zae-maydis Mukunya & Boothr., Phytopathology 63: 530. 1973 \equiv *Didymella zae-maydis* (Mukunya & Boothr.) Arx, Beih. Nova Hedwigia 87: 288. 1987.— Fig. 955.

Type — USA: New York, Aurora, Cayuga Co., Cornell Agronomy Farm. On pale spots on upper surface of living leaves of *Zea mays* (Poaceae). Mukunya, IV 1972 (IMI 196883, isotype).

Anamorph: *Phyllosticta maydis* Arny & Nelson *vide* Sivanesan (1984).

Accepted as *Didymella zae-maydis* (Mukunya & Boothr.) Arx by von Arx (op. cit.), with which the the isotype agrees well, with asci clavate, paraphysoids thin, ascospores pointed, 14-18 \times 3.5-5 μ m.

Mycosphaerella zeicola G.L. Stout, Mycologia 22: 278. 1930.— Fig. 956.

Type — USA: On pale spots on upper surface of living leaves of *Zea mays* (Poaceae).

No type material was found in any of the herbaria consulted. Material seen (India, Gokarna, Hari no. 452, VII 1966, IMI no. 122690), which is in agreement with the protologue, is *Didymella zae-maydis* (Mukunya & Boothr.) Arx, with asci cylindrical, paraphysoids thin, ascospores pointed, 14-17 \times 4-5.5 μ m.

Mycosphaerella zeina Saccas, Rev. Pathol. Vég. Entomol. Agric. France 30: 181. 1951.— Fig. 957.

Type — Central African Republic: Boukoko-Oubarqui-Chari. On upper and lower surface of dead, especially marginal, sections of living leaves of *Zea mays* (Poaceae). Saccas, VII 1950 (PC, isotype).

This is morphologically indistinguishable from *Phaeosphaeria variiseptata* (G.L. Stout) Shoemaker with paraphyses 3 μ m wide, ascospores 1-3-septate, becoming golden brown, 16-18 \times 3.5-4.5 μ m, mostly immature.

Sphaerella zeina Pass., Atti Reale Accad. Lincei, Mem. Cl. Sci. Fis., ser. 4, 3: 9. 1887.

Type — Italy: *Zea mays* (Poaceae).

No material was studied as the type was not found in any of the herbaria consulted. Cited as synonymous with *Mycosphaerella maydina* by Tomilin (1979).

Mycosphaerella zelkova Syd. & Hara, in Syd. & P. Syd., Ann. Mycol. 11: 60. 1913 [as “zelkova”] = *Sphaerella zelkova* (Syd. & Hara) Trotter, Syll. Fung. 24: 891. 1928 [as “zelkova”].— Fig. 958.

Type — Japan: Mino, Kawanye-muro. Hara, V 1912 (S, holotype). On upper and lower surface of dead leaves of *Zelkova acuminata* (Ulmaceae).

This belongs to section *Longispora*, and is morphologically indistinguishable from *M. millegrana*, with asci cylindrical, ascospores 16-21 × 2.5-3.5 µm.

Mycosphaerella zeylanica Cash. See *Mycosphaerella petchii* M. Morelet.

Mycosphaerella zilingii Petr., Hedwigia 68: 214. 1928.— Fig. 959.

Type — Russia: Siberia, Southern Altaj, Terechta. On dead stems of *Dracocephalum ruschiana* (Lamiaceae). Ziling no. P 565, VII 1926 (W, holotype).

This belongs to *Davidiella*, and is morphologically indistinguishable from *D. ammophilae*, with asci pyriform, ascospores 7-9 × 3-4 µm.

Mycosphaerella zingiberi Shirai & Hara, Bot. Mag. (Tokyo) 25: 70. 1911 = *Sphaerella zingiberi* (Shirai & Hara) Trotter, Syll. Fung. 24: 892. 1928.

Type — Japan: *Zingiber mioga* (Zingiberaceae).

No material was studied as the type was not included in a loan from TNS.

Mycosphaerella zizaniae (Schwein.) Lindau, in Engl. & Prantl, Natürlichem Pflanzenf. 1(1): 426. 1897 = *Sphaeria zizaniae* Schwein., Trans. Amer. Philos. Soc., ser. 2, 4: 223. 1832 = *Sphaerella zizaniae* (Schwein.) Ellis & Everh., N. Amer. Pyrenomyc.: 298. 1892 = *Mycosphaerella spraguei* Tomilin, Novosti Sist. Nizsh. Rast. 1967: 190. 1967, superfluous (illegitimate, Article 52) nomen novum (Article 58).

Type — USA: Philadelphia. On upper and lower surface of dead leaves of *Zizania aquatica* (Poaceae). (PH, holotype; PH, isotype; K, isotype).

This is morphologically indistinguishable from *Glomerella cingulata*, for which it would represent an older epithet, with hamathecium absent, ascospores simple, 13-16 × 7-9 µm. On the dead culms a *Septoriella* is also present. Additional material studied (USA, Wilmington, Delaware, Commons, distributed in Ellis & Everhart, North American Fungi no. 1797, VIII 1886, L) belongs to *Davidiella*, and is morphologically indistinguishable from *D. disseminata*, with asci pyriform, ascospores 12-15 × 3.5-4.5 µm.

Mycosphaerella zizaniicola (Speg.) Tomilin, Novosti Sist. Nizsh. Rast. 1967: 191. 1967 = *Sphaerella zizaniicola*

Speg., Anales Mus. Nac. Hist. Nat. Buenos Aires 6: 266. 1898.

Type — Argentina: *Zizania bonariensis* (Poaceae).

No material was studied as the type was not included in a loan from LPS.

Sphaerella ziziphi Pat., Catalogue raisonné des plantes cellulaires de la Tunisie. Fungi: 104. 1897 [as “zizyphi”].

Type — Tunisia. On branches of *Ziziphus* (Rhamnaceae). Patouillard, II 1893 (FH-Patouillard, holotype).

The type contains several coelomycetes and only one ascomycete that is in accordance with the protologue, which is, however, weakly lichenized. It should be concluded that this is morphologically indistinguishable from *Arthonia punctiformis* Ach., with ascospores 3-septate, 13-15 × 4.5-5.5 µm.

Mycosphaerella ziziphicola Petr., Hedwigia 68: 215. 1928 [as “zizyphicola”].— Fig. 960.

Type — Russia: Siberia, Southern Altaj, Kuraj. On dead stems of *Ziziphus clinopodioides* (Rhamnaceae). Antonov, III 1927 (W, holotype).

This belongs to section *Longispora*, and is morphologically indistinguishable from *M. latebrosa*, with asci cylindrical, ascospores 20-25 × 2.5-3.5 µm.

Sphaerella zonata Ellis & Everh., J. Inst. Jamaica 1: 285. 1893.— Fig. 961.

Type — Jamaica: Catherine Hall Estate, near Montego Bay. On brown spots on upper surface of living leaves of *Cocos nucifera* (Arecaceae). Sinclair no. 69 (BPI, isotype).

The isotype and additional material seen (Orange River Station, Leather, XI 1962, IMI no. 98333b; also Above Rocks, XI 1947, IMI no. 22990) all belong to a *Planistromella* species, with ascomata in dense stromata, asci pyriform, pseudoparaphyses septate, gnarled, 2-3.5 µm wide, ascospores 8-11 × 2-3.5 µm. Therefore the following new combination is proposed: **Planistromella zonata** (Ellis & Everh.) Aptroot comb. nov., **MB 500375**. **Basionym:** *Sphaerella zonata* Ellis & Everh., J. Inst. Jamaica 1: 285. 1893.

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HOST INDEX

This index gives the accepted host genera (alphabetically) and the species entries. In case of homotypic synonyms and orthographic variants, only the epithet under which the taxon is treated is listed. One listing can refer to several different entries under the same epithet, e.g. with other authors or subspecific taxa. Subspecific taxa are only listed when they are recorded from a different host genus than the nominal form, or when several, but not all, were reported from the type host.

Host genera (whether plant or fungus) are ***in bold and italic*** and start with a capital, *fungus taxa* treated are in *lower case italic*.

- Abies***
abietis
bohemica
pinsapo
Abutilon
abutilontidicola
Acacia
acaciae
acaciigena
koae
silveirae
thailandica
Acaena
acaenae
fuegiana
Acanthopanax
acanthopanax
wladiwostokensis
Acer
acerifera
acerina
aceris
acerna
alarum
brunnea
glaucescens
latebrosa
maculiformis f. *aceris*
maculiformis f. *aceris-*
pseudoplatani
maculiformis f. *pseudoplatani*
maculiformis f. *sorbi* [sic]
maculiformis f. *spermogonifera*
mycopappi
paraneura
pseudoplatani
septorioides
sphaerellula
Achillea
achilleae
ptarmicae
Aconitum
aconitorum
antonovii
Acorus
acori
Acrocomia
acrocomiicola
Actaea
actaeae
- Actinidia***
actinidiae
tamseliensis
Adenophora
adenophorae
Adenostyles
petasitidis var. *adenostylidis*
Adhatoda
adhatodae
Adiantum
rehmiana
Adonis
adonidina
adonis
Adoxa
adoxae
Aegopodium
aegopodii
podagrariae
Aeluropus
aeluropodis
Aesculus
aesculi
hippocastani
sparsa f. *aesculi*
Agapanthus
agapanthi
agapanthi-umbellati
dealbens
Agave
agaves
melanorhabdos
Agrimonia
agrimoniae
Agrostis
agrostidis
nigrificata
phyllachoroides
Agrostistachys
agrostistachydis
Ailanthus
ailanthi
ailanthina
henriquesiana
Aira
airicola
anarithma
Albizzia
albizziae
Alcea
- parasita*
Alchemilla
alchemillae
alchemillicola
Aleurites
aleuritidicola
aleuritidis
websteri
Alisma
alismatis
Alliaria
cruciferarum f. *alliariae*
Allionia
allioniae
Allium
allicina
alii-cepae
schoenoprasi
Allosorus
tiroloensis var. *allosoricola*
Alnus
alnea
alni
alnicola
alniviridis
alnobetulae
borealis
cesatiana
conglomerata
conglomeratiformis
incomperta
insularis
maculiformis f. *alni*
maculiformis var. *alni*
minutissima
Alocasia
alocasiae
Aloë
aloës
Alpinia
alpiniae
alpinicola
Alstroemeria
alstroemeriae
Althaea
althaeina
pseudosphaerioides
Alyssum
alyssi
Alyxia

alyxiae
Amelanchier
Speg.ii
Ammophila
ammophilae
perforans
psammae
pseudopsammae
stromatica
Amomum
amomi
Anacardium
anacardiicola
Anaptychia
hageniae
Andira
andirae
Andromeda
andromedae
polifoliae
wichuriana var. *andromedae-*
polifoliae
Andropogon
spilota
Androsace
midzurenensis
Andryala
andryalae
Anemia
subastoma
Anemone
anemones
pulsatillae var. *major*
Anethum
anethi
foeniculacea
foeniculi
Angelica
acilegna
angelicae
brionnensis
leptasca
punctiformis var. *angeliaria*
rubella
Angophora
angophorae
Anigozanthos
millepunctata
Anthemis
anthemidina
Anthistiria
anthistiriae
Anthurium
anthurii
Anthyllis
vulnerariae
Aphyllanthes
aphyllanthis
monserratica
Apocynum
apocynica

Apophlaea
apophlaeae
Aquilegia
aquilegiae
aquilegiae-jonesii
coerulea
sawadae
Arabis
hambergii
pachyascae
Arachis
arachidicola
arachidis
argentinensis
berkeleyi
Aralia
araliae
papyrifera
Araucaria
araucariae
Arbutus
arbuticola
unedinis
Arctium
bardanae
Arctostaphylos
arbuti
grumiformis
shawii
Ardisia
ardisiae
Arenaria
arenariae
arenariicola
densa
graeca
leptosphaerioides
tingens
Aretia
aretiae
Argania
biguttulata
Arisaema
bhandardarensis
Aristolochia
aristolochiae
circumscissa
Aristolelia
aristoleliae
Armeria
armeriae
kirschsteinii
magellanica
pachyascae var. *alpina*
staticicola
subsequens
Armoracia
armoraciae
Arnica
arnicae
Aronia

aethiops var. *aroniae*
Aronica
aronici
Artemisia
artemisiae
eriphila
ferruginea
osborniae
Arthonia
arthoniae
Arthraxon
arthraxonicola
Artocarpus
artocarpus
Aruncus
dejanira
Arundinaria
arundinariae
eumorpha
Arundo
donacis
Asarum
asarifolia
Asclepias
asclepiadis
Ascophyllum
ascophylli
Asimina
asiminae
Asparagus
asparagi
Asperella
hystrix
Asperula
asperulae
Asphodeline
montenegrina
Asphodelus
asphodelina
asteroma var. *asphodeli*
Aspidopterys
cordatae
Asplenium
asplenii
filicum
polypodii f. *asplenii*
Aster
astericola
tatarica
Astragalus
afghanica
alboi
argyrophylli
astragali
astragalina
magnusiana
megastoma
melaena
melaenodes
ruthenica
spinarum

Athamanta
athamantae
Atichia
atichiae
Atractylodes
attractylodis
Atropa
atropae
belladonnae
Aucuba
aucubae
Audibertia
audibertiae
Avicennia
pneumatophorae
Azara
campoi
Baccharis
baccharidiphila
baccharidis
Ballota
polygramma
Balsamorhiza
balsamorhizae
Bambusa
bambusae
bambusicola
bambusina
Banksia
banksiae
Baptisia
baptisiicola
granulata
leucophaea
Barnadesia
barnadesiae
Bauhinia
bauhiniae
Begonia
bagoniae
Berberis
ambiens
angulata
australis
berberidiphilum
berberidis
schelkovnikovii
Beta
sphaerosperma
tabifica
Betonica
polygramma f. *betonicae*
Betula
babajani
betulae
ditricha
harthensis
Bignonia
capreolatae
passiflorae var. *bignoniae*
Bixa

biaxe
Boehmeria
boehmeriae
Boerhavia
boureae
Bolax
patagonica
Boquila
boquilae
Borreria
borreriae
Botrychium
botrychii
Brachypodium
brachypodii
Brachyscome
barchyscomes
Brahea
braheae
Brassica
brassicicola
capsellae
napicola
Briedelia
briedeliae
Bromus
clallamensis
ignobilis f. *bromi*
longissima
nevodovskii
Bryonia
bryoniae
Bryum
bryii
Buckinghamia
buckinghamiae
Bumelia
bumeliae
Bupleurum
bupleuri
bupleurina
Buddleja
puiggarii
Buxus
briardi
buxi
buxicola
buxifolia
limbalis
patouillardii
Byrsonima
byrsonimae
Cacalia
cacaliae
Caladium
caladii
Calamagrostis
calamagrostidis
greenii
microspora
parallela

tassiana var. *macrospora*
volkartii
Callistemon
bombycina
gaubae
Calluna
callunae
Calopogon
calopogonii
Calotropis
calotropidis
Caltha
calthae
palustris
Calycanthus
calycanthi
coneglanensis
Calystegia
harknessii
Camellia
camelliae
ikedai
theae
Campanula
campanulae
intermixta
Camphorosma
camphorosmae
Canavalia
canavaliae
venezuelensis
Canna
cannae
Cannabis
cannabis
Capparis
capparidis
Caragana
jaczewskii
Carex
alpina
bulgarica
carectorum
caricicola
caricis
hranicensis
hyperopta
kandawanica
leptospora
lineolata
proximella
pusilla
saxatilis
scirrhoides
tassiana
theodulina
wichuriana
Carica
caricae
Carlina
affinis

- carlinae*
compositarum
Carpha
carphae
Carpinus
carpineae
maculiformis f. *carpini*
maculiformis var. *carpini*
millegrana
myriadea var. *carpini*
Carya
convexula
dendroides
Caryophyllus
caryophyllata
Cassia
cassiae
frauxii
guineensis
sieberiana
sordidula
Cassinopsis
cassinopsidis
Cassiope
andromedae
immersa
inconspicua
Cassytha
cassythae
Castanea
arcana
castaneae
castanicola
echinophila
etrusca
maculiformis f. *castaneae*
maculiformis f. *castaneae-vescae*
maculiformis var. *castaneae*
maculiformis var. *castaneaecola*
maculiformis var. *centigrana*
oblivia
Castanopsis
castanopsidis
janus
weiriana
Castilla
castillae
Castilleja
castillejae
vagans
Casuarina
ascoscypha
Cattleya
cattleyae
Cecropia
cecropiae
Cedrela
cedrelae
Ceiba
eriodendri
Celtis
bonaerensis
castagnei
celtidis
dominicana
Centaurea
aliena
Centranthus
galatea var. *centranthi*
radiata
Cephalaria
cephalariae
Cerastium
stellarinearum
subnivalis
Ceratonia
ceratoniae
cuprea
Cercis
cercidicola
cercidis
conglomerata f. *siliquastri*
Cereus
cerei
Chaerophyllum
morthieri
sciadophila
Chamaenerion
caulicola
chamaenerii
danica
discrepans
epilobii
jutlandica
ludwigii
minor
nivalis
Chamaerops
bicalcarata
chamaeropsis
Cheiranthus
cruciferarum
Chelidonium
chelidonii
sciadophila var. *chelidonii*
Chenopodium
chenopodii
chenopodiicola
Chimaphila
chimaphilae
chimaphilina
marginata
Chionanthus
chionanthi
Chlorogalum
chlorogali
Chondrus
chondri
Chrysanthemum
chrysanthemi
ligulicola
Chrysobalanus
chrysobalani
chrysobalanicola
Chusquea
chusqueicola
Cicer
rabiei
Cichorium
ambigua
compositarum
Cicuta
cicutae
Cimicifuga
thalictrina
Cinna
cinnafolia
Cinnamomum
cinnamomicola
Cirsium
cirsii
cirsii-arvensis
Citrullus
citrullina
cucumis
Citrus
aurantiorum
citri
citricola
gibelliana
hesperidum
horii
inflata
lageniformis
limonis
loefgreni
sicula
Cladium
cladii
Cleidion
cleidionii
Clematis
applanata
clematidina
clematidis
clematitidis
haematites
vagabunda
vitalbae
vitalbina
Clethra
clethrae
Cleyera
cleyerae
Clidemia
clidemiae
Clusia
clusiae
guttiferae
Cocculus
mauica
Cocos
cocoës

cocoina
cocophylla
gastonis
palmicola
zonata

Codonopsis
codonopsidis

Coffea
canephorae
coffeae
coffeicola
costii

Colebrookia
mysorensis

Colocasia
colocasiae

Colona
columbiae

Colutea
vesicaria

Comarum
comari
innumerella
maculiformis f. *comari-palustris*

Commelina
tetraspora

Conium
taeniographa

Convallaria
asteroma
brunneola
convallariae
subradians

Convolvulus
adusta

Coprosma
coacervata
spissa

Coptis
coptis

Cordyline
cordylinicola

Coriaria
benguetensis
verecunda

Corispermum
corispermi

Cornus
auerswaldii
corni

cornicola
cornifolia

Coronilla
ariadna
coronillae-variae
oerteliana

Corrigiola
antoniana
corrigiolae

Corylus
corylaria

corylina
maculiformis
maculiformis var. *coryli*
punctiformis f. *coryli*
punctiformis var. *corylaria*

Cotinus
coggygriae

Cotoneaster
cotoneastri

Crataegus
crataegi
crataegicola
oxyacanthae
slaptoniensis
vagabunda

Craterispermum
craterispermi

Crinodendron
pataguae

Crinum
crini

Crithmum
agostinii

Crotalaria
crotalariae

Cryptocarya
pachythecia

Cryptomeria
cryptomeriae

Cryptotaenia
cryptotaeniae

Cucumis
melonis

Cucurbita
cucumis
cucurbitae
kabocha

Cunninghamia
cunninghamiae

Cupania
cupaniae

Cussonia
cussoniae

Cuttsia
cuttsiae

Cyananthus
cyananthi

Cyanea
cyanaeae

Cydonia
cydoniae
pomacearum

Cynodon
cynodontis

Cynoglossum
asperifolii

Cyperus
cyperi

Cypripedium
calceoli
cypripedii

Cytisus
cytisi-sagittalis
nogalesii

Dacrydium
dacrydii

Dactylis
dactylidis

ignobilis
recutita
volkartii

Dahlia
dahliae

Dalbergia
dalbergiae
devia

Damnacanthus
shikaeana

Danaea
danaeae

Daphne
daphnes

gnidii
laureolae

Daphniphyllum
daphniphylli

Daucus
dauci

himantia
sagedioides f. *dauci-carotae*

Daviesia
daviesiae

daviesicola

Delphinium
delphinii

delphiniicola

Dendrobium
dendrobii-nobilis

Dendrochilum
lagunensis

Dendromecon
dendromeconis

Dennettia
dennettiae

Deschampsia
deschampsiae

Desmodium
desmodii

desmodiifolia

Deutzia
deutziae

Dianella
dianellae

dianellincola
queenslandica

Dianthus
caryophyllea

caryophylli
dianthi
lusitanica
pulviscula
sibirica

tingens f. *dianthicola***Dichrostachys***dichrostachydis***Dictamnus***dictamni***Didymopanax***didymopanax***Dieffenbachia***dieffenbachiae***Digitalis***digitalis**digitalis-ambigua**mariae**petasitidis* var. *digitalidis**variabilis***Dioscorea***contraria**dioscoreae**dioscoreicola**papuana***Diospyros***diospyri**nawae***Dipsacus***asterinoides**sagedioides***Dirca***dircae***Dodartia***dodartiae***Dodonaea***dodonaeae***Doronicum***volkartiana***Dorycnium***dorycnii***Draba***carniolica**confinis**minuta**nubigena**pashkiensis***Dracaena***dracaenae***Dracocephalum***dracocephali**dracocephalicola**zilingii***Drepanocarpus***leguminosarum***Drimys***drimydis**fuscomaculans**perconferta***Drosera***droserae***Dryas***biberwierensis**dryadicola**dryadis**octopetalae**ootheca**rhytismoides**valepensis***Drymaria***drymariae***Dryopteris***aquilina* f. *aspidiorum**aquilina* f. *polypodii-filix-mas**aspidii**filicum**polypodii* f. *aspidii***Dunbaria***dunbariae***Ecdysanthera***ecdysantherae***Echinops***ritro***Echites***discophora***Ehretia***poonensis***Elaeagnus***elaegnica***Elaeis***elaeidis***Elatostema***elatostemae***Elymus***elymi**elymifoliae**munkii***Embothrium***embothrii***Empetrum***setosa***Endocarpon***dealbens***Endospermum***endospermi***Entada***entadae***Entelea***enteleae***Ephedra***ephedrae**ephedricola***Epilobium***adusta**epilobii**epilobii-montana**fuckelii**microspila**nebulosa* var. *epilobii**polia***Epimedium***epimedi***Epiphyllum***epiphylla***Equisetum***altera**equiseti**equiseticola**equisetina***Eragrostis***eragrostidis***Erechtites***erechthitidina***Erica***ericae-ciliaris**rouxii***Erigeron***subcongregata***Eriobotrya***laburni* var. *eriobotryae***Eriogonum***pachyasca***Erlangea***erlangeae***Erodium***ciconii**tomilini***Eryngium***eryngii**eryngicola**eryngina***Erysimum***denigrans***Erythrina***erythrinae**erythrinicola**stevensii***Erythralium***creberrima**merrillii***Erythroxyllum***erythroxyli***Escallonia***escalloniae***Eucalyptus***africana**ambiphylla**aurantia**columbiensis**communis**cryptica**crystallina**delegatensis**didymelloides**ellipsoidea**endophytica**eucalypti**flexuosa**gracilis**grandis**gregaria**heimii**heimioides**intermedia**irregulariramosa**juvenis**keniensis**lateralis*

longibasalis
madeirae
marksii
martinae
mexicana
molleriana
nubilosa
ohnowa
parkii
parva
readeriellophora
suberosa
suttonii
swartii
toledana
vespa
walkeri
Eugenia
aequatoriensis
eugeniae
eugenicola
vexans
Eulalia
eulaliae
Euodia
euodiae
Euonymus
euonymi
japonica
Eupatorium
eupatorii
eupatoriicola
tungurahwana
Euphorbia
beckhausii
cyparissiae
cyparissincola
degeneri
euphorbiae
euphorbiae-canariensis
euphorbiae-exiguae
euphorbiae-spinosae
euphorbiicola
nebulosa var. *euphorbiae*
parjumanica
tithymali
Euphrasia
euphrasiae
rabenhorstii
Eurya
euryae
Evansia
evansiae
Exacum
exaci
Facchinia
venziana
Fagraea
fagraeae
Fagus
antarctica

atomus
buna
errabunda
fagi
fagicola
faginea
fallax
flageoletiana
myriadea var. *fagi*
parvimacula
punctiformis var. *fagi*
Falcaria
falcariae
Feijoa
fejjoae
Fendlera
fendlerae
Ferula
ferulae
Festuca
crastophila
festucae
keruelensis
paleicola
tassiana f. *festucae*
Ficus
bolleana
elasticae
fici-ovatae
fici-wightianae
ficophila
ficus
leucospila
maculiformis var. *fici*
pittierii
Filipendula
filipendulae-denudatae
maculans
spiraeae
ulmariae
Firmiana
firmianae
Flagellaria
flagellariae
Flavaria
starbackii
Foeniculum
foeniculi
foeniculina
togniniana
Forsythia
verna
Fragaria
earliana
fragariae
louisianae
Frankenia
frankeniae
Fraxinus
effigurata
fraxinea

fraxini
fraxinicola
maculiformis f. *fraxini-excelsioris*
manshurica
petiolicola
pterophila
sapindi
Freycinetia
freycinetiae
Fuchsia
fuchsiicola
Funkia
ornithogali f. *funkiae*
Galanthus
galanthina
Galatella
galatellae
Galega
galegae
Galeopsis
caulicola
umbrosa
Galium
cruciatae
galii
galii-elliptici
ishnosperma
molluginis
ushuvaiensis
Galtonia
galtoniae
Garcinia
garciniae
Gardenia
gardeniae
luzonensis
Gaultheria
gaultheriae
Geniostoma
geniostomatis
Gentiana
andrewsii
chlorospila
deschmannii
gentianae
Geum
geicola
koldingensis
larsenii
melanoplaca
Gladiolus
fusca
minimipuncta
Glechoma
glechomatis
Gleichenia
gelicheniae
Globularia
dalmatica
ludwigiana
vindobonensis

Glochidion

glochidionis

Glyceria

chalcographa

distincta

ignobilis

Glycine

sojae

uspenskajae

Glycosma

glycosmae

Glycyrrhiza

glycyrrhizae

Gnaphalium

bakeri

Gnetum

genticola

Goodenia

goodeniae

Goodia

goodiaefolia

Godronia

parasitica

Gordonia

gordoniae

Gossypium

areola

gossypina

Gouldia

kaduae

Grevillea

atra

grevilleae

Guarea

chardonii

Guettarda

guettardina

Gunnera

gunnerae

hawaiiensis

Gymnadenia

orchidearum

Gypsophila

gypsophilae

gypsophilicola

gypsophilina

Halimodendron

halimodendri

Hancornia

discophora var. *macrospora*

Hedera

hederae

hederae-helicis

hedericola

punctiformis var. *hederae*

verrucosa

Hedychium

hedychii

Helichrysum

maculicola

Helleboris

hellebori

hermione

lachesis

salvatorensis

Helonias

heloniaefolia

Hemerocallis

aggregata

hemerocallidicola

hemerocallidis

pales

Hepatica

hepaticae

Heracleum

heraclei

heracleina

Hertia

hertiae

Heuchera

heucherae

Hevea

heveae

heveana

hevicola

Hibiscus

hibisci

Hicorius

caryigena

Hieracium

aliena f. *hieracii*

alsophila

hieracii

hieraciophila

Hippophaë

spinicola

Holcus

melconiana

Holoptelea

holopteleae

Honckenya

halophila

honckenyae

tassiana var. *arctica*

Hordeum

hordei

hordicola

Hosackia

hosackiae

Hosta

hostae

Humulus

ceriospora

erysiphina

humuli

lupulina

superflua ssp. *humili*

Hydrangea

hydrangeae

Hydrocotyle

centellae

hydrocotyles-asiaticae

Hypericum

circumvaga var. *hyperici*

elodes

hyperici

hypericina

Hypochaeris

hypochaeridis

Ilex

ilicella

ilicicola

ilicis

ilicis-canariensis

Impatiens

impatientina

impatientis

subgregaria

yaku-insularia

Imperata

imperatae

Imperatoria

ostruthii

Inula

baudysiana

Ipomoea

bataticola

bonae-noctis

ipomoeae

Iris

desmazieri

iridis

macrospora

pseudacori

Isatis

isatidis

Isoplexis

isoplexidis

Ixanthus

ixanthi

Ixodia

ixodiae

Ixora

ixorae

Jasminum

clypeata

jasminicola

jasmini-officinalis

Juglans

convexula

juglandis

saccardoana

woronowii

Juncus

eurypotami

hariotiana

juncellina

junciginea

juncina

lamprocarpi

michotii

najas

perexigua

perimens
perparva
pheidasca
praeparva
vogesiacca
Juniperus
fructinex
guadarramica
juniperi
juniperina
sabinae
sylvatica
Jurinea
jurineae
Kalmia
colorata
haematodes
Kalopanax
araliae
Khaya
khayae
Kochia
kochiae
Koeleria
dactylidis f. *matritensis*
Krigia
krigiae
Laburnum
laburni
leguminis-cytisis
Lactuca
lactucae
praecox
Lapageria
lapageriae
Lardizabala
lardizabala
Larix
laricina
laricis-leptolepidis
Laserpitium
dolichospora
jenensis
Lasia
lasiana
Lasianthus
concentrica
Lathyrus
karakulinii
lathyri
lathyrina
melaena
nemorosa
nerviseda
ontariensis
orobi
Lavoisiera
melastomatacearum
Lecanora
dispersa
epicymatia

thallophila
Leersia
leersiae
Lepidium
cruciferarum f. *lepidii-graminifolii*
Lepidosperma
lepidospermatis
Leucadendron
konae
Leucanthemum
pascuorum
Leucopogon
singularis
Leucospermum
holualoana
waimeana
Leucothoë
leucothoës
Liabum
liabi
Libanotis
libanotidis
Libocedrus
magellanicola
Ligustrum
ligustri
Lilium
cinxia
martagonis
matura
Linaria
crebra
linariae
Linnaea
leightonii
linnaeae
Linum
drobnjakensis
lini
linicola
lini-perennis
linorum
Lippia
aloyisiae
lippiae
Liriodendron
carpogena
elatior
infuscans
liriodendri
tulipifera
Lithospermum
lithospermi
Lithrea
lithreae
Lloydia
burnati
Lobelia
lobeliae
petchii
tupae

Lolium
loliacea
Lomaria
tabularis
Lonicera
caprifoliorum
clymenia
collina
implexae
implexica
lonicerae
nuristanica
ramulorum
Loranthus
loranthi
Lunularia
hepaticarum
Lupinus
lupini
Luzula
depressa
hypostomatica
luzulae
Lychnis
lychnidicola
Lycium
andicola
lycii
Lycopodium
crepini
ditissima
lycopodii
lycopodii-annotini
lycopodiicola
lycopodina
niesslii
plegmariae
Lygeum
lygei
Lyonia
andromedae
Lysimachia
lysimachiae
lysimachiicola
Lythrum
lythri
minima
Machaerium
machaerii
Macleaya
macleayae
Maclura
maclurae
Maesa
maesae
Magnolia
annulata
apula
glauca
magnoliae
milleri

- Malus**
kawanensis
pomi
sentina
- Malva**
malvina
- Manettia**
linearis
- Mangifera**
mangiferae
tahitensis
- Manihot**
helenae
henningsii
manihotis
- Mappia**
mappiae
- Matthiola**
matthiolae
- Maximilliana**
advena
- Maytenus**
aiacu
mayteni
- Medicago**
circumvaga
destructiva
medicaginicola
medicaginis
- Melaleuca**
melaleuca
melaleucoides
- Melampyrum**
winteriana
- Melastoma**
nitidula
- Melianthus**
macowaniana
- Melica**
omphalosporoides
- Melilotus**
davisii
lethalis
linhartiana
- Meliosma**
meliosmae
meliosmae-myrianthae
- Melothria**
melothriae
- Memecylon**
multiloculata
- Mentha**
menthae
- Mercurialis**
kriegeriana
mercurialis
- Metrosideros**
metrosideri
- Miconia**
miconiae
- Micromeria**
micromeriae
- Mikania**
mikaniae
mikaniae-micranthae
- Milium**
microspora
- Mimosa**
mimosae-pigrae
mimosicola
- Minuartia**
minuartiae
- Molinia**
montellica
recutita f. *molinae*
- Monarda**
monardae
- Moquilea**
moquileae
- Moraea**
moraeae
- Moriera**
persica
- Morinda**
morindae
- Morus**
arachnoidea
indica
mori
mori-albae
moricola
morifolia
- Mouriri**
cecropiae var. *macrocarpa*
- Mucuna**
mucunae
- Muhlenbergia**
muhlenbergiae
- Mulgedium**
depressa
mulgedii-alpini
- Murraya**
kankeshwarensis
murrayae
- Musa**
eumusae
fijiensis
formosana
henriquesiana
liukuensis
musae
musicola
- Muscari**
muscari
- Mutisia**
mutisiicola
- Myginda**
mygindae
- Myoporum**
maidenii
- Myrica**
evernia
myricae
pardalota
- Myricaria**
myricariae
- Myrsine**
atichiae
conspicua
myrsines
- Myrtus**
antivarensis
lumae
myrticola
- Narthecium**
grandispora
macedonica
- Nectandra**
nectandrae
- Nemesia**
nemesiae
- Nerium**
mediterranea
nerii-odorii
oleandri
- Neuroloma**
algida
- Nicotiana**
marii
nicotianae
tabaci
- Nigella**
lachmanii
- Noaea**
engleriana
- Nothofagus**
nothofagi
subantarctica
- Nuphar**
paludosa
- Nymphaea**
pontederiae
- Nyssa**
nyssicola
- Ochrolechia**
araneosa
- Oenanthe**
oenanthicola
- Oenothera**
oenotherae
- Olea**
byliana
oleina
- Omalanthus**
omalanthi
- Onobrychis**
onobrychidis
- Ononis**
ononidis
- Ophiopogon**
ophiopogonis
- Opuntia**
opuntiae

Oreoselinum*oreoselini***Origanum***polygramma* f. *origani***Orites***mycoparasitica***Ornithogalum***hyphiseda**ornithogali***Orontium***orontii***Orthilia***pyrolae**pyrolina***Oryza***danubialis**hondai**malinverniana**oryzae**shimadae**shiraiana**usteriana***Oryzopsis***oryzopsidis***Osmunda***callistea**osmundicola***Othonnopsis***othonnopsidis***Oxalis***acetosellae**depazeaeformis**oxalidis**selene***Oxydendrum***caroliniana***Oxyria***oxyriae***Pachypleurum***pachypleuri***Pachysandra***pachysandrae***Paeonia***moutan***Paepalanthus***paepalanthi***Panax***panacis***Pandanus***pandani***Panicum***crus-galli**miliacei**panicicola**panicum***Pantacantha***pantacanthae***Papaver***arthopyrenioides**karajacensis**morphaea**papaveris***Parmeliella***lepidiotae***Parnassia***parnassiae***Paronychia***paronychiae***Passiflora***passiflorae***Patrinia***patriniae***Paulownia***paulowniae***Pavonia***pavonina***Paxystima***paxistimae***Pedicularis***lineata**pedicularidis***Pelvetia***ascophylli**pelvetiae***Pennisetum***minabensis***Penstemon***fuscata**orthospora**penstemonis***Perezia***pereziae***Pericampylus***pericampyli***Pericopsis***pericopsidis***Periploca***peroplcae***Pernettya***pernettyae***Persea***exutans**perseae***Persoonia***persooniae***Petasites***petasitidis***Petrocallis***pyrenaica***Petrorhagia***caryophylli***Peucedanum***nebulosa-veneta**thysselini**umbelliferarum***Phaca***phacae-frigidae***Phacelia***phaceliiphila***Phaeophyscia***thallina***Phagnalon***asensioi***Phalaris***chlouna**cinereo-nebulosa**phalaridis***Phaseolus***cruenta**mycopron**phaseoli**phaseolicola**phaseolorum***Philodendron***philodendronis***Phleum***rousseiana***Phlomis***bolivari**phlomicola**phlomidis***Phlox***phlogina***Phoenix***frenumbensis**phoenicis***Phragmites***maculiseda**perpusilla**phragmitis**vaginae***Phyllanthus***phyllanthi***Phyllis***phyllitis***Phyllostachys***bambusifolia**phyllostachydis***Physostegia***physostegiae***Phyteuma***phyteumatis***Phytolacca***circe***Phytophthora***theobromae***Picea***picconii***Pilea***sublibera***Piliostigma***piliostigmatis***Pimpinella***pimpinellae***Pinus***acicola**conigena**dearnessii**gibsonii**hypodermellae**pinastri**pini**pinicola*

- pinifolia*
pini-patulae
Piper
piperis
pulchella
Pistacia
pistaciae
pistaciarum
pistacina
terebinthi
Pisum
pinodes
Pithecellobium
pithecellobiicola
Pittosporum
pittospori
rubiginosum
Plantago
canariensis
columbi
gaveensis
plantaginicola
plantaginis
puttemansii
theissenii
Platanus
albocrustata
circumdans
inconspicua
platani
platanifolia
stigmina-platani
veneta
Platylobium
platylobii
Plectranthus
plectranthi
pogostemonis
Poa
arctica
badensis
gangreana
graminum
recutita f. *poae*
rousseiliana f. *poae*
Podocarpus
podocarpi
podocarpicola
Polemonium
polemonii
Polygala
polygalina
Polygonatum
asteroma var. *polygonati*
cruenta
polygonati
Polygonum
eucarpa
polygони-cuspidati
polygonorum
vivipari
- Polypodium**
asperulata
polypodii
polypodii f. *polypodii-vulgaris*
polypodii f. *spermogonifera*
subostiolica
tiroloensis
tiroloensis var. *montellica*
Polytrichum
heufleri
Pongamia
pongamiae
Pontederia
pontederiae
Populus
alba
angustifoliorum
balsamopopuli
crassa
macularis
major
mandshurica
orbicularis
populi
populicola
populifolia
populnea
populorum
salicinearum f. *populi-nigrae*
subcrassa
togashiana
tremulae
tremulicola
tremulina
Poraqueiba
poraqueibae
Porocyphus
porocyphi
Potentilla
broteriana
muelleriana
potentillae
potentillae-stipularis
pulchra
tormentillae
Poterium
minor var. *poterii*
Pourthiaea
pourthiaeae
Pouzolsia
pouzolsiae
Prasium
prasii
Premna
oculata
Prenanthes
prenanthicola
prenanthis
Primula
clandestina
primulae
- primulaeicola*
Prinus
prini
Prinsepia
prinsepiae
Protea
bellula
concentrica
fibrillosa
jonkershoekensis
microspora
proteae
proteae-arboreae
stromatosa
Proustia
leptosperma
proustiae
Prunus
cerasella
cerasicola
cerasina
cerasus
lenticula
maculata
nigerristigma
padina
pruni-persicae
Psammisia
psammisiae
Pseudotsuga
andersonii
Psora
innata
psorae
Pteridium
aquilina
indistincta
insignita
olindensis
prominula
polypodii f. *pteridinis*
pteridicola
pteridis
striola
yanagawaensis
Pteris
novae-hollandiae
Pterocarpus
pterocarpi
Pterocarya
haraeana
Puccinia
teycrui
Pueraria
keissleri
puerariae
puerariicola
Pulmonaria
pulmonariae
Pulsatilla
pulsatillae

pulsatillae var. *minor*

Punica

lythracearum

Putoria

putoriae

Puya

puyae

Pyrola

pyrolae

Pyrus

bellona

chaubattiensis

fujiensis

lucillae

pyri

pyrina

sentina f. *pyri-communis*

Quercus

aethiops

aquatica

areola

berlesiana

caespitosa

catesbeyi

cookeana

cooperta

dryophila

epiphylla

familiaris

gregaria

janus

lignicola

maculiformis f. *congesta*

maculiformis f. *dispersa*

maculiformis f. *quercus*

maculiformis f. *quercus-roboris*

maculiformis var. *quercinae*

melanophora

mobilis

myriadea

nigrita

operculata

pandurata

perexigua

phellos

pleuronervia

polystigma

psilospora

punctiformis

punctiformis f. *quercus*

punctiformis var. *ambigua*

punctiformis var. *perexigua*

punctiformis var. *peritheciis-*

minutissimis

punctoidea

quercicola

quercifolia

ravenelii

septorispora

serograptus

simulans

sparsa f. *quercus*

spleniata

succinea

suspecta

Quisqualis

luzonica

Rafinesquia

nemoseridis

Rajania

rajaniae

Ranunculus

fusispora

nivalis

ranunculi

valida

Rapanea

rapanae

Rapistrum

passeriniana

Rauvolfia

rauvolfiae

Reseda

nebulosa var. *minor*

resedae

resedicola

Retinispora

retinisporae

Rhamnus

vogelii

Rhaphithamnus

rhaphithamni

Rahponticum

sajanyca

Rhododendron

handelii

millepunctata

occulta

polyspora

rhododendri

rhodorae

Rhodostachys

rhodostacheos

Rhus

fushinoki

nigredo

rhoina

rhois

sumacis

Rhynchospora

rhynchosporae

Ribes

assimilata

aurea

curva

grossulariae

pachyasca var. *ribicola*

ribis

Riccia

ricciae

Richea

munyangica

richeae

Ricinus

ricinicola

Robinia

petiolicola var. *robiniae*

pseudacaciae

robiniae

Romulea

schoenoprasi var. *romuleae*

Rosa

intermixta

jaapiana

rhodophila

rosae

rosigena

sepincolaeformis

spinicola

Rostkovia

rostkoviae

Rottboellia

rottboelliae

Rourea

roureae

Rubia

mougeotiana

peregrina

Rubus

amphigena

chamaemori

confusa

cunninghamiae

fruticum

holmii

idaeina

innumerella var. *rubi*

joerstadii

ligea

maculiformis var. *rubi*

minoënsis

nervicola

rubi

rubicola

rubina

winteri

Rudbeckia

rudbeckiae

Rumex

chorinensis

insulana

lapathi

rumicis

stromatoidea

Ruscus

rusci

ruscicola

Ruta

algarbiensis

rhea

Sabal

incisa

sabaligena

- sabalis*
serrulata
Saccharum
sacchari
saccharoides
striatiformans
wakkeri
Sagina
saginae
Sagittaria
sagittariae
Salicornia
peruviana
salicorniae
Salix
baldensis
canescens
capronii
chlorospora
epitea
genuflexa
grossulariae var. *salicella*
hyalospora
inaequalis var. *salicis*
lapponum
maculiformis var. *herbaceae*
minor var. *reticulata*
polaris
punctiformis f. *salicis-auritae*
rhabdospora
salicicola
salicina
salicinearum
salicis
spetsbergensis
turba
Salvia
salviae
Samanea
samaneae
Sambucus
ebuli
ebulina
Sanguisorba
pseudomaculiformis
sanguisorbae
Sapindus
reyesii
sapindicola
Sapium
sodiroana
Saponaria
asteroma var. *caryophyllacearum*
saponariae
smegmatos
Sarothamnus
sarothamni
Sarracenia
sarraceniae
Sasa
inaequalis
- Sassafras**
sassafras
Saussurea
lavrovii
saussureae-alpinae
trichophila
Saxegothaea
maniwana
Saxifraga
huteriana
olenjana
saxifragae
stromatica
trichophila var. *saxifragae*
Scabiosa
columbariae
sylvatica
Scaevola
scaevolae
Scandix
pectinis
Scheuchzeria
bacillifera
Schisandra
schisandrae
Schizophragma
exigua
Schoenocaulon
schoenocauli
Scirpus
maculans
perexigua var. *minima*
scirpi-lacustris
thais
vitensis
wichuriana var. *scirpella*
Scleranthus
gypsophilae var. *scleranthi*
Scorzoneria
podperae
scorzoneriae
Scrophularia
decidua var. *scrophulariae*
grisea
scrophulariae
tardiva
Secale
basicola
leptopleura
tulasnei
Securinega
securinegae
Sedum
edelbergii
sedi
sedicola
Semele
semeles
Senecio
beaglensis
jaffueli
- maculiformis* f. *senecionis*
majuscula
sarraceniae
senecionis
subalpina
Sequoia
sequoiae
Serratula
serratulae
Sesamum
sesami
sesamicola
Sesbania
sesbaniae
Seseli
seseli
superflua f. *umbelliferarum*
Shibataea
shibataeae
Sibthorpia
maderensis
Sicyos
sicyicola
Sidalcea
sidaecola
Silene
balcanica
silenes
silenes-acaulis
silenicola
woronichinii
Siler
taeniographoides
Siphocampylus
tuerckheimii
Sisymbrium
cruciferarum
passeriniana
superflua f. *sisymbrii*
Sisyrinchium
sisyrinchiiicola
Slimmia
skimmiae
Smilax
garganica
pellucida
smilacicola
smilacifolii
smilacina
smilacis
smilacis-glabrae
vagnerae
Solanum
dubia
solani
Solidago
nebulosa
solidaginea
solidaginis
virgaureae
Solorina

schaereri
Sophora
sophorae
Sorbus
aucupariae
cinerascens
inaequalis
sorbi
topographica
Sorghum
ceres
holci
Sparganium
taediosa
typharum f. *sparganii*
Spartina
spartinae
Spartium
scopulorum
Spergularia
alsinearum
gypsophilicola
Sphaeralcea
stenospora
Spigelia
spigeliae
Spiraea
crenata
manginii
spiraeae
Spondias
mombin
Staphylea
staphyleae
staphylina
Steironema
ciliata
Stellaria
alsines
isariphora
stellarinearum var. *holosteae*
uliginosa
Stephania
cepharanthae
stephaniae
stephaniicola
Stephanorossia
stephanorossiae
Stigmaphyllon
stigmaphyllonis
Stipa
chlorina
graminis
stipina
Strychnos
strychni
Stylosanthes
melanococca
Styrax
styracis
Suaeda

suaedae-australis
Symphoricarpos
symphoricarpi
Symphyostemon
symphyostemonis
Symplocos
bhauria
Syringa
cruchetii
polycarpa
syringicola
Syzygium
marasasii
syzygii
Tabebuia
brunneomaculans
ipiranguensis
tabebuiae
Taccarum
taccari
Tagetes
allicina var. *ferlinii*
Tamarindus
tamarindi
Tamarix
tamaricis
Tanacetum
lindiana
pascuorum
Tapesia
parasitica var. *tapesiae*
Taraxacum
taraxaci
Tarzetia
persica
Taxodium
taxodii
Taxus
taxi
Tecoma
erysiphoides
tecomae
viegasii
Telopea
telopeae
Ternstroemia
ternstroemiae
Tetragastris
rotlerae
Tetroncium
magellanicola
tetroncii
Teucrium
polygramma f. *caruanae*
teucriti
Thalictrum
allescheri
fendleri
insidens
punctata
thalictri

thalictricola
thalictrina
Thallomyces
mycoparasitica
Thelypteris
perparva
thelypteridis
Theobroma
theobromae
Thermopsis
thermosidisi
Thesium
thesii
Thlaspi
andina
Thuja
canadensis
conicola
conigena
thujae
Thujopsis
thujopsidisi
Thymus
serpylli
Tilia
bracteophila
chondrospora
fennica
incanescens
maculiformis var. *tiliae*
microsora
platytheca
punctiformis f. *tiliae*
sparsa
sparsa f. *tiliae*
sparsa f. *tiliae-parvifoliae*
tiliae
Tinospora
tinosporeae
Tocoyena
tocoyenae
Todea
todeae
Tortula
tortulae
Tournefortia
tournefortiae
Trachycarpus
trachycarpi
Tragopogon
sydowii
tragopogonicola
Trapelia
cookei
Trichilia
asunciensis
Trichomanes
assurgens
trichomanes
Trifolium
balcanica

- calycicola*
carinthiaca
consociata
killianii
trifolii
Triglochin
junciginearum
Trisetum
tajmyrensis
triseti
Tristania
brunnea
tristaniae
Triticum
canifaciens
exitialis
graminicola
pusilla f. *tritici-monococci*
pusilla f. *tritici-vulgaris*
Trollius
hypsicola
Tsuga
peckii
tsugae
Tuberaria
camarae
Tussilago
peris
tussilaginis
Typha
groveana
intercellularis
microscopica
tassiana var. *rupefortensis*
typhae
typharum
typhina
Ulmus
apertiuscula
comedens
oedema
ulmi
ulmifolia
Umbellularia
arbuticola
umbellulariae
Ungnadia
ungnadiae
Urtica
nebulosa
superflua
superflua f. *urticae-dioicae*
urticae
urticae-dioicae
Vaccinium
brachytheca
gallae
leptidea
maculiformis f. *myrtilli*
myrtillina
nigromaculans
- oxycocci*
rubefaciens
stemmatea
vaccinii
vacciniicola
Valeriana
galatea
nigrita
vagans
Valerianella
galatea var. *valerianellae*
lindaviana
Veratrum
veratri
veratri-lobeliani
Verbascum
verbasicola
Verbena
verbenae
Vernonia
decidua
dummeri
vernoniae
Veronica
montana
Vesicaria
vesicariae
Viburnum
crepidophora
lantanae
lindingeri
tini
viburni
viburnicola
Vicia
coymiana
thironi
viciae
viciarum
Vinca
duchartrei
vincae
Vincetoxicum
albescens
vincetoxici
Viola
asteroma var. *violae*
violae
Vitis
angulata
bidwellii
cuboniana
fumaginea
manganottiana
mazzantioides
pampini
personata
sarmentorum
sucedanea
viticis
viticola
- vitis*
Washingtonia
washingtoniae
Weigelia
weigeliae
Weinmannia
weinmanniae
Welwitschia
welwitschii
Wisteria
wisteriae
Wollemia
wollemiae
Xanthium
xanthiicola
Xerophyllum
xerophylli
Xylomelum
xylomeli
Yucca
acervata
deightonii
sphaerelloides
yuccae
yuccina
Zea
maydina
maydis
paulula
zeae
zeae-maydis
zeicola
zeina
Zelkova
zelkovae
Zingiber
zingiberi
Zizania
zizaniae
zizantiicola
Zizia
thaspiicola
Ziziphus
ziziphi
ziziphicola

